

P2X ion channel receptors and inflammation

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Abstract Neuroinflammation limits tissue damage in response to pathogens or injury and promotes repair. There are two stages of inflammation, initiation and resolution. P2X receptors are gaining attention in relation to immunology and inflammation. The P2X7 receptor in particular appears to be an essential immunomodulatory receptor, although P2X1 and P2X4 receptors also appear to be involved. ATP released from damaged or infected cells causes inflammation by release of inflammatory cytokines via P2X7 receptors and acts as a danger signal by occupying upregulated P2X receptors on immune cells to increase immune responses. The purinergic involvement in inflammation is being explored for the development of novel therapeutic strategies.

Keywords ATP · P2X4 receptors · P2X7 receptors · Inflammasome · Cytokines

Introduction

Inflammation involves a complex haemostatic mechanism that enables the body to detect and fight foreign antigens and restore tissue integrity. ATP serves as an acute ‘danger signal’ and behaves as a mediator of inflammation and immunity [1, 2]. Purinergic signalling contributes to the fine tuning of inflammation and immune responses in such a way that the

danger to the host is eliminated efficiently with minimal damage to healthy tissues [3]. Brain inflammation occurs following responses to insults, such as bacterial and viral infection, stroke, traumatic injury, and neurodegenerative disorders. During the course of inflammation, there is upregulation of P2X purinoceptors located on immune cells (neutrophils, eosinophils, monocytes, macrophages, mast cells, and lymphocytes). ATP release from injured cells enhances the inflammatory response through increased synthesis of prostaglandin E₂ (PGE₂) [4] via P2X7 receptors [5]. P2X receptor involvement in inflammation also occurs in irritable bowel syndrome [6, 7], lung injury and fibrosis [8, 9], systemic inflammation [10], arthritis [11], fever [12], and rhinosinusitis [13]. Purinergic signalling in different inflammatory cells involves purinoceptor responses in immune cells (see [14]). Microglia are immune cells in the central nervous system (CNS) [15]. They mediate neuroinflammatory responses to insult in response to a variety of triggers, including toxic metabolites and autoimmunity by detection of pathogens [16]. In addition to microglia, astrocytes as well as perivascular monocytes and macrophages invading to sites of insult from the circulation promote neuroinflammation [17]. Neuronal activity also contributes to inflammation [18]. Activation of P2X7 receptors promotes neuroinflammation by causing the release of inflammatory cytokines, such as interleukin (IL)-1 β and tumour necrosis factor- α [19–21]. P2X3 receptors are upregulated in the colonic mucosa of humans with inflammatory bowel disease [22]. There is increased release of ATP from endothelial cells during acute inflammation [23]. ATP triggers cytokine release from inflammatory cells, acts as a chemotactic factor and, after breakdown by ectoenzymes to adenosine, is a potent immunosuppressant [24, 25]. ATP may reach a concentration of several hundred micromoles within the interstitium of inflamed tissues [26, 27]. P2X receptors play a central role in inflammation, particularly the P2X7 receptor. P2X1 receptors

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[28, 29] and P2X4 receptors [30] probably also play a role in inflammation and immunity (Fig. 1).

Multiple inflammatory mediators, including cytokines, chemokines, and prostaglandins, are elevated in the cerebrospinal fluid and in post-mortem brain tissues of patients with a history of neuroinflammatory conditions, as well as neurodegenerative diseases [31]. P2X receptors are involved in

immune-related neuroinflammatory dysfunctions, including ischaemia and neurodegenerative diseases (see [32]).

Activation of an inflammasome, a protein complex consisting of caspase-1, apoptosis-associated speck-like protein, and nod-like receptor proteins (NLRP1 or NLRP3) [33] expressed in myeloid immune precursor cells is involved. NLRP inflammasomes are activated by the recognition of pathogens-

