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Decorin as a multivalent therapeutic agent against cancer

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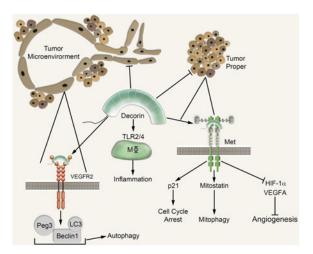
Abstract

Decorin is a prototypical small leucine-rich proteoglycan and epitomizes the multifunctional nature of this critical gene family. Soluble decorin engages multiple receptor tyrosine kinases within the target rich environment of the tumor stroma and tumor parenchyma. Upon receptor binding, decorin initiates signaling pathways within endothelial cells downstream of VEGFR2 that ultimately culminate in a Peg3/Beclin 1/LC3-dependent autophagic program. Concomitant with autophagic induction, decorin blunts capillary morphogenesis and endothelial cell migration, thereby significantly compromising tumor angiogenesis. In parallel within the tumor proper, decorin binds multiple RTKs with high affinity, including Met, for a multitude of oncosuppressive functions including growth inhibition, tumor cell mitophagy, and angiostasis. Decorin is also pro-inflammatory by modulating macrophage function and cytokine secretion. Decorin suppresses tumorigenic growth, angiogenesis, and prevents metastatic lesions in a variety of *in vitro* and *in vivo* tumor models. Therefore, decorin would be an ideal therapeutic candidate for combatting solid malignancies.

Graphical abstract

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Keywords

small leucine-rich proteoglycan; autophagy; mitophagy; angiogenesis; endothelial cells; receptor tyrosine kinases

1. Introduction

Fundamental for all facets of multicellular life and evolutionary conserved, the extracellular matrix (ECM) is a diverse network of instructional cues linking the local tissue microenvironment with the juxtaposed tumor cells [1-3]. Emerging as a critical entity in chemotherapeutics, tumorigenic progression, and predicting clinical outcome [4-6], the ECM is a nexus of signal integration for a plethora of cell-derived factors while synchronously regulating cellular behaviors [7]. This symbiotic relationship facilitates bidirectional parsing of intrinsic biological information into functionally relevant processes responsible for orchestrating tumorigenesis and angiogenesis [8-10].

The small leucine-rich proteoglycans (SLRPs) are an emerging subset of matrix-derived, soluble regulators that are inextricably woven into the fabric of the ECM. They reflect the multifactorial propensity of the matrix, and subsume crucial roles over a spectrum of homeostatic and pathological conditions [11]. This 18-member strong gene family is proving critical for restraining the development, progression, and dissemination of various solid tumors [12-14]. Decorin, the archetypical SLRP, harbors a single, covalently-attached N-terminal glycosaminoglycan (GAG) chain consisting of either dermatan or chondroitin sulfate, twelve leucine-rich tandem repeats (LRR), and a class-specific C-terminal Ear domain [15]. Although the crystal structure of decorin has been solved a head-to-tail dimer [16], it is likely that soluble decorin is active as a monomer in solution [17,18].

Decorin was originally discovered as an avid collagen-binding protein necessary for appropriate fibrillogenesis [19-22], thereby originating the eponym of decorin [15]. Akin with a role in orchestrating and ensuring proper collagen fibril network assembly, decorin regulates tissue integrity by modulating key biomechanical parameters of tendons and skin [23-26]. However, seminal work heralded a major paradigm shift in understanding the

function of SLRPs by demonstrating that soluble decorin is a high affinity, antagonistic ligand for several key receptor tyrosine kinases resulting in protracted oncostasis and angiostasis [27]. As a further mechanism for the oncosuppressive propensities of decorin, numerous growth factors-e.g. TGF- β [28,29] and CCN2/CTGF [30], to name a few-and matrix constituents are sequestered [31], and manifest as an indirect attenuation of downstream signaling apparati. More recently, decorin has emerged as a soluble pro-autophagic cue by initiating endothelial cell autophagy and evoking tumor cell mitophagy as the mechanistic basis for the documented oncostatic effects [32]. Cumulatively, decorin is a soluble tumor repressor and anti-angiogenic factor and has rightfully earned the designation of "*a guardian from the matrix*" [31].

Beyond the emerging literature regarding the role of decorin within the tumor stroma, decorin is genuinely a multifaceted signaling effector and exemplifies the growing role of SLRPs in organismal homeostasis and pathology. Germane examples include immunomodulation [33,34], cutaneous wound healing [35], proper keratinocyte function [36], diabetic nephropathies [37], fetal membrane homeostasis [38], obesity and type II diabetes [39], allergen-induced asthma [40], allergic inflammation [41], delayed hypersensitivity reactions [42], hepatic fibrosis [43], myogenesis and muscular dystrophy [44,45], post myocardial infarction remodeling [46], and mediating proper vertebrate convergent extension [47]. Moreover, decorin has been identified as a potential biomarker for ischemic stroke [48], renal and pulmonary diseases [49-51] and for maintaining hematopoietic stem cell niches [52].

In this review, we will critically evaluate decorin as a tumoricidal agent by examining the classical mechanisms of decorin-mediated oncogenic suppression and the newly discovered signaling pathways that are exploited for autophagic induction. The biofunctionality of decorin and associated mechanisms discussed herein represent novel targets for future therapeutic intervention, as derived from this versatile proteoglycan, that will satisfy a growing and unmet medical need.

1.1. General considerations: Decorin as an oncosuppressive entity

An important construct for understanding the anti-tumorigenic effects of decorin concerns the localization and corresponding expression patterns of this prototypical SLRP within the various tumorigenic compartments [53].

1.1.1. Localization and expression patterns of decorin within the tumor-

Despite a large literature describing decorin as an oncosuppressive proteoglycan [12,13,31,54], there are still several incongruencies that need to be addressed. In particular, the absence of decorin in the breast tumor stroma has been established as an important clinical prognosticator of invasive and metastatic breast cancer [10,55-57] as well as in soft tumors [58]. A similar reduction of decorin expression is seen in the microenvironments of low- and high-grade urothelial carcinoma [59] as well as in the plasma of multiple myeloma and MGUS patients [60], cases of esophageal squamous cell carcinoma [61] and instances of colon cancer [53]. An *in silico*-based query utilizing immunohistochemical arrays spanning a variety of tissues has detected a marked reduction of decorin expression in the

stroma of many solid malignancies, including breast [62]. Other studies seemingly report the opposite result inasmuch as certain tumor types, including colon and breast carcinomas [54], have elevated amounts of stromally-deposited decorin. Functionally, the increased caches of decorin within these tumors may still negatively regulate growth by physically constraining the tumor (e.g. desmoplastic-type reaction) as well as acting in a paracrine manner to downregulate the adjacent RTKs present on the tumor cell surface. As it pertains to the tumor proper, several studies have clearly demonstrated a complete loss of decorin expression in several tumor types, such as urothelial, prostate, myeloma, and hepatic carcinomas, several prominent matrix constituents were decreased, including decorin [69]. Moreover, poorly differentiated sarcomas completely lack decorin in contrast to hemangiomas which have considerable expression of decorin [66]. Therefore, the malignancy of a tumor may be linked to endogenous decorin expression.

1.1.2. Genetic and cell biological evidence for decorin as a soluble tumor

repressor—As mentioned in the preceding section (1.1.1.), decorin is found to be profusely expressed within the stroma of colon cancer. This was the very first indication of a possible connection between decorin and an oncogenic setting [70-72]. Like p53, decorin was initially perceived as an oncogene. Since this discovery, strong genetic evidence has emerged confirming the oncostatic role of decorin following the unconditional ablation of decorin from the *M. musculus* genome [73]. Mice lacking the *Dcn* gene and given a Western diet (e.g. high-fat) develop intestinal tumors [74]. Mechanistically, loss of decorin disrupts appropriate intestinal cell maturation, leading to aberrant turnover (decreased differentiation and increased proliferation consistent with suppressed p21 and p27 with elevated β -catenin) of the intestinal epithelium [74]. Moreover, the inhibition of colon carcinoma by decorin involves modulating E-cadherin levels *in vitro* and *in vivo* [75]. Moreover, when both p53 and Dcn genes are concurrently ablated, there is a genetic cooperation demonstrated by the rapid onset of aggressive T-cell lymphomas and premature death of the double mutant mice [76]. These studies indicate that genetic loss of decorin is permissive for tumorigenic initiation.

Several studies have been completed wherein decorin is potently anti-metastatic for breast carcinomas [56,57,77] while compromising otherwise rampant tumor angiogenesis [78,79]. In a murine model of osteosarcoma, decorin prevents lung metastases [80] and inhibits B16V melanoma cell migration [81]. Of clinical and therapeutic importance, re-introduction of decorin via adenoviral delivery, *de novo* ectopic expression, or systemic administration counteracts the tumorigenicity in several animal models of cancer that recapitulate solid neoplastic growth [82-88]. Notably, pre-clinical studies using infrared-labeled decorin have shown that it preferentially targets the tumor xenografts with prolonged retention of the active agent [89]. Recently, adenoviral mediated decorin expression has been shown to decrease the growth of bone metastases caused by intracardiac injections of prostate [90] and breast [91] carcinomas. Taken together, the aforementioned genetic and pre-clinical studies establish and authenticate decorin as a viable tumor repressor for combating several types of cancer.

2. Decorin structure: High-affinity interactions with several receptors

Harboring the largest known gene family of proteoglycans, decorin and related classes of SLRPs share a common core architecture [92]. They are ubiquitously expressed in all major organs during development [93], and are present within all matrix assemblies. The various members have been organized into five distinct classes based on the criteria of evolutionary conserved structural homology (including organization at the genomic and protein levels) as well as by shared functional properties [15]. The closest SLRP to decorin is biglycan, which shares more than 65% homology. These properties include the innate ability of collagen binding [20,94], growth factor binding and sequestration (predominantly those from the TGF-β superfamily) [12,31], and cell surface receptor modulation as a soluble mediator [54,95]. Moreover, a specific subclass of solubilized SLRP and matrix components can regulate autophagy [32]. Finally, these classes can be subdivided further into canonical SLRPs (classes I-III) and non-canonical SLRPs (classes IV, V) based on various structural considerations (see below). In this fashion, decorin embodies all of these principles while pioneering new functions and paradigms.

2.1. The LRR constitutes the basic unit of decorin structure and function

Leucine-rich repeats are about 24 amino acids in length and contain a conserved stretch of hydrophobic residues that form short β -sheets on the interior or internal (concave) surface of the solenoid. These short β -sheets are further arranged in a parallel conformation with the adjacent LRRs in the core (Fig. 1A). In total, there are 12 LRRs (designated with roman numerals I-XII) that constitute the protein core of decorin (Fig. 1A). Conversely, on the exterior or external (convex) surface of the solenoid, these β -sheets are flanked by and intertwined with equally short β -strands connected by several types of α -helices (Fig. 1A). Terminating each LRR at the N- and C-termini are disulfide bonds that function as a cap. The inherent structural determinants of these caps aid in further distinction among the various classes of SLRPs (e.g. classes I-III vs. classes IV, V), as discussed above [15].

This fundamental LRR architecture permits a plastic interface capable of coordinating a myriad of protein-protein interactions. Indeed, this hallmark is crucial for the widespread functionality of decorin [95], and related SLRPs, and is mediated by residues located on the internal surface of the protein [15]. Moreover, each LRR confers various functional properties for the well-established bioactivities of decorin. For example, LRR XII binds CCN2/CTGF [30], LRR V/VI aid in the binding of decorin to VEGFR2 [96], and the collagen binding sequence (SYIRIADTNIT) of LRR VII, located on the interior surface of the solenoid [97], mediates direct binding of decorin to type I collagen (Fig. 1A). A feature of decorin, also shared by Class I-III SLRPs, is the presence of an elongated (~30 amino acids) LRR known as the "ear" repeat (Fig. 1A). In decorin, this is found in the penultimate LRR, LRR XI. Interestingly, truncation or mutations arising in the ear repeat of decorin cause congenital stromal corneal dystrophy [20,98]. Mechanistically, mouse models of decorin lacking this ear repeat trigger intracellular accumulation of decorin within the endoplasmic reticulum, thereby causing ER stress, and compromising proper corneal collagen deposition and assembly [99].

