

# SCIENTIFIC REPORTS



OPEN

## An ultrasensitive label-free electrochemical immunosensor based on signal amplification strategy of multifunctional magnetic graphene loaded with cadmium ions

Received: 15 November 2015

Accepted: 20 January 2016

Published: 16 February 2016

Faying Li<sup>1</sup>, Yueyun Li<sup>1,2</sup>, Yunhui Dong<sup>1</sup>, Liping Jiang<sup>1</sup>, Ping Wang<sup>1</sup>, Qing Liu<sup>1</sup>, Hui Liu<sup>1</sup> & Qin Wei<sup>2</sup>

Herein, a novel and ultrasensitive label-free electrochemical immunosensor was proposed for quantitative detection of human Immunoglobulin G (IgG). The amino functionalized magnetic graphenes nanocomposites (NH<sub>2</sub>-GS-Fe<sub>3</sub>O<sub>4</sub>) were prepared to bond gold and silver core-shell nanoparticles (Au@Ag NPs) by constructing stable Au-N and Ag-N bond between Au@Ag NPs and -NH<sub>2</sub>. Subsequently, the Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> was applied to absorb cadmium ion (Cd<sup>2+</sup>) due to the large surface area, high conductivity and exceptional adsorption capability. The functional nanocomposites of gold and silver core-shell magnetic graphene loaded with cadmium ion (Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>) can not only increase the electrocatalytic activity towards hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) but also improve the effective immobilization of antibodies because of synergistic effect presented in Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>, which greatly extended the scope of detection. Under the optimal conditions, the proposed immunosensor was used for the detection of IgG with good linear relation in the range from 5 fg/mL to 50 ng/mL with a low detection limit of 2 fg/mL (S/N = 3). Furthermore, the proposed immunosensor showed high sensitivity, special selectivity and long-term stability, which had promising application in bioassay analysis.

Human Immunoglobulin G (IgG) is an important component in the immune system, which plays a crucial role in recognizing bacteria and viruses<sup>1,2</sup>. In addition, it has also been administered therapeutically from exogenous pooled donor sources<sup>3,4</sup>. The concentration of IgG in blood and other body fluid is direct correlation to the standard of humoral immunity, which abnormal IgG concentrations often indict the risk of disease or vulnerability to infection<sup>3,5</sup>. Therefore, it is necessary to develop method for quantitative detection IgG for immunological diagnosis.

In recent years, various detection assays have been developed for the detection of IgG, such as chemiluminescence<sup>6</sup>, colorimetric sensors<sup>7,8</sup>, fluorescence<sup>9,10</sup> and electrochemical immunosensors<sup>11,12</sup>. In comparisons, electrochemical immunoassay may be an excellent candidate for the detection of tumor markers because of fast analytical time, low detection limits, high sensitivity, simple pretreatment procedure and inexpensive instrumentation<sup>13,14</sup>. As one important branch of electrochemical immunosensors, the label-free immunosensors have become an attractive and promising approach to direct detection of tumor makers because simple procedure, ease of use and rapidity of the assay and without the use of secondary antibody compared with sandwich-type immunosensors<sup>15,16</sup>.

<sup>1</sup>School of Chemical Engineering, Shandong University of Technology, Zibo, 255049, P.R. China. <sup>2</sup>Key Laboratory of Chemical Sensing & Analysis in Universities of Shandong, School of Chemistry and Chemical Engineering, University of Jinan, Jinan, 250022, P.R. China. Correspondence and requests for materials should be addressed to Y.L. (email: liyueyun71@163.com) or Q.W. (email: sdjndxwq@163.com)

Over the past decade, a wide variety of nanomaterials have been designed as transducing materials to fabricate label-free electrochemical immunosensors, such as, graphene sheets (GS)<sup>17,18</sup>, metal oxides<sup>19,20</sup> and noble metal nanoparticles<sup>21,22</sup>. As a kind of popular transducing materials, graphene oxide (GO) possess many oxygen-containing functional groups which make the water-solubility better and make it easy to form a stable chemical bond with various materials, such as, magnetic nanostructures, metallic or catalytic<sup>23,24</sup>. To date, Fe<sub>3</sub>O<sub>4</sub> NPs have attracted a considerable interest due to its good biocompatibility<sup>25</sup> and great auxiliary catalytic activity toward the reduction of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>)<sup>20,26</sup>. Therefore, the graphene was introduced to combine with Fe<sub>3</sub>O<sub>4</sub> NPs by chemical reaction. Among the noble metal nanoparticles, gold nanoparticles (Au NPs) and silver nanoparticles (Ag NPs) have attracted a considerable interest because of its good biocompatibility<sup>27</sup> and superior auxiliary catalytic activity towards the reduction of H<sub>2</sub>O<sub>2</sub><sup>28,29</sup>. Further, Au and Ag NPs can facilitate the electron transfer because of its superior electrochemical properties. So, Au NPs are widely used in fixing antibody because of its superior biocompatibility and chemical stability<sup>30,31</sup>. Compared with Au NPs, Ag NPs exhibit excellent electro catalytic activity towards the reduction of H<sub>2</sub>O<sub>2</sub><sup>32,33</sup>. Compared with single metal NPs, bimetallic NPs with a core-shell structure show distinctly unique characteristics than their monometallic counterparts<sup>34</sup>. In addition, the Au@Ag NPs could also enable the facile conjugation of capture antibodies because of the stable Au-N<sup>35,36</sup> and Ag-N<sup>37</sup> bond between Au@Ag NPs and -NH<sub>2</sub> on antibodies. Simultaneously, the amino functionalized magnetic graphenes nanocomposites (NH<sub>2</sub>-GS-Fe<sub>3</sub>O<sub>4</sub>) was applied to absorb Cd<sup>2+</sup><sup>38,39</sup> due to the large surface area, high conductivity and exceptional adsorption capability. The adsorbed Cd<sup>2+</sup>, can further promote the redox of H<sub>2</sub>O<sub>2</sub>, which was applied to signal amplification. The signal amplification strategy, using the synergetic effect present in functional nanocomposites of gold and silver core-shell magnetic graphene loaded with cadmium ion (Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>), can further increase electron transfer efficiency on electrode surface and the reaction efficiency of the nanocomposite toward H<sub>2</sub>O<sub>2</sub> reduction to improve the detection sensitivity of the immunosensor.

In this research, a novel and ultrasensitive label-free immunosensor for the quantitative detection the IgG was prepared using Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> as a signal amplification platform. The synergetic effect between functionalized magnetic graphene nanocomposites (GS-Fe<sub>3</sub>O<sub>4</sub>), Au@Ag NPs and Cd<sup>2+</sup> can not only increase the electro-catalytic activity towards hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) but also improve the effective immobilization of antibodies, which greatly extend the scope of detection. The proposed immunosensor provides a useful technology for the quantitative detection of IgG in human serum, shows high sensitivity, good selectivity and stability for the quantitative detection of IgG, holding a great potential in clinical and diagnostic applications.

## Experimental

**Apparatus and reagents.** Scanning electron microscopy (SEM) images and energy-dispersive X-ray spectroscopy (EDS) analysis were collected using a FEI QUANTA FEG250 coupled with INCA Energy X-MAX-50. Fourier transform infrared spectroscopy (FTIR) spectrum was collected using VERTEX 70 spectrometer (Bruker, Germany). All electrochemistry measurements were performed on a CHI760E electrochemical workstation (Chenhua Instrument Shanghai Co., Ltd, China) by using a conventional three-electrode system consisted of a glassy carbon electrode (GCE, 4 mm in diameter) as working electrode, a saturated calomel electrode (SCE) as the reference electrode, and the platinum wire electrode as the counter electrode.

