

# Hairy and Groucho mediate the action of juvenile hormone receptor Methoprene-tolerant in gene repression

Tusar T. Saha<sup>a,b</sup>, Sang Woon Shin<sup>a</sup>, Wei Dou<sup>a,c</sup>, Sourav Roy<sup>a,b</sup>, Bo Zhao<sup>a,b</sup>, Yuan Hou<sup>d</sup>, Xue-Li Wang<sup>d</sup>, Zhen Zou<sup>d</sup>, Thomas Girke<sup>b,e</sup>, and Alexander S. Raikhel<sup>a,b,1</sup>

<sup>a</sup>Department of Entomology, University of California, Riverside, CA 92521; <sup>b</sup>The Institute for Integrative Genome Biology, University of California, Riverside, CA 92521; <sup>c</sup>College of Plant Protection, Southwest University, Chongqing 400715, People's Republic of China; <sup>d</sup>State Key Laboratory of Integrated Management of Pest Insects and Rodents, Institute of Zoology, Chinese Academy of Sciences, Beijing 100101, People's Republic of China; and <sup>e</sup>Department of Botany and Plant Sciences, University of California, Riverside, CA 92521

Contributed by Alexander S. Raikhel, December 10, 2015 (sent for review August 10, 2015; reviewed by David N. Arnosti and Jinsong Zhu)

The arthropod-specific juvenile hormone (JH) controls numerous essential functions. Its involvement in gene activation is known to be mediated by the transcription factor Methoprene-tolerant (Met), which turns on JH-controlled genes by directly binding to E-box-like motifs in their regulatory regions. However, it remains unclear how JH represses genes. We used the *Aedes aegypti* female mosquito, in which JH is necessary for reproductive maturation, to show that a repressor, Hairy, is required for the gene-repressive action of JH and Met. The RNA interference (RNAi) screen for *Met* and *Hairy* in the *Aedes* female fat body revealed a large cohort of Met- and Hairy-corepressed genes. Analysis of selected genes from this cohort demonstrated that they are repressed by JH, but RNAi of either *Met* or *Hairy* renders JH ineffective in repressing these genes in an in vitro fat-body culture assay. Moreover, this JH action was prevented by the addition of the translational inhibitor cycloheximide (CHX) to the culture, indicating the existence of an indirect regulatory hierarchy. The lack of Hairy protein in the CHX-treated tissue was verified using immunoblot analysis, and the upstream regions of *Met*/Hairy-corepressed genes were shown to contain common binding motifs that interact with Hairy. *Groucho* (*gro*) RNAi silencing phenocopied the effect of *Hairy* RNAi knockdown, indicating that it is involved in the JH/Met/Hairy hierarchy. Finally, the requirement of Hairy and *Gro* for gene repression was confirmed in a cell transfection assay. Thus, our study has established that Hairy and its cofactor *Gro* mediate the repressive function of JH and Met.

juvenile hormone | receptor | hairy | groucho | repressor

Arthropod-specific sesquiterpenoids, juvenile hormones (JH), control numerous essential physiological functions in immature and adult insects, including development, reproduction, pheromone production, and cast differentiation in social insects (1). Methoprene-tolerant (Met), a member of the family of basic helix-loop-helix (bHLH)-Per-Arnt-Sim (PAS) transcription factors, has been recognized as the JH receptor (1). In *Drosophila melanogaster*, JH action is mediated by two receptors, encoded by the paralogous genes *Met* and *germ-cell-expressed* (*gce*) (1). However, only a single *Met* gene, an ortholog of *Drosophila gce*, exists in the genome of the yellow fever mosquito *Aedes aegypti*, which is at present the most important vector of dengue and chikungunya viruses (2).

JH plays a critical role in reproduction of many insects by synchronizing sexual maturation as well as controlling vitellogenesis and egg production (3–5). JH III coordinates development of reproductive organs in newly eclosed female mosquitoes during the posteclosion (PE) phase which lasts 3–5 d (4, 6, 7). Ovaries become capable of receptor-mediated endocytosis of yolk protein precursors (8). Their primary follicles increase twofold in length, oocytes acquire endocytic organelles, and the follicular epithelium goes through differentiation (6, 9, 10). Similarly, the fat body, a tissue that is analogous to the vertebrate liver, undergoes JH-regulated cellular remodeling, building up secretory machinery

for massive production of yolk protein precursors (11, 12). JH action is critical for a female mosquito in attaining responsiveness to 20-hydroxyecdysone (20E), an insect steroid hormone that is a major regulator of vitellogenic events in Dipteran insects. To accomplish this, JH induces the translation of the competence factor, nuclear orphan receptor betaFTZ-F1 (*fushi tarazu* binding factor 1) (13).

Microarray analysis has shown a high level of differential gene expression during the PE phase in the *A. aegypti* female fat body (14). The hierarchical clustering identified three major gene clusters: 1,843 early (EPE) genes maximally expressed at 6 h PE, 457 mid (MPE) genes at 24 h PE, and 1,815 late (LPE) genes at 66 h PE. The microarray screen with *Met* RNAi has demonstrated Met repression of EPE and MPE and activation of LPE genes. Bioinformatics analysis revealed the consensus for a 9-mer Met-binding motif—CACG<sup>C</sup><sub>T</sub>G<sup>A</sup><sub>G</sub><sup>T</sup><sub>A</sub>G. Met-binding motif variants were overrepresented in promoters of Met up-regulated (LPE) genes, but not in Met down-regulated (EPE/MPE) genes. Electrophoretic mobility shift assays (EMSAs), using a combination of mutational and anti-Met antibody supershift analyses, confirmed binding properties of the Met consensus motif variants (14). Hence, it appears that the Met action on some activated genes is direct, whereas the repressive action of JH/Met requires additional factors, the identity of which has been unknown.

## Significance

Juvenile hormone is unique to arthropods and controls numerous essential functions in these organisms. Its receptor, Methoprene-tolerant, turns on genes by directly binding to E-box-like motifs in their regulatory regions. However, how juvenile hormone represses genes has remained unclear. Here, we used the *Aedes aegypti* female mosquito, in which juvenile hormone is necessary for reproductive maturation, to show that the repressor Hairy is required for the gene-repressive action of juvenile hormone and Methoprene-tolerant. Moreover, we demonstrate that a corepressor, Groucho, supports this Hairy function. Hence, Methoprene-tolerant recruits the evolutionarily conserved Hairy/Groucho molecular system to mediate the juvenile hormone gene-repressive function.

Author contributions: T.T.S., S.W.S., Z.Z., and A.S.R. designed research; T.T.S., S.W.S., W.D., B.Z., Y.H., X.-L.W., and Z.Z. performed research; T.T.S., S.W.S., S.R., Y.H., X.-L.W., Z.Z., and T.G. contributed new reagents/analytic tools; T.T.S., S.W.S., W.D., S.R., Z.Z., T.G., and A.S.R. analyzed data; and T.T.S. and A.S.R. wrote the paper.

Reviewers: D.N.A., Michigan State University; and J.Z., Virginia Tech.

The authors declare no conflict of interest.

See Commentary on page 1474.

<sup>1</sup>To whom correspondence should be addressed. Email: alexander.raikhel@ucr.edu.

This article contains supporting information online at [www.pnas.org/lookup/suppl/doi:10.1073/pnas.1523838113/-DCSupplemental](http://www.pnas.org/lookup/suppl/doi:10.1073/pnas.1523838113/-DCSupplemental).

Considering that this question is of general importance for understanding the action of JH at the molecular level in insects, we used *A. aegypti* female mosquitoes to investigate this question. In these mosquitoes, JH and Met strongly activate transcriptional repressors Krüppel homolog-1 (Kr-h1) and Hairy (15, 16). The former has been implicated in mediating JH and Met signaling during development and metamorphosis (1), whereas the role of Hairy has remained unclear.

*D. melanogaster hairy* (*DmHairy*) is a pair-rule gene involved in early patterning of the embryo (17, 18). It plays a key role in the segmentation gene hierarchy by repressing a pair-rule gene, *fushi tarazu* (*ftz*) (19). As a long-range repressor, *DmHairy* has the ability to inhibit the action of activators located more than 1 kb away from the gene transcription start site (20). It can induce a widespread histone deacetylation and inhibit the recruitment of basal transcription machinery without triggering chromatin compaction in the repression of *ftz* gene (21).