Importantly, the covalently attached glycosaminoglycan chain plays a pivotal role in the regulation of collagen fibrillogenesis [15]. However, in the context of controlling intracellular signaling cascades via cell surface receptors, the glycosaminoglycan chain is dispensable.

The glycosaminoglycan chain has a pivotal role in various connective tissue disorders insofar as alterations in the chain are found in congenital stromal corneal dystrophy and Ehlers-Danlos syndrome [100] as well as in cancer [12]. Improperly modified or missing chains can disrupt structural functions as mediated by decorin by compromising the architecture of the surrounding matrix. This is exemplified in the skin fragility phenotype of patients with Ehlers-Danlos syndrome, where roughly half of the secreted decorin lacks the chain [101]. Mechanistically, early stages of collagen fibril formation are impaired following the loss of the glycosaminoglycan chain. Moreover, the type and composition of the attached glycosaminoglycan can also vary, particularly in cancer (colon, ovarian, pancreatic, gastric), where it is predominantly chondroitin sulfate [10,12,72,102]. In contrast, the chemically more complex dermatan sulfate is less abundant in these types of tumors [102]. The presence of CS is postulated to facilitate cell migration, thereby increasing the malignancy of the tumor [102].

2.2. Decorin is a soluble pan-RTK inhibitor and binds multiple cell surface receptors

As discussed above (section 2.1), the overall arrangement of decorin, in conjunction with the individual composition of the LRRs, endows a rather promiscuous nature of binding multiple targets expressed within the tumor microenvironment and by the tumor proper. Of critical importance for attenuating tumorigenic progression and preventing metastases, decorin avidly binds numerous cell surface receptors [95] (Fig. 1B). Decorin can be considered an endogenous, soluble pan-RTK inhibitor, especially targeting cells enriched in EGFR, Met, and VEGFR2. These three RTKs are the most established and instrumental for transducing signals necessary for oncogenic and angiogenic suppression [31,54] (Fig. 1B). As such, this trio of receptors will be discussed in more depth in the forthcoming sections (see below, sections 3 and 4).

Decorin, non-canonically, engages IGF-IR (Fig. 1B), but does not trigger internalization nor compromise the stability of the receptor complex at the cell surface [59,103], unlike EGFR and Met (see below) [54]. Instead, decorin decreases the stability of critical downstream signaling effectors such as IRS-1 [59], thereby attenuating sufficient activation of the Akt/ MAPK/Paxillin pathway for IGF-I induced mobility [104]. Moreover, the role of decorin as an IGF-IR ligand is strictly context-dependent as decorin is an IGF-IR agonist in normal tissues, but functions as an obligate IGF-IR antagonist in cancer [103]. Adding an additional layer of complexity in modulating the IGF-IR signaling axis, decorin exerts control over discrete IR-A ligands by differentially binding and sequestering (analogous with requisitioning TGF- β members) the various IR-A ligand isoforms [105]. The role of decorin and related proteoglycans, particularly SLRP members, in mediating receptor cross-talk between EGFR and IGF-IR is emerging as a central mechanism in estrogen-responsive breast carcinomas [106].

A prime example can be made from PDGFR- α/β that will reinforce the central dogma of decorin. Screening the RTKome of two different chemically induced models of hepatocellular carcinoma (HCC), it was found that, in a Dcn null background, many RTKs are constitutively activated [68]. Indeed, the global loss of decorin permits inappropriate, basal activation of several RTKs as measured by an increase in the phospho-Tyr signal. From this screen, PDGFR- α/β emerged (Fig. 1B) as a viable candidate to which decorin engages with high affinity and suppresses the formation of HCC [68]. Importantly, these results are congruent with the finding that decorin is suppressed, at the transcriptomic level, in HCC [69]. These strong genetic data clearly demonstrate the importance of decorin in preventing aberrant and constitutive RTK activation while maintaining proper tissue homeostasis.

2.3. Decorin is pro-inflammatory by engaging TLR2/4 on the surface of macrophages

It is becoming evident that soluble decorin can regulate the innate immune response [33] via tolllike receptors 2 and 4 (Fig. 1B) and is considered a damage-associated molecular pattern member [107]. This pro-inflammatory property is analogous to that of circulating biglycan [108,109]. Via high-affinity interactions, decorin engages TLR2/4 and promotes a proinflammatory state by triggering the synthesis and secretion of TNF- α and IL-12p70 [33]. Indirectly, via the formation of decorin/TGF- β complexes, anti-inflammatory mediators (such as IL-10) are translationally suppressed by PDCD4 [33]. Thus, circulating decorin is a pro-inflammatory proteoglycan for innate immune modulation [33]. It has emerged that biglycan is a viable biomarker of inflammatory renal diseases [110]. Likewise, cancer patients have significantly increased levels of circulating decorin [33], positing decorin as a desirable therapeutic target.

3. Suppression of growth and tumor angiogenesis via EGFR and Met

Innate and distinct biological information pertinent for abrogating tumorigenic growth and suppressing tumor angiogenesis is stored within the solenoid structure of decorin [31]. This information is interpreted and transduced via engagements to a specific subset of RTKs (Fig. 2) that are amplified and enriched within the tumor parenchyma [10,12]. In the context of Met and EGFR, monomeric decorin [17] binds a narrow region that partially overlaps with that of the agonist binding cleft [111]. This binding subsequently promotes receptor dimerization, analogous to the natural ligand EGF [112], followed by a rapid and transient phosphorylation of the unstructured intracellular tails. This is followed by recruitment and activation of downstream effectors, caveosome-mediated internalization of the decorin/ receptor complex, and eventual lysosomal degradation [9,14,113,114]. The latter causes a protracted cessation of intracellular receptor signaling. Overall, this mechanism of action is a hallmark of decorin activity in the contextual framework of tumorigenic RTK signaling.

Seemingly, receptors harboring specific structural motifs, specifically members of the IgG superfamily, may provide essential docking platforms for decorin engagement [111,115]. Indeed, the ectodomains of EGFR, Met and VEGFR2 all contain multiple IgG folds [116,117]. Mechanistically, decorin binding may promote a combinatorially different phosphorylation signature than the pattern obtained with natural agonist (e.g. TGFa, EGF,

3.1. Decorin binds EGFR for tumor cell cycle arrest and apoptosis

The concept of decorin-mediated RTK-antagonism was pioneered following the discovery that EGFR is a main target [118] and that decorin represents an endogenous ligand for receptor occupancy and modulation [119]. In mouse models carrying A431 orthotopic tumor xenografts, it was established that decorin, by targeting EGFR, significantly subverts tumorigenic growth in vivo [120]. Decorin indirectly inhibits Her/ErbB2 activity [121], potentially via the titration of active ErbB1/ErbB2 dimers [54]. Decorin also directly binds and represses ErbB4/STAT3 signaling [122] in the central nervous system. Mechanistically, decorin triggers transient activation of downstream ERK1/2 signaling (following stimulation of the innate EGFR kinase) [87] concurrent with a regulated burst of cytosolic Ca^{2+} [123]. Paradoxically, positive EGFR/MAPK signaling (despite total EGFR being reduced by >50%) evokes induction of the cyclin-dependent kinase inhibitor, p21^{WAF1} with concomitant conversion of pro-caspase-3 into active caspase 3 [87]. Collectively, this promotes cell cycle arrest and induces the intrinsic apoptotic pathway, respectively (Fig. 2). Imperative for the protracted function of decorin, decorin/EGFR complexes are shuttled into caveolin-1 coated pits [31]. Specific phopho-residues are required for the association of caveolin-1 with EGFR [124] and internalized via endocytosis for degradation. This system prevents recycling of EGFR for additional rounds of signaling, in contrast to active ligands which sort EGFR into clathrin-coated pits. This leads to endosomal recycling and, ultimately, to repopulation of the cell surface with activated EGFR for additional signal transduction (Fig. 2).

3.2. Decorin evokes oncoprotein degradation and suppresses angiogenesis via Met

A major tenet of decorin-mediated suppression of oncogenesis involves transient activation of the receptor complex [31]. Using a discovery tool, such as a phosphotyrosine RTK array, it was found that a second RTK, Met or HGF receptor, is specifically activated by soluble decorin proteoglycan or decorin protein core [115] (Fig. 2). Met is the key receptor for decorin and is responsible for relaying signals applicable for anti-tumorigenesis, angiostasis and pro-mitophagic functionalities (see below, section 4.2) [54,115]. Moreover, decorin exhibits a tighter binding affinity for Met when compared with EGFR, (Kd~2 vs 87 nM, respectively). [115]. Heterodimeric decorin/Met complexes are shuttled from the cell surface into caveolin-1 positive endosomes following recruitment of the c-Cbl E3-ubiqtuin ligase to Met via Tyr1003 (Fig. 2), a residue phosphorylated and favored by decorin treatment [115]. Association of decorin/Met with caveolin-1 ensures termination of oncogenic signaling, which in parallel with decorin/EGFR is in stark contrast with HGF/Met (and EGF/EGFR) complexes localizing within clathrin-coated endocytic vescicles for proficient receptor recycling [89].

As a major consequence of inhibiting Met, two potent oncogenes, β -catenin and Myc, are targeted for unremitting degradation via the 26S proteasome [89] (Fig. 2). Decorin-evoked transcriptional suppression coupled with phosphorylation-dependent protein degradation of Myc (at Thr58, the effector kinase(s) remains unknown) permits de-repression of the

CDKN1A locus via loss of the AP4 repressor [89]. Moreover, decorin suppresses β -catenin signaling in a non-canonical fashion insofar as being independent from Axin/DSH/GSK-3 β activity [89]. In this scenario, β -catenin is phosphorylated, not for increased protein stability, and is instead targeted for degradation [125] in a manner consistent with direct phosphorylation of β -catenin by an RTK, such as Met [126-129] (Fig. 2). The observation that Myc and β -catenin signaling is governed by decorin may account for the intestinal tumor formation seen upon decorin ablation, as β -catenin is a major oncogenic driver for intestinal epithelium turnover and maturation [130]. Constitutive activation of Met is found in many cases of colon carcinoma and directly influences β -catenin signaling [131]. Therefore, as global loss of decorin relieves the basal inhibition of several RTKs [68], this could certainly contribute to Met/ β -catenin driven transformation of the intestinal epithelium and/or other solid malignancies directed by this axis.

Concomitant with the concerted suppression of two potent oncogenes, Met also serves as the primary node for angiogenic suppression in cervical and breast carcinomas [79] (Fig. 2). Positive signaling via Met non-canonically suppresses the transcription of HIF1A regardless of oxygen concentration [79]. Correspondingly, VEGFA mRNA and proteins are compromised in several in vitro studies utilizing primary endothelial cells, MDA-MB-231 triple-negative breast carcinoma cells, and *in vivo* as demonstrated with HeLa tumor xenografts [79]. Moreover, MMP2/9 (Gelatinase A and B, respectively) which liberate matrix bound VEGFA, are also significantly suppressed [79]. In parallel with a protracted suppression of pro-angiogenic effectors, decorin also evokes the expression and secretion of anti-angiogenic factors such as TIMP3 and TSP-1 [79] (Fig. 2). Further studies have indicated that decorin triggers the rapid secretion of TSP-1 from MDA-MB-231 cells in an EGFR-dependent manner by attenuating the RhoA/ROCK1 pathway [132]. Given the powerful anti-angiogenic activity of TSP-1 and the involvement in several pathophysiological processes [133-138], it is likely that this indirect activity of decorin in malignant cells could have a protective role against cancer growth and metabolism. Taken together, decorin differentially regulates potent angiokines [139] that favor silencing rampant tumor neovascularization, thereby contributing further to the ascribed antitumorigenic and anti-metastatic properties.

