

Extreme positive allometry of animal adhesive pads and the size limits of adhesion-based climbing

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Organismal functions are size-dependent whenever body surfaces supply body volumes. Larger organisms can develop strongly folded internal surfaces for enhanced diffusion, but in many cases areas cannot be folded so that their enlargement is constrained by anatomy, presenting a problem for larger animals. Here, we study the allometry of adhesive pad area in 225 climbing animal species, covering more than seven orders of magnitude in weight. Across all taxa, adhesive pad area showed extreme positive allometry and scaled with weight, implying a 200-fold increase of relative pad area from mites to geckos. However, allometric scaling coefficients for pad area systematically decreased with taxonomic level and were close to isometry when evolutionary history was accounted for, indicating that the substantial anatomical changes required to achieve this increase in relative pad area are limited by phylogenetic constraints. Using a comparative phylogenetic approach, we found that the departure from isometry is almost exclusively caused by large differences in size-corrected pad area between arthropods and vertebrates. To mitigate the expected decrease of weight-specific adhesion within closely related taxa where pad area scaled close to isometry, data for several taxa suggest that the pads' adhesive strength increased for larger animals. The combination of adjustments in relative pad area for distantly related taxa and changes in adhesive strength for closely related groups helps explain how climbing with adhesive pads has evolved in animals varying over seven orders of magnitude in body weight. Our results illustrate the size limits of adhesion-based climbing, with profound implications for large-scale bio-inspired adhesives.

scaling | adhesion | evolution | bio-inspired adhesives

The evolution of adaptive traits is driven by selective pressures but can be bound by phylogenetic, developmental, and physical constraints (1). Integrating evolution and biomechanics provides a powerful tool to unravel this complex interaction, because physical constraints can often be predicted easily from first principles (2). The influence of physical constraints is especially evident in comparative studies across organisms that differ substantially in size (3–6). For example, Fick's laws of diffusion state that diffusive transport becomes increasingly insufficient over large distances, explaining the development of enlarged surfaces for gas and nutrient exchange (e.g., leaves, roots, lungs, gills, and guts) and integrated long-distance fluid transport systems (e.g., xylem/phloem and circulatory systems) in larger animals and plants. How these systems change with size is determined by physical constraints (7–9). Although “fractal” surface enlargements are possible without disrupting other body functions, strong positive allometry can conflict with anatomical constraints. For example, structural stability demands that animals should increase the cross-sectional area of their bones in proportion to their body weight, but excessively thick leg bones can compromise other physiological functions and hamper locomotion (3, 10, 11).

Adhesive pads are another example of an adaptive trait subject to size-dependent physical constraints. These systems allow

animals to climb smooth vertical or inverted surfaces, thereby opening up new habitats. Adhesive pads have evolved multiple times independently within arthropods, reptiles, amphibians, and mammals and show impressive performance: They are rapidly controllable, can be used repeatedly without any loss of performance, and function on rough, dirty, and flooded surfaces (12). This performance has inspired a considerable amount of work on technical adhesives that mimic these properties (13). A key challenge for both biological and bio-inspired adhesive systems is to achieve size-independent performance (14–16), that is, the maximum sustainable adhesion force, F , should be proportional to the mass to be supported, m . For vertically climbing animals, F is the product of the maximum adhesive stress, σ , and the adhesive pad area, A , each of which may change with mass ($A \propto m^a$ and $\sigma \propto m^b$), so that constant size-specific attachment performance requires

$$m \propto A\sigma \propto m^a m^b \rightarrow a + b \approx 1, \quad [1]$$

where a and b are the scaling coefficients for σ and A in relation to body mass, respectively. If animals maintain geometric similarity when increasing in size, A would scale as $m^{2/3}$, so that the adhesion per body weight for large geckos ($m \approx 100$ g) is expected to be approximately $10^{7/3} \approx 200$ times smaller than for tiny mites ($m \approx 10$ μ g) if the pads' adhesive strength σ remained unchanged ($b = 0$). Large animals can only circumvent this problem by (i) developing disproportionately large adhesive pads ($a > 2/3$) and/or (ii) systematically increasing the maximum force per unit pad area ($b > 0$). How do large climbing animals achieve adhesive forces equivalent to their body weight?

Significance

How adhesive forces can be scaled up from microscopic to macroscopic levels is a central problem for biological and bio-inspired adhesives. Here, we elucidate how animals with sticky footpads cope with large body sizes. We find an extreme positive allometry of footpad area across all 225 species studied, implying a 200-fold increase of relative pad area from mites to geckos. Within groups, however, pads were almost isometric, but their adhesive strength increased with size, inconsistent with existing models. Extrapolating the observed scaling, we show that to support a human's body weight, an unrealistic 40% of the body surface would have to be covered with adhesive pads, suggesting that anatomical constraints may prohibit the evolution of adhesion-based climbers larger than geckos.

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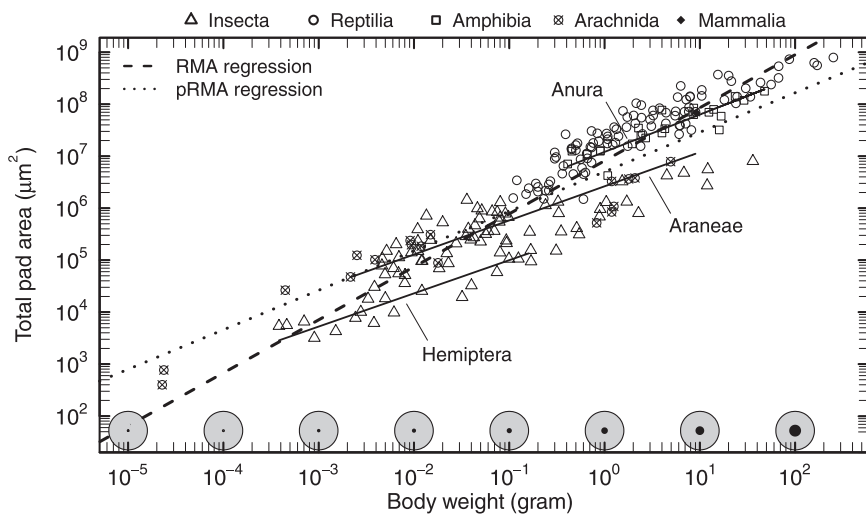