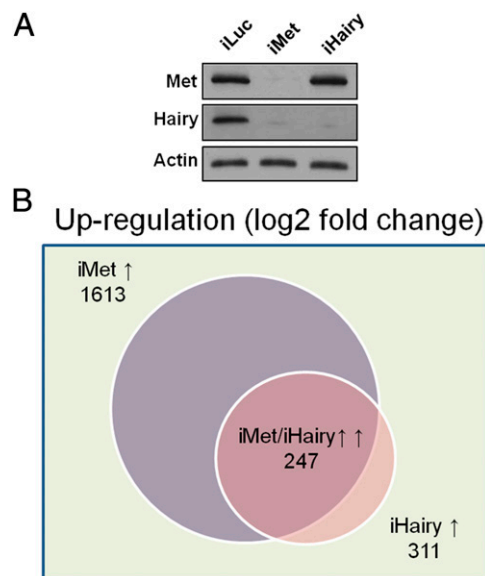
In this work, we have shown that Hairy is an integral part of the JH/Met hierarchy, down-regulating genes in adult insects. Moreover, a corepressor Groucho (Gro) is required for this action.

## Results

**Identification of Genes Corepressed by Met and Hairy.** To investigate the role of Hairy as a possible intermediate factor downstream of Met, we performed RNAi-based transcriptomic screens for *A. aegypti* *Met* (*AaMet*) and *Hairy* (*AaHairy*). In preparation for this study, we investigated the endogenous levels of Met and Hairy in the *A. aegypti* female fat body at the protein levels by immunoblotting with anti-Met and anti-Hairy antibodies, respectively. This analysis showed that Met was constitutively present throughout the PE period (Fig. S1A). In contrast, the Hairy level raised significantly during the PE period; it was undetectable at 6 h PE, but was intensified between 24- and 72 h PE (Fig. S1A). The latter suggests that Hairy was regulated during the PE period. Indeed, the immunoblots revealed the absence of Hairy in fat-body samples from mosquitoes with RNAi-mediated depletions of *Met* (iMet), supporting the hypothesis that it is a factor downstream of Met (Fig. 1A). The level of Met was not altered in fat-body samples from mosquitoes with RNAi-mediated depletions of *Hairy* (iHairy), ruling out the presence of any feedback control loop (Fig. 1A). The Met and Hairy protein levels were negligible in fat-body samples with their respective RNAi depletions, indicating high efficiency of RNAi (Fig. 1A).

The ovarian maturation of newly eclosed female mosquito is governed by JH and is manifested by the increase in length of the primary ovarian follicles to about 100–120  $\mu\text{m}$  in length by 72 h PE (6), and this increase in follicular length is mediated by Met, providing a direct evidence for its involvement in reproductive maturation in adult female mosquito (14, 15). To investigate a possible effect of Hairy on this stage of reproduction, we evaluated the length of iHairy ovarian follicle compared with those of iMet and controls, iLuc and uninjected UGAL mosquitoes at 72 h PE. The average length of iMet primary follicles was 55  $\mu\text{m}$ , whereas that of iLuc was 109  $\mu\text{m}$  and that of wild-type UGAL was 112  $\mu\text{m}$  (Fig. S1B). A more modest but significant decrease in follicular length was observed in ovaries of iHairy mosquitoes with an average length of 90  $\mu\text{m}$  (Fig. S1B). This experiment suggests importance of Hairy for PE reproductive events.

Illumina RNA-seq technology was used to generate the transcriptional profiles of genes affected by RNAi-mediated depletions of *Met* (iMet) and *Hairy* (iHairy) in the fat body of *A. aegypti* female mosquitoes. RNAi for *luciferase* (iLuc) served as a control. Three biological replicates of RNA-seq libraries were constructed for each RNAi treatment. Each RNA-seq library contained between 62 and 241 million reads (read length, 101 nucleotides for first and second replicates; and 51 nucleotides for third) (Table S1). The reads were aligned with Bowtie2 against *A. aegypti* tran-



**Fig. 1.** Illumina RNA-seq analysis of Met and Hairy RNAi-depleted *A. aegypti* mosquito fat body. (A) WBs showing the presence of Met and Hairy proteins in iLuc, iMet, and iHairy samples. WBs also provided proof of the efficiency of dsRNA-mediated knockdowns. Anti- $\beta$ -actin antibody was used as a loading control. (B) Venn diagram of Met and Hairy RNAi fat-body transcriptome analysis showing that 79% of iHairy up-regulated genes are also up-regulated by iMet. iMET $\uparrow$ , more than twofold up-regulation by dsRNA depletion of Met (AKA, Met-repressed); iHairy $\uparrow$ , more than twofold up-regulation by dsRNA depletion of Hairy (AKA, Hairy-repressed); iMet/iHairy $\uparrow\uparrow$ , more than twofold up-regulation by dsRNA depletion of Met and Hairy (AKA, Met/Hairy-repressed). All comparisons were made with the control iLuc fat-body transcriptome.

script sequence database (*AaeGL1.3* geneset; Vectorbase). The resulting count table was transformed into fragments per kilobase pair of transcript per million fragments mapped (FPKM) values. Relative transcript abundance of iLuc, iMet, and iHairy, defined as the sum of FPKM values of the three treatment samples, was sorted from highest to lowest.

From the first set of RNA-seq experiments, 10,000 transcripts with the highest abundance were selected for further analysis (Dataset S1). The differentially expressed transcripts were defined by a twofold increase or decrease of FPKM values. Among these 10,000 abundantly present transcripts, a total of 2,151 transcripts was identified as having differential expression between iMet and control iLuc mosquitoes, with 1,613 transcripts being activated in iMet (Fig. 1B and Dataset S2). A total of 455 transcripts showed a more than twofold differential expression between iHairy and iLuc treatments, with 311 being up-regulated (Fig. 1B and Dataset S2). Significantly, 80% of iHairy-activated genes were also activated by iMet. In total, 247 common genes were activated by RNAi knockdowns of either *AaMet* or *AaHairy*. This large overlap between the two transcriptomes suggests that Met and Hairy are involved in the same hierarchy mediating the JH-repressive action (Fig. 1B). However, a relatively small number of Met-repressed genes corepressed by Hairy suggests the existence of an additional Hairy-independent mechanism for JH/Met gene repression. The reproducibility of results was verified by comparing genes coactivated by iMet and iHairy in three biological replicates. This comparison revealed that 185 genes were coactivated by iMet and iHairy in all three replicates (Fig. S2 and Dataset S3).

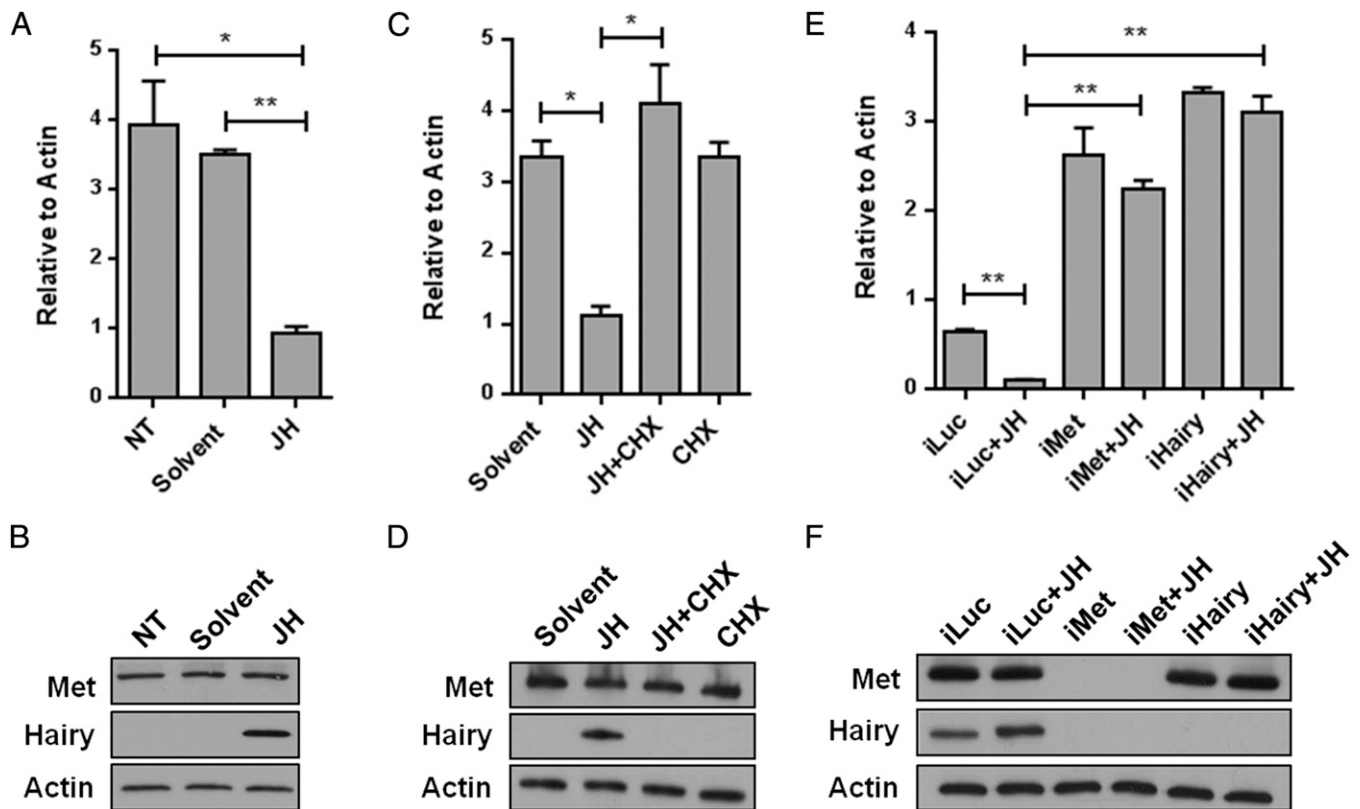
**Ontology Analysis of Genes Corepressed by Met and Hairy.** The analysis of orthologous groups (OGs) based on adjusted nonsupervised orthologous groups (NOG) (Table S2 and Dataset S4) revealed