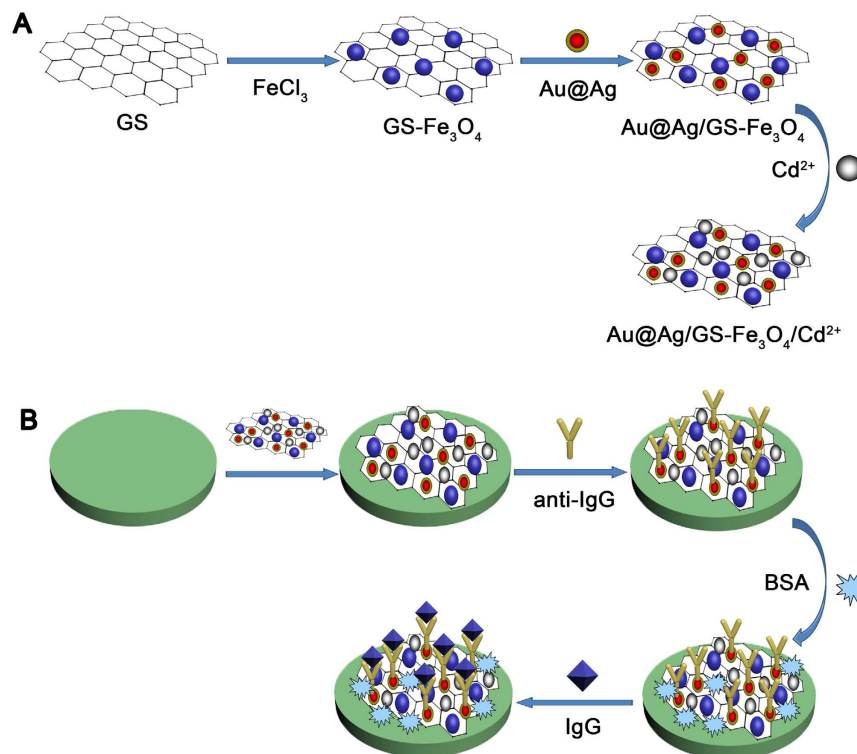
HAuCl<sub>4</sub>·4H<sub>2</sub>O was purchased from Sinopharm Chemical Reagent Co., Ltd (Shanghai, China). FeCl<sub>3</sub>·6H<sub>2</sub>O was purchased from Damao Chemical Reagent Co., Ltd (Tianjin, China). 3-aminopropyl triethoxysilane (APTES) was purchased from XiBao biological technology co., Ltd. (Shanghai, China). Human immunoglobulin G (IgG), goat Anti-Human IgG and bovine serum albumin (BSA, 96–99%) were obtained from Dingguo Biochemical Reagents (Beijing, China). All other reagents were at analytical grade and ultrapure water was used throughout the study.

**Preparation of the NH<sub>2</sub>-GS-Fe<sub>3</sub>O<sub>4</sub>.** Graphene oxide (GO) was synthesized according an improved Hummer's method<sup>40</sup>. In brief, graphite flakes (0.6 g) and KMnO<sub>4</sub> (3.6 g) were dispersed in a mixture of concentrated H<sub>2</sub>SO<sub>4</sub> (72 mL) and H<sub>3</sub>PO<sub>4</sub> (8 mL). Subsequently, the reaction was heated to 50 °C and maintained at this temperature for 12 h with stirred. The mixture was cooled to room temperature after reaction and poured onto ice (80 mL) with 30% H<sub>2</sub>O<sub>2</sub> (0.6 mL). Then, the mixture was centrifuged and removed the supernatant. After that, the remaining solid material was thoroughly washed with water, 0.2 mol/L HCl (60 mL), ethanol and ether. Finally, the obtained solid material was dried in vacuum overnight.

GS-Fe<sub>3</sub>O<sub>4</sub> was synthesized according to a protocol described previously<sup>39</sup>. FeCl<sub>3</sub>·6H<sub>2</sub>O (0.5 g) was dissolved in ethylene glycol (10 mL) to form a clear solution, then, NaAc (1.5 g), ethanediamine (5 mL) and GO (0.5 g) was added into the mixture orderly and dissolved under stirred vigorously for 30 min. Subsequently, the mixture was transferred into the teflon-lined stainless steel autoclave. The autoclave was heated to and maintained at 200 °C for 8 h and cooled down to room temperature after reaction. The prepared compound sample was thoroughly washed to remove the impurities and separated via a strong magnet. The resulting GS-Fe<sub>3</sub>O<sub>4</sub> was dried under high vacuum overnight. It should be noted that the GO was translated into the graphene sheet (GS) in the process of reaction.

The amino-functionalized GS-Fe<sub>3</sub>O<sub>4</sub> (NH<sub>2</sub>-GS-Fe<sub>3</sub>O<sub>4</sub>) was synthesized by an improved method<sup>41</sup>. Briefly, GS-Fe<sub>3</sub>O<sub>4</sub> (0.1 g) was dispersed in a solution of ethanol (10 mL) containing APTES (0.1 mL). Subsequently, the solution was heated to 70 °C and kept for 1.5 h. Finally, the NH<sub>2</sub>-GS-Fe<sub>3</sub>O<sub>4</sub> was obtained by magnetic separation and dried under high vacuum overnight.

**Preparation of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>.** The preparation of Au NPs was referred to the classical Frens method<sup>42</sup>. In brief, Sodium citrate (1.5 mL, 10 mg/mL) was added to the aqueous solution (100 mL) containing HAuCl<sub>4</sub> (1 wt%, 1 mL). Then, the mixture was heated to reflux and kept for 15 min. A wine red solution of Au NPs was obtained after being cooled to room temperature and stored at 4 °C. Au@Ag NPs was synthesized according



**Figure 1.** (A) The prepare process of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>; (B) Schematic diagram for fabrication of the label-free electrochemical immunosensor.

an to the literature previously<sup>43</sup>. 1.0 mL of ascorbic acid (100 mmol/L) and 0.5 mL of AgNO<sub>3</sub> (10 mmol/L) were added into 20 mL of hexadecyl trimethyl ammonium bromide (CTAB) solution (50 mmol/L). After that, 2 mL of the Au NPs solution was added into the mixed solution. Then, 0.1 mL of NaOH aqueous solution (1 mol/L) was added drop-wise to the above solution under stirred vigorously. In this way, the color of the solution changed from red to bright golden yellow, indicating the success preparation of Au@Ag NPs.

The prepared NH<sub>2</sub>-GS-Fe<sub>3</sub>O<sub>4</sub> (10 mg) was dispersed in the Au@Ag NPs (40 mL) solution. The suspension was stirred for 24 h and magnetic separation. Au@Ag NPs could bind with amino groups on the surface of NH<sub>2</sub>-GS-Fe<sub>3</sub>O<sub>4</sub>. The sediment was dried and the obtained powder was designated as Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>.