Fig. 1. Allometry of pad area for 225 species, ranging from $\approx 20 \mu\text{g}$ to $\approx 200 \text{ g}$ in body weight. Across all taxa, pad area was directly proportional to body weight (RMA, dashed line). The increase in the fraction (black circles) of the total available surface area (gray circles) required to accommodate this disproportionate change is schematically illustrated along the bottom axis, assuming that a climbing animal of 80 kg has a total surface area of 2 m^2 , comparable to a human of 80 kg and 180 cm, and that body surface area is approximately isometric. Strikingly, scaling relationships were significantly less steep within closely related groups (representative solid lines from RMA regressions) and closer to isometry when phylogenetic relationships were accounted for (pRMA regression, dotted line).

Using the simple biomechanics argument outlined above as a framework, we here provide a comparative analysis of the allometry of adhesive pad area across 225 species, covering more than seven orders of magnitude in weight—almost the entire weight range of animals climbing with adhesive pads—and including representatives from all major groups of adhesion-based climbers.

Results and Discussion

Scaling of Adhesive Pad Area. Across all taxa, adhesive pad area showed extreme positive allometry and scaled as $A \propto m^{1.02}$ [reduced major axis regression (RMA); see Fig. 1 and Table S1 for detailed statistics], an increase sufficient to compensate for the predicted loss of weight-specific adhesion, even if adhesive strength remained unchanged. Thus, adhesive pads occupy a larger fraction of the body surface area in larger animals. Within closely related taxonomic groups, however, pad area grew more slowly with body mass, indicating a strong phylogenetic signal (Figs. 1 and 2).

When evolutionary relationships were accounted for, the observed scaling coefficient decreased dramatically and was almost consistent with isometry (Figs. 1 and 2 and Table S1). This systematic change of allometric coefficients with taxonomic rank suggests that phylogenetic inertia impedes a disproportionate increase of pad area within closely related groups (Figs. 2 and 3A). Our results thus add to a body of evidence suggesting that the evolutionary flexibility of allometric slopes is low and larger changes in particular traits are mainly achieved by shifts of the allometric elevation (17, 18).

Removal of the influence of body size by analyzing the residuals (termed “relative pad area” in the following) of a phylogenetic reduced major axis regression (pRMA) allowed us to further investigate at what taxonomic level major shifts in relative pad area occurred, separating the effects of size and ancestry. Relative pad area differed strongly between vertebrates and arthropods, but comparatively little variation existed within these groups (Fig. 3A). More than 58% of the variation in residual pad area was explained by differences between vertebrates and arthropods (nested ANOVA, $F_{1,173} = 845$, $P < 0.001$; Table S2), so that body weight and phylum alone accounted for more than 90% of the total variation in pad area. Rather than being driven solely by variation in body size, differences in relative pad area seem to be tied to characteristic features of the corresponding phyla, such as, for example, the presence or absence of multiple distal pads (toes) per leg (Fig. 3B and C). However, we also found evidence for differences in relative pad area within lower taxonomic ranks. For example, members of the gecko genus *Sphaerodactylus* had considerably smaller pads than other Gekkotan lizards, whereas *Gekko* lizards had particularly well-developed

pads (based on their relative pad area; Fig. 3A). In insects, hemimetabolous orders had smaller relative pad areas than holometabolous orders (Fig. 3A). Adhesive pads allow access to arboreal habitats (19–21), but they may come at the cost of reduced locomotor performance in situations where no adhesion is required (22, 23). Thus, the multiple independent losses, gains, and reductions of adhesive pads in amphibians, insects, lizards, and spiders (24–27) likely reflect the ecological, behavioral, and taxonomic diversity within these groups (28, 29).

The Size Limits of Adhesion-Based Climbing. Strong positive allometry of nonconvoluted body structures in organisms ranging in size over many orders of magnitude is difficult to achieve, owing to simple anatomical constraints. For example, bone mass in terrestrial animals is predicted to increase with mass^{4/3} to maintain constant bone stress levels, but this would require unrealistic relative bone masses for larger mammals (scaling up an 8-g shrew with *ca.* 4% bone mass would produce a rather unfortunate 8-ton elephant with 400% bone mass). The actual scaling coefficient is inevitably smaller (≈ 1.1) (10, 30), and alternative strategies have evolved to limit bone stresses (11).

Maintaining a pad area proportional to body weight in animals ranging from 10^{-5} to 10^2 g requires extraordinary morphological changes: Assuming otherwise isometric animals, the proportion of the total body surface area specialized as adhesive pad needs to increase by a factor of $10^{7/3} \approx 200$. This extreme shape change may impose a size limit for adhesion-based climbing. Scaling up the relative pad area of arthropods and small vertebrates to a human of 180 cm body length and 80 kg body mass would result in an adhesive pad area of $\approx 10^{6.91} \cdot 80,000^{1.02} \approx 0.81 \text{ m}^2$, approximately two-fifths of the total available body surface area [$\approx 2 \text{ m}^2$ (31)]. The required morphological changes, if at all possible, would thus be enormous, and difficult to achieve over short evolutionary timescales. Our results therefore indicate that phylogenetic inertia restricts the “design space” for evolution at least for closely related taxa. Larger animals within closely related taxa must therefore either cope with a size-related decrease in their attachment ability or develop alternative strategies to compensate for it. Recent studies on tree frogs and ants revealed that pad adhesive strength can vary systematically with size, resulting in an almost body size-independent attachment abilities despite a near-isometric growth of pad area (16, 32). Here, we extend these studies to investigate whether such adaptations are also present above the species or genus level.

