

Estrogen Receptor α Regulates Dlx3-Mediated Osteoblast Differentiation

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Estrogen receptor α (ER- α), which is involved in bone metabolism and breast cancer, has been shown to have transcriptional targets. Dlx3 is essential for the skeletal development and plays an important role in osteoblast differentiation. Various osteogenic stimulators and transcription factors can induce the protein expression of Dlx3. However, the regulatory function of ER- α in the Dlx3 mediated osteogenic process remains unknown. Therefore, we investigated the regulation of Dlx3 and found that ER- α is a positive regulator of Dlx3 transcription in BMP2-induced osteoblast differentiation. We also found that ER- α interacts with Dlx3 and increases its transcriptional activity and DNA binding affinity. Furthermore, we demonstrated that the regulation of Dlx3 activity by ER- α is independent of the ligand (estradiol) binding domain. These results indicate that Dlx3 is a novel target of ER- α , and that ER- α regulates the osteoblast differentiation through modulation of Dlx3 expression and/or interaction with Dlx3.

INTRODUCTION

The Distal-less (Dlx) family is composed of six members, Dlx1-Dlx6, and they have a highly conserved homeobox domain related to that of Distal-less (Dll) in *Drosophila* (Panganiban and Rubenstein, 2002). Dlx2 is essential for craniofacial development and mesenchymal condensation (Robinson and Mahon, 1994). Dlx5 and Dlx6 play important roles in axial, craniofacial, and appendicular skeletal development (Depew et al., 2002; Robledo et al., 2002). The role of Dlx3 is demonstrated in the development of osteoblasts and chondrocytes from osteogenic cells (Choi et al., 2008; 2009; Ghoul-Mazgar et al., 2005; Hassan et al., 2004).

Estrogen signaling in bone occurs mainly through two estrogen receptors, estrogen receptor alpha (ER- α) and ER- β . Although both receptors are important in bone physiology and development, a point mutation in ER- α caused unfused growth

plates and osteoporosis (Smith et al., 1994). Since then, the role of ER- α on bone metabolism has become a major focus of research (Khosla, 2013; Price et al., 2011; Vico and Vanacker, 2010).

Recent studies suggest that ER- α in osteoblast associates with and regulates the activity of transcription factors or signal transduction pathways (Kousteni et al., 2007; McCarthy et al., 2003; 2011). Hormone-activated ER- α interacts directly with transcription factor Runx2 and increases its transcriptional activity in osteoblast differentiation (McCarthy et al., 2003), whereas ER- α might indirectly modulate the expression and activity of other osteogenic transcription factors as regulators through the regulation of specific components with selected kinase (Kousteni et al., 2007). However, the molecular mechanisms by which ER- α regulates osteoblast differentiation remain to be elucidated.

In this study, we examined the functional role of ER- α in Dlx3 modulation. We found that ER- α increases BMP-2-induced osteoblast differentiation. In addition, ER- α interacts with Dlx3 and increases the transcriptional activity of Dlx3 in a ligand-independent manner. We also found that ER- α increases the DNA binding affinity of Dlx3. Collectively, our results indicate that ER- α -induced Dlx3 activity may induce the osteoblast differentiation in a ligand independent manner.

MATERIALS AND METHODS

Materials

Estradiol (E2) was purchased from Sigma-Aldrich. Antibodies against the following epitopes were used: Myc (9E10) and HA (12CA5) from Roche Applied Science (Germany), GFP (G1544) from Santa Cruz Biotechnology (USA), and α -Tubulin (B-5-1-2) from Sigma-Aldrich (USA).

Cell culture

C2C12 mouse myoblast cells and HEK293 cells were maintained in DMEM containing 10% heat-inactivated FBS and antibiotic-antimycotic solution at 37°C in a humidified atmosphere of 5% CO₂. DMEM, FBS, and antibiotic-antimycotic solution were purchased from Life Technologies (USA).