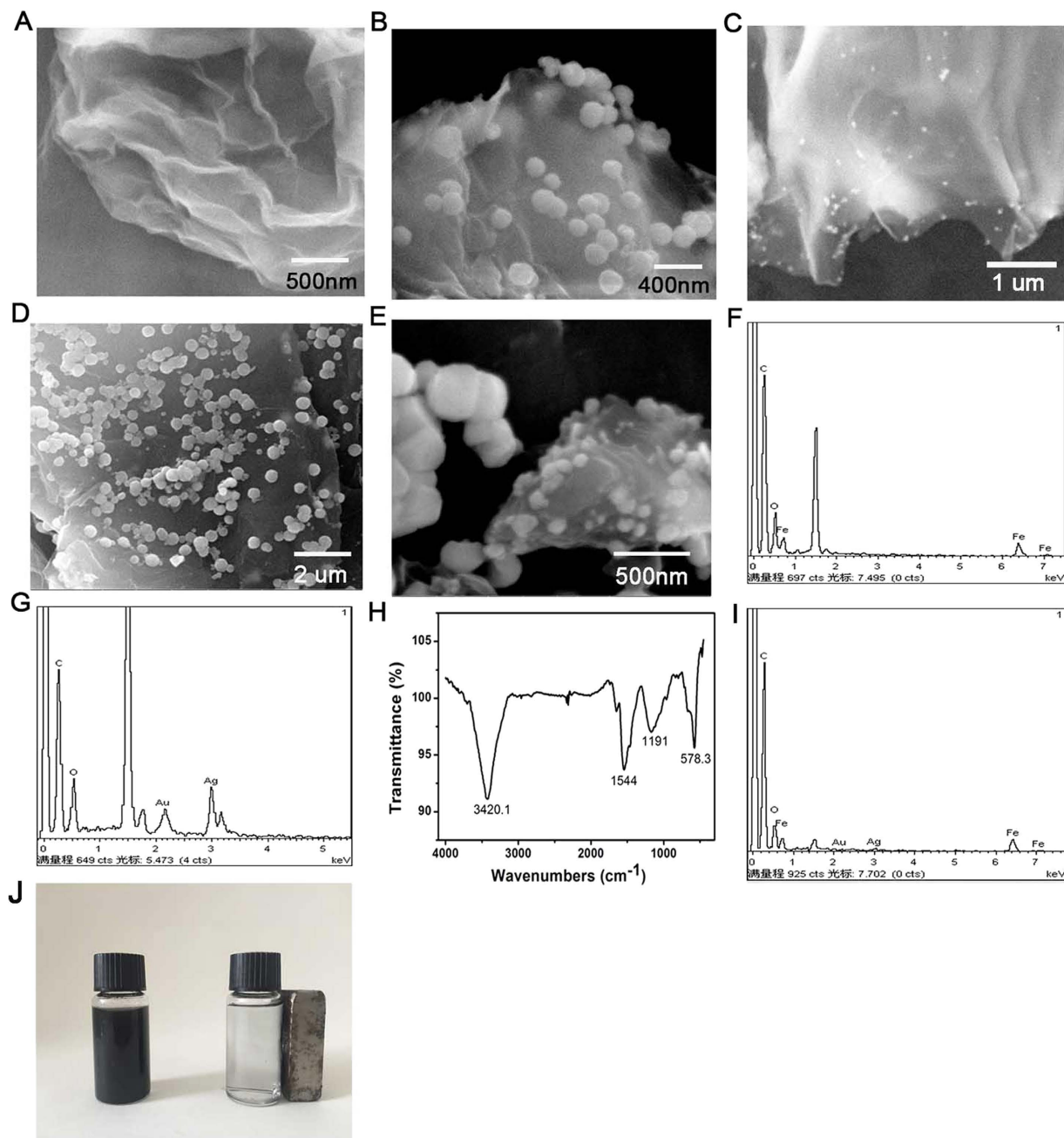
**Preparation of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>.** Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> (10 mg) was dispersed into cadmium sulphate solution (10 mL, 2 mg/mL). The solution had been oscillated for 24 h to ensure that Cd<sup>2+</sup> could be fully absorbed onto the Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>. The Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> was obtained for further use after magnetic separation. Figure 1A shows the preparation process of the Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>.

**Fabrication of the immunosensor.** The schematic diagram of the stepwise self-assembly procedure of the proposed label-free immunosensor is shown in Fig. 1B. Generally, GCE was polished to a mirror-like, and washed thoroughly with ultrapure water. Firstly, the aqueous solution of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> (2 mg/mL, 6  $\mu$ L) was coated onto the surface of GCE and dried at room temperature. After drying for 1 h, the resultant Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>/GCE was incubated with anti-IgG (10  $\mu$ g/mL, 6  $\mu$ L) by the chemical bonding between Au@Ag NPs and available amine groups of anti-IgG. After incubated for another 1 h at 4 °C, BSA solution (1 wt%, 3  $\mu$ L) was added onto the electrode to eliminate nonspecific binding sites. After 1 h incubation, the BSA/anti-IgG/Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>/GCE was washed with ultrapure water and incubated with a varying concentration of IgG (5 fg/mL to 50 ng/mL, 6 mL) for 1 h at room temperature, and then the IgG/BSA/anti-IgG/Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>/GCE was washed extensively to remove unbounded IgG molecules. Ultimately, the proposed immunosensor was stored at 4 °C for further usage.

**Detection of IgG.** Phosphate buffered saline (PBS, pH = 6.8) were prepared by mixing Na<sub>2</sub>HPO<sub>4</sub> and KH<sub>2</sub>PO<sub>4</sub> stock solution and used as the electrolyte in the process of electrochemical measurements. Amperometric *i*-*t* curve was used to record the amperometric response by scanning the potential at -0.4 V. 5 mM H<sub>2</sub>O<sub>2</sub> was added into the PBS (10 mL) after the back ground current was stabilized. The cyclic voltammetry (CV) experiments were recorded in 5 mM K<sub>3</sub>[Fe(CN)<sub>6</sub>] by scanning the potential from -1 V to 1 V. For A.C. impedance measurements, a frequency range of 0.1 kHz to 100 Hz and AC voltage amplitude of 5 mV were used.