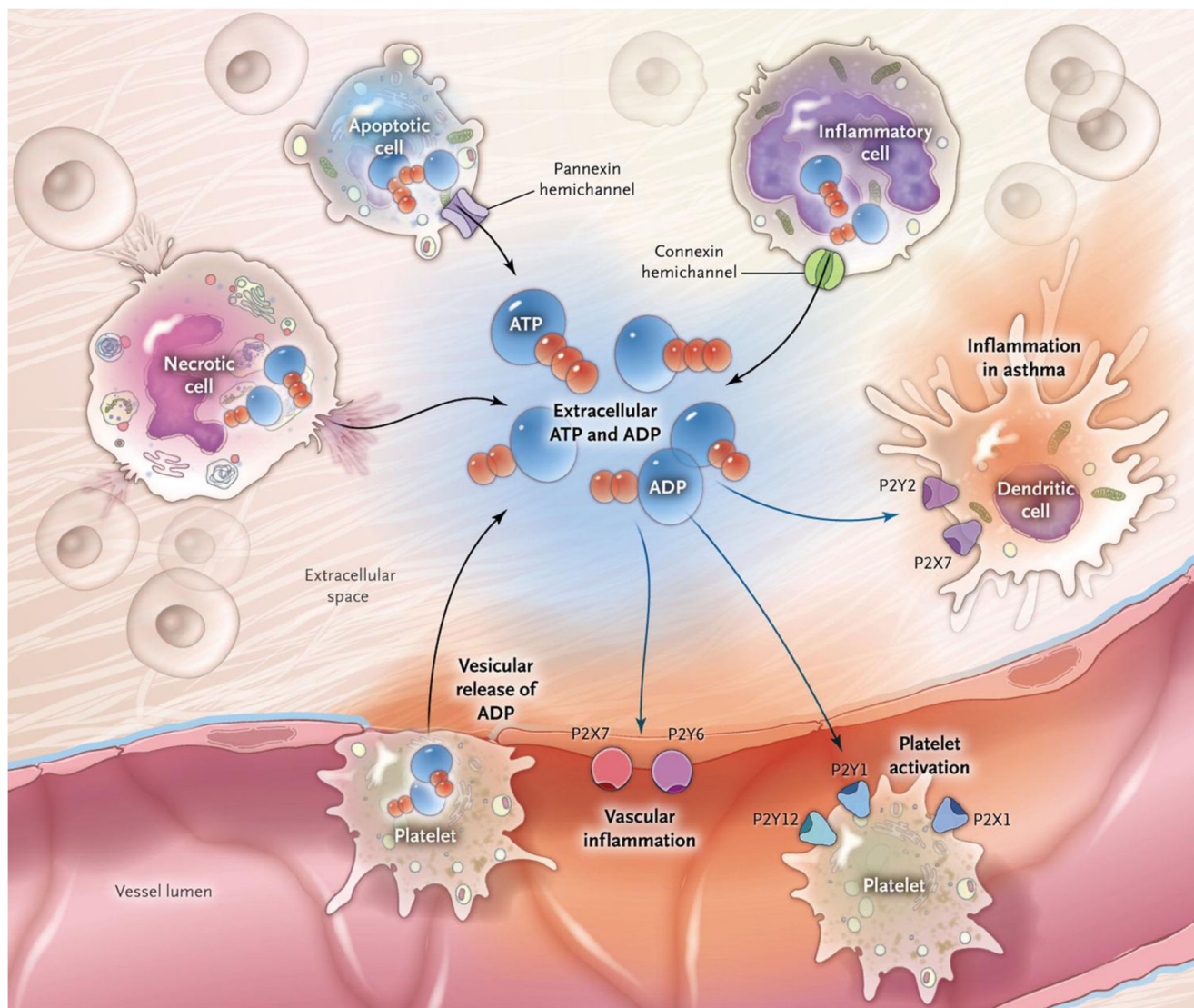


Fig. 1 Release of extracellular adenosine triphosphate (ATP) and adenosine diphosphate (ADP) and activation of ATP (P2) receptors during inflammation. During inflammatory conditions that occur in vascular thrombosis, hypoxia, ischemia, inflammatory bowel disease, and acute lung injury, multiple cell types release nucleotides, typically in the form of ATP or ADP, from the intracellular compartment into the extracellular space. The release of nucleotides includes release of ATP from necrotic cells, pannexin-hemichannel-dependent release of ATP during apoptosis, and release of ATP through connexin hemichannels from activated inflammatory cells such as polymorphonuclear granulocytes (neutrophils). In addition, release of extracellular ATP has been shown to occur through vesicular exocytosis or connexin hemichannels from endothelial and urothelial cells, osteoblasts, and astrocytes, as well as nerves (not shown). An additional source of

extracellular nucleotides in inflammatory conditions is provided by activated platelets, which release ATP and ADP through the release of granules and exocytosis. In the extracellular space, these nucleotides function as signalling molecules that can activate P2Y receptors (G protein-coupled receptors) or P2X receptors (ligand-gated ion channels). Examples of nucleotide-receptor signalling in inflammatory conditions include P2Y6- or P2X7-receptor signalling, which mediates vascular inflammation, and P2Y1-, P2X1-, and P2Y12-receptor signalling, which mediate platelet activation. Activation of P2 receptors of the P2Y2 and P2X7 family that are expressed on dendritic cells is thought to play a role in promoting lung inflammation in chronic lung diseases such as asthma (reproduced from [9], with permission from the Massachusetts Medical Society)

associated molecular patterns or damage-associated molecular patterns (DAMPs) [34]. Inflammasomes are involved in P2X7 receptor coupling to IL-1 β release [19].

ATP release occurs from damaged cells at the site of injury and from activated immune cells, glial cells, and endothelial cells. ATP release *in vivo* has also been shown in response to contact allergens [35], irradiation, allograft rejection [36], and intraperitoneal lipopolysaccharide administration [4], as well as mechanical distortion [37, 38]. ATP released during viral infection is an important inflammatory regulator that activates the inflammasome pathway and regulates inflammatory responses [39]. P2X4 receptors were claimed to influence inflammasome activation after spinal cord injury [40]. The secretion of ATP by bacteria infected macrophages leads to activation of P2X7 receptors [41]. Prevention of cell death or ATP release through p38 or AKT activation interfered with inflammasome activation and IL-1 β production. Overexpression of P2X7 receptors was reported in the intestinal mucosa of Crohn's (inflammatory) disease patients [42].

Reviews focussing on nucleotide signalling during inflammation are available [14, 43–47].

Inflammation and P2X receptors

Changes in P2X receptor subtype expression in neuroinflammatory conditions in various *in vitro* and *in vivo* models have been reported. P2X4 receptors are associated with an early inflammatory mediator, PGE₂ [30]. P2X4 receptors, similar to P2X7, form a large conductance pore on the cell membrane, facilitating ion efflux and subsequent inflammasome activation [5]. The P2X4 receptor may act as an initial trigger, while the P2X7 receptor, in concert with pannexin 1, may amplify the signal [47]. The P2X4 receptor contribution to PGE₂ release in mice is of minor relevance when compared to that of P2X7 receptors [4].

Of the seven P2X subtypes, the P2X7 receptor is the most important for involvement in mediating neuroinflammation [20]. Activation of P2X7 receptors results in DAMP, initiating neuroinflammatory cascades [5]. Further, the formation of the P2X7 receptor pore appears to be necessary for activating the inflammasome [48]. The P2X7 receptor is one of the most potent plasma membrane receptors responsible for the release of inflammatory cytokines of the IL-1 family, IL2, IL6, and IL18 [45, 49, 50]. P2X7 receptor activation is a strong stimulus for IL-18 as well as IL-1 β [51, 52] and IL-1 α secretion [53]. Microglia are the main source of IL-1 β release, but it has also been claimed that IL-1 β release from neurons is important [40]. IL-2 synthesis in lymphocytes requires functional P2X receptors [54], probably P2X7 [55, 56]. P2X7 receptors also mediate biglycan-stimulated IL-1 β release from mouse macrophages [57]. A P2X7 receptor-P2X4 receptor interaction in the process of IL-1 β and IL-18 release has been

identified in bone marrow-derived dendritic cells [58]. Smoking contributes to the pro-inflammatory status of perivascular visceral adipose tissue by enhancing the expression and activity of the P2X7 receptor-inflammasome complex [59]. Stimulation of P2X7 receptors drives release of both exosomes and microvesicles from several different cell types relevant to inflammation.

P2X7 receptors are expressed on glial and immune cells of monocyte-macrophage origin and on presynaptic terminals on neurons, with the highest levels on microglia [60–63]. P2X7 receptors, acting via different pathways, play a major role in the promotion as well as in the suppression of inflammation in different pathophysiological conditions ([64, 65] and see [46]). P2X7 receptors mediate transforming growth factor β secretion. P2X7 receptor activation also releases a potent immunosuppressive agent, HLA-G [66, 67] and vascular endothelial growth factor, another major player in inflammation [68]. Another function of P2X7 receptors in inflammation is the activation of transcription factors such as NF κ B and NFAT [69, 70]. P2X7 receptor activation opens a cation-specific channel activating several pathways, including the inflammasome, leading to the stress-activated protein kinase pathway that results in apoptosis, and the mitogen-activated protein kinase pathway. Ectonucleotidases control P2X7 receptor function, including the resolution as well as the initial phases of inflammation (see [71]). [¹¹C]-A-740003, a P2X7 receptor antagonist, has been used as a novel tracer of neuroinflammation [72].

P2X7 receptors trigger the activation of the NLRP3 inflammasome, the main intracellular complex involved in the transduction of danger signals and in the initiation of inflammation [73]. The role of the NLRP3 inflammasome in pro-IL-1 β processing and pyroptosis places the P2X7 receptor at the centre of cytokine immunology. Since the discovery of the inflammasome [33], whether activation by pathogen- and damage-associated molecular patterns requires a direct, physical, interaction with the scaffold NLR inflammasome proteins has been discussed. Ca²⁺ might be a suitable second messenger responsible for inflammasome activation [39], and this would be consistent with the role of the P2X7 receptor as a trigger of the NLRP3 inflammasome since P2X7 receptor opening drives a large Ca²⁺ influx from the extracellular space.