Plasmids and transfection

The N-terminal epitope-tagged human Dlx3 expression plasmids were constructed in a CMV promoter-derived mammalian expression vector (pCS4). The HA-tagged full length-ER- α and ER- α - Δ LBD were kindly provided by Dr. Keesook Lee (Chonnam National University, Korea). The polyethyleneimine (PEI)

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(Polysciences Inc., USA)-mediated method was used for transient transfection. Total amounts of transfected plasmids in each condition were equalized with an empty vector.

Immunoblotting (IB) and immunoprecipitation (IP)

HEK293 cells were co-transfected with the indicated plasmids and then lysed with an ice cold lysis buffer previously described (Lee et al., 2013). After centrifugation, supernatants containing 30 μ g of total protein were subjected to SDS-PAGE. The separated proteins were transferred to a polyvinylidene fluoride membrane for 120 min at 70 V. After exposure for 1 h to TBST containing 0.05% Tween 20 and 5% dried nonfat milk, the membranes were incubated with the appropriate primary antibodies for overnight at 4°C, and then with HRP-conjugated secondary antibodies for 1 h at room temperature. The proteins were visualized using ECL reagent (Millipore Corporation). For IP, centrifuged lysate supernatants were precipitated using appropriate antibodies and Protein A-conjugated Sepharose beads. The immunoprecipitated proteins were resolved using SDS-PAGE and visualized using IB.

Semi-quantitative RT-PCR

Total RNA was extracted from C2C12 cells with the use of RNAso Plus (Takara Bio Inc., Japan) according to the manufacturer's instructions. Random-primed cDNAs were synthesized from 1 μ g of total RNA by using the GoScript Reverse Transcription System (Promega, USA). The following conditions were used for PCR: initial denaturation at 94°C for 5 min, 25-30 cycles of denaturation at 94°C for 1 min, annealing at a temperature optimized for each primer pair for 1 min, extension at 72°C for 1 min, and a final extension at 72°C for 10 min. The primer sequences used are as follows: mouse Bone sialoprotein (BSP) forward 5'-ACA CTT ACC GAG CTT ATG AGG-3' and reverse 5'-TTG CGC AGT TAG CAA TAG CAC-3' (Jeong et al., 2010); mouse Collagen type I α 1 (COL1A1) forward 5'-TCT CCA CTC TTC TAG GTT CCT-3' and reverse 5'-TTG GGT CAT TTC CAC ATG C-3' (Choi et al., 2012; Dao et al.,

2011); mouse Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) forward 5'-ACC ACA GTC CAT GCC ATC AC-3' and reverse 5'-TCC ACC ACC CTG TTG CTG TA-3' (Dao et al., 2011).

Luciferase reporter assay

Luciferase reporter assays were conducted using alkaline phosphatase (ALP) promoter reporter, BSP promoter reporter, Dlx-response element (DRE) reporter, and estrogen-response element (ERE) reporter plasmids. HEK293 cells were transfected with the indicated plasmids and reporter plasmids, and pCMV- β -gal for 36 h. All the luciferase assays were performed using a Luciferase Reporter Assay Kit (Promega). Data were normalized by corresponding β -galactosidase activity to compensate for transfection efficiency.

ALP staining

C2C12 cells were transfected and then stimulated with BMP2 (10 ng/ml) for 3 days. Cells were fixed with 4% paraformaldehyde (Sigma-Aldrich) for 10 min and stained with 300 μ g/ml BCIP/NBT solution (Sigma-Aldrich) for 15 min at room temperature. ALP-positive cells were stained a blue/purple color.

DNA affinity precipitation assay (DAPA)

The DNA binding assay was performed by mixing whole cell lysates with a biotinylated oligomers containing three tandem repeats of the Dlx3-responsive element, and the DNA-protein complexes were precipitated using streptavidin-coated beads. The binding proteins were eluted using loading buffer and resolved using 12% SDS-PAGE followed by IB. The following biotinylated DRE sequences were used: DRE forward 5'-biotin-GCG ATA ATT GCG GCG ATA ATT GCG-3' and reverse 5'-biotin-CGC AAT TAT CGC CGC AAT TAT CGC CGC AAT TAT CGC-3' (Li et al., 2014).