Fig. 4 shows whole-body adhesion per pad area plotted against body weight for 17 frog species from 4 families and 12 genera (33–36; see Supporting Information). All adhesion measurements were conducted using a tilting platform and are thus comparable across studies. Over two orders of magnitude in body weight, adhesion force per unit area increased with $m^{0.3}$ [RMA slope 0.3, 95%

sputter; Quorum Technologies) and examined with a field emission gun SEM at a beam voltage of 5 kV (Leo Gemini 1530VP; Carl-Zeiss NTS GmbH).

Data on toepad-bearing gecko species were collected from live animals kept in the D.J.I. laboratory [under an Institutional Animal Care and Use (IACUC) protocol 2012–0064 from the University of Massachusetts, Amherst to D.J.I.] and preserved specimens from the American Museum of Natural History and the Museum of Comparative Zoology at Harvard University. For each specimen, photos of one forefoot were obtained by pressing it tightly against the glass plate of an Epson Perfection V500 Photo Scanner (Seiko Epson Corp.) next to a ruler and taking a digital scan. The total toepad area across all digits was measured using ImageJ, version 1.49r (88). We also measured snout-vent length (± 1 mm) from each individual using a clear plastic ruler. Where possible, we measured multiple conspecific individuals and used the mean as the species value.

Literature data were taken from the papers' text or tables or were extracted from figures using WebPlotDigitizer 3.3 (WPD, developed by Ankit Rohatgi; arohatgi.info/WebPlotDigitizer) or ImageJ, version 1.49m. We tested the performance of WPD with an x - y plot of 20 random numbers between 1 and 1,000 (x) and 0.01 and 10,000 (y) on a log-log scale and found an accuracy of $\approx 0.6\%$ for the raw data.

We used live body weights where available and interpolated live weight from body length where necessary, using established scaling relationships (38). A list of the species included can be found in the [Supporting Information](#).

Adhesive Pad Area. Animals attach to smooth surfaces by using specialized attachment pads on their legs. These pads are either covered with dense arrays of fibrils (hairy pads) or are macroscopically unstructured (smooth pads). To compare pad areas across different taxa and pad morphologies, the following assumptions were made:

- i) "Projected" pad area is the most meaningful measure of contact area in comparative studies. Projected pad area is the surface area of the foot specialized specifically for generating adhesion and friction (89). In a fibrillar pad, inevitably only a fraction of this area comes into surface contact (i.e., the "real" contact area is significantly smaller than the projected contact area).
- ii) All animals use a similar fraction of their available pad area when maximum performance is required. It is unclear what fraction of the available pad area is used by animals of different size (16, 90, 91), and systematic studies are lacking. However, the small number of direct contact area observations available strongly suggest that animals often use the entire area of their adhesive pads (89, 92), that is, that they are not "overbuilt." We thus assume that all climbing pads are designed so that their whole area can be used in critical situations.
- iii) Adhesive performance is dominated by distal pads. Insects can have several attachment pads per leg. There is strong evidence that these pad types differ in their morphology, as well as in their performance and function during locomotion (16, 42, 63, 68, 89). Many insects do not use their distal pads when no adhesion is required (42, 63, 93–96), whereas during inverted climbing only distal pads are in surface contact (81). Accordingly, insects with ablated distal pads cannot cling upside-down to smooth surfaces (42, 96). Distal pads thus seem to be true adhesive pads (81). Proximal pads, in contrast, can lack strong adhesion and may be designed as nonadhesive friction pads (97, 98). The distal pads are usually part of the pretarsus, but some insects lack a pretarsal pad. In these insects, the distal tarsal pad can show morphological specializations similar to those of pretarsal pads (99, 100). Proximal pads are mainly found in arthropods, but they may also be present in frogs (33, 101). As the contribution of proximal pads to adhesion is unclear and likely variable, we exclude them for this study. In most insects, the total proximal pad area is between three to five times larger than the distal pad area, and pad area is still positively allometric even when proximal pads are included (reduced major axis regression slope between 0.8–0.9).
- iv) The variation of pad area/adhesive strength between different legsto/es and sexes of the same species and the variation introduced by the animals' ecology is independent and randomly distributed with respect to body weight. Several studies have shown that the size, morphology and performance of attachment devices can depend on the ecological niche occupied by the animals (19, 21, 28, 57, 102). Variation can also occur between sexes (68, 103, 104), different legs or toes (19, 82), or even between populations of the same species occupying different habitats (23). For this study, we assume that because of the large number of samples and the wide range of body sizes included, any bias introduced by these factors can be ignored.
- v) Adhesive performance of "wet" and "dry" pads is comparable. Dynamic adhesive pads are frequently categorized as wet or dry. However, there

is no evidence for a functional difference between these two pad types (105), and indeed maximal adhesive stresses are comparable (16).

Because some of the data used in this study originate from different groups, we quantified the consistency of pad area measurements among researchers. A selection of SEM images (three hairy and seven smooth pads) were given to 10 scientists who independently measured nominal pad area. We found an average coefficient of variation of $17 \pm 9\%$, which was independent of the animals' body weight (ANOVA, $F_{1,8} = 0.067$, $P = 0.8$). Scaling relationships calculated with this dataset did not vary significantly across scientists (slope: likelihood ratio statistic = 0.38; elevation: Wald statistic = 1.21, both $df = 9$ and $P > 0.9$).