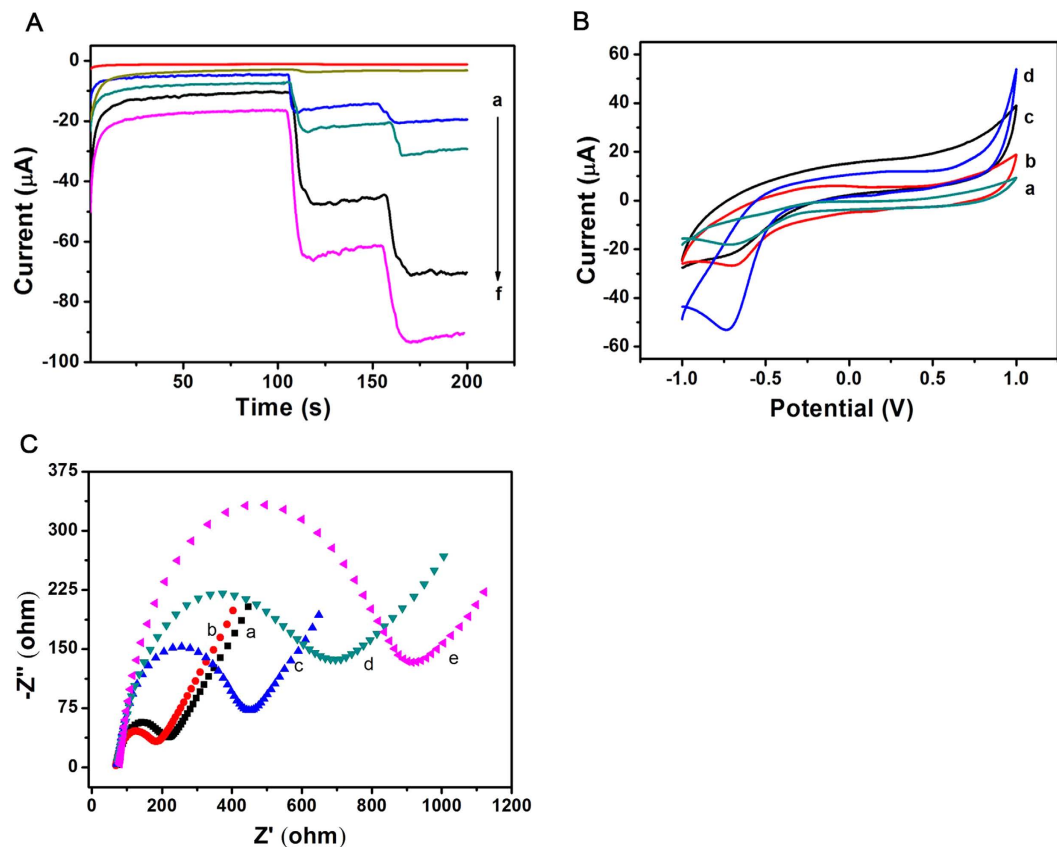
## Results and Discussion

**Characterization of Au/Ag@GS-Fe<sub>3</sub>O<sub>4</sub>.** As shown in (Fig. 2A), the surface of GO exhibits wrinkled, paper-like structure. After magnetization, GS was loaded with a lot of nearly monodisperse microspheres Fe<sub>3</sub>O<sub>4</sub>



**Figure 2.** SEM image: GO (A), GS-Fe<sub>3</sub>O<sub>4</sub> (B); Au@Ag/GS (C); Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> (D,E). EDX spectrum: GS-Fe<sub>3</sub>O<sub>4</sub> (F); EDX spectrum of the Au@Ag/GS (G); FT-IR spectrometer analysis of GS-Fe<sub>3</sub>O<sub>4</sub> (H); comparison of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> in the absence and presence of a magnet (I).

(Fig. 2B). It was observed that these Fe<sub>3</sub>O<sub>4</sub> NPs own quasi-monodisperse size with an average grain diameter of 230 nm by the SEM. Obvious Fe, C, and O elements were observed which prove Fe<sub>3</sub>O<sub>4</sub> was loaded on the GS successfully (Fig. 2F). Furthermore, FT-IR spectra of GS-Fe<sub>3</sub>O<sub>4</sub> were recorded to prove that GS was loaded successfully with Fe<sub>3</sub>O<sub>4</sub> (Fig. 2H). As shown in Fig. 2G, the peak at 3420 cm<sup>-1</sup> corresponding to the hydroxyl group stretching vibration (O-H), two peaks at 1544 and 1191 cm<sup>-1</sup> corresponded to the carboxyl (-COOH) and carbonyl (C=O) stretching vibrations. A peak at 573 cm<sup>-1</sup> was typical Fe-O stretching vibration of the prepared GS-Fe<sub>3</sub>O<sub>4</sub> composites<sup>44</sup>. This result indicated that Fe<sub>3</sub>O<sub>4</sub> NPs was loaded successfully on the surface of GO. When the Au@Ag NPs were coated on the surface of NH<sub>2</sub>-GS, the surface morphology was greatly alternated. Au@Ag NPs are well monodispersed and uniformly spherical in shape. As shown in (Fig. 2C), many small particles were loaded on the NH<sub>2</sub>-GS by constructing stable Au-N and Ag-N bond between Au@Ag NPs and -NH<sub>2</sub>. Subsequently, EDX spectrum of Au@Ag-GS was recorded which clearly confirmed the presence of Au@Ag NPs attached on the surface of the NH<sub>2</sub>-GS. As shown in (Fig. 2G), obvious Au, Ag, C, and O elements were observed, in which the signal of Au, Ag are assigned to the Au@Ag NPs, and that of C and O elements are belonged to the

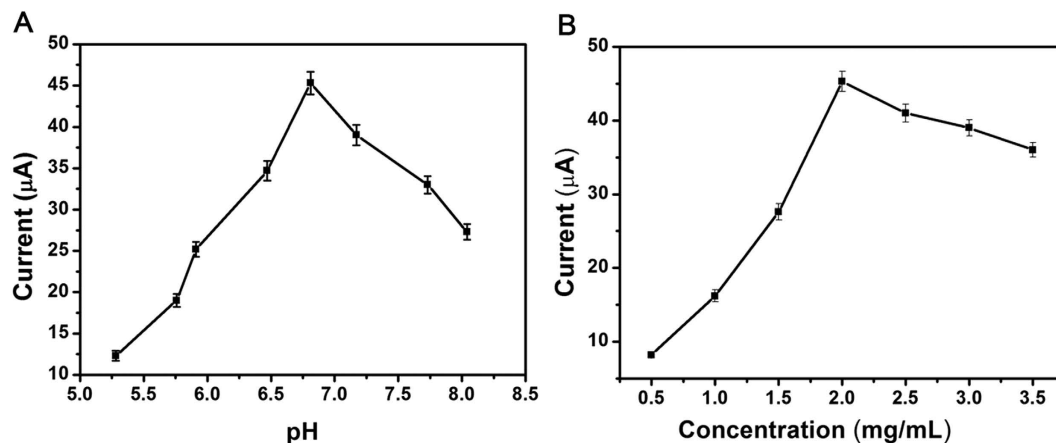


**Figure 3.** (A) Amperometric response: (a) bare GCE, (b) GO, (c) GS-Fe<sub>3</sub>O<sub>4</sub>, (d) Au@Ag NPs, (e) Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>, (f) Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>; (B) CV of the immunosensor: a bare GCE was scanned in 0.1 mg/mL of Cd<sup>2+</sup> from -1.0 V to 1.0 V (a); using Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> as signal amplification platform (c); using Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> as signal amplification platform in PBS at pH = 6.8 before (b) and after (d) the addition of 5 mM H<sub>2</sub>O<sub>2</sub>; (C) Nyquist plots of the EIS method: GCE (a), Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>/GCE (b), anti-IgG/Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>/GCE (c), BSA/anti-IgG/Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>/GCE (d), IgG/BSA/anti-IgG/Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>/GCE (e).

GS. Figure 2D,E shows the SEM image of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> in different scale respectively, which contains two kinds of size of particle. It is the combination of the Fig. 2B,C, suggesting the synthesized of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> successfully. Furthermore, obvious Au, Ag, C, and O and Fe elements were observed in Fig. 2I, which also suggest the synthesized of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> successfully. Figure 2J also proved that the magnetic field presents significant effect on the dispersion.

**The mechanism of multiple signal amplification strategy.** The sensitivity of the label-free immunosensor was affected directly by the superior electrocatalytic properties of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> towards the reduction of H<sub>2</sub>O<sub>2</sub>. To comparative analysis, the amperometric i-t curve was employed to record the catalytic properties of bare GCE, GO, GS-Fe<sub>3</sub>O<sub>4</sub>, Au@Ag NPs, Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>, Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> to H<sub>2</sub>O<sub>2</sub>. As shown in Fig. 3A, there no catalytic effect for bare GCE towards the reduction of H<sub>2</sub>O<sub>2</sub> (curve a). In addition, a very weak signal was detected when GS (curve b) were loaded on the electrode. Fe<sub>3</sub>O<sub>4</sub> possess excellent auxiliary catalytic activity towards the reduction of H<sub>2</sub>O<sub>2</sub><sup>20</sup>. When GS-Fe<sub>3</sub>O<sub>4</sub> was applied as the signal amplification platform, the current response was significantly increased (curve c). Simultaneously, a much larger current response was observed when Au@Ag NPs was applied to modify the bare GCE due to the synergistic effect of Au NPs<sup>28</sup> and Ag NPs<sup>45</sup> towards the reduction of H<sub>2</sub>O<sub>2</sub> (curve d). Subsequently, the current response was further increased (curve e) when the electrode was modified with Au/Ag/GS-Fe<sub>3</sub>O<sub>4</sub>. As expected, using Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> as signal amplification platform displayed the highest current change due to synergistic effect (curve f). The result indicated that Fe<sub>3</sub>O<sub>4</sub> NPs, Au@Ag NPs and Cd<sup>2+</sup> promote the multiple signal amplification toward the reduction of H<sub>2</sub>O<sub>2</sub> as an analytical signal. Hence, the resultant nanocomposites (Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>) was adopted as signal amplification platform due to excellent electrochemical performance to improve the sensitivity.