Statistical analysis

The data are representative of three independent experiments.

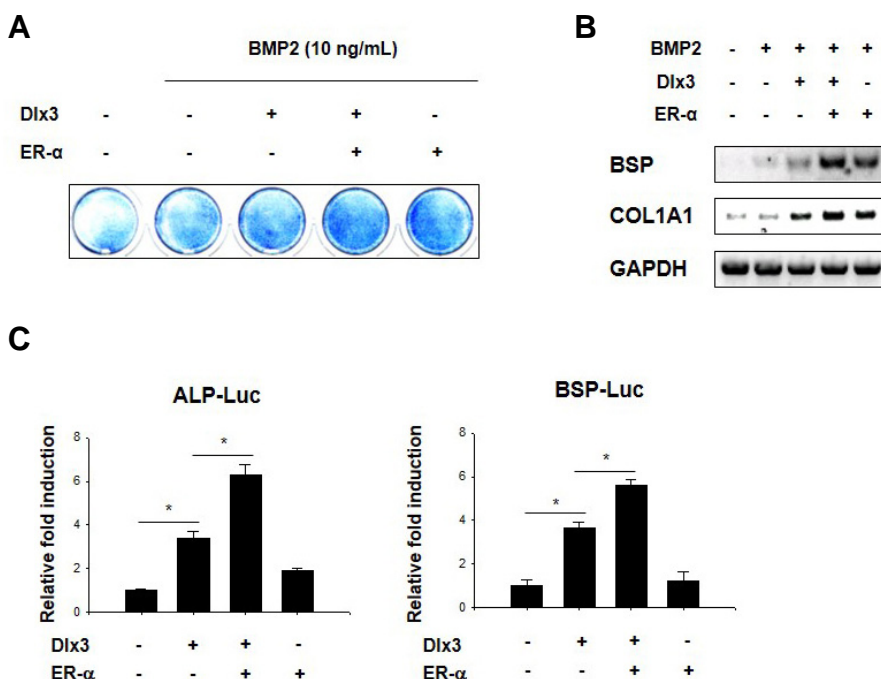


Fig. 1. ER- α induces Dlx3-enhanced osteoblast differentiation. (A) C2C12 cells were transfected with an empty vector or the indicated combinations of Dlx3 and ER- α , stimulated with BMP2, and then stained for ALP activity. (B) C2C12 cells were transfected with an empty vector or the indicated combinations of Dlx3 and ER- α , stimulated with BMP2, and then harvested for RT-PCR. GAPDH is the loading control. (C) C2C12 cells were transfected with pCMV- β -gal, ALP-Luc, or BSP-Luc reporter vector along with the indicated combinations of Dlx3 and ER- α . Luciferase activity was then measured after 36 h. Data are expressed as means \pm SEM of at least three experiments. * p < 0.05 by Student's t -test.