that Met- and/or Hairy-repressed transcripts were categorized mostly into two broad groups, one of Cellular Process and Signaling and another of Metabolism (Fig. S34). The significantly over-represented OGs ( $P < 0.01$  in a hypergeometric distribution) in Hairy-repressed transcripts were as follows: three in the Cellular Process and Signaling category—[V] defense mechanisms, [W] extracellular structure, and [O] posttranslational modification, protein turnover, chaperones; and three in the Metabolism category—[G] carbohydrate transport and metabolism, [E] amino acid transport and metabolism, and [Q] secondary metabolite biosynthesis, transport, and metabolism (Fig. S3B). All of these OGs were also categorized as Met-repressed. Of these six overrepresented OGs, all but one—[E] Amino acid transport and metabolism—were also over-represented among *AaMet/AaHairy*-coregulated transcripts, suggesting the existence of the Met–Hairy hierarchy in gene repression (Fig. S3B). Further, Met- and Hairy-coregulated genes were manually annotated, during which 54 isoform-specific and gene-duplicated transcripts were removed, resulting in 193-curated Met/Hairy-repressed genes (Dataset S5).

**Hairy Acts as an Intermediate Component of the JH/Met Gene-Repressive Hierarchy.** To test the hypothesis that Hairy is an intermediate component of the JH/Met gene-repressive hierarchy, we selected

several highly up-regulated genes from 193 genes activated by both *iMet* and *iHairy* and cross-examined them using quantitative RT-PCR (qPCR). Compared with *iLuc* controls, the genes encoding AAEL000044 (Ornithine decarboxylase), AAEL005004 (Unknown), and AAEL002655 (Matrix metalloproteinase) were up-regulated in both *iMet* and *iHairy* samples (Fig. S44). Next, we analyzed the effect of JH on these Met–Hairy–coregulated genes. JH was topically applied on female mosquitoes 3 h PE, when the JH titer is very low, and their fat bodies were collected 8 h later, as described previously (14). qPCR showed that treatment with JH dramatically repressed transcript levels of all of the tested genes (Fig. 2A and Fig. S4B). Immunoblots revealed the presence of a Hairy protein band only in fat-body samples from the JH-treated mosquitoes but not in controls treated with a solvent (acetone). Met protein, however, was detected in both JH- and solvent-treated samples (Fig. 2B).

To establish the requirement of intermediate factor(s) and the necessity of de novo translation in the JH-dependent repression, we used an approach using the protein synthesis inhibitor cycloheximide (CHX). Fat bodies of newly eclosed female mosquitoes (6 h PE) were incubated in the JH-containing culture medium in the presence or absence of CHX. Transcript levels of tested genes were high after incubation with medium containing solvent (acetone), but were significantly less in the presence of



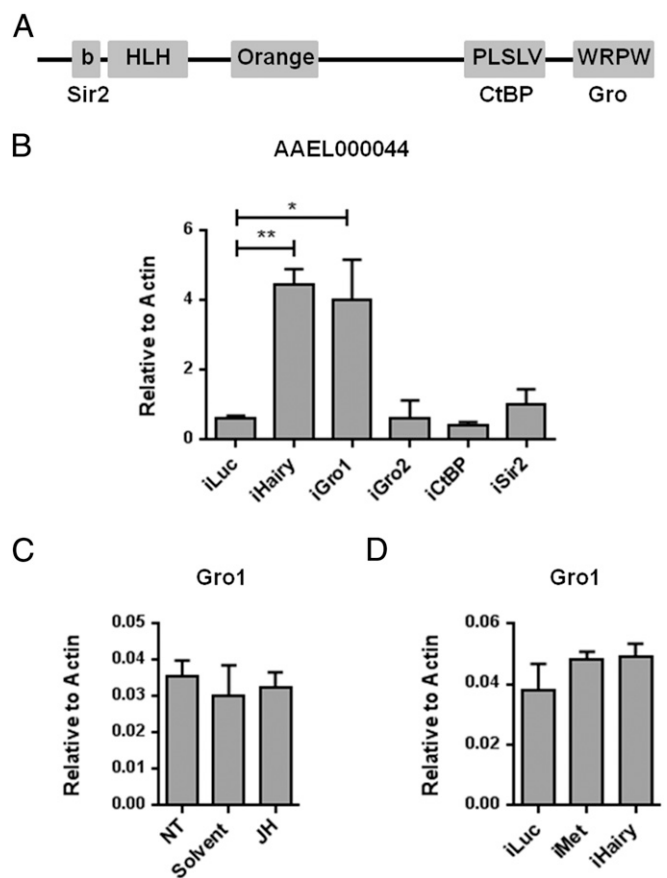
**Fig. 2.** Hairy acts as an intermediate component of JH/Met gene repression. (A) Effect of JH on the expression of the *AAEL000044* gene. JH was applied topically onto the abdomen of newly eclosed female mosquitoes at 6 h PE, and the effect was determined by means of qPCR. NT, no treatment. (B) WB showing the levels of endogenous Met and Hairy proteins in the fat bodies under the same conditions as that of A. (C) *In vitro* fat-body culture experiments showing the effect of cycloheximide (CHX) on the JH-mediated induction of the expression of the *AAEL000044* gene. The effect of JH on the *AAEL000044* gene expression was compromised by the addition of CHX to the JH-containing tissue culture medium, as measured by qPCR. Minimal medium without JH and CHX (solvent) and CHX only (CHX) were used as controls. (D) WB showing the effect of JH and CHX on Met and Hairy protein levels under the same conditions as that of C. A clear induction of Hairy but not Met protein by JH is observed. CHX treatment results in the loss of Hairy but not Met protein. (E) Hairy is required for the JH-mediated repression of the *AAEL000044* gene. Luc, Met, and Hairy dsRNA-treated fat bodies were cultured in medium with or without JH. The *AAEL000044* gene was repressed by JH treatment in *iLuc* (control), but this repression was compromised in *iMet*+JH samples. The knockdown of Hairy phenocopied the effects of *iMet* samples, indicating that Hairy is a downstream component of the JH/Met-mediated gene repression hierarchy. (F) WB for Met and Hairy protein levels in the same sample sets described in E. All experiments were performed in triplicate, with similar results. Error bars represent  $\pm$  SD. \* $P < 0.05$ ; \*\* $P < 0.01$ . Additional genes are shown in Fig. S3.

JH. Addition of CHX to the culture medium rendered these genes unresponsive to the repressive action of JH (Fig. 2C and Fig. S4C). *AaKr-h1* (AAEL002390), which is directly controlled by Met (1, 16), served as control in this experiment. CHX was ineffective in preventing the activating effect of JH on this gene (Fig. S4C). Western blot (WB) analysis showed that *Hairy* was reduced to an undetectable level as a result of CHX addition to the culture media (Fig. 2D). However, *Met* was not affected by this treatment. This result clearly established the role of intermediate factors downstream of *Met* in JH-mediated repression of the tested genes. The absence of *Hairy* in CHX-treated fat bodies strongly supports our hypothesis that JH/*Met* gene repression functions through *Hairy* de novo translation for a subset of *Met*/*Hairy*-corepressed genes.