ATP, the extracellular messenger of cellular injury, accumulates to hundred micromolar levels at sites of injury and inflammation [26, 27]. In the presence of inflammation or stress, there is a fast increase of extracellular ATP to near millimolar levels quickly mediating stimulation of pro-inflammatory pathways [74]. Some P2X7 receptor polymorphisms appear to protect against infection, but others increase the risk of developing chronic inflammatory diseases [75].

Immune cells and inflammation

The participation of P2X receptors in inflammation and immunity is gaining attention probably because of the role played by P2X7 receptors in IL-1 β processing and release. All immune cells, whether of the myeloid or lymphoid lineage, express at least one P2X receptor subtype, and many express all seven subtypes [2, 45, 76–81]. Mononuclear phagocytes are the inflammatory cell type where P2X receptor expression has been best characterized [82]. Monocyte/macrophage and myeloid dendritic cells express P2X1, P2X4, and P2X7 receptors [45]. P2X5 receptors are expressed by T lymphocytes. The function of P2X5 receptors in inflammation is not clear. Neutrophils and eosinophils express P2X1, P2X4, and P2X7 receptors, although the level of expression is different in the quiescent or activated state [83–85]. P2X1, P2X4, and P2X7 receptors are expressed on T and B lymphocytes and natural killer cells [45]. ATP released by tissue damage, acts as a danger signal by acting on P2 receptors on immune cells to stimulate the immune response [86]. P2 receptors are present on immune cells and their expression is modulated by inflammatory cytokines [87]. P2X receptors have been implicated in the participation of the immune system in inflammatory pain [88, 89]. P2X1, P2X4, P2X7, and perhaps P2X3 receptors are expressed by mast cells [90, 91]. Mast cells were the cell type in which the properties of the P2X7 receptor were initially observed and characterized by Cockcroft and Gomperts [92]. P2X receptor expression is also present on microglia, both in vitro and in vivo, particularly P2X4 and P2X7 receptors [93, 94]. P2X receptors on mast cells are involved in the pathogenesis of chronic airway allergic inflammation [91].

Inflammatory pain

P2X7 receptors are involved in inflammatory pain [95–99]. There is reduced inflammation-induced hyperalgesia in rats following treatment with oxidized ATP, a P2X7 receptor antagonist [100]. P2X7 receptors play a transductional role in the development of inflammatory pain [101].

A review includes a discussion of the role of P2X3 receptors in inflammatory pain [102]. During the inflammatory process in peripheral tissue, neither prostaglandins nor sympathetic amines can sensitize primary afferent neurons by themselves; they depend on previous neuronal P2X3 receptor activation [103]. Spontaneous and evoked responses of spinal nociceptive neurons are attenuated by P2X3 receptor antagonism in inflamed rats [104]. Data has been presented to indicate that antagonism of spinal P2X3/P2X2/3 receptors regulates an indirect activation of the opioid system to alleviate inflammatory hyperalgesia [105]. P2X4 receptors probably

also participate because of their involvement in neuropathic pain [106, 107], which is relevant for inflammation. Mice lacking P2X4 receptors show impaired inflammasome activation [40] and do not develop pain hypersensitivity in response to inflammatory agents, and this is paralleled by a complete absence of PGE₂ in inflammatory exudates [30]. There is also a suggestion that P2X4 receptors might modulate P2X7 receptor activity [56]. P2X7 receptor antagonists reduce inflammatory pain in rats [100, 108–110]. Chronic inflammatory pain was abolished in P2X7 receptor knockout mice [95]. Central sensitization of nociceptive neurons in medullary dorsal horn of rats involves P2X7 receptors [111]. P2X7 receptor antagonism of chronic pain is likely mediated through immunoneural interactions that affect the release of inflammatory cytokines [112]. Inflammatory pain involved in dressing changes of burn patients was relieved by puerarin, an isoflavonoid derived from a Chinese herb [113]. The effects were correlated with the decreased expression of P2X7 receptor mRNA and protein in peripheral blood mononuclear cells in burn patients.

Pathology and inflammation

Purinergic contributions to neuroinflammation in relation to disorders of the CNS are being explored. Pathological neuroinflammation, promoting apoptosis and necrosis, and influencing the synaptic and intrinsic membrane properties of neurons contributes to CNS pathologies [114]. A role for neuroinflammation occurs in neurodegenerative diseases, such as Alzheimer's disease, Parkinson's disease, multiple sclerosis, amyotrophic lateral sclerosis, Huntington's disease, stroke, and epilepsy [115]. Neuroinflammation is also a pathological factor in psychiatric mood disorders [116, 117]. The NLRP3 inflammasome is a central mediator of systemic inflammation and a link between psychological stress and the emergence of depression and other psychiatric illnesses [118] and ATP, accumulated following insult, induces NLRP-mediated IL-1 β processing [93]. Epidemiological and gene-linking studies have implicated P2X7 in a host of CNS diseases [119, 120]. Neuroimmunological changes occur in psychiatric disorders, including major depressive disorder, bipolar disorder, obsessive compulsive disorder, and schizophrenia. Chronic inflammation associated with diabetes, obesity, or autoimmune diseases increases the risk of psychiatric disorders (see [47]). These disorders are characterized by chronic, low grade, or intermittent inflammation, in contrast to neurodegenerative diseases, where there is acute inflammation in the brain parenchyma. Schizophrenia is considered to be a neurodevelopmental disorder, and foetal neuroinflammation, resulting from maternal infection, is implicated [116]. Enhanced levels of pro-inflammatory cytokines in the brain

and enhanced microglial activation occur in foetal neuroinflammation, leading to abnormal brain maturation.

Associations between susceptibility or resistance to parasites and bacteria and loss- or gain-of-function polymorphisms in the P2X7 receptor indicate that it is important in infectious disease [121]. ATP activation of the NLRP3 inflammasome protects mice against bacterial infection [122]. The P2X7 receptor plays a role in acute and chronic stages of infection as well as a ‘danger signal’ in the initial stages of inflammation (see [71]).

Therapeutic potential

Purinergic-based therapies may be useful to halt excessive inflammation and promote repair of neuroinflammatory disorders [4, 80, 123, 124].

P2X7 receptor antagonists are promising targets for anti-inflammatory therapy [125, 126], including inflammation in the CNS [11, 95, 100, 127, 128]. In view of its potent pro-inflammatory effect, the analgesic activity of P2X7 receptor blockers is of interest for therapeutic implications [99]. Blockade of P2X7 receptors reduced nociception in animal models of chronic inflammatory pain [96, 125, 129–132]. Relief of inflammatory pain was produced by the P2X7 receptor antagonist, oxidized ATP, in arthritic rats [11]. Blockade by the selective P2X7 receptor antagonist, A-839977, was lost in IL-1 α knockout mice [133].

P2X3 receptor antagonists have also been suggested to be a therapeutic target for pain therapy [134]. Application of apyrase to CD39-deficient mice prior to ischaemia reduced infarct volumes and neutrophil counts [135, 136]. Kinase inhibitors have been recommended for the treatment of inflammatory and autoimmune disorders, such as rheumatoid arthritis, psoriasis, organ transplantation, and autoimmune diseases [137, 138].