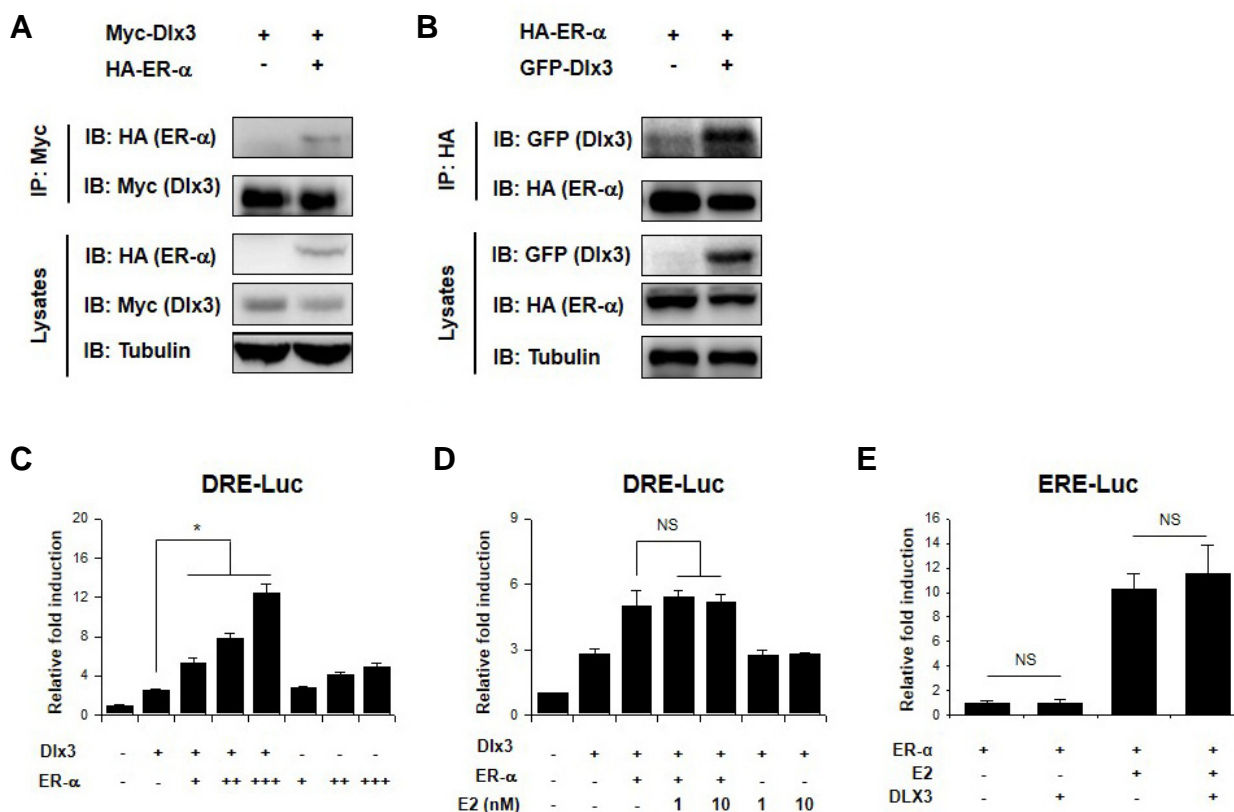


Fig. 2. ER- α physically associates with and induces the transcriptional activity of Dlx3. HEK293 cells were transfected with Myc-tagged Dlx3 and HA-tagged ER- α (A) or with HA-tagged ER- α and GFP-tagged Dlx3 (B), and the cell lysates were subjected to immunoprecipitation using an anti-Myc antibody and Protein A-Sepharose beads. The immunoprecipitated proteins were then analyzed using immunoblotting. (C) HEK293 cells were transfected with pCMV- β -gal and a DRE-Luc reporter vector along with the indicated combinations of Dlx3 and ER- α . Luciferase activity was then measured after 36 h. Data are expressed as means \pm SEM of at least three experiments. * p < 0.05 by Student's t -test. (D and E) HEK293 cells were transfected with pCMV- β -gal, DRE-Luc (D), or ERE-Luc (E) reporter vectors along with the indicated combinations of Dlx3 and ER- α . Transfected cells were treated for 12 h with estradiol (E2) and luciferase activity was then measured. Data are expressed as means \pm SEM of at least three experiments. NS, not significant by Student's t -test.

The Results are expressed as means \pm standard error of the mean. Analyses were performed with Student's t -test. P -values of less than 0.05 were considered significant.