4. Decorin ameliorates tumorigenesis by evoking stromal autophagy and tumor mitophagy

A major breakthrough in deciphering the *in vivo* bioactivity of decorin came from a preclinical screen that sought novel decorin-regulated genes [88]. With this goal, triple-negative breast carcinoma orthotopic xenografts were established and treated systemically with decorin, for downstream utilization on a high-resolution transcriptomic platform [88]. Unlike traditional microarrays, this chip was designed for the simultaneous analysis and detection of species-specific genes modulated within the host stroma (*Mus musculus*) and those originating from the tumor xenograft (*Homo sapiens*) [88]. Following validated bioinformatics approaches, it was found that decorin regulates a small subset of genes; however, this signature showed differential regulation exclusively within the tumor microenvironment derived from the murine host [88], with minimal transcriptomic changes

in the tumor cells of human origin [88]. The transcriptomic profile obtained implies that exogenous decorin treatment reprograms the tumor stroma in a fashion that disfavors tumorigenic growth, consistent with the function of decorin acting as a soluble tumor repressor from the outside.

4.1. Decorin evokes endothelial cell autophagy in a Peg3-dependent manner

Using the decorin-treated breast carcinoma xenografts described above, several novel tumorderived genes were discovered [88]. Among these genes, the genomically-imprinted zincfinger transcription factor, *PEG3* [140-143] was selected [88]. Previously, Peg3 has been implicated in regulating stem cell progenitors [144,145], mediating p53-dependent apoptosis of myogenic and neural lineages [146-150], and maternal/paternal behavioral patterns [151,152]. Peg3 has been implicated in the pathogenesis of cervical and ovarian carcinoma as its expression is frequently lost via promoter hypermethylation and/or loss of heterozygosity [153-156]. Thus, Peg3 is considered a *bona fide* tumor suppressor [157]. Importantly, Peg3 represents another tumor suppressor induced by decorin in addition to mitostatin and Beclin 1 (see below). Moreover, in analogy to decorin bioactivity in cancer cells, Peg3 non-canonically suppresses the Wnt/β-catenin pathway [158].

As a proxy for the tumor stroma, we investigated the function of Peg3 within endothelial cells, as this particular cell type conveys major angiogenic advantages for a growing tumor and constitutes the primary cell type involved in capillary morphogenesis and patent vessel formation. Moreover, these cells are significantly responsive to soluble decorin, which suppresses the expression of VEGFA, a major survival factor [79]. Serendipitously, we found that Peg3 mobilizes into large intracellular structures reminiscent of autophagosomes [159] in primary endothelial cells (HUVEC). Co-immunocolocalization and coimmunoprecipitation studies of canonical autophagic markers, e.g., Beclin 1 and LC3 [160,161], and Peg3 have clearly demonstrated that decorin evokes a novel gene involved in autophagy initiation [159] (Fig. 3, *left*). Intriguingly, Peg3 is required for decorin-mediated BECN1 and MAP1LC3A expression and is responsible for maintaining basal levels of Beclin 1 [159,162]. Mechanistically, decorin promotes a competent pro-autophagic signaling composed of Peg3, Beclin 1 and LC3 while combinatorially precluding Bcl-2, a known autophagic inhibitor [163]. At the endothelial cell surface, decorin engages VEGFR2, the central receptor for endothelial cells, for autophagic induction [159] (Fig. 3, left). Pharmacological inhibition with the small molecule inhibitor (SU5416) abrogates the autophagic response, suggesting that decorin requires the VEGFR2 kinase for successful autophagy [159,162]. Downstream of stimulated VEGFR2, decorin differentially regulates decisive signaling complexes by activating pro-autophagic modules (e.g. AMPKa and Vps34) while concurrently attenuating, in a protracted fashion, anti-autophagic nodes (e.g. PI3K/Akt/mTOR) [164] (Fig. 3, left). Concomitant with autophagic initiation, decorin also impairs capillary morphogenesis [78,79,159]. Therefore, it is plausible that decorin evokes autophagy as the molecular underpinning for suppressing tumor angiogenesis from the perspective of endothelial cell-driven angiogenesis (Fig. 3, left).

4.2. Decorin induces tumor cell mitophagy in a mitostatin-dependent manner

As a novel constituent of the multi-pronged approach for curtailing tumorigenesis and halting angiogenesis (differential modulation of pro- and anti-angiogenic factors and induction of endothelial cell autophagy) decorin directly influences catabolic programs and organelle turnover within the tumor proper (Fig.3, right). Induction of tumor cell mitochondrial autophagy (mitophagy) [165] may functionally reconcile the canonical tumoricidal effects of decorin with the emerging biology of matrix-mediated autophagic induction for retarding tumorigenic and angiogenic progression. In a mechanism analogous to that of VEGFR2, decorin requires the kinase activity of Met for proper mitophagic induction in breast carcinoma cells [165] (Fig. 3, right). Both forms of autophagic induction require the presence of a cell surface receptor (VEGFR2 or Met) and the intrinsic kinase activity of referenced receptor. At the nexus of decorin-evoked mitophagy is a poorly characterized tumor suppressor gene known as mitostatin or trichoplein (mitostatin has the HuGO gene symbol, TCHP, and is located on chromosome 12q24.1). Mitostatin was originally identified as a decorin-inducible gene using subtractive hybridization and probes from decorin-transfected (and thereby, growth suppressed) cells [166]. Notably, mitostatin is downregulated in bladder and breast carcinomas [166,167], suggesting that it might represent a potential tumor suppressor gene. Mitostatin primarily resides at the outer mitochondrial membrane [167] and at specialized membrane:membrane contact sites at the juxtaposition of the endoplasmic reticulum and mitochondria where it interacts with mitofusion-2 [168]. Hence the given eponym for mitostatin, mitochondrial protein with oncostatic activity.

During the early stages of mitophagy, downstream of Met, a master regulator of mitochondrial homeostasis and biogenesis, PGC-1 α [169] is mobilized into the nucleus and binds *TCHP* mRNA directly for rapid stabilization coincident with mitostatin protein accumulation [165] (Fig. 3, *right*). Mediating the interaction of PGC-1 α with *TCHP* mRNA via the C-terminal RNA recognition motif [165] is critical for stabilization. Truncating this domain or silencing PRMT1, for appropriate arginine methylation, compromises mitostatin mRNA stabilization [165]. The delineation of this pathway has revealed a unique cooperation between a novel mitophagic effector and a known oncogenic driver. PGC-1 α mediates B-Raf mediated oxidative metabolism [170] while defining a subset of aggressive melanoma characterized by an augmented mitochondrial capacity for increased resistance to oxidative stress [171].

The process of decorin-evoked mitophagy depends on the presence and yet-to-beelucidated-function of mitostatin [165] (Fig. 3, *right*). RNAi-mediated silencing of mitostatin prevents turnover of respiratory chain components (OXPHOS), decreased mtDNA content, VDAC clearance, and collapse of the mitochondrial network [165], all established markers of mitophagy [172]. Moreover, failure of mitophagic induction precludes the ability of decorin in suppressing VEGFA expression and protein [165] (Fig. 3, *right*), suggesting that mitophagy is key for understanding a fundamental hallmark of decorin biology. Subsequent to the collapse and aggregation of the tubular mitochondrial network, decorin triggers mitochondrial depolarization [165], with an activity comparable to that of an established depolarization agent (FCCP). This loss of membrane potential across

the outer and inner mitochondrial membrane is a harbinger for mitochondrial dysfunction and eventual turnover [173,174]. Depolarized mitochondria may be the end product of increased Ca^{+2} levels as occur downstream from decorin/EGFR interactions [123]. As mitostatin is positioned at mitochondrial-associated membrane and interacts with Mfn-2, it may permit an efflux of Ca^{+2} from the ER directly into the mitochondria as the initial event for decorin-evoked mitophagy.

In either scenario, depolarization of the mitochondria triggers recruitment of the PINK1/ Parkin complex for eventual clearance of the damaged organelle. The E3-ubiquitin ligase, Parkin is strictly required for proper mitochondrial homeostasis, as recessive mutations in Parkin are found in the neurodegenerative disease, Parkinson's [173,175,176]. It remains plausible that mitostatin may interact with or facilitate the conscription of PINK1/Parkin for mitochondrial turnover (Fig. 3, *right*). Alternatively, mitostatin may directly stimulate the inherent PINK1 kinase activity for proper recruitment, ubiquitin activation [177,178], and/or Parkin-mediated ubiquitination of target mitochondrial proteins [179-181]. Indeed, this axis is key for recycling respiratory chain complexes [182,183].

Collectively, the above findings imply that decorin transduces biological information via the Met kinase for mitophagic stimulation, in a mitostatin-dependent manner, within the tumor parenchyma of breast and prostate carcinomas [90]. This conserved catabolic process, coupled with the induction of endothelial cell autophagy, may form the molecular basis for the various outputs of decorin-mediated RTK regulation. Indeed, this newly-found activity may lie at the crossroads of controlling tumorigenic growth and unchecked tumor vascularization.

5. Gene and protein therapy in various preclinical tumor studies

Delivery of decorin via adenovirus (Ad) vectors together with the systemic administration of decorin proteoglycan or protein core, has been tested in a variety of preclinical studies. In Table 1 we summarize past and current studies utilizing these two approaches focused exclusively on cancer treatment and delivery. Although the therapeutic efficacy varies among these studies, it is clear that decorin has a deleterious effect on growth, apoptosis, metabolism and angiogenesis.

This concept was established by initial studies demonstrating that ectopically expressing decorin for the rapid neutralization and inhibition of tumorigenic growth from various histogenetically distinct origins held potential clinical relevance [84]. These studies provided further evidence that administering decorin, either decorin proteoglycan or protein core, in a systemic fashion prevented growth and metastases of orthotopic tumor xenografts [87]. Several studies (Table 1), have subsequently evaluated the feasibility of delivering decorin via adenovirus in several tumor types including breast and prostate carcinoma. Collectively, these studies have reaffirmed the *in vivo* applicability of utilizing decorin as a therapeutic modality for the prevention of metastatic lesions as well as suppressing the oncogenic and angiogenic properties of tumors.

6. Conclusions

The extracellular matrix is rapidly emerging as a crucial component for better understanding fundamental cellular processes and behaviors as well as providing novel therapeutic targets for combating complex pathological conditions [6] after these pathways have gone awry. Our pursuit of comprehending the varied intricacies and subtleties of reciprocal cell:matrix signaling for homeostatic and tumorigenic processes has been facilitated by an exhaustive proteomics approach, organized into an invaluable resource accessible for query [184]. As this database will undoubtably aid research concerning the contributions of matrix in various pathologies, the plenary discoveries of decorin mediated RTK-antagonism have revealed heretofore unknown signaling roles encoded within members of the soluble matrix. Since this pioneering breakthrough, similar mechanisms have been proposed as the underlying molecular explanation for a variety of biological phenomena [15] across diverse tissues and microenvironments. Indeed, the ever-expanding decorin interactome [31] encompasses a plethora of critical matrix-bound and cell-localized binding partners that substantially attenuate pro-tumorigenic and pro-angiogenic signaling cues [54] while simultaneously inducing conserved, intracellular catabolic processes [32,95]. In summation, this manifests as patent and long-lasting oncosuppression [88,89] that is efficacious and clinically-relevant in a variety of solid tumors.

Structure always determines function; this axiom is epitomized within the leucine rich repeats composing the protein core of decorin. This regularly patterned structure inherently provides for a high affinity and multivalent interface capable of binding and interacting with a large number of effector proteins to potentiate probable cellular outcomes. As such, decorin requires and depends on this proclivity for binding multiple partners for competently executing downstream events under a variety of conditions. This concept is exemplified in the context of RTK binding. Canonically, decorin is characterized as an unwavering and unbridled antagonistic ligand for the EGFR and Met receptor, resulting in the inhibition of potent oncoproteins and pro-angiogenic factors. The mechanistic perspective for decorin (at the receptor level) has shifted after identifying a decorin-specific transcriptomic signature exclusively within the tumor stroma, and the subsequent discovery of endothelial cell autophagy in which VEGFR2 kinase activity is required. Therefore, decorin acts as a partial receptor agonist. A similar requirement is operational in Met kinase activity during the process of mitophagic initiation in breast carcinoma cells [165]. These findings support the hypothesis that decorin could engage a receptor for autophagic induction as a basis for oncostasis. Indeed, the oncogenic EGFR/Akt signaling suppresses Beclin 1 for increased chemo-resistance and tumorigenicity [185,186]. Moreover, a novel mechanism detailing the transcriptional induction and enhanced secretion of decorin from cardiac tissue and isolated mouse embryonic fibroblasts following a 25-hour fast has been recently identified [187]. Notably, the global ablation of decorin attenuates autophagic responses and blunts autophagic flux, further underscoring the critical importance of decorin as a soluble, in vivo pro-autophagic regulator [187]. This study may wield clinical relevance as a starting point for drug development towards molecules targeting *Dcn* induction and secretion for organismal-wide autophagic regulation and tumor suppression [188].