To address specifically whether *Hairy* is involved as an intermediate factor in the JH/*Met* gene repression hierarchy, we carried out experiments combining RNAi silencing and in vitro fat-body culture, as previously described (14). Fat bodies from mosquitoes with RNAi-depleted *Luc*, *Met*, and *Hairy* were incubated in the presence or absence of JH. JH decreased transcript levels of the analyzed genes in *Luc* RNAi mosquito fat bodies, whereas RNAi depletion of *Met* rendered these genes insensitive to JH (Fig. 2E and Fig. S4D). *Hairy* RNAi silencing had a similar effect to that of *Met* RNAi on the expression of the tested genes (Fig. 2E and Fig. S4D). WB analysis using *Met* and *Hairy* antibodies showed that *Hairy* was absent after *Met* RNAi treatment, regardless of whether JH was present (Fig. 2F). This experiment establishes *Hairy* as the factor downstream of *Met* in the JH/*Met* repression hierarchy.

**A Corepressor Gro1 Is Involved in the *Met*/*Hairy*-Mediated Gene Repression.** To act as a repressor, *Hairy* requires recruitment of corepressors (22–25). *Hairy* carries binding domains for three corepressors—silent information regulator 2 (*Sir2*), C terminus binding protein (*CtBP*), and *Gro* (Fig. 3A). There are two homologs of *Drosophila gro* in the *A. aegypti* genome—*gro1* (AAEL010433) and *gro2* (AAEL005009). To investigate the possible involvement of *CtBP* (AAEL005400), *Sir2* (AAEL005816), and the two *Gro* in the *Met*/*Hairy*-mediated gene repression, we used their respective RNAi knockdowns and monitored the response of the *Met*/*Hairy*-regulated gene *AAEL000044*. RNAi silencing of *gro1*, but not *gro2*, *CtBP* or *Sir2*, phenocopied the effect of *Hairy* RNAi depletion on the expression of the *AAEL000044* gene, indicating a possible corepressor role for *Gro1* (Fig. 3B). Next, we tested the dependency of *Gro1* expression on the JH/*Met*/*Hairy* cascade. The *gro1* gene expression was insensitive to the JH topical application (Fig. 3C). Moreover, RNAi knockdowns of *Met* and *Hairy* had no effect on *gro1* transcript levels (Fig. 3D). Taken together, these experiments suggest a corepressor role for *Aedes Gro1* in the *Hairy*-mediated JH/*Met* gene repression hierarchy.

The tetra-amino acid WRPW (Trp-Arg-Pro-Trp) sequence that is located at the *Hairy* C terminus serves as a protein–protein interaction domain with *Gro* (26). To test physical interaction between *Hairy* and *Gro1* from *A. aegypti*, we used coimmunoprecipitation (co-IP) experiments in a cell culture system. Flag-tagged, full-length *Hairy* (*Hairy*-Flag) and V5-tagged, full-length *Gro1* (*Gro1*-V5) were cloned and expressed in *D. melanogaster* S2 cells. After immunoprecipitation (IP) with anti-Flag antibody, *Gro1*-V5 was detected as a co-IP product in WBs with anti-V5 antibody only when both fusion proteins were coexpressed in the cell culture system (Fig. S5). The protein–protein interaction between *A. aegypti* *Hairy* and *Gro1* was further confirmed by additional co-IP experiments (IP:V5, WB:Flag; IP:*Hairy*, WB:V5; IP:V5, WB:Flag), where a clear band could only be detected when both proteins were present (Fig. S5). To test the domain specificity of this binding, a truncated *Hairy* fusion protein without the *Gro1* interacting C terminus WRPW tail (*Hairy*ΔWRPW-Flag) was cloned and transfected to *D. melanogaster* S2 cells in the presence or ab-



**Fig. 3.** *AaGro* is a corepressor of *AaHairy* in the JH/*Met* gene repression hierarchy. (A) Schematic diagram showing the domain composition of *Hairy* protein and the potential binding sites for various cofactors (based on refs. 23–26). *Hairy* carries binding domains for three corepressors—silent information regulator 2 (*Sir2*), C-terminal binding protein (*CtBP*), and *Gro* (*Gro*). (B) Involvement of putative *Hairy* corepressors was tested by their respective dsRNA-mediated knockdowns. *Gro1* RNAi silencing phenocopied the effect of the *Hairy* RNAi depletion on the expression of the *AAEL000044* gene. (C and D) The *Gro1* expression is independent of the JH/*Met*/*Hairy* cascade. *Gro1* is not regulated by JH as revealed by in vivo hormonal treatment assays (C). Depletions of either *Met* or *Hairy* do not affect the expression of *Gro1* (D). Expression analyses were performed using qPCR. All experiments were done in triplicate with similar results. Error bars represent  $\pm$  SD. \* $P < 0.05$ ; \*\* $P < 0.01$ .

sence of *Gro1*-V5. The loss of *Hairy* WRPW domain completely compromised the *Hairy*–*Gro1* interaction, as evidenced by the lack of a band in co-IP experiments (Fig. S5). Together, these experiments confirm the interaction between *A. aegypti* *Hairy* and *Gro1* in a domain-specific manner.

#### Identification of *Hairy*-Binding Motifs in *Met*/*Hairy*-Repressed Genes.

*Hairy* belongs to the *Hairy*/Enhancer of split/*Deadpan* (HES) subclass of repressor bHLH proteins (27). bHLH proteins bind to a canonical E-box motif (CANNTG) that occurs in two classes—class A with consensus sequence  $CA^G/CCTG$  and class B with consensus sequence  $CAC^G/ATG$  (28). *D. melanogaster* *Hairy* binds specifically to a canonical class B site CACGTG, but also interacts with non-canonical site CGCGTG, resulting in a *Hairy*-binding consensus sequence  $C^A/GCGTG$  (29, 30). Additionally, previous studies have shown that bHLH proteins interacting with CACGTG site can recognize and bind to an E-box variant with an adenine in place of guanine at the fourth position (28). Based on this information, we used EMSA to test the interaction of in vitro-translated *AaHairy* with these motifs.

The C<sup>A</sup>/<sub>G</sub>CGTG motifs along with their flanking sequences from Met/Hairy-regulated genes *AAEL013984* and *AAEL013703* interacted specifically with *AaHairy* (Fig. S6). Hairy formed a band with the CACGTG core and its flanks from the 5' upstream region of the *AAEL013984* gene (Fig. S6A). The binding was abolished after a 2-nt mutation in the E-box sequence to CAGCTG. Excess of the unlabeled specific probe, but not the nonspecific one, competed out the labeled probe resulting in the abolition of the band. A supershift was observed using anti-Hairy polyclonal antibodies, indicating the presence of Hairy in the complex (Fig. S6A). Hairy also formed a band interacting with the CACGTG (AAEL013703A) and CGCGTG (AAEL013703B) cores and their respective flanking sequences from the *AAEL013703* gene promoter (Fig. S6B). However, a 2-base mutation in the core CGCGTG to CGGCTG (AAEL013703Bm) abolished the binding, indicating specificity of the E-box-like motif (Fig. S6B).

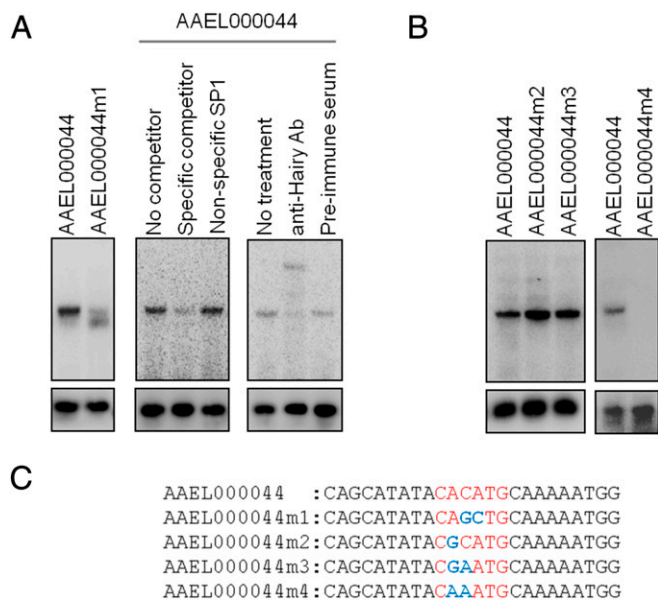
To test *AaHairy* binding to C<sup>A</sup>/<sub>G</sub>CATG motifs, we used oligonucleotide sequences from the promoter region of the Met/Hairy-repressed gene *AAEL000044* in EMSA. A strong binding was observed between the CACATG along with its flanking sequence and in vitro-translated *Aedes* Hairy protein (Fig. 4A and C). The mutation of the CACATG sequence of *AAEL000044* to CAGCTG (AAEL000044m1) almost completely abolished this binding, indicating the binding specificity and verifying the fact that *AaHairy* does not recognize class A E-box motif (C<sup>A</sup>/<sub>G</sub>CTG). Competition assay with unlabeled specific probe, but not with a nonspecific competitor SP1, resulted in elimination of the band, providing further evidence for specificity of this interaction (Fig. 4A). Addition of anti-Hairy antibody resulted in the supershift of the band, showing that Hairy is a component of the DNA-protein complex (Fig. 4A). Then, we tested the binding of the E-box-like motif CGCATG to *AaHairy* by mutating the *AAEL000044* (CACATG) to *AAEL000044m2* (CGCATG), which also resulted in a clear