Activation of the purinergic pathway may be implicated in transplantation-related injuries. Following transplantation, ATP, the pro-inflammatory danger signal, is released from damaged cells to promote proliferation of immune cells, T cell activation, and inflammation. Targeting purinoceptors may promote immunosuppression and reduce inflammation. The ectonucleotidases, CD39 and CD73, hydrolyze ATP to the anti-inflammatory mediator adenosine, which suppresses pro-inflammatory cytokine production leading to improved graft survival. The mechanisms of action of several immunosuppressive drugs, such as calcineurin and mTOR inhibitors, involve purinergic signalling. Targeting the purinergic signalling pathway by increasing ectonucleotidase activity and/or boosting short term adenosine-mediated immunosuppression have potential in preventing allograft vascular injury, ameliorating rejection, and promoting tolerance.

Conclusion

P2X receptors mediate the responses to ATP, one of the most ancient evolutionary extracellular messengers (see [139]). ATP is an intracellular molecule, so its release is suited to signal cell distress or injury. This ‘danger signal role’ of ATP became more and more relevant in multicellular animals. P2X receptors play important roles in pathophysiology (see [140, 141]) and P2X7 receptors, in particular, are vitally involved in inflammation.

References

1. Di Virgilio F, Ferrara N, Idzko M, Panther E, Norgauer J, La Sala A, Girolomoni G (2003) Extracellular ATP, receptors and inflammation. *Drug Dev Res* 59:171–174
2. Di Virgilio F, Boeynaems JM, Robson SC (2009) Extracellular nucleotides as negative modulators of immunity. *Curr Opin Pharmacol* 9:507–513
3. Bours MJ, Swennen EL, Di Virgilio F, Cronstein BN, Dagnelie PC (2006) Adenosine 5'-triphosphate and adenosine as endogenous signaling molecules in immunity and inflammation. *Pharmacol Ther* 112:358–404
4. Barberà-Cremades M, Baroja-Mazo A, Gomez AI, Machado F, Di Virgilio F, Pelegrín P (2012) P2X7 receptor-stimulation causes fever via PGE2 and IL-1 β release. *FASEB J* 26:2951–2962
5. Fiebich BL, Akter S, Akundi RS (2014) The two-hit hypothesis for neuroinflammation: role of exogenous ATP in modulating inflammation in the brain. *Front Cell Neurosci* 8:260
6. De Man JG, Seerden TC, De Winter BY, Van Marck EA, Herman AG, Pelckmans PA (2003) Alteration of the purinergic modulation of enteric neurotransmission in the mouse ileum during chronic intestinal inflammation. *Br J Pharmacol* 139:172–184
7. Kurashima Y, Kiyono H, Kunisawa J (2015) Pathophysiological role of extracellular purinergic mediators in the control of intestinal inflammation. *Mediators Inflamm* 2015:427125
8. Riteau N, Gasse P, Fauconnier L, Gombault A, Couegnat M, Fick L, Kanellopoulos J, Quesniaux VF, Marchand-Adam S, Crestani B, Ryffel B, Couillin I (2010) Extracellular ATP is a danger signal activating P2X₇ receptor in lung inflammation and fibrosis. *Am J Respir Crit Care Med* 182:774–783
9. Eltzschig HK, Sitkovsky MV, Robson SC (2012) Purinergic signaling during inflammation. *N Engl J Med* 367:2322–2333
10. Cauwels A, Rogge E, Vandendriessche B, Shiva S, Brouckaert P (2014) Extracellular ATP drives systemic inflammation, tissue damage and mortality. *Cell Death Dis* 5:e1102
11. Dell'Antonio G, Quattrini A, Dal Cin E, Fulgenzi A, Ferrero ME (2002) Antinociceptive effect of a new P_{2Z}/P2X₇ antagonist, oxidized ATP, in arthritic rats. *Neurosci Lett* 327:87–90
12. Gourine AV, Dale N, Gourine VN, Spyer KM (2004) Fever in systemic inflammation: roles of purines. *Front Biosci* 9:1011–1022
13. Kim IS, Rhee CS, Lee JH, Heo JH, Park J, Lee CH (2007) Effects of purinergic stimulation on ciliary beat frequency and chloride secretion in sinusitis. *Laryngoscope* 117:1677–1682
14. Jacob F, Pérez Novo C, Bachert C, Van CK (2013) Purinergic signaling in inflammatory cells: P2 receptor expression, functional effects, and modulation of inflammatory responses. *Purinergic Signal* 9:285–306