Furthermore, the clinical efficacy of decorin as a novel therapeutic is exemplified by the diverse array of studies employing decorin as a potent soluble tumor repressor.

In conclusion, the work on decorin provides a new paradigm in the more general scheme of matrix-dependent regulation of cancer growth: soluble ECM constituents can act as proautophagic factors by interacting with various cell surface receptors for the proficient modulation of the intracellular catabolic network. This new function integrates well with the traditional oncosuppressive properties of decorin exerted on RTKs. Thus, decorin and related SLRPs, including soluble ECM fragments derived from larger parental molecules [95,189], hold great therapeutic potential and clinical benefit for combating cancer.

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References

- DeBerardinis RJ, Lum JJ, Hatzivassiliou G, Thompson CB. The biology of cancer: Metabolic reprogramming fuels cell growth and proliferation. Cell Metab. 2008; 7:11–20. [PubMed: 18177721]
- Hahn WC, Weinberg RA. Rules for making human tumor cells. N Engl J Med. 2002; 347:1593– 1603. [PubMed: 12432047]
- 3. Hanahan D, Winberg RA. Hallmarks of cancer: The next generation. Cell. 2011; 144:646–674. [PubMed: 21376230]
- 4. Weigelt B, Bissell MJ. Unraveling the micronevironment influences on the normal mammary gland and breast cancer. Semin Cancer Biol. 2008; 18:311–321. [PubMed: 18455428]
- 5. Bissell MJ, Hines WC. Why don't we get more cancer? A proposed role of the microenvironment in restraining cancer progression. Nat Med. 2011; 17:320–329. [PubMed: 21383745]
- Rønnov-Jessen L, Bissell MJ. Breast cancer by proxy: can the microenvironment be both the cause and consequence? Trends Mol Med. 2009; 15:5–13. [PubMed: 19091631]
- Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. Cancer Cell. 2012; 21:309–322. [PubMed: 22439926]
- 8. Grahovac J, Wells A. Matrikine and matricellular regulators of EGF receptor signaling on cancer cell migration. Lab Invest. 2014; 94:31–40. [PubMed: 24247562]
- Merline, R.; Nastase, MV.; Iozzo, RV.; Schaefer, L. Small Leucine-rich proteoglycans: multifunctional signaling effectors. In: Karamanos, N., editor. Extracellular Matrix: Pathobiology and signaling. Walter de Gruytier Gmbh and Co.; Berlin: 2012. p. 185-196.
- Goldoni S, Iozzo RV. Tumor microenvironment: Modulation by decorin and related molecules harboring leucine-rich tandem motifs. Int J Cancer. 2008; 123:2473–2479. [PubMed: 18798267]
- Iozzo RV, Sanderson RD. Proteoglycans in cancer biology, tumour microenvironment and angiogenesis. J Cell Mol Med. 2011; 15:1013–1031. [PubMed: 21155971]
- Theocharis AD, Skandalis SS, Neill T, Multhaupt HA, Hubo M, Frey H, Gopal S, Gomes A, Afratis N, Lim HC, Couchman JR, Filmus J, Ralph DS, Schaefer L, Iozzo RV, Karamanos NK. Insights into the key roles of proteoglycans in breast cancer biology and translational medicine. Biochim Biophys Acta. 2015; 1855:276–300. [PubMed: 25829250]
- Schaefer L. Proteoglycans, key regulators of cell-matrix dynamics. Matrix Biol. 2014; 35:1–2. [PubMed: 24871042]
- 14. Schaefer L, Iozzo RV. Small leucine-rich proteoglycans, at the crossroad of cancer growth and inflammation. Curr Opin Genet Dev. 2012; 22:56–57. [PubMed: 22326829]

- Iozzo RV, Schaefer L. Proteoglycan form and function: A comprehensive nomenclature of proteoglycans. Matrix Biol. 2015; 42:11–55. [PubMed: 25701227]
- Scott PG, McEwan PA, Dodd CM, Bergmann EM, Bishop PN, Bella J. Crystal structure of the dimeric protein core of decorin, the archetypal small leucine-rich repeat proteoglycan. Proc Natl Acad Sci USA. 2004; 101:15633–15638. [PubMed: 15501918]
- Goldoni S, Owens RT, McQuillan DJ, Shriver Z, Sasisekharan R, Birk DE, Campbell S, Iozzo RV. Biologically active decorin is a monomer in solution. J Biol Chem. 2004; 279:6606–6612. [PubMed: 14660661]
- Islam M, Gor J, Perkins SJ, Ishikawa Y, Bãchinger HS, Hohenester E. The concave face of decorin mediates reversible dimerization and collagen binding. J Biol Chem. 2013; 288:35526–35533. [PubMed: 24169694]
- Chen S, Young MF, Chakravarti S, Birk DE. Interclass small leucine-rich repeat proteoglycan interactions regulate collagen fibrillogenesis and corneal stromal assembly. Matrix Biol. 2014; 35:103–111. [PubMed: 24447998]
- Chen S, Birk DE. The regulatory roles of small leucine-rich proteoglycans in extracellular matrix assembly. FEBS J. 2013; 280:2120–2137. [PubMed: 23331954]
- 21. Zhang G, Chen S, Goldoni S, Calder BW, Simpson HC, Owens RT, McQuillan DJ, Young MF, Iozzo RV, Birk DE. Genetic evidence for the coordinated regulation of collagen fibrillogenesis in the cornea by decorin and biglycan. J Biol Chem. 2009; 284:8888–8897. [PubMed: 19136671]
- Reese SP, Underwood CJ, Weiss JA. Effects of decorin proteoglycan on fibrillogenesis, ultrastructure, and mechanics of type I collagen gels. Matrix Biol. 2013; 32:414–423. [PubMed: 23608680]
- Robinson PS, Huang TF, Kazam E, Iozzo RV, Birk DE, Soslowsky LJ. Influence of decorin and biglycan on mechanical properties of multiple tendons in knockout mice. J Biomechanical Eng. 2005; 127:181–185.
- 24. Zhang G, Ezura Y, Chervoneva I, Robinson PS, Beason DP, Carine ET, Soslowsky LJ, Iozzo RV, Birk DE. Decorin regulates assembly of collagen fibrils and acquisition of biomechanical properties during tendon development. J Cell Biochem. 2006; 98:1436–1449. [PubMed: 16518859]
- 25. Dunkman AA, Buckley MR, Mienaltowski MJ, Adams SM, Thomas SJ, Kumar A, Beason DP, Iozzo RV, Birk DE, Soslowsky LJ. The injury response of aged tendons in the absence of biglycan and decorin. Matrix Biol. 2014; 35:232–238. [PubMed: 24157578]
- 26. Dunkman AA, Buckley MR, Mienaltowski MJ, Adams SM, Thomas SJ, Satchell L, Kumar A, Pathmanathan L, Beason DP, Iozzo RV, Birk DE, Soslowsky LJ. Decorin expression is important for age-related changes in tendon structure and mechanical properties. Matrix Biol. 2013; 32:3–13. [PubMed: 23178232]
- Järveläinen H, Sainio A, Wight TN. Pivotal role for decorin in angiogenesis. Matrix Biology. 2015; 43:15–26. [PubMed: 25661523]
- Droguett R, Cabello-Verrugio C, Riquelme C, Brandan E. Extracellular proteoglycans modify TGF-β bio-availability attenuating its signaling during skeletal muscle differentiation. Matrix Biol. 2006; 25:332–341. [PubMed: 16766169]
- 29. Curran CS, Keely PJ. Breast tumor and stromal cell response to TGF-β and hypoxia in matrix deposition. Matrix Biol. 2013; 32:95–105. [PubMed: 23262216]
- Vial C, Gutierrez J, Santander C, Cabrera D, Brandan E. Decorin interacts with connective tissue growth factor (CTGF)/CCN2 by LRR12 inhibiting its biological activity. J Biol Chem. 2011; 286:24242–24252. [PubMed: 21454550]
- Neill T, Schaefer L, Iozzo RV. Decorin, a guardian from the matrix. Am J Pathol. 2012; 181:380– 387. [PubMed: 22735579]
- Neill T, Schaefer L, Iozzo RV. Instructive roles of extracellular matrix on autophagy. Am J Pathol. 2014; 184:2146–2153. [PubMed: 24976620]
- 33. Merline R, Moreth K, Beckmann J, Nastase MV, Zeng-Brouwers J, Tralhão JG, Lemarchand P, Pfeilschifter J, Schaefer RM, Iozzo RV, Schaefer L. Signaling by the matrix proteoglycan decorin controls inflammation and cancer through PDCD4 and microRNA-21. Sci Signal. 2011; 4:ra75. [PubMed: 22087031]

- Frey T, Schroeder N, Manon-Jensen T, Iozzo RV, Schaefer L. Biological interplay between proteoglycans and their innate immune receptors in inflammation. FEBS J. 2013; 280:2165–2179. [PubMed: 23350913]
- 35. Järveläinen HJ, Puolakkainen P, Pakkanen S, Brown EL, Höök M, Iozzo RV, Sage H, Wight TN. A role for decorin in cutaneous wound healing and angiogenesis. Wound Rep Reg. 2006; 14:443–452.
- Nikolovska K, Renke JK, Jungmann O, Grobe K, Iozzo RV, Zamfir AD, Seidler DG. A decorindeficient matrix affects skin chondroitin/dermatan sulfate levels and keratinocyte function. Matrix Biol. 2014; 35:91–102. [PubMed: 24447999]
- 37. Merline R, Lazaroski S, Babelova A, Tsalastra-Greul W, Pfeilschifter J, Schluter KD, Gunther A, Iozzo RV, Schaefer RM, Schaefer L. Decorin deficiency in diabetic mice: aggravation of nephropathy due to overexpression of profibrotic factors, enhanced apoptosis and mononuclear cell infiltration. J Physiol Pharmacol. 2009; 60(suppl 4):5–13. [PubMed: 20083846]
- 38. Wu Z, Horgan CE, Carr O, Owens RT, Iozzo RV, Lechner BE. Biglycan and decorin differentially regulate signaling in the fetal membranes. Matrix Biol. 2014; 35:266–275. [PubMed: 24373743]
- Bolton K, Segal D, McMillan J, Jowett J, Heilbronn L, Abberton K, Zimmet P, Chisholm D, Collier G, Walder K. Decorin is a secreted protein associated with obesity and type 2 diabetes. Int J Obes. 2008; 32:1113–1121.
- Marchica CL, Pinelli V, Borges M, Zummer J, Narayanan V, Iozzo RV, Ludwig MS. A role for decorin in a murine model of allergen-induced asthma. Am J Physiol Lung Cell Mol Physiol. 2011; 300:863–873.
- Bocian C, Urbanowitz AK, Owens RT, Iozzo RV, Gotte M, Seidler DG. Decorin potentiates interferon-gamma activity in a model of allergic inflammation. J Biol Chem. 2013; 288:12699– 12711. [PubMed: 23460644]
- Seidler DG, Mohamed NA, Bocian C, Stadtmann A, Hermann S, Schäfers K, Schäfers M, Iozzo RV, Zarbock A, Götte M. The role for decorin in delayed-type hypersensitivity. J Immunol. 2011; 187:6108–6199. [PubMed: 22043007]
- 43. Baghy K, Dezsó K, László V, Fullár A, Péterfia B, Paku S, Nagy P, Schaff Z, Iozzo RV, Kovalszky I. Ablation of the decorin gene enhances experimental hepatic fibrosis and impairs hepatic healing in mice. Lab Invest. 2011; 91:439–451. [PubMed: 20956977]
- Brandan E, Cabello-Verrugio C, Vial C. Novel regulatory mechanisms for the proteoglycans decorin and biglycan during muscle formation and muscular dystrophy. Matrix Biol. 2008; 27:700–708. [PubMed: 18694824]
- 45. Brandan E, Gutierrez J. Role of skeletal muscle proteoglycans during myogenesis. Matrix Biol. 2013; 32:289–297. [PubMed: 23583522]
- Weis SM, Zimmerman SD, Shah M, Covell JW, Omens JH, Ross J Jr, Dalton N, Jones Y, Reed CC, Iozzo RV, McCulloch AD. A role for decorin in the remodeling of myocardial infarction. Matrix Biol. 2005; 24:313–324. [PubMed: 15949932]
- Zoeller JJ, Pimtong W, Corby H, Goldoni S, Iozzo AE, Owens RT, Ho SY, Iozzo RV. A central role for decorin during vertebrate convergent extension. J Biol Chem. 2009; 284:11728–11737. [PubMed: 19211552]
- Xu YZ, Zhang YH, Zhang YW, Hong B, Liu JM. Dynamic reduction of plasma decorin following ischemic stroke: A pilot study. Neurochem Res. 2012; 37:1843–1848. [PubMed: 22678721]
- Schaefer L. Small leucine-rich proteoglycans in kidney disease. J Am Soc Nephrol. 2011; 22:1200–1207. [PubMed: 21719787]
- Nastase MV, Iozzo RV, Schaefer L. Key roles for the small leucine-rich proteoglycans in renal and pulmonary pathophysiology. Biochim Biophys Acta. 2014; 1840:2460–2470. [PubMed: 24508120]
- Schaefer L. Extracellular matrix molecules: endogenous danger signals as new drug targets in kidney diseases. Curr Opin Pharmacol. 2010; 10:185–190. [PubMed: 20045380]
- 52. Ichii M, Frank MB, Iozzo RV, Kincade PW. The canonical Wnt pathway shapes niches supportive of hematopoietic stem/progenitor cells. Blood. 2012; 119:1683–1692. [PubMed: 22117039]