Phylogenetic and Statistical Analyses. To account for the nonindependence of data from related species, we first formed groups within which adhesive pads are likely homologous, based on their position on the leg and their structure (i.e., hairy vs. smooth pads). These groups are (i) Squamata, (ii) Anura, (iii) Araneae, (iv) mites with smooth pads, (v) mites with hairy pads, (vi) insects with tarsal hair fields (e.g., some Coleoptera and Raphidioptera), (vii) insects with smooth pulvilli (e.g., some Hemiptera), (viii) insects with hairy pulvilli (e.g., some Diptera), (ix) insects with unfoldable arolia (some Hymenoptera), (x) insects with nonreversible arolia (e.g., some Polyneoptera, Hemiptera and Lepidoptera), (xi) insects with specialized distal euplantula (some Polyneoptera), and (xii) insects with tibial pads (e.g., some aphids). These groups were all connected directly to the root of the tree, so that analogous structures share no branch, and thus the respective elements in the error covariance matrix of our linear models are zero (106). Within these groups, we assembled a tree topology from phylogenies published for the constituent groups [Anura (107), Squamata (108), Araneae (27), Insecta (109), Blattodea (110), Coleoptera (111), Diptera (112), Hemiptera (113, 114), and Hymenoptera (115, 116)]. Because statistically supported branch lengths were not available, comparative phylogenetic procedures that allow for more complex evolutionary models, such as Ornstein–Uhlenbeck models (117, 118), were not feasible. Instead, we performed phylogenetic generalized least squares on 10,000 trees with randomized branch lengths that were rendered ultrametric via a correlated rate model (119). To account for the uncertainty of the phylogenetic error covariance structure, Pagel's λ was estimated simultaneously via a maximum likelihood optimization (120, 121). The fitted coefficients were normally distributed, with a coefficient of variation below 1% (Fig. S1). For simplicity, we report results for an ultrametric tree calculated from a tree with all branch lengths set to 1.

All analyses involving pad area and weight were performed on \log_{10} -transformed values, and with the routines implemented in the R packages nlme v.3.1–118, ape v.3.1–1, and phytools v.0.3–93 in R v.3.1.3 (122, 123). There is some controversy as to whether (reduced) major axis or ordinary least squares regression is more appropriate for estimating allometric coefficients from data containing both "measurement" and "biological" error

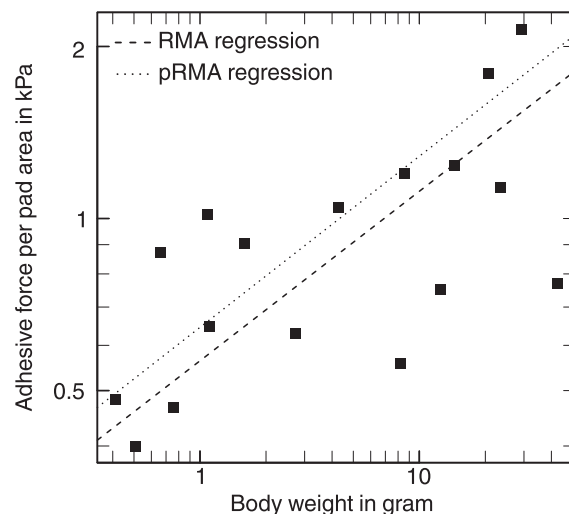


Fig. 4. Adhesive force per area in 17 tree frog species increased with body mass, indicating that larger species possess more efficient pads. In contrast to the allometry of pad area, this result was not significantly influenced by phylogeny.

(17, 18, 124–126). We thus report results for both techniques and note that the key results of this study hold independent of what regression model is applied.

An alternative method to account for relatedness is to monitor the change of the estimated parameters as one moves up in taxonomic rank. We performed multiple reduced major axis regressions across all taxa and separately within the taxonomic levels class, order, family, and genus (for example, all Hymenoptera provide one allometric slope data point within the level “order”). Within the taxonomic levels, groups were only included if the weight range of the available species exceeded a factor of 3, and if data for at least four different groups from the next sublevel were available (e.g., an order was included if at least four families were represented).

To visualize the effect of evolutionary history on body size-corrected pad area, residuals from a phylogenetic reduced major axis regression were used to estimate maximum likelihood ancestral states for all nodes and along the branches via the method described in ref. 127. These values are only rough estimates, first because we do not have statistically supported branch lengths,

second because the species sampling in our phylogeny is incomplete, third because we do not account for the influence of relative pad area on diversification rate, and fourth because only shared ancestors among lower taxonomic ranks possessed adhesive pads. Ancestral state estimates are solely used to visualize systematic differences at the tip level, and no conclusions are based on them.