15. Streit WJ, Mrak RE, Griffin WS (2004) Microglia and neuroinflammation: a pathological perspective. *J Neuroinflammation* 1:14
16. Gendelman HE (2002) Neural immunity: friend or foe? *J Neurovirol* 8:474–479
17. Yamasaki R, Lu H, Butovsky O, Ohno N, Rietsch AM, Cialic R, Wu PM, Doykan CE, Lin J, Coteleur AC, Kidd G, Zorlu MM, Sun N, Hu W, Liu L, Lee JC, Taylor SE, Uehlein L, Dixon D, Gu J, Floruta CM, Zhu M, Charo IF, Weiner HL, Ransohoff RM (2014) Differential roles of microglia and monocytes in the inflamed central nervous system. *J Exp Med* 211:1533–1549
18. Xanthos DN, Sandkuhler J (2014) Neurogenic neuroinflammation: inflammatory CNS reactions in response to neuronal activity. *Nat Rev Neurosci* 15:43–53
19. Di Virgilio F (2007) Liaisons dangereuses: P2X₇ and the inflammasome. *Trends Pharmacol Sci* 28:465–472
20. Lister MF, Sharkey J, Sawatzky DA, Hodgkiss JP, Davidson DJ, Rossi AG, Finlayson K (2007) The role of the purinergic P2X₇ receptor in inflammation. *J Inflamm (Lond)* 4:5
21. Tschopp J, Schroder K (2010) NLRP3 inflammasome activation: the convergence of multiple signalling pathways on ROS production? *Nat Rev Immunol* 10:210–215
22. Yiangou Y, Facer P, Baecker PA, Ford AP, Knowles CH, Chan CL, Williams NS, Anand P (2001) ATP-gated ion channel P2X₃ is increased in human inflammatory bowel disease. *Neurogastroenterol Motil* 13:365–369
23. Bodin P, Burnstock G (1998) Increased release of ATP from endothelial cells during acute inflammation. *Inflamm Res* 47:351–354
24. Hasko G, Linden J, Cronstein B, Pacher P (2008) Adenosine receptors: therapeutic aspects for inflammatory and immune diseases. *Nat Rev Drug Discov* 7:759–770
25. Rayah A, Kanellopoulos JM, Di Virgilio F (2012) P2 receptors and immunity. *Microbes Infect* 14:1254–1262
26. Pellegatti P, Raffaghello L, Bianchi G, Piccardi F, Pistoia V, Di Virgilio F (2008) Increased level of extracellular ATP at tumor sites: in vivo imaging with plasma membrane luciferase. *PLoS One* 3:e2599
27. Falzoni S, Donvito G, Di Virgilio F (2013) Detecting adenosine triphosphate in the pericellular space. *Interface Focus* 3:20120101
28. Chvatchko Y, Valera S, Aubry JP, Renno T, Buell G, Bonnefoy JY (1996) The involvement of an ATP-gated ion channel, P_{2X1}, in thymocyte apoptosis. *Immunity* 5:275–283
29. Woehrle T, Yip L, Elkhali A, Sumi Y, Chen Y, Yao Y, Insel PA, Junger WG (2010) Pannexin-1 hemichannel-mediated ATP release together with P2X₁ and P2X₄ receptors regulate T-cell activation at the immune synapse. *Blood* 116:3475–3484
30. Ulmann L, Hirbec H, Rassendren F (2010) P2X₄ receptors mediate PGE₂ release by tissue-resident macrophages and initiate inflammatory pain. *EMBO J* 29:2290–2300
31. Chakraborty S, Kaushik DK, Gupta M, Basu A (2010) Inflammasome signaling at the heart of central nervous system pathology. *J Neurosci Res* 88:1615–1631
32. Apolloni S, Montilli C, Finocchi P, Amadio S (2009) Membrane compartments and purinergic signalling: P2X receptors in neurodegenerative and neuroinflammatory events. *FEBS J* 276:354–364
33. Martinon F, Burns K, Tschopp J (2002) The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL-beta. *Mol Cell* 10:417–426
34. Bernier LP (2012) Purinergic regulation of inflammasome activation after central nervous system injury. *J Gen Physiol* 140:571–575
35. Weber FC, Esser PR, Müller T, Ganesan J, Pellegatti P, Simon MM, Zeiser R, Idzko M, Jakob T, Martin SF (2010) Lack of the purinergic receptor P2X₇ results in resistance to contact hypersensitivity. *J Exp Med* 207:2609–2619
36. Wilhelm K, Ganesan J, Muller T, Durr C, Grimm M, Beilhack A, Krempl CD, Sorichter S, Gerlach UV, Juttner E, Zerweck A, Gartner F, Pellegatti P, Di Virgilio F, Ferrari D, Kambham N, Fisch P, Finke J, Idzko M, Zeiser R (2010) Graft-versus-host disease is enhanced by extracellular ATP activating P2X_{7R}. *Nat Med* 16:1434–1438
37. Burnstock G (1999) Release of vasoactive substances from endothelial cells by shear stress and purinergic mechanosensory transduction. *J Anat* 194:335–342
38. Bodin P, Burnstock G (2001) Purinergic signalling: ATP release. *Neurochem Res* 26:959–969
39. Lee BH, Hwang DM, Palaniyar N, Grinstein S, Philpott DJ, Hu J (2012) Activation of P2X₇ receptor by ATP plays an important role in regulating inflammatory responses during acute viral infection. *PLoS One* 7:e35812
40. de Rivero Vaccari JP, Bastien D, Yurcisin G, Pineau I, Dietrich WD, De Koninck Y, Keane RW, Lacroix S (2012) P2X₄ receptors influence inflammasome activation after spinal cord injury. *J Neurosci* 32:3058–3066
41. Ali SR, Timmer AM, Bilgrami S, Park EJ, Eckmann L, Nizet V, Karin M (2011) Anthrax toxin induces macrophage death by p38 MAPK inhibition but leads to inflammasome activation via ATP leakage. *Immunity* 35:34–44
42. Neves AR, Castelo-Branco MT, Figliuolo VR, Bernardazzi C, Buongusto F, Yoshimoto A, Nanini HF, Coutinho CM, Carneiro AJ, Coutinho-Silva R, de Souza HS (2014) Overexpression of ATP-activated P2X₇ receptors in the intestinal mucosa is implicated in the pathogenesis of Crohn's disease. *Inflamm Bowel Dis* 20:444–457
43. Di Virgilio F, Falzoni S, Mutini C, Sanz JM, Chiozzi P (1998) Purinergic P2X₇ receptor: a pivotal role in inflammation and immunomodulation. *Drug Dev Res* 45:207–213
44. Bours MJ, Dagnelie PC, Giuliani AL, Wesselius A, Di Virgilio F (2011) P2 receptors and extracellular ATP: a novel homeostatic pathway in inflammation. *Front Biosci (Schol Ed)* 3:1443–1456
45. Idzko M, Ferrari D, Eltzschig HK (2014) Nucleotide signalling during inflammation. *Nature* 509:310–317
46. Di Virgilio F (2015) P2X receptors and inflammation. *Curr Med Chem* 22:866–877
47. Beamer E, Göllöncsér F, Horváth G, Bekö K, Otrokocsi L, Koványi B, Sperlágh B (2015) Purinergic mechanisms in neuroinflammation: an update from molecules to behavior. *Neuropharmacology* In Press
48. Monif M, Reid CA, Powell KL, Smart ML, Williams DA (2009) The P2X₇ receptor drives microglial activation and proliferation: a trophic role for P2X_{7R} pore. *J Neurosci* 29:3781–3791
49. Solini A, Chiozzi P, Morelli A, Fellin R, Di Virgilio F (1999) Human primary fibroblasts in vitro express a purinergic P2X₇ receptor coupled to ion fluxes, microvesicle formation and IL-6 release. *J Cell Sci* 112:297–305
50. Ferrari D, Pizzirani C, Adinolfi E, Lemoli RM, Curti A, Idzko M, Panther E, Di Virgilio F (2006) The P2X₇ receptor: a key player in IL-1 processing and release. *J Immunol* 176:3877–3883
51. Perregaux DG, McNiff P, Laliberte R, Conklyn M, Gabel CA (2000) ATP acts as an agonist to promote stimulus-induced secretion of IL-1β and IL-18 in human blood. *J Immunol* 165:4615–4623
52. Mehta VB, Hart J, Wewers MD (2001) ATP-stimulated release of interleukin (IL)-1β and IL-18 requires priming by lipopolysaccharide and is independent of caspase-1 cleavage. *J Biol Chem* 276:3820–3826
53. Pelegrin P, Barroso-Gutierrez C, Surprenant A (2008) P2X₇ receptor differentially couples to distinct release pathways for IL-1β in mouse macrophage. *J Immunol* 180:7147–7157
54. Yip L, Woehrle T, Corriden R, Hirsh M, Chen Y, Inoue Y, Ferrari V, Insel PA, Junger WG (2009) Autocrine regulation of T-cell