- Nyman MC, Sainio AO, Pennanen MM, Lund RJ, Vuorikoski S, Sundström JT, Järveläinen HT. Decorin in Human Colon Cancer: Localization In Vivo and Effect on Cancer Cell Behavior In Vitro. J Histochem Cytochem. 2015; 63:710–720. [PubMed: 26001829]
- 54. Neill T, Schaefer L, Iozzo RV. An oncosuppressive role for decorin. Mol Cell Oncol. 2015; 2:e975645.
- 55. Troup S, Njue C, Kliewer EV, Parisien M, Roskelley C, Chakravarti S, Roughley PJ, Murphy LC, Watson PH. Reduced expression of the small leucine-rich proteoglycans, lumican, and decorin is associated with poor outcome in node-negative invasive breast cancer. Clin Cancer Res. 2003; 9:207–214. [PubMed: 12538471]
- 56. Goldoni S, Seidler DG, Heath J, Fassan M, Baffa R, Thakur ML, Owens RA, McQuillan DJ, Iozzo RV. An anti-metastatic role for decorin in breast cancer. Am J Pathol. 2008; 173:844–855. [PubMed: 18688028]
- 57. Reed CC, Waterhouse A, Kirby S, Kay P, Owens RA, McQuillan DJ, Iozzo RV. Decorin prevents metastatic spreading of breast cancer. Oncogene. 2005; 24:1104–1110. [PubMed: 15690056]
- Matsumine A, Shintani K, Kusuzaki K, Matsubara T, Satonaka H, Wakabayashi T, Iino T, Uchida A. Expression of decorin, a small leucine-rich proteoglycan, as a prognostic factor in soft tissue tumors. J Surg Oncol. 2007; 96:411–418. [PubMed: 17579351]
- Iozzo RV, Buraschi S, Genua M, Xu SQ, Solomides CC, Peiper SC, Gomella LG, Owens RT, Morrione A. Decorin antagonizes IGF receptor I (IGF-IR) function by interfering with IGF-IR activity and attenuating downstream signaling. J Biol Chem. 2011; 286:34712–34721. [PubMed: 21840990]
- 60. Kristensen IB, Pedersen L, Ro TD, Christensen JH, Lyng MB, Rasmussen LM, Ditzel HJ, Borset M, Abildgaard N. Decorin is down-regulated in multiple myeloma and MGUS bone marrow plasma and inhibits HGF-induced myeloma plasma cell viability and migration. Eur J Haematol. 2013; 91:196–200. [PubMed: 23607294]
- Wu IC, Wu DC, Huang CC, Lin HS, Chen YK, Tsai HJ, Lu CY, Chou SH, Chou YP, Li LH, Tai SY, Wu MT. Plasma decorin predicts the presence of esophageal squamous cell carcinoma. Int J Cancer. 2010; 127:2138–2146. [PubMed: 20143390]
- 62. Bozoky B, Savchenko A, Guven H, Ponten F, Klein G, Szekely L. Decreased decorin expression in the tumor microenvironment. Cancer Med. 2014; 3:485–491. [PubMed: 24634138]
- Li X, Pennisi A, Yaccoby S. Role of decorin in the antimyeloma effects of osteoblasts. Blood. 2008; 112:159–168. [PubMed: 18436739]
- 64. Sainio A, Nyman M, Lund R, Vuorikoski S, Boström P, Laato M, Boström PJ, Järveläinen H. Lack of decorin expression by human bladder cancer cells offers new tools in the therapy of urothelial malignancies. PLoS ONE. 2013; 8:e76190. [PubMed: 24146840]
- 65. Boström P, Sainio A, Kakko T, Savontaus M, Söderström M, Järveläinen H. Localization of decorin gene expression in normal human breast tissue and in benign and malignant tumors of the human breast. Histochem Cell Biol. 2013; 139:161–171. [PubMed: 23007289]
- 66. Salomäki HH, Sainio AO, Söderström M, Pakkanen S, Laine J, Järveläinen HT. Differential expression of decorin by human malignant and benign vascular tumors. J Histochem Cytochem. 2008; 56:639–646. [PubMed: 18413650]
- 67. Henke A, Grace OC, Ashley GR, Stewart GD, Riddick ACP, Yeun H, O'Donnell M, Anderson RA, Thomson AA. Stromal expression of decorin, semaphorin6D, SPARC, Sprouty 1 and Tsukushi in developing prostate and decreased levels of decorin in prostate cancer. PLoS ONE. 2012; 7:e4251.
- Horvath Z, Kovalszky I, Fullar A, Kiss K, Schaff Z, Iozzo RV, Baghy K. Decorin deficiency promotes hepatic carcinogenesis. Matrix Biol. 2014; 35:194–205. [PubMed: 24361483]
- 69. Duncan MB. Extracellular matrix transcriptome dynamics in hepatocellular carcinoma. Matrix Biol. 2013; 32:393–398. [PubMed: 23727079]
- Iozzo RV, Bolender RP, Wight TN. Proteoglycan changes in the intercellular matrix of human colon carcinoma. Lab Invest. 1982; 47:124–138. [PubMed: 7109538]
- Adany R, Iozzo RV. Hypomethylation of the decorin proteoglycan gene in human colon cancer. Biochem J. 1991; 276:301–306. [PubMed: 1710888]

- 72. Adany R, Heimer R, Caterson B, Sorrell JM, Iozzo RV. Altered expression of chondroitin sulfate proteoglycan in the stroma of human colon carcinoma. Hypomethylation of PG-40 gene correlates with increased PG-40 content and mRNA levels. J Biol Chem. 1990; 265:11389–11396. [PubMed: 2162845]
- Danielson KG, Baribault H, Holmes DF, Graham H, Kadler KE, Iozzo RV. Targeted disruption of decorin leads to abnormal collagen fibril morphology and skin fragility. J Cell Biol. 1997; 136:729–743. [PubMed: 9024701]
- 74. Bi X, Tong C, Dokendorff A, Banroft L, Gallagher L, Guzman-Hartman G, Iozzo RV, Augenlicht LH, Yang W. Genetic deficiency of decorin causes intestinal tumor formation through disruption of intestinal cell maturation. Carcinogenesis. 2008; 29:1435–1440. [PubMed: 18550571]
- 75. Bi X, Pohl NM, Yang GR, Gou Y, Guzman G, Kajdacsy-Balla A, Iozzo RV, Yang W. Decorinmediated inhibition of colorectal cancer growth and migration is associated with E-cadherin *in vitro* and in mice. Carcinogenesis. 2012; 33:326–330. [PubMed: 22159220]
- 76. Iozzo RV, Chakrani F, Perrotti D, McQuillan DJ, Skorski T, Calabretta B, Eichstetter I. Cooperative action of germline mutations in decorin and p53 accelerates lymphoma tumorigenesis. Proc Natl Acad Sci USA. 1999; 96:3092–3097. [PubMed: 10077642]
- 77. Araki K, Wakabayashi H, Shintani K, Morikawa J, Matsumine A, Kusuzaki K, Sudo A, Uchida A. Decorin suppresses bone metastasis in a breast cancer cell line. Oncology. 2009; 77:92–99. [PubMed: 19590249]
- Grant DS, Yenisey C, Rose RW, Tootell M, Santra M, Iozzo RV. Decorin suppresses tumor cellmediated angiogenesis. Oncogene. 2002; 21:4765–4777. [PubMed: 12101415]
- Neill T, Painter H, Buraschi S, Owens RT, Lisanti MP, Schaefer L, Iozzo RV. Decorin antagonizes the angiogenic network. Concurrent inhibition of Met, hypoxia inducible factor-1a and vascular endothelial growth factor A and induction of thrombospondin-1 and TIMP3. J Biol Chem. 2012; 287:5492–5506. [PubMed: 22194599]
- Shintani K, Matsumine A, Kusuzaki K, Morikawa J, Matsubara T, Wakabayashi T, Araki K, Satonaka H, Wakabayashi H, Lino T, Uchida A. Decorin suppresses lung metastases of murine osteosarcoma. Oncology Reports. 2008; 19:1533–1539. [PubMed: 18497961]
- Stock C, Jungmann O, Seidler DG. Decorin and chondroitin-6 sulfate inhibit B16V melanoma cell migration and invasion by cellular acidification. J Cell Physiol. 2011; 226:2641–2650. [PubMed: 21792923]
- Santra M, Skorski T, Calabretta B, Lattime EC, Iozzo RV. *De novo* decorin gene expression suppresses the malignant phenotype in human colon cancer cells. Proc Natl Acad Sci USA. 1995; 92:7016–7020. [PubMed: 7624361]
- De Luca A, Santra M, Baldi A, Giordano A, Iozzo RV. Decorin-induced growth suppression is associated with upregulation of p21, an inhibitor of cyclin-dependent kinases. J Biol Chem. 1996; 271:18961–18965. [PubMed: 8702560]
- 84. Santra M, Mann DM, Mercer EW, Skorski T, Calabretta B, Iozzo RV. Ectopic expression of decorin protein core causes a generalized growth suppression in neoplastic cells of various histogenetic origin and requires endogenous p21, an inhibitor of cyclin-dependent kinases. J Clin Invest. 1997; 100:149–157. [PubMed: 9202067]
- Reed CC, Gauldie J, Iozzo RV. Suppression of tumorigenicity by adenovirus-mediated gene transfer of decorin. Oncogene. 2002; 21:3688–3695. [PubMed: 12032837]
- 86. Tralhão JG, Schaefer L, Micegova M, Evaristo C, Schönherr E, Kayal S, Veiga-Fernandes H, Danel C, Iozzo RV, Kresse H, Lemarchand P. In vivo selective and distant killing of cancer cells using adenovirus-mediated decorin gene transfer. FASEB J. 2003; 17:464–466. [PubMed: 12631584]
- 87. Seidler DG, Goldoni S, Agnew C, Cardi C, Thakur ML, Owens RA, McQuillan DJ, Iozzo RV. Decorin protein core inhibits *in vivo* cancer growth and metabolism by hindering epidermal growth factor receptor function and triggering apoptosis via caspase-3 activation. J Biol Chem. 2006; 281:26408–26418. [PubMed: 16835231]
- 88. Buraschi S, Neill T, Owens RT, Iniguez LA, Purkins G, Vadigepalli R, Evans B, Schaefer L, Peiper SC, Wang Z, Iozzo RV. Decorin protein core affects the global gene expression profile of

the tumor microenvironment in a triple-negative orthotopic breast carcinoma xenograft model. PLoS ONE. 2012; 7:e45559. [PubMed: 23029096]