CV was applied to further verify that the Cd<sup>2+</sup> was adsorbed successfully onto the Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> and have better electrocatalytic properties toward the reduction of H<sub>2</sub>O<sub>2</sub> (Fig. 3B). A reduction peak at -0.7 V is obvious when a bare GCE was scanned in 0.1 mg/mL of Cd<sup>2+</sup> solution (curve a). Simultaneously, a reduction peak (curve b) was also observed at -0.7 V when a bare GCE was scanned in PBS (pH = 6.8) using Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> as signal amplification platform. Hence, it inferred that Cd<sup>2+</sup> was adsorbed successfully onto the Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>. Furthermore, there was no current response (curve c) when the electrode was scanned in PBS (pH = 6.8) using



**Figure 4.** (A) The optimization of experimental conditions with pH, (B) Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> concentration, Error bar = RSD (n = 5).

Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub> as signal amplification platform. The comparison clearly suggests that the observed current response was directly associated with absorbed Cd<sup>2+</sup>. In order to verify that Cd<sup>2+</sup> possess better electrocatalytic properties toward the reduction of H<sub>2</sub>O<sub>2</sub>. As shown in Fig. 3B, a bare GCE was modified with Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> in PBS (pH = 6.8) with the addition of 5 mM H<sub>2</sub>O<sub>2</sub>. After the addition of H<sub>2</sub>O<sub>2</sub>, a dramatic increase of the reduction current (curve d) was observed at -0.7 V, which further verify the Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> has a good electrocatalytic performance towards the reduction of H<sub>2</sub>O<sub>2</sub>.

#### Electrochemical impedance spectroscopy (EIS) characterization of immunosensor.

Electrochemical impedance spectroscopy (EIS) was regarded as an effective method to characterize the fabrication process of the proposed immunosensor by monitoring the interfacial properties<sup>46</sup> and the changes of the electron-transfer resistance (Ret)<sup>47</sup>. The Nyquist plots of EIS were recorded in a solution which consist of 0.1 M KCl and 2.5 mmol/L Fe(CN)<sub>6</sub><sup>3-</sup>/Fe(CN)<sub>6</sub><sup>4-</sup>, the frequencies from 0.1 to 10<sup>5</sup> Hz and the potentiostatic at 0.188 V.

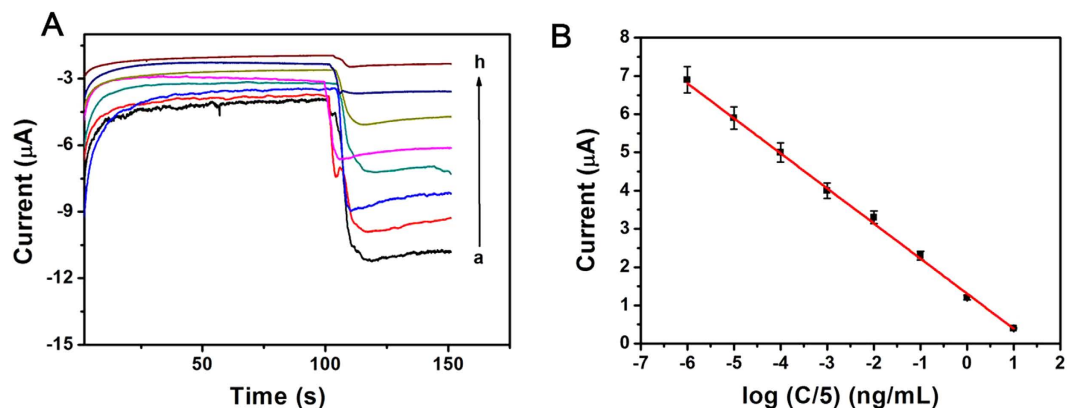
As shown in Fig. 3, the bare GCE exhibited a smaller Ret (curve a). After the Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> was modified on the electrode, the semicircle is much smaller (curve b) due to the high electrical transport properties of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>. After incubation with anti-IgG, the Ret (curve c) was significantly increased due to the anti-IgG is protein which can hinder electron transfer, which indicates anti-IgG was immobilized on the electrode successfully. Followed by blocking the nonspecific binding spots with BSA, the Ret (curve d) was further increased due to the blocking effect on electron transferring by the modified protein molecules on the surface of the electrode. Additionally, the Ret further increased with the addition of IgG (curve e), which indicates the successful capture of IgG and the formation of immunocomplex layer hinder the electron transfer. As a result, we can conclude that the proposed immunosensor was fabricated successfully.

**Optimization of experimental conditions.** In order to detect optimal electrocatalytic signal, it was necessary to optimize the experimental conditions including pH and the concentration of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>. The pH of the PBS was investigated with same concentration of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> (2.0 mg/mL). As shown in Fig. 4A, the current signal increases with the variation of pH from 5.6 to 6.8, and then decreases with the variation of pH from 6.8 to 8.1. The optimal amperometric response was obtained at pH = 6.8. It was inferred that the pH value obviously affected the electrocatalytic process of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> toward the reduction of H<sub>2</sub>O<sub>2</sub>. Simultaneously, the highly acidic or alkaline surroundings would influence the activity of the antigens, antibodies and damage the immobilized protein<sup>48,49</sup>. By contrast, the antigens and antibodies could keep their bioactivity in this approximate neutral conditions of pH<sup>50</sup>.

The concentration of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> would affect the amperometric response by accelerating the electron transfer efficiency and immobilize of antibodies. As shown in Fig. 4B, with the increasing of the concentration from 0.5 mg/mL to 2.0 mg/mL, the current signal increased, but the current signal decreased with the concentration from 2.0 mg/mL to 3.5 mg/mL. The optimal amperometric response was obtained at 2.0 mg/mL. It was inferred that the increasing of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> film thickness might lead to an increase of interface electron transfer resistance, and the electron transfer become more difficult. Simultaneously, the higher or lower concentrations of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> influenced the catalytic performance for the reduction of H<sub>2</sub>O<sub>2</sub><sup>51</sup>. Therefore, the concentration of 2.0 mg/mL was used as the optimal concentration for this study.

In addition, the rest experiment conditions were also required strictly. For example, the concentration of antibodies was 10 µg/mL, the incubation time of 1 h, which was enough to capture antigen (ng/mL) and achieve the specific recognition between antigens and antibodies<sup>52</sup>. Under the optimal conditions, the proposed immunosensor will obtain an optimal electrochemical response for quantitative detection of IgG.