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- Gould SJ (2002) *The Structure of Evolutionary Theory* (Harvard Univ Press, Cambridge, MA).
- Taylor G, Thomas A (2014) *Evolutionary Biomechanics: Selection, Phylogeny, and Constraint* (Oxford Univ Press, Oxford).
- McMahon T (1973) Size and shape in biology. *Science* 179(4079):1201–1204.
- Schmidt-Nielsen K (1975) Scaling in biology: The consequences of size. *J Exp Zool* 194(1):287–307.
- McMahon TA, Bonner JT, Freeman W (1983) *On Size and Life* (Scientific American Library, New York).
- Schmidt-Nielsen K (1984) *Scaling: Why Is Animal Size So Important?* (Cambridge Univ Press, Cambridge, UK).
- LaBarbera M (1990) Principles of design of fluid transport systems in zoology. *Science* 249(4972):992–1000.
- Weibel ER (1991) Fractal geometry: A design principle for living organisms. *Am J Physiol* 261(6 Pt 1):L361–L369.
- West GB, Brown JH, Enquist BJ (1999) The fourth dimension of life: Fractal geometry and allometric scaling of organisms. *Science* 284(5420):1677–1679.
- Alexander R, Jayes A, Malojo G, Wathuta E (1979) Allometry of the limb bones of mammals from shrews (*Sorex*) to elephant (*Loxodonta*). *J Zool* 189(3):305–314.
- Biewener AA (1989) Scaling body support in mammals: Limb posture and muscle mechanics. *Science* 245(4913):45–48.
- Autumn K, Niewiarowski PH, Puthoff JB (2014) Gecko adhesion as a model system for integrative biology, interdisciplinary science, and bioinspired engineering. *Annu Rev Ecol Syst* 45:445–470.
- Jagota A, Hui C (2011) Adhesion, friction, and compliance of bio-mimetic and bio-inspired structured interfaces. *Mater Sci Eng Rep* 72(12):253–292.
- Bartlett MD, et al. (2012) Looking beyond fibrillar features to scale gecko-like adhesion. *Adv Mater* 24(8):1078–1083.
- Hawkes EW, Eason EV, Asbeck AT, Cutkosky MR (2013) The gecko's toe: Scaling directional adhesives for climbing applications. *IEEE/ASME Trans Mechatron* 18(2): 518–526.
- Labonte D, Federle W (2015) Scaling and biomechanics of surface attachment in climbing animals. *Philos Trans R Soc Lond B Biol Sci* 370(1661):20140027.
- Egset CK, et al. (2012) Artificial selection on allometry: Change in elevation but not slope. *J Evol Biol* 25(5):938–948.
- Pélabon C, et al. (2014) Evolution of morphological allometry. *Ann N Y Acad Sci* 1320:58–75.
- Macrini TE, Irschick DJ, Losos JB (2003) Ecomorphological differences in toepad characteristics between mainland and island anoles. *J Herpetol* 37(1):52–58.
- Betz O (2003) Structure of the tarsi in some *Stenus* species (Coleoptera, Staphylinidae): External morphology, ultrastructure, and tarsal secretion. *J Morphol* 255(1):24–43.
- Wolff JO, Gorb SN (2015) Adhesive foot pads: An adaptation to climbing? An ecological survey in hunting spiders. *Zoology (Jena)* 118(1):1–7.
- Russell AP, Higham TE (2009) A new angle on clinging in geckos: Incline, not substrate, triggers the deployment of the adhesive system. *Proc Biol Sci* 276(1673): 3705–3709.
- Collins CE, Russell AP, Higham TE (2014) Subdigital adhesive pad morphology varies in relation to structural habitat use in the Namib Day Gecko. *Funct Ecol* 29(1):66–77.
- Düllman W, Trüb L (1997) *Biology of Amphibians* (Johns Hopkins Univ Press, Baltimore).
- Beutel RG, Gorb SN (2001) Ultrastructure of attachment specializations of Hexapods (Arthropoda): Evolutionary patterns inferred from a revised ordinal phylogeny. *J Zoological Syst Evol Res* 39(4):177–207.
- Gamble T, Greenbaum E, Jackman TR, Russell AP, Bauer AM (2012) Repeated origin and loss of adhesive toepads in geckos. *PLoS One* 7(6):e39429.
- Wolff JO, Nentwig W, Gorb SN (2013) The great silk alternative: Multiple co-evolution of web loss and sticky hairs in spiders. *PLoS One* 8(5):e62682.
- Johnson MK, Russell AP (2009) Configuration of the setal fields of Rhothropus (Gekkota: Gekkonidae): Functional, evolutionary, ecological and phylogenetic implications of observed pattern. *J Anat* 214(6):937–955.
- Higham TE, Birn-Jeffery AV, Collins CE, Hulsev CD, Russell AP (2015) Adaptive simplification and the evolution of gecko locomotion: Morphological and biomechanical consequences of losing adhesion. *Proc Natl Acad Sci USA* 112(3):809–814.