- Buraschi S, Pal N, Tyler-Rubinstein N, Owens RT, Neill T, Iozzo RV. Decorin antagonizes Met receptor activity and downregulates β-catenin and Myc levels. J Biol Chem. 2010; 285:42075– 42085. [PubMed: 20974860]
- 90. Xu W, Neill T, Yang Y, Hu Z, Cleveland E, Wu Y, Hutten R, Xiao X, Stock SR, Shevrin D, Kaul K, Brendler C, Iozzo RV, Seth P. The systemic delivery of an oncolytic adenovirus expressing decorin inhibits bone metastasis in a mouse model of human prostate cancer. Gene Therapy. 2015; 22:31–40.
- 91. Yang Y, Xu WW, Neill T, Hu Z, Wang CH, Xiao X, Stock S, Guise T, Yun CO, Brendler CB, Iozzo RV, Seth P. Systemic Delivery of an Oncolytic Adenovirus Expressing Decorin for the Treatment of Breast Cancer Bone Metastases. Hum Gene Ther. 2015 In Press.
- Jozzo, RV.; Goldoni, S.; Berendsen, A.; Young, MF. Small leucine-rich proteoglycans. In: Mecham, RP., editor. Extracellular Matrix: An overview. Springer; 2011. p. 197-231.
- 93. Scholzen T, Solursh M, Suzuki S, Reiter R, Morgan JL, Buchberg AM, Siracusa LD, Iozzo RV. The murine decorin. Complete cDNA cloning, genomic organization, chromosomal assignment and expression during organogenesis and tissue differentiation. J Biol Chem. 1994; 269:28270– 28281. [PubMed: 7961765]
- 94. Fernandez-Zapico ME, Ellenrieder V. NFAT transcription factors, the potion mediating "Dr.Jekyll-Mr. Hyde" transformation of the TGFp pathway in cancer cells. Cell Cycle. 2010; 9:3838–3839. [PubMed: 20935481]
- Neill T, Schaefer L, Iozzo RV. Decoding the Matrix: Instructive Roles of Proteoglycan Receptors. Biochemistry. 2015; 54:4583–4598. [PubMed: 26177309]
- 96. Khan GA, Girish GV, Lala N, DiGuglielmo GM, Lala PK. Decorin is a novel VEGFR-2-binding antagonist for the human extravillous trophoblast. Mol Endocrinol. 2011; 25:1431–1443. [PubMed: 21659473]
- Kalamajski S, Aspberg A, Oldberg Å. The decorin sequence SYIRIADTNIT binds collagen type I. J Biol Chem. 2007; 282:16062–16067. [PubMed: 17426031]
- 98. Chen S, Sun M, Meng X, Iozzo RV, Kao WWY, Birk DE. Pathophysiological mechanisms of autosomal dominant congenital stromal corneal dystrophy. C-terminal-truncated decorin results in abnormal matrix assembly and altered expression of small leucine-rich proteoglycans. Am J Pathol. 2011; 179:2409–2419. [PubMed: 21893019]
- 99. Chen S, Sun M, Iozzo RV, Kao WW, Birk DE. Intracellularly-retained decorin lacking the Cterminal ear repeat causes ER stress: a cell-based etiological mechanism for congenital stromal corneal dystrophy. Am J Pathol. 2013; 183:247–256. [PubMed: 23685109]
- 100. Seidler DG. The galactosaminoglycan-containing decorin and its impact on diseases. Curr Opin Struct Biol. 2012; 22:578–582. [PubMed: 22877511]
- 101. Rühland C, Schönherr E, Robenek H, Hansen U, Iozzo RV, Bruckner P, Seidler DG. The glycosaminoglycan chain of decorin plays an important role in collagen fibril formation at the early stages of fibrillogenesis. FEBS J. 2007; 274:4246–4255. [PubMed: 17651433]
- 102. Theocharis AD, Tzanakakis G, Karamanos NK. Proteoglycans in health and disease: Novel proteoglycan roles in malignancy and their pharmacological targeting. FEBS J. 2010; 277:3904– 3923. [PubMed: 20840587]
- Morrione A, Neill T, Iozzo RV. Dichotomy of decorin activity on the insulin-like growth factor-I system. FEBS J. 2013; 280:2138–2149. [PubMed: 23351020]
- 104. Metalli D, Lovat F, Tripodi F, Genua M, Xu SQ, Spinelli M, Alberghina L, Vanoni M, Baffa R, Gomella LG, Iozzo RV, Morrione A. The insulin-like growth factor receptor I promotes motility and invasion of bladder cancer cells through Akt- and mitogen-activated protein kinasedependent activation of paxillin. Am J Pathol. 2010; 176:2997–3006. [PubMed: 20395438]
- 105. Morcavallo A, Buraschi S, Xu SQ, Belfiore A, Schaefer L, Iozzo RV, Morrione A. Decorin differentially modulates the activity of insulin receptor isoform A ligands. Matrix Biol. 2014; 35:82–90. [PubMed: 24389353]
- 106. Skandalis SS, Afratis N, Smirlaki G, Nikitovic D, Theocharis AD, Tzanakakis GN, Karamanos NK. Cross-talk between estradiol receptor and EGFR/IGF-IR signaling pathways in estrogen-

responsive breast cancers: focus on the role and impact of proteoglycans. Matrix Biol. 2014; 35:182–193. [PubMed: 24063949]

- 107. Schaefer L. Complexity of danger: the diverse nature of damage-associated molecular patterns. J Biol Chem. 2014; 289:35237–35245. [PubMed: 25391648]
- 108. Moreth K, Frey H, Hubo M, Zeng-Brouwers J, Nastase MV, Hsieh LT, Haceni R, Pfeilschifter J, Iozzo RV, Schaefer L. Biglycan-triggered TLR-2- and TLR-4-signaling exacerbates the pathophysiology of ischemic acute kidney injury. Matrix Biol. 2014; 35:143–151. [PubMed: 24480070]
- 109. Zeng-Brouwers J, Beckmann J, Nastase MV, Iozzo RV, Schaefer L. De novo expression of circulating biglycan evokes an innate inflammatory tissue response via MyD88/TRIF pathways. Matrix Biol. 2014; 35:132–142. [PubMed: 24361484]
- 110. Hsieh LT, Nastase MV, Zeng-Brouwers J, Iozzo RV, Schaefer L. Soluble biglycan as a biomarker of inflammatory renal diseases. Int J Biochem Cell Biol. 2014; 54C:223–235. [PubMed: 25091702]
- 111. Santra M, Reed CC, Iozzo RV. Decorin binds to a narrow region of the epidermal growth factor (EGF) receptor, partially overlapping with but distinct from the EGF-binding epitope. J Biol Chem. 2002; 277:35671–35681. [PubMed: 12105206]
- Schlessinger J. Ligand-induced, receptor-mediated dimerization and activation of EGF receptor. Cell. 2002; 110:669–672. [PubMed: 12297041]
- 113. Zhu JX, Goldoni S, Bix G, Owens RA, McQuillan D, Reed CC, Iozzo RV. Decorin evokes protracted internalization and degradation of the EGF receptor via caveolar endocytosis. J Biol Chem. 2005; 280:32468–32479. [PubMed: 15994311]
- Moreth K, Iozzo RV, Schaefer L. Small leucine-rich proteoglycans orchestrate receptor crosstalk during inflammation. Cell Cycle. 2012; 11:2084–2091. [PubMed: 22580469]
- 115. Goldoni S, Humphries A, Nyström A, Sattar S, Owens RT, McQuillan DJ, Ireton K, Iozzo RV. Decorin is a novel antagonistic ligand of the Met receptor. J Cell Biol. 2009; 185:743–754. [PubMed: 19433454]
- Schlessinger J. Common and distinct elements in cellular signaling via EGF and FGF receptors. Science. 2004; 306:1506–1507. [PubMed: 15567848]
- Lemmon MA, Schlessinger J. Cell signaling by receptor tyrosine kinases. Cell. 2010; 141:1117– 1134. [PubMed: 20602996]
- 118. Moscatello DK, Santra M, Mann DM, McQuillan DJ, Wong AJ, Iozzo RV. Decorin suppresses tumor cell growth by activating the epidermal growth factor receptor. J Clin Invest. 1998; 101:406–412. [PubMed: 9435313]
- Iozzo RV, Moscatello D, McQuillan DJ, Eichstetter I. Decorin is a biological ligand for the epidermal growth factor receptor. J Biol Chem. 1999; 274:4489–4492. [PubMed: 9988678]
- 120. Csordás G, Santra M, Reed CC, Eichstetter I, McQuillan DJ, Gross D, Nugent MA, Hajnóczky G, Iozzo RV. Sustained down-regulation of the epidermal growth factor receptor by decorin. A mechanism for controlling tumor growth *in vivo*. J Biol Chem. 2000; 275:32879–32887. [PubMed: 10913155]
- 121. Santra M, Eichstetter I, Iozzo RV. An anti-oncogenic role for decorin: downregulation of ErbB2 leads to growth suppression and cytodifferentiation of mammary carcinoma cells. J Biol Chem. 2000; 275:35153–35161. [PubMed: 10942781]
- 122. Minor KH, Bournat JC, Toscano N, Giger RJ, Davies SJA. Decorin, erythroblastic leukaemia viral oncogene homologue B4 and signal transducer and activator of transcription 3 regulation of semaphorin 3A in central nervous system scar tissue. Brain. 2011; 134:1140–1155. [PubMed: 21115466]
- 123. Patel S, Santra M, McQuillan DJ, Iozzo RV, Thomas AP. Decorin activates the epidermal growth factor receptor and elevates cytosolic Ca²⁺ in A431 cells. J Biol Chem. 1998; 273:3121–3124. [PubMed: 9452417]
- 124. Abulrob A, Giuseppin S, Andrade MF, McDermid A, Moreno M, Stanimirovic D. Interactions of EGFR and caveolin-1 in human glioblastoma cells: evidence that tyrosine phosphorylation regulates EGFR association with caveolae. Oncogene. 2004; 23:6967–6979. [PubMed: 15273741]