RESULTS

ER- α affects Dlx3 in BMP2-stimulated osteoblast differentiation

First, we examined the effect of ER- α on Dlx3-driven induction of osteoblast differentiation. C2C12 cells were transfected with Dlx3 and ER- α and then treated with BMP2. ALP staining showed that ER- α increases Dlx3-induced activation of osteoblast differentiation (Fig. 1A). Next, we examined whether ER- α affects the expression of osteogenic marker genes. ER- α increased the expression of BSP, and COL1A1 and co-transfection of Dlx3 and ER- α further increased their expression (Fig. 1B). We also examined the effects of ER- α on the transcriptional activities of Dlx3-induced activation of ALP and BSP promoter reporter genes. C2C12 cells were transfected with ALP-Luc and BSP-Luc along with combinations of Dlx3 and ER- α . Dlx3 alone increased the ALP-Luc and BSP-Luc activity, which was further enhanced by ER- α (Fig. 1C). Collec-

tively, these results indicate that ER- α plays a positive role in osteoblast differentiation and reinforces the Dlx3-induced activation of osteoblast differentiation.

ER- α physically associates with and induces the transcriptional activity of Dlx3

To determine whether a physical interaction between Dlx3 and ER- α affected BMP2-induced osteoblast differentiation, HEK293 cells were transfected with Myc-Dlx3 and HA-ER- α , followed by IP and IB. Additionally, HEK293 cells were transfected with HA-ER- α and GFP-Dlx3 and then analyzed using IP and IB. These experiments showed that Dlx3 was bound to ER- α (Figs. 2A and 2B). To determine whether ER- α can regulate the transcriptional activity of Dlx3, HEK293 cells were transfected with a DRE-Luc reporter gene with different combinations of Dlx3 and ER- α . Dlx3 by itself increased reporter activity compared to control group, and co-transfection with ER- α further increased reporter activity in a dose-dependent manner (Fig. 2C). To determine whether its regulation is related to the ligand, we treated the cells with E2. However, the transcriptional activity of Dlx3 was not influenced by E2 (Fig. 2D). Since the nature of the interactions between Dlx3 and ER- α predicted