- Schmidt-Nielsen K (1970) Energy metabolism, body size, and problems of scaling. *Fed Proc* 29(4):1524–1532.
- Du Bois D, Du Bois EF (1916) Clinical calorimetry. Tenth paper. A formula to estimate the approximate surface area if height and weight be known. *Arch Intern Med (Chic)* 17(6_2):863–871.
- Smith JM, Barnes WJP, Downie J, Ruxton G (2006) Adhesion and allometry from metamorphosis to maturation in hylid tree frogs: A sticky problem. *J Zool (Lond)* 270(11):372–383.
- Hanna G, Barnes WJP (1991) Adhesion and detachment of the toe pads of tree frogs. *J Exp Biol* 155(1):103–125.
- Barnes WJP, Olines C, Smith JM (2006) Whole animal measurements of shear and adhesive forces in adult tree frogs: Insights into underlying mechanisms of adhesion obtained from studying the effects of size and scale. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol* 192(11):1179–1191.
- Smith JM, Barnes WJP, Downie JR, Ruxton GD (2006) Structural correlates of increased adhesive efficiency with adult size in the toe pads of hylid tree frogs. *J Comp Physiol A* 192(11):1193–1204.
- Crawford N, Endlein T, Barnes WJP (2012) Self-cleaning in tree frog toe pads; a mechanism for recovering from contamination without the need for grooming. *J Exp Biol* 215(22):3965–3972.
- Arzt E, Gorb S, Spolenak R (2003) From micro to nano contacts in biological attachment devices. *Proc Natl Acad Sci USA* 100(19):10603–10606.
- Peattie AM, Full RJ (2007) Phylogenetic analysis of the scaling of wet and dry biological fibrillar adhesives. *Proc Natl Acad Sci USA* 104(47):18595–18600.
- Gorb S, Gorb E, Kastner V (2001) Scale effects on the attachment pads and friction forces in syrphid flies (Diptera, Syrphidae). *J Exp Biol* 204(8):1421–1431.
- Federle W (2006) Why are so many adhesive pads hairy? *J Exp Biol* 209(14): 2611–2621.
- Arnold SJ (1983) Morphology, performance and fitness. *Am Zool* 23(1):347–361.
- Roth L, Willis E (1952) Tarsal structure and climbing ability of cockroaches. *J Exp Zool* 119(3):483–517.
- Emerson S, Diehl D (1980) Toe pad morphology and mechanisms of sticking in frogs. *Biol J Linn Soc Lond* 13(3):199–216.
- Stork N (1980) Experimental analysis of adhesion of *Chrysolina polita* (Chrysomelidae: Coleoptera) on a variety of surfaces. *J Exp Biol* 88(1):91–107.
- Green D, Alberch P (1981) Interdigital webbing and skin morphology in the neotropical salamander genus *Bolitoglossa* (Amphibia; Plethodontidae). *J Morphol* 170(3):273–282.
- Walker G, Yulf AB, Ratcliffe J (1985) The adhesive organ of the blowfly, *Calliphora vomitoria*: A functional approach (Diptera: Calliphoridae). *J Zool* 205(2):297–307.
- Bauer AM, Good D (1986) Scaling of scansorial surface area in the genus *Gekko*. *Studies in Herpetology*, ed Roček Z (Charles Univ, Prague), pp 363–366.
- Ishii S (1987) Adhesion of a leaf-feeding ladybird *Epilachna vigintioctomaculata* (Coleoptera, Coccinellidae) on a vertically smooth surface. *Appl Entomol Zool (Jpn)* 22(2):222–228.
- Lees AD, Hardie J (1988) The organs of adhesion in the aphid *Megoura viciae*. *J Exp Biol* 136(1):209–228.
- Dixon AFG, Croghan P, Gowing R (1990) The mechanism by which aphids adhere to smooth surfaces. *J Exp Biol* 152:243–253.
- Walker G (1993) Adhesion to smooth surfaces by insects—a review. *Int J Adhes Adhes* 13(1):3–7.
- Irschick DJ, et al. (1996) A comparative analysis of clinging ability among pad-bearing lizards. *Biol J Linn Soc Lond* 59(1):21–35.
- Eisner T, Aneshansley DJ (2000) Defense by foot adhesion in a beetle (*Hemiphaerota cyanea*). *Proc Natl Acad Sci USA* 97(12):6568–6573.
- Federle W, Rohrseitz K, Hölldobler B (2000) Attachment forces of ants measured with a centrifuge: Better ‘wax-runners’ have a poorer attachment to a smooth surface. *J Exp Biol* 203(3):505–512.
- Gorb SN, et al. (2002) Structural design and biomechanics of friction-based releasable attachment devices in insects. *Integr Comp Biol* 42(6):1127–1139.
- Kesel A, Martin A, Seidl T (2004) Getting a grip on spider attachment: An AFM approach to microstructure adhesion in arthropods. *Smart Mater Struct* 13(3):512–518.