- activation by ATP release and P2X₇ receptors. *FASEB J* 23:1685–1693
55. Newbolt A, Stoop R, Virginio C, Surprenant A, North RA, Buell G, Rassendren F (1998) Membrane topology of an ATP-gated ion channel (P2X receptor). *J Biol Chem* 273:15177–15182
 56. Antonio LS, Stewart AP, Xu XJ, Varanda WA, Murrell-Lagnado RD, Edwardson JM (2011) P2X₄ receptors interact with both P2X₂ and P2X₇ receptors in the form of homotrimeric. *Br J Pharmacol* 163:1069–1077
 57. Babelova A, Moreth K, Tsalastra-Greul W, Zeng-Brouwers J, Eickelberg O, Young MF, Bruckner P, Pfeilschifter J, Schaefer RM, Grone HJ, Schaefer L (2009) Biglycan, a danger signal that activates the NLRP3 inflammasome via toll-like and P2X receptors. *J Biol Chem* 284:24035–24048
 58. Sakaki H, Tsukimoto M, Harada H, Moriyama Y, Kojima S (2013) Autocrine regulation of macrophage activation via exocytosis of ATP and activation of P2Y₁₁ receptor. *PLoS One* 8:e59778
 59. Rossi C, Santini E, Chiarugi M, Salvati A, Comassi M, Vitolo E, Madec S, Solini A (2014) The complex P2X₇ receptor/inflammasome in perivascular fat tissue of heavy smokers. *Eur J Clin Invest* 44:295–302
 60. Choi HB, Ryu JK, Kim SU, McLarnon JG (2007) Modulation of the purinergic P2X₇ receptor attenuates lipopolysaccharide-mediated microglial activation and neuronal damage in inflamed brain. *J Neurosci* 27:4957–4968
 61. Matute C, Torre I, Pérez-Cerdá F, Pérez-Samartín A, Alberdi E, Etxebarria E, Arranz AM, Ravid R, Rodríguez-Antiguedad A, Sánchez-Gómez M, Domercq M (2007) P2X₇ receptor blockade prevents ATP excitotoxicity in oligodendrocytes and ameliorates experimental autoimmune encephalomyelitis. *J Neurosci* 27:9525–9533
 62. Skaper SD, Debetto P, Giusti P (2010) The P2X₇ purinergic receptor: from physiology to neurological disorders. *FASEB J* 24:337–345
 63. Weisman GA, Camden JM, Peterson TS, Ajit D, Woods LT, Erb L (2012) P2 receptors for extracellular nucleotides in the central nervous system: role of P2X₇ and P2Y₂ receptor interactions in neuroinflammation. *Mol Neurobiol* 46:96–113
 64. Solle M, Labasi J, Perregaux DG, Stam E, Petrushova N, Koller BH, Griffiths RJ, Gabel CA (2001) Altered cytokine production in mice lacking P2X₇ receptors. *J Biol Chem* 276:125–132
 65. Labasi JM, Petrushova N, Donovan C, McCurdy S, Lira P, Payette MM, Brissette W, Wicks JR, Audoly L, Gabel CA (2002) Absence of the P2X₇ receptor alters leukocyte function and attenuates an inflammatory response. *J Immunol* 168:6436–6445
 66. Rizzo R, Ferrari D, Melchiorri L, Stignani M, Gulinelli S, Baricordi OR, Di Virgilio F (2009) Extracellular ATP acting at the P2X₇ receptor inhibits secretion of soluble HLA-G from human monocytes. *J Immunol* 183:4302–4311
 67. Rizzo R, Bortolotti D, Bolzani S, Fainardi E (2014) HLA-G molecules in autoimmune diseases and infections. *Front Immunol* 5:592
 68. Adinolfi E, Raffaghello L, Giuliani AL, Cavazzini L, Capece M, Chiozzi P, Bianchi G, Kroemer G, Pistoia V, Di Virgilio F (2012) Expression of P2X₇ receptor increases in vivo tumor growth. *Cancer Res* 72:2957–2969
 69. Ferrari D, Chiozzi P, Falzoni S, dal Susino M, Melchiorri L, Baricordi OR, Di Virgilio F (1997) Extracellular ATP triggers IL-1 β release by activating the purinergic P2Z receptor of human macrophages. *J Immunol* 159:1451–1458
 70. Ferrari D, Stroth C, Schulze-Osthoff K (1999) P2X₇/P2Z purinoreceptor-mediated activation of transcription factor NFAT in microglial cells. *J Biol Chem* 274:13205–13210
 71. Morandini AC, Savio LE, Coutinho-Silva R (2014) The role of P2X₇ receptor in infectious inflammatory diseases and the influence of ectonucleotidases. *Biomed J* 37:169–177
 72. Janssen B, Vugts DJ, Funke U, Spaans A, Schuit RC, Kooijman E, Rongen M, Perk LR, Lammertsma AA, Windhorst AD (2014) Synthesis and initial preclinical evaluation of the P2X₇ receptor antagonist [¹¹C]A-740003 as a novel tracer of neuroinflammation. *J Labelled Comp Radiopharm* 57:509–516
 73. Lopez-Castejon G, Pelegrin P (2012) Current status of inflammasome blockers as anti-inflammatory drugs. *Expert Opin Investig Drugs* 21:995–1007
 74. North RA (2002) Molecular physiology of P2X receptors. *Physiol Rev* 82:1013–1067
 75. Caseley EA, Muench SP, Roger S, Mao HJ, Baldwin SA, Jiang LH (2014) Non-synonymous single nucleotide polymorphisms in the P2X receptor genes: association with diseases, impact on receptor functions and potential use as diagnosis biomarkers. *Int J Mol Sci* 15:13344–13371
 76. Haag F, Adriouch S, Brass A, Jung C, Möller S, Scheuplein F, Bannas P, Seman M, Koch-Nolte F (2007) Extracellular NAD and ATP: partners in immune cell modulation. *Purinergic Signal* 3:71–81
 77. Myrtek D, Idzko M (2007) Chemotactic activity of extracellular nucleotides on human immune cells. *Purinergic Signal* 3:5–11
 78. Martinon F (2008) Detection of immune danger signals by NALP3. *J Leukoc Biol* 83:507–511
 79. Wewers MD, Sarkar A (2009) P2X₇ receptor and macrophage function. *Purinergic Signal* 5:189–195
 80. Ferrero ME (2011) Purinoceptors in inflammation: potential as anti-inflammatory therapeutic targets. *Front Biosci (Landmark Ed)* 16:2172–2186
 81. Junger WG (2011) Immune cell regulation by autocrine purinergic signalling. *Nat Rev Immunol* 11:201–212
 82. Di Virgilio F, Chiozzi P, Ferrari D, Falzoni S, Sanz JM, Morelli A, Torboli M, Bolognesi G, Baricordi OR (2001) Nucleotide receptors: an emerging family of regulatory molecules in blood cells. *Blood* 97:587–600
 83. Koshi R, Coutinho-Silva R, Cascabulho CM, Henrique-Pons A, Knight GE, Loesch A, Burnstock G (2005) Presence of the P2X₇ purinergic receptor on immune cells that invade the rat endometrium during oestrus. *J Reprod Immunol* 66:127–140
 84. Ferrari D, Idzko M, Dichmann S, Purlis D, Virchow C, Norgauer J, Chiozzi P, Di Virgilio F, Luttmann W (2000) P2 purinergic receptors of human eosinophils: characterization and coupling to oxygen radical production. *FEBS Lett* 486:217–224
 85. Lucattelli M, Cicko S, Muller T, Lommatzsch M, De Cunto G, Cardini S, Sundas W, Grimm M, Zeiser R, Durk T, Zissel G, Sorichter S, Ferrari D, Di Virgilio F, Virchow JC, Lungarella G, Idzko M (2011) P2X₇ receptor signaling in the pathogenesis of smoke-induced lung inflammation and emphysema. *Am J Respir Cell Mol Biol* 44:423–429
 86. Vitiello L, Gorini S, Rosano G, La Sala A (2012) Immunoregulation through extracellular nucleotides. *Blood* 120:511–518
 87. Di Virgilio F, Ferrari D, Chiozzi P, Falzoni S, Sanz JM, dal Susino M, Mutini C, Hanau S, Baricordi OR (1996) Purinoceptor function in the immune system. *Drug Dev Res* 39:319–329
 88. Marchand F, Perretti M, McMahon SB (2005) Role of the immune system in chronic pain. *Nat Rev Neurosci* 6:521–532
 89. Moalem G, Tracey DJ (2006) Immune and inflammatory mechanisms in neuropathic pain. *Brain Res Rev* 51:240–264
 90. Wareham K, Vial C, Wykes RC, Bradding P, Seward EP (2009) Functional evidence for the expression of P2X₁, P2X₄ and P2X₇ receptors in human lung mast cells. *Br J Pharmacol* 157:1215–1224
 91. Bulanova E, Bulfone-Paus S (2010) P2 receptor-mediated signaling in mast cell biology. *Purinergic Signal* 6:3–17
 92. Cockcroft S, Gomperts BD (1980) The ATP₄-receptor of rat mast cells. *Biochem J* 188:789–798