**Analytical performance of the immunosensor.** Under the optimal experimental conditions, amperometric i-t curve was used to detect different concentrations of IgG (Fig. 5A) using Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup> as a signal amplification platform in pH 6.8 PBS. The amperometric response towards the reduction of 5 mmol/L



**Figure 5.** (A) Amperometric response of the immunosensor to different concentration of IgG, from a to h: 5 fg/mL, 50 fg/mL, 500 fg/mL, 5 pg/mL, 50 pg/mL, 500 pg/mL, 5 ng/mL, 50 ng/mL; (B) Calibration curves of immunosensor to different concentrations of IgG, error bar = RSD (n = 5).

$\text{H}_2\text{O}_2$  and IgG concentrations were shown in Fig. 5A. As can be seen, the *i-t* current response change was linearly related to the logarithmic values of the IgG concentration within the range from 5 fg/mL to 50 ng/mL, and with an extremely low limit of detection of 2 fg/mL ( $S/N = 3$ ). The regression equation of the calibration curve as:  $I = 1.301 - 0.9168 \log C$  with correlation coefficient of 0.9970 (Fig. 5B). The results demonstrate the proposed method have an acceptable quantitative performance for the IgG detection. The performance of the immunosensor was compared with previously reported methods for the detection of IgG. As shown in Supplementary Table S1, the proposed immunosensor has a wider linear range and lower detection limit than previously report. The low detection limit was attributed to several factors. Firstly,  $\text{Au@Ag/GS-Fe}_3\text{O}_4/\text{Cd}^{2+}$ , a novel material, is a combination of  $\text{Fe}_3\text{O}_4$ -GS,  $\text{Au@Ag}$  NPs and  $\text{Cd}^{2+}$  through covalent bonding, which has high electrocatalytic performance towards the reduction of  $\text{H}_2\text{O}_2$  leading to higher sensitivity. Secondly,  $\text{Au@Ag}$  NPs not only possess good biocompatibility which firmly conjugated with a relatively large amount of antibodies, but also in assisting the electron transfer process to amplify the signal. In addition,  $\text{Cd}^{2+}$  possess good electrocatalytic activity towards the reduction of  $\text{H}_2\text{O}_2$ , which can be used to amplify the detection signal and led to the high sensitivity of the designed immunosensor. Consequently, the synergic effect between  $\text{Au@Ag/GS-Fe}_3\text{O}_4/\text{Cd}^{2+}$  can not only increase the electrocatalytic activity towards  $\text{H}_2\text{O}_2$  but also improve the effective immobilization of antibodies, which could greatly improve the probability of antibody-antigen recognition and extended the scope of detection. Hence, high sensitivity is one of the characteristics of the proposed immunosensor.

**Reproducibility, selectivity and stability.** To evaluate the reproducibility of the immunosensor, a series of five different electrodes were prepared for the detection of 0.05 ng/mL of IgG (Fig. 6A). The relative standard deviation (RSD) of the measurements for the five electrodes was 3.6%, indicating the precision and reproducibility of the immunosensor is acceptable.

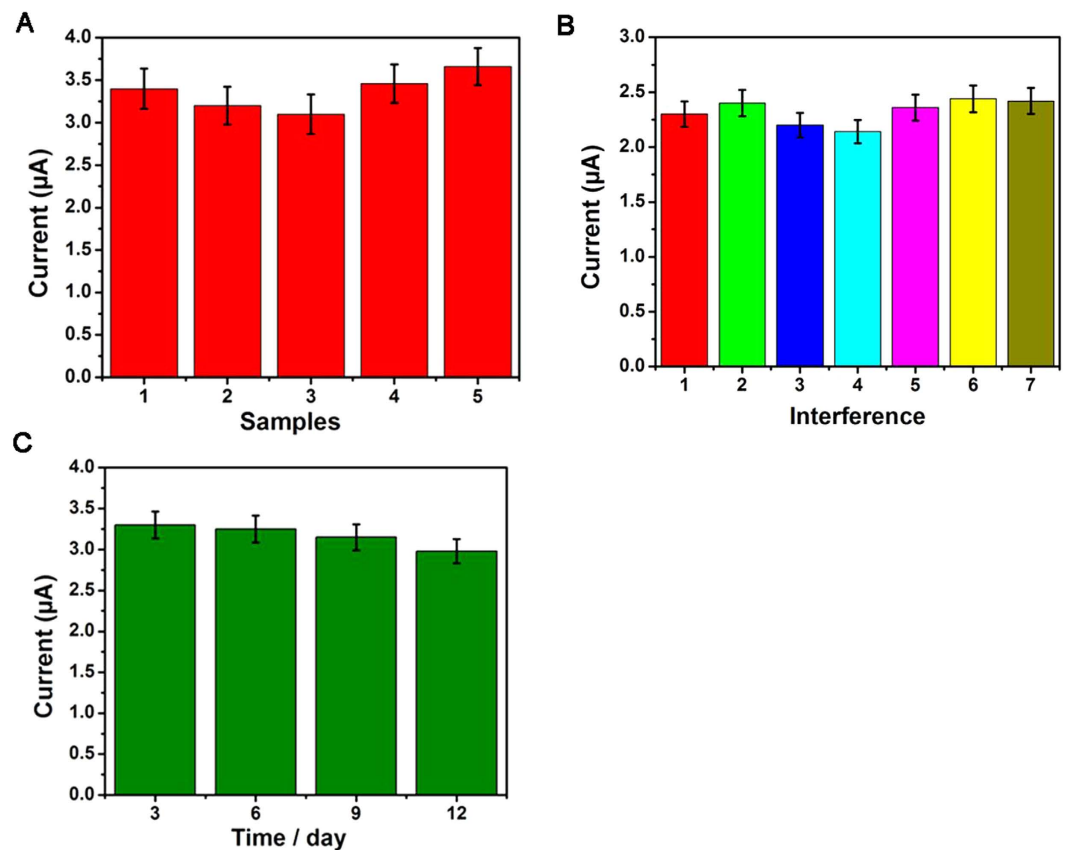
To investigate the selective of the proposed immunosensor, interferences study was performed using (alpha fetoprotein) AFP, BSA, carcinoembryonic antigen (CEA), and glucose. IgG solution (0.5 ng/mL) containing 50 ng/mL of interfering substances were measured by the proposed immunosensor (Fig. 6B). The electrocatalytic current response variation was 4.2%, less than 5% of that without interferences, suggesting the selectivity of the immunosensor is good.

The stability was investigated at 0.05 ng/mL of IgG. As shown in Fig. 6C, the current response of the immunosensor kept constant after 3 days, the current response of the immunosensor only had a minor change of 4.2% after 2 weeks. Subsequently, the current response retained 90.5% of their initial current after 3 weeks. It could be found that current responses to same concentration of IgG has no apparent change compared to the immunosensor freshly prepared which was used to directly detect the same concentration of IgG without being stored, suggesting the stability of the immunosensors was also acceptable. The better stability can be attributed to the good biocompatibility of transducing materials. The reproducibility, selectivity and stability were all acceptable, indicating that the proposed immunosensor was suitable for quantitative detection of IgG in real samples.

**Real sample analysis.** In order to test the feasibility and precision of the proposed label-free immunosensor, standard addition method was used to detect the recoveries of different concentrations of IgG in human serum samples. As shown in Supplementary Table S2, the RSD was in the range from 1.40% to 3.01% and the recovery was in the range from 99.53% to 100.4%. The results imply that the proposed immunosensor could be effectively applied to the quantitative determination of IgG in human serums.