57. Elstrott J, Irschick DJ (2004) Evolutionary correlations among morphology, habitat use and clinging performance in caribbean Anolis lizards. *Biol J Linn Soc Lond* 83(3): 389–398.
58. Federle W, Baumgartner W, Hölldobler B (2004) Biomechanics of ant adhesive pads: Frictional forces are rate- and temperature-dependent. *J Exp Biol* 207(1):67–74.
59. Frantsevich L, Gorb S (2004) Structure and mechanics of the tarsal chain in the hornet, *Vespa crabro* (Hymenoptera: Vespidae): Implications on the attachment mechanism. *Arthropod Struct Dev* 33(1):77–89.
60. Zahn M (2006) Allometrie der Haftaarmorphologie bei Arthropoden. Master's thesis (Univ of Würzburg, Würzburg, Germany).
61. Bohn HF (2007) Biomechanik von Insekten-Pflanzen-Interaktionen bei Nepenthes-Kannenpflanzen. PhD thesis (Univ of Würzburg, Würzburg, Germany).
62. Voigt D, Gorb E, Gorb S (2007) Plant surface-bug interactions: *Dicyphus errans* stalking along trichomes. *Arthropod-Plant Interact* 1(4):221–243.
63. Clemente CJ, Federle W (2008) Pushing versus pulling: Division of labour between tarsal attachment pads in cockroaches. *Proc Biol Sci* 275(1640):1329–1336.
64. Drechsler P (2008) Mechanics of adhesion and friction in stick insects and tree frogs. PhD thesis (Univ of Würzburg, Würzburg, Germany).
65. Niewiarowski PH, Lopez S, Ge L, Hagan E, Dhinojwala A (2008) Sticky gecko feet: The role of temperature and humidity. *PLoS One* 3(5):e2192.
66. Frantsevich L, et al. (2008) Adhesive properties of the arolium of a lantern-fly, *Lycorma delicatula* (Auchenorrhyncha, Fulgoroidea). *J Insect Physiol* 54(5):818–827.
67. Al Bitar L, Voigt D, Zebitz CP, Gorb SN (2009) Tarsal morphology and attachment ability of the codling moth *Cydia pomonella* L. (Lepidoptera, Tortricidae) to smooth surfaces. *J Insect Physiol* 55(11):1029–1038.
68. Bullock JMR, Federle W (2009) Division of labour and sex differences between fibrillar, tarsal adhesive pads in beetles: Effective elastic modulus and attachment performance. *J Exp Biol* 212(12):1876–1888.
69. Geiselhardt SF, et al. (2009) Impact of chemical manipulation of tarsal liquids on attachment in the Colorado potato beetle, *Leptinotarsa decemlineata*. *J Insect Physiol* 56(4):398–404.
70. Lüken D, Voigt D, Gorb S, Zebitz C (2009) Die Tarsenmorphologie und die Haftfähigkeit des Schwarzen Batatenkäfers *Cylas puncticollis* (Boheman) auf glatten Oberflächen mit unterschiedlichen physiko-chemischen Eigenschaften. *Mitt Dtsch Ges Allg Angew Entomol* 17:109–113. German.
71. Webster NB, Johnson MK, Russell AP (2009) Ontogenetic scaling of scansorial surface area and setal dimensions of *Chondrodactylus bibronii* (Gekkota: Gekkonidae): Testing predictions derived from cross-species comparisons of gekkotans. *Acta Zool* 90(1):18–29.
72. Gorb EV, Hosoda N, Miksch C, Gorb SN (2010) Slippery pores: Anti-adhesive effect of nanoporous substrates on the beetle attachment system. *J R Soc Interface* 7(52): 1571–1579.
73. Riskin D, Racey P (2010) How do sucker footed bats hold on, and why do they roost head up? *Biol J Linn Soc Lond* 99(12):233–240.
74. Varenberg M, Pugno N, Gorb S (2010) Spatulate structures in biological fibrillar adhesion. *Soft Matter* 6(13):3269–3272.
75. Orchard MJ, Kohonen M, Humphries S (2012) The influence of surface energy on the self-cleaning of insect adhesive devices. *J Exp Biol* 215(2):279–286.
76. Orchard MJ (2012) The functional morphology of insect adhesive devices and its implications for ecology. PhD thesis (Univ of Hull, Kingston upon Hull, UK).
77. Voigt D, Schweikart A, Fery A, Gorb S (2012) Leaf beetle attachment on wrinkles: Isotropic friction on anisotropic surfaces. *J Exp Biol* 215(11):1975–1982.
78. Wolff JO, Gorb SN (2012) Surface roughness effects on attachment ability of the spider *Philodromus dispar* (Araneae, Philodromidae). *J Exp Biol* 215(1):179–184.
79. Anyon MJ (2013) A multi-disciplinary study of insect adhesion: Functional biomechanics and applications. PhD thesis (Univ of Hull, Kingston upon Hull, UK).
80. Endlein T, et al. (2013) Sticking under wet conditions: The remarkable attachment abilities of the torrent frog, *Staurois guttatus*. *PLoS One* 8(9):e73810.
81. Labonte D, Federle W (2013) Functionally different pads on the same foot allow control of attachment: stick insects have load-sensitive “heel” pads for friction and shear-sensitive “toe” pads for adhesion. *PLoS One* 8(12):e81943.
82. Crandell KE, Herrel A, Sasa M, Losos JB, Autumn K (2014) Stick or grip? Co-evolution of adhesive toepads and claws in Anolis lizards. *Zoology (Jena)* 117(6):363–369.
83. Grohmann C, Blankenstein A, Koops S, Gorb SN (2014) Attachment of *Galerucella nymphalae* (Coleoptera, Chrysomelidae) to surfaces with different surface energy. *J Exp Biol* 217(23):4213–4220.
84. Lapinski W, Walther P, Tschapka M (2015) Morphology reflects microhabitat preferences in an assemblage of neotropical wandering spiders. *Zoomorphology* 134(2): 219–236.
85. Luff M (2007) *The Carabidae (Ground Beetles) of Britain and Ireland*. RES Handbooks for the Identification of British Insects (Field Studies Council, Shrewsbury, UK), Vol 4.
86. Biedermann R, Niedringhaus R (2009) *The Plant- and Leafhoppers of Germany: Identification Key to All Species* (Wissenschaftlich Akademischer Buchvertrieb-Fründ, Scheessel, Germany).
87. Kunz G, Nickel H, Niedringhaus R (2011) *Fotoatlas der Zikaden Deutschlands* (Wissenschaftlich Akademischer Buchvertrieb-Fründ, Scheessel, Germany).
88. Schneider CA, Rasband WS, Eliceiri KW (2012) NIH Image to ImageJ: 25 years of image analysis. *Nat Methods* 9(7):671–675.
89. Bullock JMR, Drechsler P, Federle W (2008) Comparison of smooth and hairy attachment pads in insects: Friction, adhesion and mechanisms for direction-dependence. *J Exp Biol* 211(20):3333–3343.
90. Autumn K, Gravish N (2008) Gecko adhesion: Evolutionary nanotechnology. *Philos Trans A Math Phys Eng Sci* 366(1870):1575–1590.
91. Bullock JMR, Federle W (2011) Beetle adhesive hairs differ in stiffness and stickiness: In vivo adhesion measurements on individual setae. *Naturwissenschaften* 98(5):381–387.