93. Burnstock G (2008) Purinergic signalling and disorders of the central nervous system. *Nat Rev Drug Disc* 7:575–590
94. Inoue K (2008) Purinergic systems in microglia. *Cell Mol Life Sci* 65:3074–3080
95. Chessell IP, Hatcher JP, Bountra C, Michel AD, Hughes JP, Green P, Egerton J, Murfin M, Richardson J, Peck WL, Grahames CB, Casula MA, Yiangou Y, Birch R, Anand P, Buell GN (2005) Disruption of the P2X₇ purinoceptor gene abolishes chronic inflammatory and neuropathic pain. *Pain* 114:386–396
96. Hughes JP, Hatcher JP, Chessell IP (2007) The role of P2X₇ in pain and inflammation. *Purinergic Signal* 3:163–169
97. Chu YX, Zhang Y, Zhang YQ, Zhao ZQ (2010) Involvement of microglial P2X₇ receptors and downstream signaling pathways in long-term potentiation of spinal nociceptive responses. *Brain Behav Immun* 24:1176–1189
98. Alves LA, Bezerra RJ, Faria RX, Ferreira LG, da Silva FV (2013) Physiological roles and potential therapeutic applications of the P2X₇ receptor in inflammation and pain. *Molecules* 18:10953–10972
99. Burnstock G (2013) Purinergic mechanisms and pain—an update. *Eur J Pharmacol* 716:24–40
100. Dell’Antonio G, Quattrini A, Cin ED, Fulgenzi A, Ferrero ME (2002) Relief of inflammatory pain in rats by local use of the selective P2X₇ ATP receptor inhibitor, oxidized ATP. *Arthritis Rheum* 46:3378–3385
101. Clark AK, Staniland AA, Marchand F, Kaan TK, McMahon SB, Malcangio M (2010) P2X₇-dependent release of interleukin-1 β and nociception in the spinal cord following lipopolysaccharide. *J Neurosci* 30:573–582
102. Wirkner K, Sperlágh B, Illes P (2007) P2X₃ receptor involvement in pain states. *Mol Neurobiol* 36:165–183
103. Prado FC, Araldi D, Vieira AS, Oliveira-Fusaro MC, Tambeli CH, Parada CA (2013) Neuronal P2X₃ receptor activation is essential to the hyperalgesia induced by prostaglandins and sympathomimetic amines released during inflammation. *Neuropharmacology* 67:252–258
104. Xu J, Chu KL, Brederson JD, Jarvis MF, McGaraughty S (2012) Spontaneous firing and evoked responses of spinal nociceptive neurons are attenuated by blockade of P2X₃ and P2X_{2/3} receptors in inflamed rats. *J Neurosci Res* 90:1597–1606
105. McGaraughty S, Honore P, Wismer CT, Mikusa J, Zhu CZ, McDonald HA, Bianchi B, Faltynek CR, Jarvis MF (2005) Endogenous opioid mechanisms partially mediate P2X₃/P2X_{2/3}-related antinociception in rat models of inflammatory and chemogenic pain but not neuropathic pain. *Br J Pharmacol* 146:180–188
106. Tsuda M, Shigemoto-Mogami Y, Koizumi S, Mizokoshi A, Kohsaka S, Salter MW, Inoue K (2003) P2X₄ receptors induced in spinal microglia gate tactile allodynia after nerve injury. *Nature* 424:778–783
107. Beggs S, Trang T, Salter MW (2012) P2X₄R+ microglia drive neuropathic pain. *Nat Neurosci* 15:1068–1073
108. Honore P, Donnelly-Roberts D, Namovic MT, Hsieh G, Zhu CZ, Mikusa JP, Hernandez G, Zhong C, Gauvin DM, Chandran P, Harris R, Medrano AP, Carroll W, Marsh K, Sullivan JP, Faltynek CR, Jarvis MF (2006) A-740003 [N-(1-{{[cyanoimino}(5-quinolinylamino) methyl]amino}-2,2-dimethylpropyl)-2-(3,4-dimethoxyphenyl)acetamide], a novel and selective P2X₇ receptor antagonist, dose-dependently reduces neuropathic pain in the rat. *J Pharmacol Exp Ther* 319:1376–1385
109. King BF (2007) Novel P2X₇ receptor antagonists ease the pain. *Br J Pharmacol* 151:565–567
110. Teixeira JM, Oliveira MC, Parada CA, Tambeli CH (2010) Peripheral mechanisms underlying the essential role of P2X₇ receptors in the development of inflammatory hyperalgesia. *Eur J Pharmacol* 644:55–60
111. Itoh K, Chiang CY, Li Z, Lee JC, Dostrovsky JO, Sessle BJ (2011) Central sensitization of nociceptive neurons in rat medullary dorsal horn involves purinergic P2X₇ receptors. *Neuroscience* 192:721–731
112. McGaraughty S, Chu KL, Namovic MT, Donnelly-Roberts DL, Harris RR, Zhang XF, Shieh CC, Wismer CT, Zhu CZ, Gauvin DM, Fabiyi AC, Honore P, Gregg RJ, Kort ME, Nelson DW, Carroll WA, Marsh K, Faltynek CR, Jarvis MF (2007) P2X₇-related modulation of pathological nociception in rats. *Neuroscience* 146:1817–1828
113. Zhang J, Li X, Gao Y, Guo G, Xu C, Li G, Liu S, Huang A, Tu G, Peng H, Qiu S, Fan B, Zhu Q, Yu S, Zheng C, Liang S (2013) Effects of puerarin on the inflammatory role of burn-related procedural pain mediated by P2X₇ receptors. *Burns* 39:610–618
114. Yirmiya R, Goshen I (2011) Immune modulation of learning, memory, neural plasticity and neurogenesis. *Brain Behav Immun* 25:181–213
115. Frank-Cannon TC, Alto LT, McAlpine FE, Tansey MG (2009) Does neuroinflammation fan the flame in neurodegenerative diseases? *Mol Neurodegener* 4:47
116. Meyer U (2013) Developmental neuroinflammation and schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* 42:20–34
117. Najjar S, Pearlman DM, Alper K, Najjar A, Devinsky O (2013) Neuroinflammation and psychiatric illness. *J Neuroinflammation* 10:43
118. Iwata M, Ota KT, Duman RS (2013) The inflammasome: pathways linking psychological stress, depression, and systemic illnesses. *Brain Behav Immun* 31:105–114
119. Hansen T, Jakobsen KD, Fenger M, Nielsen J, Krane K, Fink-Jensen A, Lublin H, Ullum H, Timm S, Wang AG, Jorgensen NR, Werge T (2008) Variation in the purinergic P2RX₇ receptor gene and schizophrenia. *Schizophr Res* 104:146–152
120. Ursu D, Ebert P, Langron E, Ruble C, Munsie L, Zou W, Fijal B, Qian YW, McNamey TA, Mogg A, Grubisha O, Merchant K, Sher E (2014) Gain and loss of function of P2X₇ receptors: mechanisms, pharmacology and relevance to diabetic neuropathic pain. *Mol Pain* 10:37
121. Miller CM, Boulter NR, Fuller SJ, Zakrzewski AM, Lees MP, Saunders BM, Wiley JS, Smith NC (2011) The role of the P2X₇ receptor in infectious diseases. *PLoS Pathog* 7:e1002212
122. Xiang Y, Wang X, Yan C, Gao Q, Li SA, Liu J, Zhou K, Guo X, Lee W, Zhang Y (2013) Adenosine-5'-triphosphate (ATP) protects mice against bacterial infection by activation of the NLRP3 inflammasome. *PLoS One* 8:e63759
123. Fumagalli M, Lecca D, Abbracchio MP (2011) Role of purinergic signalling in neuro-immune cells and adult neural progenitors. *Front Biosci (Landmark Ed)* 16:2326–2341
124. Ochoa-Cortes F, Liñán-Rico A, Jacobson KA, Christofi FL (2014) Potential for developing purinergic drugs for gastrointestinal diseases. *Inflamm Bowel Dis* 20:1259–1287
125. Sorge RE, Trang T, Dorfman R, Smith SB, Beggs S, Ritchie J, Austin JS, Zaykin DV, Vander MH, Costigan M, Herbert TA, Yarkoni-Abitbul M, Tichauer D, Livneh J, Gershon E, Zheng M, Tan K, John SL, Slade GD, Jordan J, Woolf CJ, Peltz G, Maixner W, Diatchenko L, Seltzer Z, Salter MW, Mogil JS (2012) Genetically determined P2X₇ receptor pore formation regulates variability in chronic pain sensitivity. *Nat Med* 18:595–599
126. Mehta N, Kaur M, Singh M, Chand S, Vyas B, Silakari P, Bahia MS, Silakari O (2014) Purinergic receptor P2X₇: a novel target for anti-inflammatory therapy. *Bioorg Med Chem* 22:54–88
127. Friedle SA, Curet MA, Watters JJ (2010) Recent patents on novel P2X₇ receptor antagonists and their potential for reducing central nervous system inflammation. *Recent Pat CNS Drug Discov* 5:35–45
128. Takenouchi T, Sekiyama K, Sekigawa A, Fujita M, Waragai M, Sugama S, Iwamaru Y, Kitani H, Hashimoto M (2010) P2X₇