binding between this motif-variant and *AaHairy* (Fig. 4B and C). Interestingly, a mutated core-binding motif CGAATG could also be recognized by Hairy. However, the CAAATG sequence was incapable of interaction with Hairy, as evidenced by the lack of DNA-protein complex in EMSA (Fig. 4B and C). Thus, we have identified previously unknown Hairy-interacting motifs, C<sup>A</sup>/<sub>G</sub>CATG and CGAATG, in addition to the canonical C<sup>A</sup>/<sub>G</sub>CGTG motif in *A. aegypti*. The detailed number of Hairy-interacting motifs found in upstream regions of Met/Hairy-repressed genes is provided in Dataset S6.

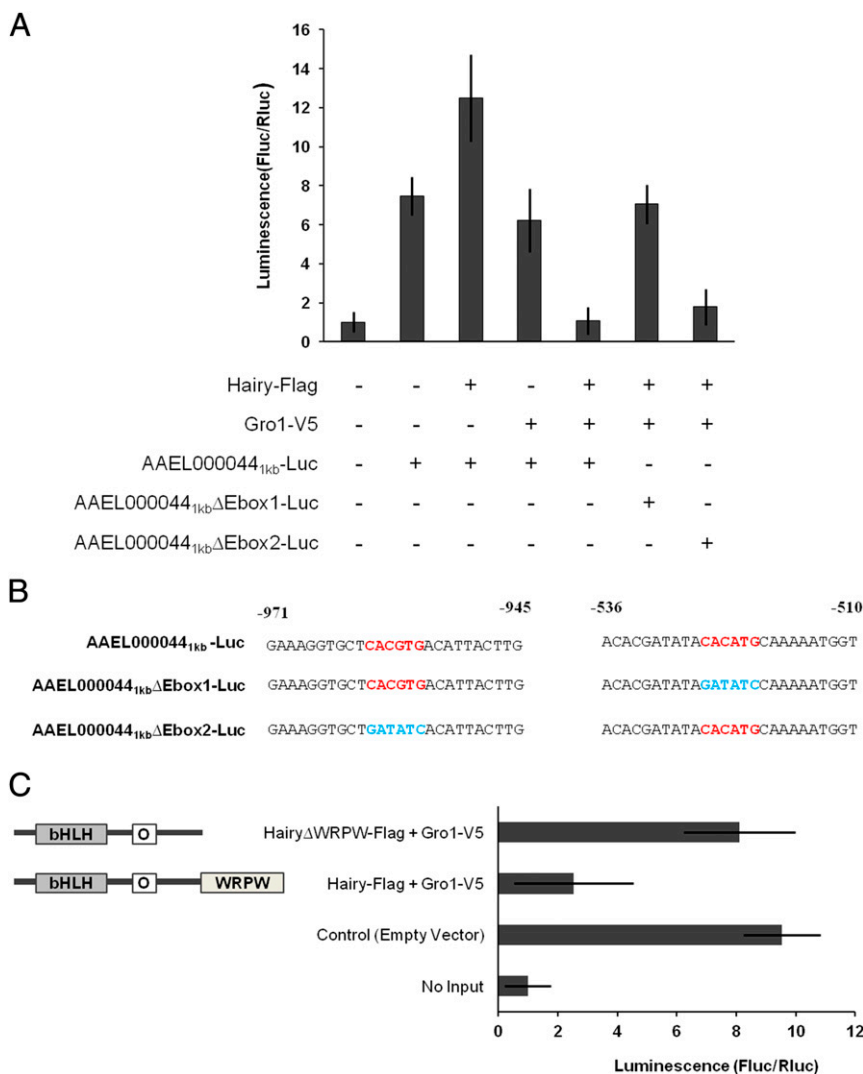
**Analysis of Gene Promoters Containing *AaHairy*-Interacting Motifs in the Cell Transfection Assay.** To test whether Hairy can transrepress gene expression, a 1-kb 5' upstream regulatory region of the Met/Hairy-repressed *AAEL000044* gene harboring the Hairy binding site was cloned into the luciferase reporter vector pGL3basic (AAEL000044<sub>1kb</sub>-Luc). A sixfold to eightfold basal induction in the reporter activity was observed when AAEL000044<sub>1kb</sub>-Luc was transfected into *D. melanogaster* S2 cells (Fig. 5A), rendering this experimental system feasible to address gene repression. Surprisingly, cotransfection of AAEL000044<sub>1kb</sub>-Luc and the Hairy-Flag expression vector into the cell culture system resulted in an induction of luciferase activity, which was elevated with increasing concentrations of the Hairy vector (Fig. S7A). This result suggested that Hairy by itself was not sufficient to repress the promoter activity and raised the possibility of involvement of additional cofactors. Therefore, the *AaGro1-V5* expression vector was cotransfected along with *AaHairy-Flag* into S2 cells. The dual-luciferase assay revealed a clear repression of the promoter activity in the presence of 200 ng of *AaHairy* and of *AaGro1* expression vectors (Fig. 5A). However, no decrease in luciferase activity by Hairy/Gro1 was observed after the Hairy binding E-box-like motif CACATG in *AAEL000044* promoter (AAEL000044<sub>1kb</sub>ΔEbox1-luc) was mutated, demonstrating the necessity of the DNA-protein interaction for this repression (Fig. 5A and B). Interestingly, mutation of E-box CACGTG keeping the E-box-like motif CACATG intact (AAEL000044<sub>1kb</sub>ΔEbox2-luc) did not alter the reporter activity when supplied with Hairy-Flag and Gro1-V5 (Fig. 5A and B). There could be several possible reasons for this observation including the distance of the "CACGTG" site from the core promoter and the lack of sufficient number of nucleotides on the 5' end of the motif. No change in the reporter activity was observed when only *AaGro1-V5* was transfected in the cell culture system, indicating that *AaGro1* by itself is incapable of gene repression (Fig. S7A). Furthermore, the gene repression was compromised when the expression vector with the truncated Hairy lacking a C-terminal Gro-binding WPRW domain was used (HairyΔWPRW-Flag) along with Gro1-V5 instead of the full-length Hairy (Fig. 5C). Taken together, these experiments prove that *AaHairy* and *AaGro1* are necessary for repression of luciferase activity by the 1-kb promoter of the *AAEL000044* gene harboring the Hairy-binding E-box-like motif CACATG.

Similarly, the 1-kb promoter region of the *AAEL002655* gene harboring the Hairy-binding motif variant "CGAATG" was also cloned into a reporter vector and transfected to S2 cells along with increasing concentrations of Hairy-Flag, Gro1-V5, or both. A clear repression could only be observed when both factors were present (Fig. S7B). The repression of luciferase activity was lost when CGAATG was mutated to CGGCTG, indicating this motif's capacity to act as a functional Hairy-binding element leading to gene repression (Fig. S7B). The expression of the tagged proteins *AaHairy-Flag*, *AaGro1-V5*, and *AaHairyΔWPRW-Flag* in the cell culture system was confirmed by WBs using anti-Flag and anti-V5 antibodies (Fig. S7C and D).