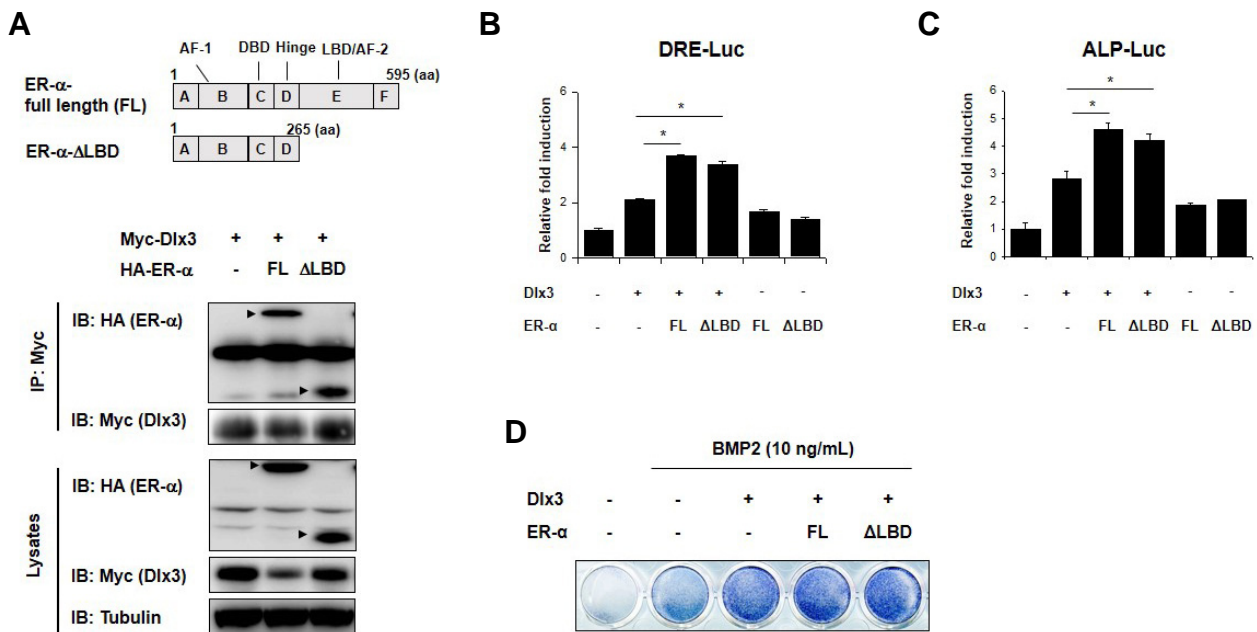


Fig. 3. ER- α - Δ LBD interacts with and affects Dlx3 in BMP2-stimulated osteoblast differentiation. (A) A schematic of the ER- α structure. FL, full length; AF-1, activation function 1; DBD, DNA binding domain; LBD, ligand binding domain; AF-2, activation function 2. HEK293 cells were transfected with the indicated combinations of plasmids and 48 h later cell lysates were harvested and analyzed using immunoprecipitation and immunoblotting. (B) HEK293 cells were transfected with pCMV- β -gal, and DRE-Luc reporter vector along with the indicated combinations of Dlx3, ER- α -FL or ER- α - Δ LBD. Luciferase activity was measured after 36 h. Data are expressed as mean \pm SEM of at least three experiments. * p < 0.05 by Student's t -test. (C) C2C12 cells were transfected with pCMV- β -gal, ALP-Luc reporter vector along with the indicated combinations of Dlx3, ER- α -FL or ER- α - Δ LBD. Luciferase activity was measured after 36 h. Data are expressed as mean \pm SEM of at least three experiments. * p < 0.05 by Student's t -test. (D) C2C12 cells were transfected with an empty vector or the indicated combinations of Dlx3 and ER- α -FL or ER- α - Δ LBD, stimulated with BMP2, and then stained for ALP activity.

that they might also regulate each other, next we examined whether Dlx3 can affect ER-transcriptional activity. HEK293 cells were transfected an ERE-Luc with different combinations of Dlx3 and ER- α and then treated with E2 for 12 h. We found that Dlx3 had no effect on ER- α -dependent reporter gene activity (Fig. 2E). These results suggest that ER- α interacts with Dlx3 and increases its transcriptional activity.

ER- α - Δ LBD interacts with and affects Dlx3 in BMP2-stimulated osteoblast differentiation

To determine whether the regulation of Dlx3 by ER- α is independent of the ligand (E2), we fragmented ER- α based on the characteristics in the previous report (Kumar et al., 1987). In particular, we used ER- α -FL and the A/B/C/D fragment (ER- α - Δ LBD). HEK293 cells were transfected with Myc-Dlx3 and HA-ER- α -FL or HA-ER- α - Δ LBD and analyzed using IP and IB. We found that Dlx3 was bound to both HA-ER- α -FL and HA-ER- α - Δ LBD (Fig. 3A). Next, we examined whether HA-ER- α - Δ LBD also can modulate the transcriptional activity of Dlx3. HEK293 cells were transfected with a DRE-Luc reporter gene along with combinations of Dlx3 and ER- α -FL or ER- α - Δ LBD. We found that ER- α - Δ LBD also increased the transcriptional activity of Dlx3 (Fig. 3B). Then we examined whether HA-ER- α - Δ LBD affects BMP2-induced osteoblast differentiation. We found that HA-ER- α - Δ LBD enhanced ALP-Luc activity (Fig. 3C) and ALP staining (Fig. 3D). These results indicate that the regulation of Dlx3 by ER- α is independent of the ligand (E2), and that un-

liganded ER- α interacts with Dlx3 and enhances BMP2-induced osteoblast differentiation.

ER- α increases DNA affinity binding of Dlx3 in a ligand-independent manner

As ER- α interacts with Dlx3, we examined whether ER- α has an effect on Dlx3 binding to DRE in experiments using a DAPA assay, in which short biotinylated DNA probes containing the DRE sequences were hybridized with nuclear proteins obtained from HEK293 cells transfected with combinations of Dlx3 and ER- α -FL or ER- α - Δ LBD. We found that both ER- α -FL and ER- α - Δ LBD induced efficient Dlx3 association with the DRE motif (Figs. 4A and 4B). These results imply that ER- α increases the affinity of Dlx3 for the DRE sequence in a ligand-independent manner.