## Conclusions

In this paper, a novel and ultrasensitive label-free electrochemical immunosensor for quantitative detection of IgG was prepared using  $\text{Au@Ag/GS-Fe}_3\text{O}_4/\text{Cd}^{2+}$  as a signal amplification platform. To ensure a high-performance electrochemical immunosensor,  $\text{Au@Ag/GS-Fe}_3\text{O}_4/\text{Cd}^{2+}$  was immobilized on the electrode, which can improve the electronic transmission rate and increase the surface area to capture a larger amount of antibodies. The



**Figure 6.** (A) Amperometric change response of biosensor to different electrodes treated in same way; (B) Current responses of the immunosensor to 0.5 ng/mL IgG (1), 0.5 ng/mL IgG + 50 ng/mL AFP (2), 0.5 ng/mL IgG + 50 ng/mL BSA (3), 0.5 ng/mL IgG + 50 ng/mL CEA (4), 0.5 ng/mL IgG + 50 ng/mL glucose (5), 0.5 ng/mL IgG + 50 ng/mL BSA + 50 ng/mL AFP (6), 0.5 ng/mL IgG + 50 ng/mL BSA + 50 ng/mL CEA (7). Error bar = RSD (n = 5).

proposed label-free immunosensor displayed wide linear range, low detection limit, high sensitivity, good reproducibility, long-term stability and acceptable selectivity. Herein, this proposed method will not only expand the application of Au@Ag/GS-Fe<sub>3</sub>O<sub>4</sub>/Cd<sup>2+</sup>, but also provide an attractive way to detect other biomolecules.

## References

- Kapur, R., Einarsdottir, H. K. & Vidarsson, G. IgG-effector functions: “The Good, The Bad and The Ugly”. *Immunol. Lett.* **160**, 139–144 (2014).
- Liu, L. *et al.* A Label-Free Voltammetric Immunoassay Based on 3D-Structured RGO–MWCNT–Pd for Detection of Human Immunoglobulin G. *Sensor. Actuat B: Chem.* **211**, 170–176 (2015).
- Hou, S. *et al.* Exploration of attenuated total reflectance mid-infrared spectroscopy and multivariate calibration to measure immunoglobulin G in human sera. *Talanta* **142**, 110–119 (2015).
- Schwab, I. & Nimmerjahn, F. Intravenous immunoglobulin therapy: how does IgG modulate the immune system? *Nat. Rev. Immunol.* **13**, 176–189 (2013).
- Sitcharungsri, R. *et al.* Immunoglobulin values in healthy Thai children aged ≤24 months determined by nephelometry. *Asian. Pac. J. Allergy.* **31**, 307 doi: 310.12932/AP10306. 12931.12934. 12013 (2013).
- Li, Z. P., Wang, Y. C., Liu, C. H. & Li, Y. K. Development of chemiluminescence detection of gold nanoparticles in biological conjugates for immunoassay. *Anal. Chim. Acta* **551**, 85–91 (2005).
- Shi, H., Yuan, L., Wu, Y. & Liu, S. Colorimetric immunosensing via protein functionalized gold nanoparticle probe combined with atom transfer radical polymerization. *Biosens. Bioelectron.* **26**, 3788–3793 (2011).
- Lai, K. K., Renneberg, R. & Mak, W. C. Multifunctional protein particles with dual analytical channels for colorimetric enzymatic bioassays and fluorescent immunoassays. *Biosens. Bioelectron.* **32**, 169–176 (2012).
- Luo, Y. *et al.* Interference-free determination of ischemia-modified albumin using quantum dot coupled X-ray fluorescence spectroscopy. *Biosens. Bioelectron.* **51**, 136–142 (2014).
- Tan, Y. *et al.* Proximity-dependent protein detection based on enzyme-assisted fluorescence signal amplification. *Biosens. Bioelectron.* **51**, 255–260 (2014).
- Cao, X., Liu, S., Feng, Q. & Wang, N. Silver nanowire-based electrochemical immunoassay for sensing immunoglobulin G with signal amplification using strawberry-like ZnO nanostructures as labels. *Biosens. Bioelectron.* **49**, 256–262 (2013).
- Qiu, L.-P. *et al.* A label-free electrochemical immunoassay for IgG detection based on the electron transfer. *Talanta* **83**, 42–47 (2010).
- Ahirwal, G. K. & Mitra, C. K. Gold nanoparticles based sandwich electrochemical immunosensor. *Biosens. Bioelectron.* **25**, 2016–2020 (2010).
- Zhao, Y. *et al.* Dual amplified electrochemical immunosensor for highly sensitive detection of *Pantoea stewartii* sbsp. *stewartii*. *ACS. Appl. mater. inter.* **6**, 21178–21183 (2014).