**The Met/Hairy/Gro Repression Hierarchy Is Conserved in *Anopheles gambiae* Mosquitoes.** We investigated whether the Met/Hairy/Gro signaling cascade was conserved. To test this, we used another major



**Fig. 4.** Characterization of Hairy-interacting motifs in *A. aegypti* JH/Met-repressed genes. (A) EMSAs showing the binding of in vitro-translated Hairy with E-box-like motif "CACATG" and its flanks from the promoter region of the Met/Hairy-repressed gene *AAEL000044*. The specificity of the binding was confirmed by competition with unlabeled specific probe; the presence of Hairy in the DNA-protein complex was verified by a supershift with anti-*AaHairy* polyclonal antibody. (B) The binding capacity of in vitro-translated Hairy to variants of E-box-like motif was tested by mutating the CACATG core. Hairy binds to "CGCATG" and CGAATG, but not to "CAAATG" sites. (C) The motifs along with corresponding flanking sequences used for the EMSA assays.

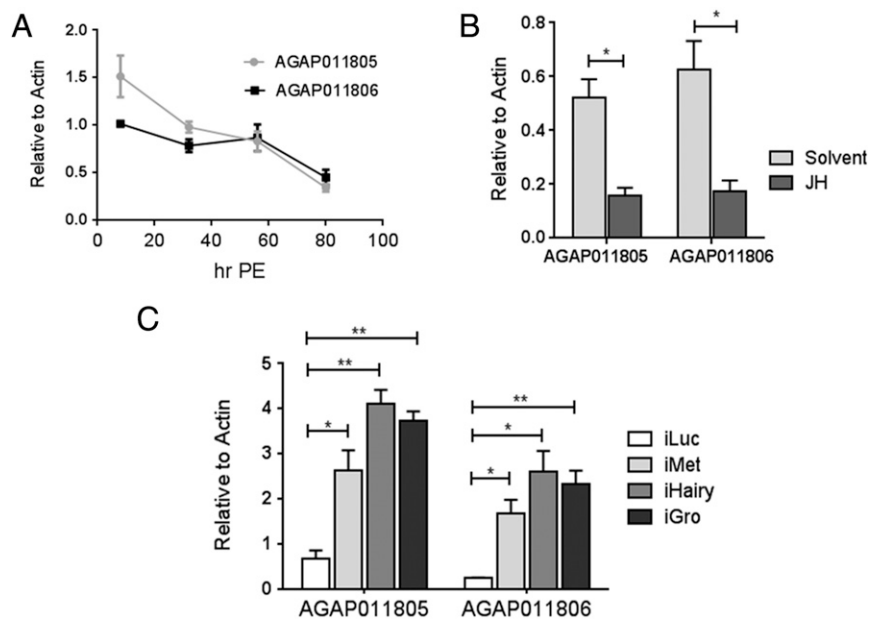


**Fig. 5.** Gro1 is required for the Hairy-mediated gene repression from the *AEEL000044* promoter. (A) Luciferase reporter assays after cotransfection of expression vectors Hairy-Flag and/or Gro1-V5 along with the desired reporter constructs. Treatments with no input DNA and empty expression vector served as controls. (B) The putative Hairy binding sites along with flanking regions harbored within the 1-kb region of the *AEEL000044* promoter and the mutated sequences for experiments in A are indicated. (C) Luciferase reporter assay with truncated Hairy lacking the C-terminal Gro binding domain. The experimental conditions are similar to above. A clear loss of repression from the 1-kb *AEEL000044* promoter was observed when HairyΔWRPW-Flag and Gro1-V5 were cotransfected into the cell culture system. Empty expression vector pAc5.1 was added to normalize the final concentration of DNA. In the graph, readings from no input DNA are represented as 1, with corresponding adjustments for other sample readings. Luminescence represents Firefly to *Renilla* luciferase readings. Data represent means  $\pm$  SD ( $n = 3$ ).

vector mosquito, *A. gambiae*, which transmits malaria pathogens. First, we characterized whether Hairy is a downstream factor of the JH/Met pathway in *A. gambiae*. We examined temporal expression patterns of *AgHairy*, *AgMet*, and *AgGro* during the PE development of female *A. gambiae* mosquitoes. We found that expression patterns of these factors were similar to those of *A. aegypti*; the *AgHairy* transcript showed a gradual increase in expression throughout the PE phase, reaching a peak at 60 h PE, whereas transcript levels of *AgMet* and *AgGro* remained constant during this phase (Fig. S8A). JH induced expression of *AgHairy* but had no effect on *AgMet* or *AgGro* (Fig. S8B). RNAi-mediated *AgMet* knockdown resulted in a dramatic decrease in the abundance of *AgHairy* mRNA (Fig. S8C). *AgGro* showed no change in expression in either iMet or iHairy mosquitoes (Fig. S8D). These results reveal that *AgHairy* is a downstream factor of the JH/Met pathway.

Next, we probed whether this *AgMet/AgHairy* cascade was involved in the repression of JH-regulated genes. *A. gambiae*

orthologs of the gene encoding *A. aegypti* *Ornithine decarboxylase* (*AEEL000044*) was used to address this question. There are two paralogs of this gene in the *A. gambiae* genome (*AGAP0011805* and *AGAP0011806*), both of which were assayed in this study. Transcripts of both these genes showed a similar pattern of expression, being highly up-regulated after adult eclosion and gradually decreasing to basal levels by late PE (Fig. 6A). Expression of both genes was down-regulated by the ectopic JH application in a manner similar to that of their *A. aegypti* ortholog (Fig. 6B). Thus, these duplicated genes were good candidates to test the existence of the Met/Hairy-Gro gene repression hierarchy in *A. gambiae*. Both *AGAP0011805* and *AGAP0011806* were transcriptionally up-regulated in *A. gambiae* iMet mosquitoes (Fig. 6C). RNAi-mediated knockdown of either *AgHairy* or *AgGro* phenocopied this induction, demonstrating conservation of the role of Hairy and Gro in mediating the JH/Met signaling in gene repression during reproductive development of adult female mosquitoes (Fig. 6C).



**Fig. 6.** JH/Met/Hairy-Gro signaling pathway regulates *Anopheles* ortholog of *Aedes* AAEL000044 gene. (A) Expression profiles of AGAP011805 and AGAP011806 during PE development, as measured using qPCR. (B) Hormonal application experiments showing that both AGAP011805 and AGAP011806 genes are repressed by JH. (C) The effect of dsRNA-mediated depletion of AgMet or AgHairy or AgGro on transcript levels of AGAP011805 and AGAP011806 compared with iLuc controls. All expression analyses were done using qPCR. All experiments were done in triplicate with similar results. Error bars represent  $\pm$  SD. \* $P < 0.05$ ; \*\* $P < 0.01$ .

## Discussion

In this study, we found that Hairy and its corepressor Gro mediate the action of JH and Met in gene repression. The initial evidence of Hairy involvement originated from the RNAi screen for *Met* and *Hairy* showing that 80% of Hairy-repressed genes were also found in the *Met*-repressed transcriptome. Additional support came from the immunoblot analysis using anti-*AaMet* and anti-*AaHairy* antibodies that revealed the absence of Hairy in iMet-treated fat bodies, suggesting that Hairy is a factor downstream of *Met*. Using topical application of JH onto female mosquitoes, we demonstrated that this hormone repressed selected genes from the gene cohort, which were corepressed by *Met* and Hairy. Similar to *AaMet* RNAi, *AaHairy* RNAi silencing rendered this JH action ineffective in the in vitro fat-body culture assay, once again showing that Hairy mediates JH action along with *Met*. A classic approach using CHX, described by Ashburner et al. (31) for the ecdysone gene regulatory hierarchy, established the requirement of an intermediate factor in the JH-dependent repression in female mosquitoes. JH repression of tested genes was prevented by CHX in the fat-body culture, and Hairy protein was reduced to an undetectable level as a result of the CHX treatment. This further substantiates our hypothesis that Hairy serves as a downstream factor in the JH/Met gene repression hierarchy.

As mentioned above, *D. melanogaster* Hairy binds to a canonical E-box class B site CACGTG but also interacts with a noncanonical site CGCGTG, resulting in a Hairy-binding consensus sequence  $C^A/G$ CGTG (29, 30). Using EMSA, we have identified different Hairy-interacting motifs,  $C^A/G$ CATG and CGAATG, in addition to the canonical  $C^A/G$ CGTG motif, in *A. aegypti*. Functionality of the newly identified Hairy-interacting motifs was confirmed in the cell transfection assay.