DISCUSSION

Estrogen is an important hormone for bone development, growth, and metabolism. Although many studies have focused on estrogen as an inhibitor of bone resorption, others suggest additional effects on bone formation. Estrogen may act in part on osteoblasts to inhibit apoptosis (Gohel et al., 1999; Kousteni et al., 2001; 2002; Zhou et al., 2001) or to regulate AP-1-related transcription factors by specific kinase-dependent pathways (Chen et al., 1996; Kelly and Levin, 2001; Kousteni et al., 2002; Zhou et al., 2001). Interactions have been reported between

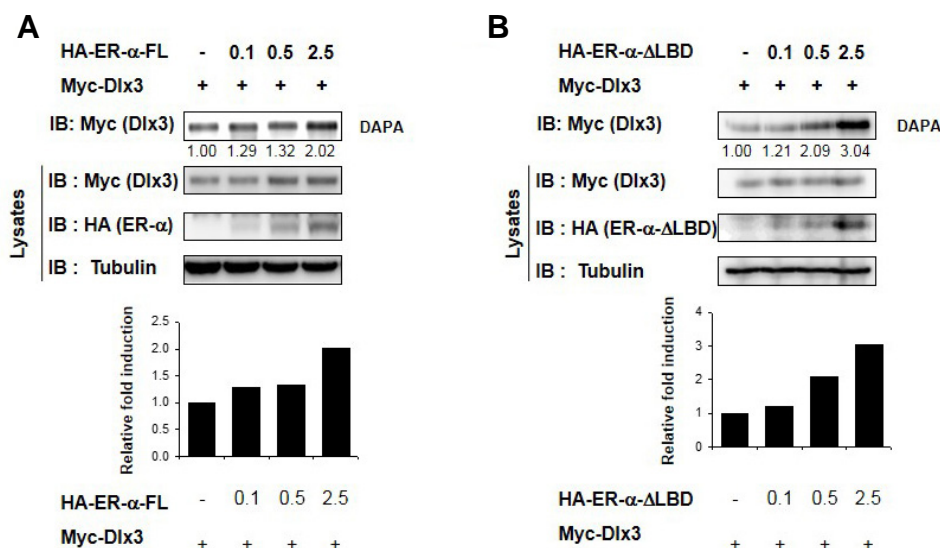


Fig. 4. ER- α increases DNA binding affinity of Dlx3 in a ligand-independent manner. (A, B) HEK293 cells were transfected with an empty vector or indicated combinations of Dlx3 and ER- α -FL (A) or ER- α - Δ LBD (B). The cell lysates were incubated with biotin-labeled oligonucleotide Dlx3-responsive elements, and the DNA-bound proteins were analyzed by immunoblotting using an anti-Dlx3 antibody.

ER- α and several other transcription factors, including retinoid receptors (Lee et al., 1998), forkhead transcription factors (Schoor et al., 2001), Smads (Matsuda et al., 2001; Yamamoto et al., 2002), Stat5 (Bjornstrom and Sjoberg, 2002), GATA4 (Guemes et al., 2014), AP-1 (Lambertini et al., 2008), C/EBP δ (Chang et al., 2005), and Runx2 (Lambertini et al., 2007; McCarthy et al., 2003). However, the regulatory mechanism of estrogen and the ER in osteoblasts remains to be elucidated. In this study, we examined the effect of estrogen and ER- α on Dlx3 (Hassan et al., 2004; Robinson and Mahon, 1994).