- 125. Aberle H, Bauer A, Stappert J, Kispert A, Kemler R. β-catenin is a target for the ubiquitinproteasome pathway. EMBO J. 1997; 16:3797–3804. [PubMed: 9233789]
- 126. Danilkovitch-Miagkova A, Miagkov A, Skeel A, Nakaigawa N, Zbar B, Leonard EJ. Oncogenic mutants of RON and MET receptor tyrosine kinases cause activation of the β-catenin pathway. Mol Cell Biol. 2001; 21:5857–5868. [PubMed: 11486025]
- 127. Clevers H. Wnt/β-catenin signaling in development and disease. Cell. 2006; 127:469–480. [PubMed: 17081971]
- 128. Monga SPS, Mars WM, Pediaditakis P, Bell A, Mulé K, Bowen WC, Wang X, Zarnegar R, Michalopoulos GK. Hepatocyte growth factor induces Wnt-independent nuclear translocation of β-catenin after MET-β-catenin dissociation in hepatocytes. Cancer Res. 2008; 62:2064–2071. [PubMed: 11929826]
- 129. Rasola A, Fassetta M, De Bacco F, D'Alessandro L, Gramaglia D, Di Renzo MG, Comoglio PM. A positive feedback loop between hepatocyte growth factor receptor and β-catenin sustains colorectal cancer cell invasive growth. Oncogene. 2007; 26:1078–1087. [PubMed: 16953230]
- 130. Finch AJ, Soucek L, Junttila MR, Swigart LB, Evan GI. Acute overexpression of Myc in intestinal epithelium recapitulates some but not all the changes elicited by Wnt/β-catenin pathway activation. Mol Cell Biol. 2009; 29:5306–5315. [PubMed: 19635809]
- 131. Herynk MH, Tsan R, Radinsky R, Gallick GE. Activation of c-Met in colorectal carcinoma cells leads to constitutive association of tyrosine-phosphorylated β-catenin. Clin Exp Metastasis. 2003; 20:291–300. [PubMed: 12856716]
- 132. Neill T, Jones HR, Crane-Smith Z, Owens RT, Schaefer L, Iozzo RV. Decorin induces rapid secretion of thrombospondin-1 in basal breast carcinoma cells via inhibition of Ras homolog gene family, member A/Rho-associated coiled-coil containing protein kinase 1. FEBS J. 2013; 280:2353–2368. [PubMed: 23350987]
- 133. Murphy-Ullrich JE, Sage EH. Revisiting the matricellular concept. Matrix Biol. 2014; 37:1–14. [PubMed: 25064829]
- 134. Wallace DM, Murphy-Ullrich JE, Downs JC, O'Brien CJ. The role of matricellular proteins in glaucoma. Matrix Biol. 2014; 37:174–182. [PubMed: 24727033]
- 135. Resovi A, Pinessi D, Chiorino G, Taraboletti G. Current understanding of the thrombospondin-1 interactome. Matrix Biol. 2014; 37:83–91. [PubMed: 24476925]
- 136. Rogers NM, Sharifi-Sanjani M, Csanyi G, Pagano PJ, Isenberg JS. Thrombospondin-1 and CD47 regulation of cardiac, pulmonary and vascular responses in health and disease. Matrix Biol. 2014; 37:92–101. [PubMed: 24418252]
- 137. Duquette M, Nadler M, Okuhara D, Thompson J, Shuttleworth T, Lawler J. Members of the thrombospondin gene family bind stromal interaction molecule 1 and regulate calcium channel activity. Matrix Biol. 2014; 37:15–24. [PubMed: 24845346]
- 138. Soto-Pantoja DR, Shih HB, Maxhimer JB, Cook KL, Ghosh A, Isenberg JS, Roberts DD. Thrombospondin-1 and CD47 signaling regulate healing of thermal injury in mice. Matrix Biol. 2014; 37:25–34. [PubMed: 24840925]
- 139. DeCarvalho S. Angiokines, angiogenesis and angiolymphoproliferative syndromes (ALPS). Angiology. 1983; 34:231–243. [PubMed: 6188389]
- 140. Relaix F, Weng X, Marazzi G, Yang E, Copeland N, Jenkins N, Spence SE, Sassoon D. Pw1, a novel zinc finger gene implicated in the myogenic and neuronal lineages. Dev Biol. 1996; 177:383–396. [PubMed: 8806818]
- 141. Kim J, Ashworth L, Branscomb E, Stubbs L. The human homolog of a mouse-imprinted gene, *Peg3*, maps to a zinc finger gene-rich region of human chromosome 19q13.4. Genome Res. 1997; 7:532–540. [PubMed: 9149948]
- 142. Kuroiwa Y, Kaneko-Ishino T, Kagitani F, Kohda T, Li LL, Tada M, Suzuki R, Yokoyama M, Shiroishi T, Wakana S, Barton SC, Ishino F, Surani MA. *Peg3* imprinted gene on proximal chromosome 7 encodes for a zinc finger protein. Nature Genet. 1996; 12:186–190. [PubMed: 8563758]
- 143. Thiaville MM, Huang JM, Kim H, Ekram MB, Roh TY, Kim J. DNA-binding motif and target genes of the imprinted transcription factor PEG3. Gene. 2013; 512:314–320. [PubMed: 23078764]

- 144. Besson V, Smeriglio P, Wegener A, Relaix F, Nait Oumesmar B, Sassoon DA, Marazzi G. *PW1* gene/paternally expressed gene 3 (PW1/Peg3) identifies multiple adult stem and progenitor cell populations. Proc Natl Acad Sci USA. 2011; 108:11470–11475. [PubMed: 21709251]
- 145. Bonfanti C, Rossi G, Tedesco FS, Giannotta M, Benedetti S, Tonlorenzi R, Antonini S, Marazzi G, Dejana E, Sassoon D, Cossu G, Messina G. PW1/Peg3 expression regulates key properties that determine mesoangioblast stem cell competence. Nat Commun. 2015; 6:6364. [PubMed: 25751651]
- 146. Relaix F, Wei X, Li W, Pan J, Lin Y, Bowtell DD, Sasoon DA, Wu X. Pw1/Peg3 is a potential cell death mediator and cooperates with Siah1a in p53-mediated apoptosis. Proc Natl Acad Sci USA. 2000; 97:2105–2110. [PubMed: 10681424]
- 147. Johnson MD, Wu X, Aithmitti N, Morrison RS. Peg3/Pw1 is a mediator between p53 and Bax in DNA damage-induced neuronal death. J Biol Chem. 2002; 277:23000–23007. [PubMed: 11943780]
- 148. Yamaguchi A, Taniguchi M, Hori O, Ogawa S, Tojo N, Matsuoka N, Miyake S, Kasai K, Sugimoto H, Tamatani M, Yamashita T, Yamashita T, Tohyama M. Peg3/Pw1 is involved in p53-mediated cell death pathway in brain ischemia/hypoxia. J Biol Chem. 2002; 277:623–629. [PubMed: 11679586]
- Deng Y, Wu X. Peg3/Pw1 promotes p53-mediated apoptosis by inducing Bax translocation from cytosol to mitochondria. Proc Natl Acad Sci USA. 2000; 97:12050–12055. [PubMed: 11050235]
- 150. Coletti D, Yang E, Marazzi G, Sassoon D. TNFalpha inhibits skeletal myogenesis through a PW1-dependent pathway by recruitment of caspase pathways. EMBO J. 2002; 21:631–642. [PubMed: 11847111]
- 151. Li LL, Keverne EB, Aparicio SA, Ishino F, Barton SC, Surani MA. Regulation of maternal behavior and offspring growth by paternally expressed *Peg3*. Science. 1999; 284:330–333. [PubMed: 10195900]
- 152. Champagne FA, Curley JP, Swaney WT, Hasen NS, Keverne EB. Paternal influence on female behavior: the role of Peg3 in exploration, olfaction, and neuroendocrine regulation of maternal behavior of female mice. Behav Neurosci. 2009; 123:469–480. [PubMed: 19485553]
- 153. Nye MDHC, Huang Z, Vidal AC, Wang F, Overcash F, Smith JS, Vasquez B, Hernandez B, Swai B, Oneko O, Mlay P, Obure J, Gammon MD, Bartlett JA, Murphy SK. Association between methylation of *paternally expressed gene 3 (PEG3)*, cervical intraepithelial neoplasia and invasive cervical cancer. PLoS ONE. 2013; 8:e56325. [PubMed: 23418553]
- 154. Dowdy SC, Gostout BS, Shridhar V, Wu X, Smith DI, Podratz KC, Jiang SW. Biallelic methylation and silencing of paternally expressed gene 3 (*PEG3*) in gynecologic cancer cell lines. Gynecol Oncol. 2005; 99:126–134. [PubMed: 16023706]
- 155. Feng W, Marquez RT, Lu Z, Liu J, Lu KH, Issa JPJ, Fishman DM, Yu Y, Bast RC. Imprinted tumor suppressor genes ARHI and PEG3 are the most frequently down-regulated in human ovarian cancers by loss of heterozygosity and promoter methylation. Cancer. 2008; 112:1489– 1502. [PubMed: 18286529]
- 156. Maegawa S, Yoshioka H, Itaba N, Kubota N, Nishihara S, Shirayoshi Y, Nanba E, Oshimura M. Epigenetic silencing of PEG3 gene expression in human glioma cell lines. Mol Carcinogenesis. 2001; 31:1–9.
- 157. Kohda T, Asai A, Kuroiwa Y, Kobayashi S, Aisaka K, Nagashima G, Yoshida MC, Kondo Y, Kagiyama N, Kirino T, Kaneko-Ishino T, Ishino F. Tumour suppressor activity of human imprinted gene *PEG3* in a glioma cell line. Genes Cells. 2001; 6:237–247. [PubMed: 11260267]
- 158. Jiang X, Yu Y, Yang HW, Agar NYR, Frado L, Johnson MD. The imprinted gene *PEG3* inhibits Wnt signaling and regulates glioma growth. J Biol Chem. 2010; 285:8472–8480. [PubMed: 20064927]
- 159. Buraschi S, Neill T, Goyal A, Poluzzi C, Smythies J, Owens RT, Schaefer L, Torres A, Iozzo RV. Decorin causes autophagy in endothelial cells via Peg3. Proc Natl Acad Sci USA. 2013; 110:E2582–E2591. [PubMed: 23798385]
- 160. Mizushima N, Levine B. Autophagy in mammalian development and differentiation. Nat Cell Biol. 2010; 12:823–830. [PubMed: 20811354]

- 161. He C, Klionsky DJ. Regulation mechanisms and signaling pathways of autophagy. Annu Rev Genet. 2009; 43:67–93. [PubMed: 19653858]
- 162. Neill T, Torres AT, Buraschi S, Iozzo RV. Decorin has an appetite for endothelial cell autophagy. Autophagy. 2013; 9:1626–1628. [PubMed: 23989617]
- 163. Pattingre S, Levine B. Bcl-2 inhibition of autophagy: A new route to cancer? Cancer Res. 2006; 66:2885–2888. [PubMed: 16540632]
- 164. Goyal A, Neill T, Owens RT, Schaefer L, Iozzo RV. Decorin activates AMPK, an energy sensor kinase, to induce autophagy in endothelial cells. Matrix Biol. 2014; 34:46–54. [PubMed: 24472739]
- 165. Neill T, Torres A, Buraschi S, Owens RT, Hoek J, Baffa R, Iozzo RV. Decorin induces mitophagy in breast carcinoma cells via peroxisome proliferator-activated receptor y coactivator-1α (PGC-1α) and mitostatin. J Biol Chem. 2014; 289:4952–4968. [PubMed: 24403067]
- 166. Vecchione A, Fassan M, Anesti V, Morrione A, Goldoni S, Baldassarre G, Byrne D, D'Arca D, Palazzo JP, Lloyd J, Scorrano L, Gomella LG, Iozzo RV, Baffa R. *MITOSTATIN*, a putative tumor suppressor on chromosome 12q24.1, is downregulated in human bladder and breast cancer. Oncogene. 2009; 28:257–269. [PubMed: 18931701]
- 167. Fassan M, D'Arca D, Letko J, Vecchione A, Gardiman MP, McCue P, Wildemore B, Rugge M, Shupp-Byrne D, Gomella LG, Morrione A, Iozzo RV, Baffa R. Mitostatin is down-regulated in human prostate cancer and suppresses the invasive phenotype of prostate cancer cells. PLoS ONE. 2011; 6:e19771. [PubMed: 21573075]
- 168. Cerqua C, Anesti V, Pyakurel A, Liu D, Naon D, Wiche G, Baffa R, Dimmer KS, Scorrano L. Trichoplein/mitostatin regulates endoplasmic reticulum-mitochondria juxtaposition. EMBO Reports. 2010; 11:854–860. [PubMed: 20930847]
- 169. Ventura-Clapier R, Garnier A, Veksler W. Transcriptional control of mitochondrial biogenesis: the central role of PGC-1α. Cardiovasc Res. 2008; 79:208–217. [PubMed: 18430751]
- 170. Haq R, Shoag J, Andreu-Perez P, Yokoyama S, Edelman H, Rowe GC, Frederick DT, Hurley AD, Nellore A, Kung AL, Wargo JA, Song JS, Fisher DE, Arany Z, Widlund HR. Oncogenic BRAF regulates oxidative metabolism via PGC1α and MITF. Cancer Cell. 2013; 23:302–315. [PubMed: 23477830]
- 171. Vazquez F, Lim JH, Chim H, Bhalla K, Girnun G, Pierce K, Clish CB, Granter SR, Widlund HR, Spiegelman BM, Puigserver P. PGC1a expression defines a subset of human melanoma tumors with increased mitochondrial capacity and resistance to oxidative stress. Cancer Cell. 2013; 23:287–301. [PubMed: 23416000]
- 172. Levine B, Kroemer G. Autophagy in the pathogenesis of disease. Cell. 2008; 132:27–42. [PubMed: 18191218]
- 173. Dagda R, Cherra SJI, Kulich SM, Tandon A, Park D, Chu CT. Loss of PINK1 function promotes mitophagy through effects on oxidative stress and mitochondrial fission. J Biol Chem. 2009; 284:13843–13855. [PubMed: 19279012]
- 174. Kubli DA, Gustafsson ÅB. Mitochondria and mitophagy: the yin and yang of cell death control. Circ Res. 2012; 111:1208–1221. [PubMed: 23065344]
- 175. Staropoli JF, McDermott C, Martinat C, Schulman B, Demireva E, Abeliovich A. Parkin is a component of an SCF-like ubiquitin ligase complex and protects postmitotic neurons from kainate excitotoxicity. Neuron. 2003; 37:735–749. [PubMed: 12628165]
- 176. Narendra D, Walker JE, Youle R. Mitochondrial quality control mediated by PINK1 and Parkin: links to parkinsonism. Cold Spring Harb Perspect Biol. 2012; 4
- 177. Kane LA, Lazarou M, Fogel AI, Li Y, Yamano K, Sarraf SA, Banerjee S, Youle RJ. PINK1 phosphorylates ubiquitin to activate Parkin E3 ubiquitin ligase activity. J Cell Biol. 2014; 205:143–153. [PubMed: 24751536]
- 178. Koyano F, Okatsu K, Kosako H, Tamura Y, Go E, Kimura M, Kimura Y, Tsuchiya H, Yoshihara H, Hirokawa T, Endo T, Fon EA, Trempe JF, Saeki Y, Tanaka K, Matsuda N. Ubiquitin is phosphorylated by PINK1 to activate parkin. Nature. 2014; 510:162–166. [PubMed: 24784582]
- 179. Matsuda N, Sato S, Shiba K, Okatsu K, Saisho K, Gautier CA, Sou YS, Saiki S, Kawajiri S, Sato F, Kimura M, Komatsu M, Hattori N, Tanaka K. PINK1 stabilized by mitochondrial