15. Joungh, C. K. *et al.* A nanoporous membrane-based impedimetric immunosensor for label-free detection of pathogenic bacteria in whole milk. *Biosens. Bioelectron.* **44**, 210–215 (2013).
16. Li, N. *et al.* A label-free electrochemical immunosensor based on Au@Pd/Ag yolk-bimetallic shell nanoparticles and amination graphene for detection of nuclear matrix protein 22. *Sensor. Actuat. B: Chem.* **202**, 67–73 (2014).
17. Jiang, X., Chai, Y., Wang, H. & Yuan, R. Electrochemiluminescence of luminol enhanced by the synergetic catalysis of hemin and silver nanoparticles for sensitive protein detection. *Biosens. Bioelectron.* **54**, 20–26 (2014).
18. Liu, L. *et al.* A Label-Free Voltammetric Immunoassay Based on 3D-Structured rGO-MWCNT-Pd for Detection of Human Immunoglobulin G. *Sensor. Actuat. B: Chem.* **211**, 170–176 (2015).
19. Ansari, A. A., Kaushik, A., Solanki, P. R. & Malhotra, B. D. Sol-gel derived nanoporous cerium oxide film for application to cholesterol biosensor. *Electrochem. Commun.* **10**, 1246–1249 (2008).
20. Zarei, H., Ghourchian, H., Eskandari, K. & Zeinali, M. Magnetic nanocomposite of anti-human IgG/COOH–multiwalled carbon nanotubes/Fe<sub>3</sub>O<sub>4</sub> as a platform for electrochemical immunoassay. *Anal. Biochem.* **421**, 446–453 (2012).
21. Li, S., Liu, F., Ge, S., Yu, J. & Yan, M. Application of nanoporous Pd as catalytically promoted nanolabels for ultrasensitive electrochemiluminescence immunosensor based on Ag/graphene nanocomposite. *Sensor. Actuat. B: Chem.* **210**, 460–467 (2015).
22. Lu, J. *et al.* Ultrasensitive electrochemical immunosensor based on Au nanoparticles dotted carbon nanotube–graphene composite and functionalized mesoporous materials. *Biosens. Bioelectron.* **33**, 29–35 (2012).
23. Kamat, P. V. Graphene-based nanoarchitectures. Anchoring semiconductor and metal nanoparticles on a two-dimensional carbon support. *J. Phys. Chem. Lett.* **1**, 520–527 (2009).
24. Yang, X. *et al.* Superparamagnetic graphene oxide–Fe<sub>3</sub>O<sub>4</sub> nanoparticles hybrid for controlled targeted drug carriers. *J. Mater. Chem.* **19**, 2710–2714 (2009).
25. Guo, S., Li, D., Zhang, L., Li, J. & Wang, E. Monodisperse mesoporous superparamagnetic single-crystal magnetite nanoparticles for drug delivery. *Biomaterials* **30**, 1881–1889 (2009).
26. Wei, H. & Wang, E. Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles as peroxidase mimetics and their applications in H<sub>2</sub>O<sub>2</sub> and glucose detection. *Anal. Chem.* **80**, 2250–2254 (2008).
27. Liu, L. *et al.* A label-free amperometric immunosensor for detection of zearalenone based on trimetallic Au-core/AgPt-shell nanorattles and mesoporous carbon. *Anal. Chim. Acta* **847**, 29–36 (2014).
28. Meng, F., Yan, X., Liu, J., Gu, J. & Zou, Z. Nanoporous gold as non-enzymatic sensor for hydrogen peroxide. *Electrochim. Acta* **56**, 4657–4662 (2011).
29. Kurowska, E., Brzózka, A., Jarosz, M., Sulka, G. D. & Jaskuła, M. Silver nanowire array sensor for sensitive and rapid detection of H<sub>2</sub>O<sub>2</sub>. *Electrochim. Acta* **104**, 439–447 (2013).
30. Cao, Y. *et al.* Ultrasensitive luminol electrochemiluminescence for protein detection based on *in situ* generated hydrogen peroxide as coreactant with glucose oxidase anchored AuNPs@MWCNTs labeling. *Biosens. Bioelectron.* **31**, 305–309 (2012).
31. Kong, F. Y., Xu, B. Y., Xu, J. J. & Chen, H. Y. Simultaneous electrochemical immunoassay using CdS/DNA and PbS/DNA nanochains as labels. *Biosens. Bioelectron.* **39**, 177–182 (2013).
32. Zhao, W. *et al.* A novel nonenzymatic hydrogen peroxide sensor based on multi-wall carbon nanotube/silver nanoparticle nanohybrids modified gold electrode. *Talanta* **80**, 1029–1033 (2009).
33. Qin, X. *et al.* One-step synthesis of Ag nanoparticles-decorated reduced graphene oxide and their application for H<sub>2</sub>O<sub>2</sub> detection. *Electrochim. Acta* **79**, 46–51 (2012).
34. Cao, X., Wang, N., Jia, S., Guo, L. & Li, K. Bimetallic AuPt nanochains: Synthesis and their application in electrochemical immunosensor for the detection of carcinoembryonic antigen. *Biosens. Bioelectron.* **39**, 226–230 (2013).
35. Gao, Z. D., Guan, F. F., Li, C. Y., Liu, H. F. & Song, Y. Y. Signal-amplified platform for electrochemical immunosensor based on TiO<sub>2</sub> nanotube arrays using a HRP tagged antibody–Au nanoparticles as probe. *Biosens. Bioelectron.* **41**, 771–775 (2013).
36. Wang, Y. *et al.* Ultrasensitive sandwich-type electrochemical immunosensor based on a novel signal amplification strategy using highly loaded toluidine blue/gold nanoparticles decorated KIT-6/carboxymethyl chitosan/ionic liquids as signal labels. *Biosens. Bioelectron.* **61**, 618–624 (2014).
37. Wang, X. *et al.* An ultrasensitive electrochemical immunosensor based on the catalytic activity of MoS<sub>2</sub>-Au composite using Ag nanospheres as labels. *Sensor. Actuat. B: Chem.* **206**, 30–36 (2015).
38. Xu, T., Jia, X., Chen, X. & Ma, Z. Simultaneous electrochemical detection of multiple tumor markers using metal ions tagged immunocolloidal gold. *Biosens. Bioelectron.* **56**, 174–179 (2014).
39. Guo, X. *et al.* Synthesis of amino functionalized magnetic graphenes composite material and its application to remove Cr(VI), Pb(II), Hg(II), Cd(II) and Ni(II) from contaminated water. *J. Hazard. Mater.* **278**, 211–220 (2014).
40. Marcano, D. C. *et al.* Improved synthesis of graphene oxide. *ACS nano* **4**, 4806–4814 (2010).
41. Yamaura, M. *et al.* Preparation and characterization of (3-aminopropyl) triethoxysilane-coated magnetite nanoparticles. *J. Magn. Mater.* **279**, 210–217 (2004).
42. Frens, G. Controlled nucleation for the regulation of the particle size in monodisperse gold suspensions. *Nature* **241**, 20–22 (1973).
43. Liu, F. K., Tsai, M. H., Hsu, Y. C. & Chu, T. C. Analytical separation of Au/Ag core/shell nanoparticles by capillary electrophoresis. *J. Chromatogr. A* **1133**, 340–346 (2006).
44. Zhang, S. *et al.* Metal ions-based immunosensor for simultaneous determination of estradiol and diethylstilbestrol. *Biosens. Bioelectron.* **52**, 225–231 (2014).
45. Kurowska, E., Brzózka, A., Jarosz, M., Sulka, G. & Jaskuła, M. Silver nanowire array sensor for sensitive and rapid detection of H<sub>2</sub>O<sub>2</sub>. *Electrochim. Acta* **104**, 439–447 (2013).
46. Lu, W. *et al.* A novel label-free amperometric immunosensor for carcinoembryonic antigen based on Ag nanoparticle decorated infinite coordination polymer fibres. *Biosens. Bioelectron.* **57**, 219–225 (2014).
47. Bonora, P. L., Deflorian, F. & Fedrizzi, L. Electrochemical impedance spectroscopy as a tool for investigating underpaint corrosion. *Electrochim. Acta* **41**, 1073–1082 (1996).
48. Han, J. *et al.* Electrochemical immunoassay for thyroxine detection using cascade catalysis as signal amplified enhancer and multifunctionalized magnetic graphene sphere as signal tag. *Anal. Chim. Acta* **790**, 24–30 (2013).
49. Katz, E. & Willner, I. Switching of directions of bioelectrocatalytic currents and photocurrents at electrode surfaces by using hydrophobic magnetic nanoparticles. *Angew. Chem.* **117**, 4869–4872 (2005).
50. Wang, Y. *et al.* Novel signal amplification strategy for ultrasensitive sandwich-type electrochemical immunosensor employing Pd–Fe<sub>3</sub>O<sub>4</sub>-GS as the matrix and SiO<sub>2</sub> as the label. *Biosens. Bioelectron.* **74**, 59–65 (2015).
51. Jiang, L. *et al.* A sandwich-type electrochemical immunosensor based on multiple signal amplification for  $\alpha$ -fetoprotein labeled by platinum hybrid multiwalled carbon nanotubes adhered copper oxide. *Electrochim. Acta* **160**, 7–14 (2015).
52. Wang, Y. *et al.* Ultrasensitive sandwich-type electrochemical immunosensor based on dual signal amplification strategy using multifunctional graphene nanocomposites as labels for quantitative detection of tissue polypeptide antigen. *Sensor. Actuat. B: Chem.* **214**, 124–131 (2015).

## Acknowledgements

This study was financially supported by the Natural Science Foundation of China (Nos 21375047, 21575050, 21405095 and 21575079), All of the authors express their deep thanks.

### Author Contributions

FL., Y.L. and Q.W. conceived and designed the experiments. FL. and Y.L. performed the experiments, analyzed the data and wrote the first draft of the manuscript. Y.D., L.J., P.W., Q.L., H.L. and Q.W. contributed substantially to revisions.

### Additional Information

**Supplementary information** accompanies this paper at <http://www.nature.com/srep>

**Competing financial interests:** The authors declare no competing financial interests.

**How to cite this article:** Li, F. *et al.* An ultrasensitive label-free electrochemical immunosensor based on signal amplification strategy of multifunctional magnetic graphene loaded with cadmium ions. *Sci. Rep.* **6**, 21281; doi: 10.1038/srep21281 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>