92. Peattie AM, Dirks JH, Henriques S, Federle W (2011) Arachnids secrete a fluid over their adhesive pads. *PLoS One* 6(5):e20485.
93. Arnold J (1974) Adaptive features on the tarsi of cockroaches (Insecta: Dictyoptera). *Insect Morphol Embryol* 3(3-4):317–334.
94. Frazier SF, et al. (1999) Elasticity and movements of the cockroach tarsus in walking. *J Comp Physiol* 185(2):157–172.
95. Beutel R, Gorb S (2006) A revised interpretation of attachment structures in Hexapoda with special emphasis on Mantophasmatodea. *Arthropod Syst Phylogeny* 64(1):3–25.
96. Eberhard MJ, et al. (2009) Structure and function of the arolium of Mantophasmatodea (Insecta). *J Morphol* 270(10):1247–1261.
97. Clemente CJ, Dirks JH, Barbero DR, Steiner U, Federle W (2009) Friction ridges in cockroach climbing pads: Anisotropy of shear stress measured on transparent, microstructured substrates. *J Comp Physiol A* 195(9):805–814.
98. Labonte D, Williams JA, Federle W (2014) Surface contact and design of fibrillar ‘friction pads’ in stick insects (*Carausius morosus*): Mechanisms for large friction coefficients and negligible adhesion. *J R Soc Interface* 11(94):20140034.
99. Gorb S, Scherge M (2000) Biological microtribology: Anisotropy in frictional forces of orthopteran attachment pads reflects the ultrastructure of a highly deformable material. *Proc Biol Sci* 267(1449):1239–1244.
100. Grohmann C, Henze MJ, Nørgaard T, Gorb SN (2015) Two functional types of attachment pads on a single foot in the Namibia bush cricket *Acanthoproctus diadematus* (Orthoptera: Tettigoniidae). *Proc Biol Sci* 282(1809):20142976.
101. Drotleff DM, et al. (2015) Morphological studies of the toe pads of the rock frog, *Staurois parvus* (family: Ranidae) and their relevance to the development of new biomimetically inspired reversible adhesives. *Interface Focus* 5(1):20140036.
102. Emerson SSB (1991) The ecomorphology of bornean tree frogs (family Rhacophoridae). *Zool J Linn Soc* 101(4):337–357.
103. Voigt D, Schuppert JM, Dattinger S, Gorb SN (2008) Sexual dimorphism in the attachment ability of the Colorado potato beetle *Leptinotarsa decemlineata* (Coleoptera: Chrysomelidae) to rough substrates. *J Insect Physiol* 54(5):765–776.
104. Al Bitar L, Voigt D, Zebitz CP, Gorb SN (2010) Attachment ability of the codling moth *Cydia pomonella* L. to rough substrates. *J Insect Physiol* 56(12):1966–1972.
105. Labonte D, Federle W (2015) Rate-dependence of ‘wet’ biological adhesives and the function of the pad secretion in insects. *Soft Matter* 11(44):8661–8673.
106. Schluter D (1996) Adaptive radiation along genetic lines of least resistance. *Evolution* 50(5):1766–1774.
107. Pyron RA, Wiens JJ (2011) A large-scale phylogeny of Amphibia including over 2800 species, and a revised classification of extant frogs, salamanders, and caecilians. *Mol Phylogenet Evol* 61(2):543–583.
108. Pyron RA, Burbrink FT, Wiens JJ (2013) A phylogeny and revised classification of Squamata, including 4161 species of lizards and snakes. *BMC Evol Biol* 13(1):93.
109. Misof B, et al. (2014) Phylogenomics resolves the timing and pattern of insect evolution. *Science* 346(6210):763–767.
110. Grandcolas P (1996) The phylogeny of cockroach families: A cladistic appraisal of morpho-anatomical data. *Can J Zool* 74(3):508–527.
111. Bocak L, et al. (2014) Building the coleoptera tree-of-life for > 8000 species: Composition of public DNA data and fit with Linnean classification. *Syst Entomol* 39(1):97–110.
112. Wiegmann BM, et al. (2011) Episodic radiations in the fly tree of life. *Proc Natl Acad Sci USA* 108(14):5690–5695.
113. Hua J, et al. (2008) Comparative and phylogenomic studies on the mitochondrial genomes of Pentatomomorpha (Insecta: Hemiptera: Heteroptera). *BMC Genomics* 9(1): 610.
114. Cryan JR, Urban JM (2012) Higher-level phylogeny of the insect order Hemiptera: Is Auchenorrhyncha really paraphyletic? *Syst Entomol* 37(1):7–21.
115. Astruc C, Julien JF, Errard C, Lenoir A (2004) Phylogeny of ants (Formicidae) based on morphology and DNA sequence data. *Mol Phylogenet Evol* 31(3):880–893.
116. Maddison DR, Schulz K-S, eds (2007) The Tree of Life Web Project. Available at tolweb.org. Accessed March 2015.
117. Hansen TF, Pienaar J, Orzack SH (2008) A comparative method for studying adaptation to a randomly evolving environment. *Evolution* 62(8):1965–1977.
118. Vojte KL, Hansen TF (2013) Evolution of static allometries: Adaptive change in allometric slopes of eye span in stalk-eyed flies. *Evolution* 67(2):453–467.
119. Paradis E (2013) Molecular dating of phylogenies by likelihood methods: A comparison of models and a new information criterion. *Mol Phylogenet Evol* 67(2): 436–444.
120. Pagel M (1999) Inferring the historical patterns of biological evolution. *Nature* 401(6756):877–884.
121. Revell LJ (2010) Phylogenetic signal and linear regression on species data. *Methods Ecol Evol* 1(4):319–329.
122. Paradis E, Claude J, Strimmer K (2004) APE: Analyses of phylogenetics and evolution in R language. *Bioinformatics* 20(2):289–290.
123. Revell LJ (2012) phytools: an R package for phylogenetic comparative biology (and other things). *Methods Ecol Evol* 3(2):217–223.
124. Harvey PH, Pagel MD (1991) *The Comparative Method in Evolutionary Biology* (Oxford Univ Press, Oxford), Vol 239.
125. Warton DI, Wright IJ, Falster DS, Westoby M (2006) Bivariate line-fitting methods for allometry. *Biol Rev Camb Philos Soc* 81(2):259–291.
126. Hansen TF, Bartoszek K (2012) Interpreting the evolutionary regression: The interplay between observational and biological errors in phylogenetic comparative studies. *Syst Biol* 61(3):413–425.
127. Revell LJ (2013) Two new graphical methods for mapping trait evolution on phylogenies. *Methods Ecol Evol* 4(8):754–759.
128. Powell GL, Russell AP (1992) Locomotor correlates of ecomorph designation in Anolis: An examination of three sympatric species from Jamaica. *Can J Zool* 70(4):725–739.