depolarization recruits Parkin to damaged mitochondria and activates latent Parkin for mitophagy. J Cell Biol. 2010; 189:211–221. [PubMed: 20404107]

- 180. Iguchi M, Kujuro Y, Okatsu K, Koyano F, Kosako H, Kimura M, Suzuki N, Uchiyama S, Tanaka K, Matsuda N. Parkin-catalyzed ubiquitin-ester transfer is triggered by PINK1-dependent phosphorylation. J Biol Chem. 2013; 288:22019–22032. [PubMed: 23754282]
- 181. Geisler S, Holmstrom KM, Skujat D, Fiesel FC, Rothfuss OC, Kahle PJ, Springer W. PINK1/ Parkin-mediated mitophagy is dependent on VDAC1 and p62/SQSTM1. Nat Cell Biol. 2010; 12:119–131. [PubMed: 20098416]
- 182. Vincow ES, Merrihew G, Thomas RE, Shulman NJ, Beyer RP, MacCoss MJ, Pallanck LJ. The PINK1-Parkin pathway promotes both mitophagy and selective respiratory chain turnover in vivo. Proc Natl Acad Sci U S A. 2013; 110:6400–6405. [PubMed: 23509287]
- 183. Vincow ES, Merrihew G, Thomas RE, Shulman NJ, Beyer RP, MacCoss MJ, Pallanck LJ. The PINK1-Parkin pathway promotes both mitophagy and selective respiratory chain turnover in vivo. Proc Natl Acad Sci U S A. 2013; 110:6400–6405. [PubMed: 23509287]
- 184. Naba A, Clauser KR, Ding H, Whittaker CA, Carr SA, Hynes RO. The extracellular matrix: Tools and insights for the "omics" era. Matrix Biol. 2015
- 185. Wang RC, Wei Y, An Z, Zou Z, Xiao G, Bhagat G, White M, Reichelt J, Levine B. Akt-mediated regulation of autophagy and tumorigenesis through Beclin 1 phosphorylation. Science. 2012; 338:956–959. [PubMed: 23112296]
- 186. Wei Y, Zou Z, Becker N, Anderson M, Sumpter R, Xiao G, Kinch L, Koduru P, Christudass CS, Veltri RW, Grishin NV, Peyton M, Minna J, Bhagat G, Levine B. EGFR-mediated beclin 1 phosphorylation in autophagy suppression, tumor progression, and tumor chemoresistance. Cell. 2013; 154:1269–1284. [PubMed: 24034250]
- 187. Gubbiotti MA, Neill T, Frey H, Schaefer L, Iozzo RV. Decorin is an autophagy-inducible proteoglycan and is required for proper in vivo autophagy. Matrix Biol. 2015 In Press.
- 188. Gubbiotti MA, Iozzo RV. Proteoglycans regulate autophagy via outside-in signaling: An emerging new concept. Matrix Biol. 2015 In Press.
- 189. Douglass S, Goyal A, Iozzo RV. The role of perlecan and endorepellin in the control of tumor angiogenesis and endothelial cell autophagy. Connect Tissue Res. 2015; 19:1–11.
- 190. Ständer M, Naumann U, Dumitrescu L, Heneka M, Löschmann P, Gulbins E, Dichgans J, Weller M. Decorin gene transfer-mediated suppression of TGF-β synthesis abrogates experimental malignant glioma growth in vivo. Gene Therapy. 1998; 5:1187–1194. [PubMed: 9930319]
- 191. Biglari A, Bataille D, Naumann U, Weller M, Zirger J, Castro MG, Lowenstein PR. Effects of ectopic decorin in modulating intracranial glioma progression *in vivo*, in a rat syngeneic model. Cancer Gene Therapy. 2004; 11:721–732. [PubMed: 15475879]

Abbreviations

АМРКа	AMP-activated protein kinase, alpha			
AP4	Activating enhancer binding protein 4			
ATG	Autophagy related gene			
Bcl2	B-cell CLL/lymphoma 2			
BRAF	proto-oncogene B-Raf			
ECM	extracellular matrix			
EGFR	Epidermal growth factor receptor			
ERK	extracellular regulated kinase			
GSK-3β	glycogen synthase kinase 3β			

HGF	hepatocyte growth factor		
HIF-1a	Hypoxia inducible factor-1a		
IGF-I	insulin-like growth factor 1		
IGF-IR	insulin-like growth factor 1 receptor		
IgG	immunoglobulin G-like folds		
IR-A	Insulin receptor isoform A		
IRS	insulin receptor substrate 1		
LC3	Microtubule-associated protein 1A/1B-light chain 3		
LRR	leucine-rich repeat		
MAPK	Mitogen activated protein kinase		
MMP	Matrix metalloproteinase		
mTOR	mechanistic target of rapamycin		
OXPHOS	oxidative phosphorylation		
p70S6K	Ribosomal Protein S6 Kinase, 70kDa		
PDGFR	Platelet derived growth factor receptor		
Peg3	Paternally expressed gene 3		
PGC-1a	Peroxisome proliferator activated receptor γ co-activator-1 $\!\alpha$		
PI3K	phosphoinositide 3 kinase		
PINK1	PTEN-induced putative kinase-1		
PKB/Akt	Protein kinase B		
Rheb	Ras homolog enriched in brain		
RhoA	Ras homolog gene family, member A		
ROCK1	Rho-associated, coiled -coil-containing protein kinase 1		
RRM	RNA recognition motif		
RTK	receptor tyrosine kinase		
SLRP	small leucine-rich proteoglycan		
TGF-β1	Transforming growth factor beta 1		
TIMP3	Tissue inhibitor of metalloproteinases 3		
TSP1	Thrombospondin 1		
VDAC	Voltage-dependent anion channel		
VEGFA	vascular endothelial growth factor A		

Vps34 Vacuolar Protein Sorting 34

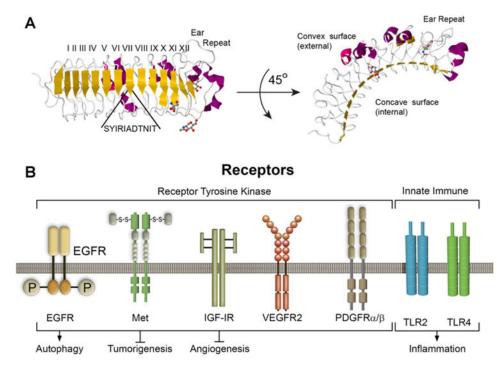


Fig. 1.

The solved crystal structure of decorin permits association with a multitude of cell surface receptors. (A) Cartoon ribbon diagram of monomeric bovine decorin rendered with PyMol v1.8. (PDB accession #: 1XKU). Vertical arrows designate β -strands whereas coiled ribbons indicate α -helices. Roman numerals situated above the diagram define each LRR from left to right, by convention. The type I collagen binding sequence has been included and is shaded yellow. (B). Schematic depiction of the various RTKs and innate immune receptors that decorin engages. Please, consult the text for additional information.

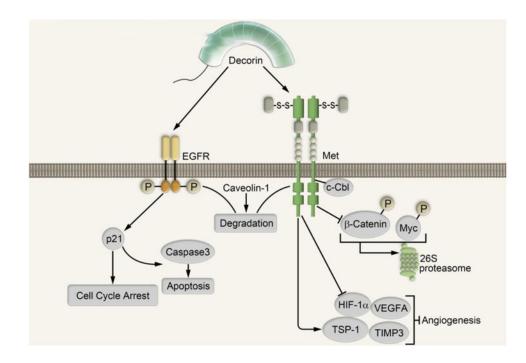


Fig. 2.

EGFR and Met coordinate growth inhibition, apoptosis, and angiostasis. Schematic representation of the signaling pathways modulated in response to decorin binding. Please, consult the text for additional information.

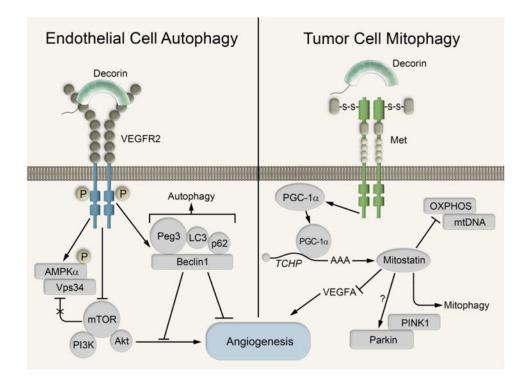


Fig. 3.

VEGFR2 and Met evoke endothelial cell autophagy and tumor cell mitophagy. Schematic representation delineating the signaling pathways active in response to decorin acting as a partial agonist of VEGFR2 or Met for endothelial cell autophagy or tumor cell mitophagy induction, respectively. Please, consult the manuscript for additional information concerning these pathways.

Table 1

Pre-clinical studies exploiting several delivery mechanisms for decorin as a therapeutic modality against cancer and across multiple species.

Tumor type	Origin	Delivery system	Reference(s)
Orthotopic squamous cell carcinoma	Human	Ectopic expression	Santra et al [84]
Orthotopic squamous cell carcinoma	Human	Recombinant decorin proteoglycan or protein core	Seidler et al [87]
Orthoptopic breast carcinoma	Human	Ad-Decorin	Reed et al [85]
Lung adenocarcinoma	Human	Ad-Decorin	Tralhão <i>et al</i> [86]
Breast metastases	Human	Ad-Decorin	Reed et al [57]
Breast metastases	Human, Rat	Ad-Decorin	Goldoni et al [56]
Multiple myeloma	Human	Rercombinant decorin proteoglycan	Li et al [63]
Orthotopic glioma	Rat	Ectopic expression	Stander et al [190]
Orthotopic glioma	Rat	Ectopic expression	Biglari et al [191]
Orthotopic breast carcinoma	Human	Recombinant decorin proteoglycan or protein core	Buraschi et al [88]
Bone metastases of prostate carcinoma	Human	Ad-Decorin	Xu et al [90]
Bone metastases of breast carcinoma	Human	Ad-Decorin	Yang et al [91]