- receptor signaling pathway as a therapeutic target for neurodegenerative diseases. *Arch Immunol Ther Exp (Warsz)* 58:91–96
129. Donnelly-Roberts DL, Jarvis MF (2007) Discovery of P2X₇ receptor-selective antagonists offers new insights into P2X₇ receptor function and indicates a role in chronic pain states. *Br J Pharmacol* 151:571–579
130. Donnelly-Roberts D, McGaraughty S, Shieh CC, Honore P, Jarvis MF (2008) Painful purinergic receptors. *J Pharmacol Exp Ther* 324:409–415
131. Carroll WA, Donnelly-Roberts D, Jarvis MF (2009) Selective P2X₇ receptor antagonists for chronic inflammation and pain. *Purinergic Signal* 5:63–73
132. Brumfield S, Matasi JJ, Tulshian D, Czarniecki M, Greenlee W, Garlisi C, Qiu H, Devito K, Chen SC, Sun Y, Bertorelli R, Ansell J, Geiss W, Le VD, Martin GS, Vellekoop SA, Haber J, Allard ML (2011) Synthesis and SAR development of novel P2X₇ receptor antagonists for the treatment of pain: part 2. *Bioorg Med Chem Lett* 21:7287–7290
133. Honore P, Donnelly-Roberts D, Namovic M, Zhong C, Wade C, Chandran P, Zhu C, Carroll W, Perez-Medrano A, Iwakura Y, Jarvis MF (2009) The antihyperalgesic activity of a selective P2X₇ receptor antagonist, A-839977, is lost in IL-1 α knockout mice. *Behav Brain Res* 204:77–81
134. Wang C, Gu Y, Li GW, Huang LY (2007) A critical role of the cAMP sensor Epac in switching protein kinase signalling in prostaglandin E₂-induced potentiation of P2X₃ receptor currents in inflamed rats. *J Physiol* 584:191–203
135. Hyman MC, Petrovic-Djergovic D, Visovatti SH, Liao H, Yanamadala S, Bouis D, Su EJ, Lawrence DA, Broekman MJ, Marcus AJ, Pinsky DJ (2009) Self-regulation of inflammatory cell trafficking in mice by the leukocyte surface apyrase CD39. *J Clin Invest* 119:1136–1149
136. Rhett JM, Fann SA, Yost MJ (2014) Purinergic signaling in early inflammatory events of the foreign body response: modulating extracellular ATP as an enabling technology for engineered implants and tissues. *Tissue Eng Part B Rev* 20:392–402
137. Bhagwat SS (2007) MAP kinase inhibitors in inflammation and autoimmune disorders. In: Macor JE (ed) *Annual Reports in Medicinal Chemistry*, Volume 42. Academic Press, Amsterdam, pp 265–278
138. Bhagwat SS (2009) Kinase inhibitors for the treatment of inflammatory and autoimmune disorders. *Purinergic Signal* 5:107–115
139. Burnstock G, Verkhratsky A (2009) Evolutionary origins of the purinergic signalling system. *Acta Physiologica* 195:415–447
140. Burnstock G (2006) Pathophysiology and therapeutic potential of purinergic signaling. *Pharmacol Rev* 58:58–86
141. Burnstock G (2007) Physiology and pathophysiology of purinergic neurotransmission. *Physiol Rev* 87:659–797