Recent studies show that unliganded ER- α regulates various cell signaling and the expressions of target genes. Unliganded ER- α is required for the osteogenic response to mechanical loading in a ligand-independent manner (Windahl et al., 2013). Sirtuin 1 represses estrogen-signaling, ligand-independent ER- α -mediated transcription, and cell proliferation in estrogen-responsive breast cells (Moore and Fallor, 2013). Unliganded ER- α promotes PC12 cell survival during serum starvation (Ferriere et al., 2013). Unliganded ER stimulates BSP gene expression (Takai et al., 2014). In our study, we found that ER- α overexpression induced Dlx3-transcriptional activities (Fig. 2C), whereas the activities were not influenced by E2 (Fig. 2D). These results suggest that unliganded ER- α is required for the regulation of Dlx3.

The internal structure of the ER contains six distinct regions which are labeled A through F as shown in Fig. 3A. The N-

terminal region (A/B) contains activation function 1 (AF-1) which is responsible for constitutive ligand-independent activation of the receptor (Weigel, 1996). Previous study revealed that A/B region of the ER especially on serine/threonine residue can be phosphorylated by mitogen-activated protein kinase (Kato et al., 1995). The C domain is a DNA-binding domain (DBD) that binds to specific sequences of DNA called hormone response elements (HRE), or in this case, EREs (Schwabe et al., 1993). The LBD located in the E region of ERs attributes to ligand-dependent activation of the receptor, in turn it induces the transcriptional activity by the action of AF-2 located at the C-terminal portion of the E region (Wurtz et al., 1996; 1998). As we found that unliganded ER- α regulates Dlx3 function (Fig. 2), we hypothesized that AF-1 might be sufficient for regulation of Dlx3 by ER- α but not AF-2. As expected, we found that ER- α - Δ LBD interacts with Dlx3 and regulates its transcriptional activity (Figs. 3A and 3B) by increasing its DNA binding affinity (Fig. 4B). The interaction between ER- α and Runx2 or Stat5 maps to the ER- α DBD (Bjornstrom and Sjoberg, 2002; McCarthy et al., 2003). We expect that ER- α and Dlx3 interact through the DBD of ER- α , although we cannot rule out other sites that may be involved in this interaction. Further study of this interaction will enhance our understanding of the regulatory mechanisms of ER- α function and the regulation of Dlx3 in osteoblast differentiation. Therefore, determination of the interaction site(s) to understand the significance of such regulation between Dlx3

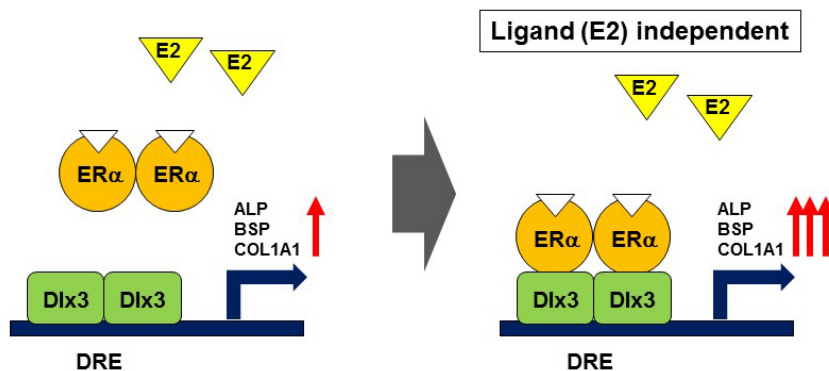


Fig. 5. The proposed model for regulation of Dlx3 by ER- α . Dlx3 binds to DRE site on osteogenic genes such as ALP, BSP, COL1A1 and subsequently increases their activities. ER- α interacts and consequently reinforces Dlx3-mediated regulation in a ligand independent manner.

and ER- α is needed.

In conclusion, as shown in Fig. 5, we have found that unliganded ER- α physically interacts with Dlx3 *in vivo* and functions as a coactivator in regulating Dlx3-mediated signaling in osteoblast differentiation. These findings may provide insight into the understanding of ER- α function in osteoblast differentiation.

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