Hairy recruits corepressors essential for the down-regulation of target genes during *D. melanogaster* embryo and larval development. The best-studied Hairy corepressor is Gro, which directly interacts with Hairy through its conserved C-terminal tetrapeptide, WPRW domain (22, 26). Other than Gro, alternative Hairy-related proteins can also interact with another cofactor, CtBP,

through its PLSLV domain situated immediately upstream of the WPRW Gro binding domain (23). Studies in *D. melanogaster* indicate that CtBP and Gro regulate separate sets of Hairy-mediated genes affecting different pathways of transcriptional repression (32, 33). Here, we provide evidence that implicates *AaGro1* as a corepressor in the *Met*/Hairy-mediated gene hierarchy, required for Hairy to function. We established this Gro1 role using RNAi knockdowns for Sir2, CtBP, and Gro1 and Gro2, and monitoring a response of the *Met*/Hairy-regulated gene AAEL000044. Additionally, the cell transfection assay demonstrated that Gro1 is required for the Hairy repressive action. Moreover, we demonstrated by means of co-IP and cell transfection assays that *AaHairy* interacts with Gro1 via the conserved C terminus WPRW domain. During *D. melanogaster* development, Gro is broadly present and it is the availability of Hairy that dictates the target gene repression (34). When Hairy is expressed, it associates with Gro resulting in the repression of a target gene. We found that the *AaHairy* level increased by the end of the female mosquito PE phase under the control of JH and *Met*, whereas *AaGro* is constitutively present, suggesting existence of the similar Hairy/Gro regulatory system in *A. aegypti*.

In addition to its role as a pair-rule gene that is involved in early patterning of the *D. melanogaster* embryo (17–19), Hairy also regulates other developmental processes: sex determination (22), peripheral nervous system development during larval development (22, 29, 30), and morphogenetic furrow progression in the developing eye (35). It has likewise been implicated in metabolic suppression of hypoxia tolerance in *D. melanogaster* (36). This Hairy–Gro repression molecular axis is conserved in vertebrates, in which Gro homologs, known as Transducin-like enhancers (TLEs), physically interact with the Hairy homolog HES (37).

In this work, we have identified another role of Hairy as a downstream factor of *Met* in the JH gene repression hierarchy in *A. aegypti* mosquitoes. We also uncovered that this role is conserved in *A. gambiae* mosquitoes. The *A. gambiae* orthologs of the AAEL000044 gene are repressed by JH/AgMet indirectly through AgHairy. Furthermore, Gro is involved as a Hairy corepressor in

this hierarchy in *A. gambiae*. Thus, our findings indicate that Met employs an evolutionarily conserved Hairy–Gro molecular system to mediate the JH action in gene repression. Future work should establish how widely this system occurs in insects of other orders.

## Experimental Procedures

**Experimental Animals.** *A. aegypti* and *A. gambiae* mosquito larvae were cultured at 27 °C in water supplemented with a mixture of yeast and rat chow (1:1 ratio). Adult mosquitoes were maintained in specifically designed cages at 27 °C, 86% humidity, and supplied with unlimited access to 10% (wt/vol) sucrose solution and water. All dissections were performed in *Aedes* physiological solution (38). Four-day-old female *A. aegypti* and *A. gambiae* mosquitoes were blood fed on white rats and white Leghorn chickens, respectively. All procedures for the use of vertebrate animals were approved by University of California, Riverside Institutional Animal Care and Use Committee.

**dsRNA-Mediated Gene Silencing.** dsRNA synthesis was performed using the MEGAscript kit (Ambion), as previously described (14). The bacterial luciferase gene was used to generate control iLuc dsRNA. Microinjection of dsRNA into the thorax of CO<sub>2</sub>-anesthetized, 1-d-old female mosquitoes PE was performed using Picospritzer II (General Valve). Mosquito fat bodies were collected 4 d postinjection. Primers used for dsRNA synthesis are listed in [Dataset S7](#).

**Illumina RNA-seq Library Preparation.** At 4 d postinjection, 10 fat bodies were collected from female mosquitoes injected with Luc (control), Met, or Hairy dsRNA. Total RNA was extracted using TRIzol (Gibco/BRL) and RNA-seq libraries prepared using TruSeq RNA Library Preparation Kit (Illumina). Three independent biological replicates of the experiment were performed, resulting in three RNA-seq libraries per treatment.

**Ontology Analyses of *Aedes* Transcriptome.** To examine the ontology of the genes, Evolutionary Genealogy of Genes: Nonsupervised Orthologous Groups (eggNOG) database, version 3.0, was used, which is constructed through identification of reciprocal best BLAST matches and triangular linkage clustering (39). The OGs are annotated with functional descriptions along with functional categories, which were derived from the original Clusters of Orthologous Groups/Eukaryotic Orthologous Groups categories.

**In Vivo Hormonal Treatment.** A 0.3- $\mu$ L aliquot of 1  $\mu$ g/mL JH III (Sigma) or solvent (acetone) was topically applied to the abdomen of newly emerged female mosquitoes (3 h PE). Samples were collected 8 h posttreatment and subjected to qPCR and WB analysis. The efficiency of the hormonal treatment was tested by induction of established JH-activated genes.

**In Vitro Fat-Body Culture.** Fat-body tissue culture experiments were performed as previously described (16). The complete medium was supplemented with 10 nM JH III or solvent (acetone). For CHX treatments, 20  $\mu$ M CHX (dissolved in DMSO) was added to the culture medium, with or without JH. The controls were treated with DMSO only. All incubations lasted 6 h, after which samples were collected for qPCR and WB analysis.

**RNA Extraction and qPCR Analysis.** RNA extraction, cDNA synthesis, and qPCR analysis were performed as previously described (14). All of the primers used for qPCR are listed in [Dataset S7](#).

**WB Analysis.** cDNAs encoding the full-length AaHairy (AAEL005480) and C-terminal region of AaMet (AAEL001746) were subcloned into the prokaryotic expression vector pGEX-4T-1. The plasmids were expressed in an *Escherichia coli*

expression system and, following protein purification, antibodies were produced in rabbits. Antisera against both AaHairy and AaMet were purified by affinity chromatography using the ImmunoPure IgG purification kit (Pierce). Fat-body protein sample collections and WB analysis were performed as previously described (40, 41).  $\beta$ -Actin was used as a loading control; it was detected by the specific mouse monoclonal antibody (Sigma).

**Coimmunoprecipitation.** *Aedes* Hairy and Gro1 cDNA were PCR amplified and cloned into pAc5.1 vectors (Invitrogen) with a C-terminal Flag (Hairy-Flag) and V5 (Gro1-V5) tag, respectively. These two plasmids were transfected separately or together in *Drosophila* S2 cells using FuGENE reagent (Promega). Cells were harvested after 48 h incubation, and co-IP experiments were performed using Protein G Immunoprecipitation Kit (Roche), as previously described (16). IPs and WBs were performed with various combinations of the following antibodies: mouse monoclonal anti-Flag (GeneScript), mouse monoclonal anti-V5 (Invitrogen), and rabbit polyclonal anti-AaHairy. To address the domain specificity of the Hairy–Gro interaction, Flag-tagged truncated Hairy lacking the Gro binding WRPW domain (Hairy $\Delta$ WRPW-Flag) was also cloned, transfected with or without Gro1-V5, in S2 cells. For input control, Hairy-Flag, Gro1-V5, and Hairy $\Delta$ WRPW-Flag were transfected separately or together in *D. melanogaster* S2 cells and detected using WB with the antibodies mentioned above.

**EMSA.** Full-length *A. aegypti* Hairy cDNA was cloned into pcDNA3.1/Zeo(+) (Invitrogen). Hairy protein was synthesized using a coupled in vitro transcription–translation system (Promega). The specific synthesis of Hairy protein was confirmed by labeling it with <sup>35</sup>S-methionine followed by detection of the appropriate-sized band in autoradiography. EMSAs were performed with [ $\gamma$ -<sup>32</sup>P]ATP-labeled annealed oligonucleotide and the in vitro-translated Hairy using a gel shift assay system (Promega). The DNA–protein complex was separated on 5% (wt/vol) TBE Criterion Precast Gel (Bio-Rad) and visualized by means of autoradiography. For competition assays, 50-fold unlabeled specific motif or unlabeled SP1 motif was incubated with Hairy for 10 min and then further incubated with labeled motif for 20 min. For supershift assay, polyclonal antibodies against AaHairy were used.

**Luciferase Reporter Assay.** The 1-kb promoter regions of AAEL000044 and AAEL002655 were PCR amplified and subcloned into reporter vector PGL3basic (Promega), and their sequences were confirmed. The putative Hairy-binding site was mutated by incorporating a restriction site in place of E-box or E-box-like sequence in reporter plasmids. Expression plasmids Hairy-Flag, Gro1-V5, and Hairy $\Delta$ WRPW-Flag were used. Transient transfections were carried out in *Drosophila* S2 cells, as mentioned above. Samples of 200 ng of desired reporter plasmid along with 50 ng of the control *Renilla* luciferase reporter vector pRLCMV (Promega). *Renilla* luciferase reporter was cotransfected with 200 ng of Hairy-Flag and Gro1-V5, either separately or together. For concentration gradients of Hairy and Gro1, increasing amounts of the expression vectors (50, 100, and 200 ng) were transfected. For validation of the necessity of Hairy–Gro1 interaction for gene repression, 200 ng of expression plasmid with truncated Hairy that lacks Gro-interacting motif (Hairy $\Delta$ WRPW-Flag) was used, keeping all experimental conditions the same as above. Accordingly, the empty expression vector pAc5.1 was added to normalize the final concentration of DNA in each well. Luciferase assays were performed using the Dual Luciferase Assay Kit (Promega), as described previously (42).

**ACKNOWLEDGMENTS.** This work was supported by National Institutes of Health Grant R01 AI036959 (to A.S.R.), Strategic Priority Research Program of the Chinese Academy of Sciences XDB11030600, National Basic Research Program of China 2014CB138405 (to Z.Z.), and Fellowship 201208500073 from China Scholarship Council, Chinese Ministry of Education (to W.D.).

- Jindra M, Palli SR, Riddiford LM (2013) The juvenile hormone signaling pathway in insect development. *Annu Rev Entomol* 58:181–204.
- Baumann A, Fujiwara Y, Wilson TG (2010) Evolutionary divergence of the paralogs *Methoprene tolerant* (*Met*) and *germ cell expressed* (*gce*) within the genus *Drosophila*. *J Insect Physiol* 56(10):1445–1455.
- Wyatt GR, Davey KG (1996) Cellular and molecular actions of juvenile hormone. II. Roles of juvenile hormone in adult insects. *Adv Insect Physiol* 26:1–155.
- Raikhel AS, Brown MR, Belles X (2005) Hormonal control of reproductive processes. *Comprehensive Molecular Insect Science*, eds Gilbert LI, Iatrou K, Gill SS (Elsevier, Boston), Vol 3, pp 433–491.
- Bilen J, Atallah J, Azanchi R, Levine JD, Riddiford LM (2013) Regulation of onset of female mating and sex pheromone production by juvenile hormone in *Drosophila melanogaster*. *Proc Natl Acad Sci USA* 110(45):18321–18326.
- Gwadz RW, Spielman A (1973) Corpus allatum control of ovarian development in *Aedes aegypti*. *J Insect Physiol* 19(7):1441–1448.
- Clements AN (1992) *The Biology of Mosquitoes: Development, Nutrition, and Reproduction* (Chapman and Hall, London), Vol 1.
- Raikhel AS, Dhadialla TS (1992) Accumulation of yolk proteins in insect oocytes. *Annu Rev Entomol* 37:217–251.
- Raikhel AS, Lea AO (1983) Preatellogenesis development and vitellogenin synthesis in the fat body of a mosquito: An ultrastructural and immunocytochemical study. *Tissue Cell* 15(2):281–299.
- Raikhel AS, Lea AO (1991) Control of follicular epithelium development and vitelline envelope formation in the mosquito; role of juvenile hormone and 20-hydroxyecdysone. *Tissue Cell* 23(4):577–591.
- Raikhel AS, Lea AO (1985) Hormone-mediated formation of the endocytic complex in mosquito oocytes. *Gen Comp Endocrinol* 57(3):422–433.
- Raikhel AS, Lea AO (1990) Juvenile hormone controls preatellogenesis proliferation of ribosomal RNA in the mosquito fat body. *Gen Comp Endocrinol* 77(3):423–434.



13. Zhu J, Chen L, Raikhel AS (2003) Posttranscriptional control of the competence factor betaFTZ-F1 by juvenile hormone in the mosquito *Aedes aegypti*. *Proc Natl Acad Sci USA* 100(23):13338–13343.
14. Zou Z, et al. (2013) Juvenile hormone and its receptor, methoprene-tolerant, control the dynamics of mosquito gene expression. *Proc Natl Acad Sci USA* 110(24):E2173–E2181.
15. Zhu J, Busche JM, Zhang X (2010) Identification of juvenile hormone target genes in the adult female mosquitoes. *Insect Biochem Mol Biol* 40(1):23–29.
16. Shin SW, Zou Z, Saha TT, Raikhel AS (2012) bHLH-PAS heterodimer of methoprene-tolerant and Cycle mediates circadian expression of juvenile hormone-induced mosquito genes. *Proc Natl Acad Sci USA* 109(41):16576–16581.
17. Carroll SB, Laughon A, Thalley BS (1988) Expression, function, and regulation of the hairy segmentation protein in the *Drosophila* embryo. *Genes Dev* 2(7):883–890.
18. Hooper KL, Parkhurst SM, Ish-Horowicz D (1989) Spatial control of hairy protein expression during embryogenesis. *Development* 107(3):489–504.
19. Ingham PW, Pinchin SM, Howard KR, Ish-Horowicz D (1985) Genetic analysis of the hairy locus in *Drosophila melanogaster*. *Genetics* 111(3):463–486.
20. Barolo S, Levine M (1997) hairy mediates dominant repression in the *Drosophila* embryo. *EMBO J* 16(10):2883–2891.
21. Li LM, Arnosti DN (2011) Long- and short-range transcriptional repressors induce distinct chromatin states on repressed genes. *Curr Biol* 21(5):406–412.
22. Paroush Z, et al. (1994) Groucho is required for *Drosophila* neurogenesis, segmentation, and sex determination and interacts directly with hairy-related bHLH proteins. *Cell* 79(5):805–815.
23. Poortinga G, Watanabe M, Parkhurst SM (1998) *Drosophila* CtBP: A Hairy-interacting protein required for embryonic segmentation and hairy-mediated transcriptional repression. *EMBO J* 17(7):2067–2078.
24. Phippen TM, et al. (2000) *Drosophila* C-terminal binding protein functions as a context-dependent transcriptional co-factor and interferes with both mad and groucho transcriptional repression. *J Biol Chem* 275(48):37628–37637.
25. Rosenberg MI, Parkhurst SM (2002) *Drosophila* Sir2 is required for heterochromatic silencing and by euchromatic Hairy/E(Spl) bHLH repressors in segmentation and sex determination. *Cell* 109(4):447–458.
26. Fisher AL, Ohsako S, Caudy M (1996) The WRPW motif of the hairy-related basic helix-loop-helix repressor proteins acts as a 4-amino-acid transcription repression and protein-protein interaction domain. *Mol Cell Biol* 16(6):2670–2677.
27. Rushlow CA, et al. (1989) The *Drosophila* hairy protein acts in both segmentation and bristle patterning and shows homology to N-myc. *EMBO J* 8(10):3095–3103.
28. Dang CV, Dolde C, Gillison ML, Kato GJ (1992) Discrimination between related DNA sites by a single amino acid residue of Myc-related basic-helix-loop-helix proteins. *Proc Natl Acad Sci USA* 89(2):599–602.
29. Ohsako S, Hyer J, Panganiban G, Oliver I, Caudy M (1994) Hairy function as a DNA-binding helix-loop-helix repressor of *Drosophila* sensory organ formation. *Genes Dev* 8(22):2743–2755.
30. Van Doren M, Bailey AM, Esnayra J, Ede K, Posakony JW (1994) Negative regulation of proneural gene activity: Hairy is a direct transcriptional repressor of achaete. *Genes Dev* 8(22):2729–2742.
31. Ashburner M, Chihara C, Meltzer P, Richards G (1974) Temporal control of puffing activity in polytene chromosomes. *Cold Spring Harb Symp Quant Biol* 38:655–662.
32. Bianchi-Frias D, et al. (2004) Hairy transcriptional repression targets and cofactor recruitment in *Drosophila*. *PLoS Biol* 2(7):E178.
33. Zhang H, Levine M (1999) Groucho and dCtBP mediate separate pathways of transcriptional repression in the *Drosophila* embryo. *Proc Natl Acad Sci USA* 96(2):535–540.
34. Cinnamon E, Paroush Z (2008) Context-dependent regulation of Groucho/TLE-mediated repression. *Curr Opin Genet Dev* 18(5):435–440.
35. Brown NL, et al. (1996) daughterless is required for *Drosophila* photoreceptor cell determination, eye morphogenesis, and cell cycle progression. *Dev Biol* 179(1):65–78.
36. Zhou D, et al. (2008) Mechanisms underlying hypoxia tolerance in *Drosophila melanogaster*: Hairy as a metabolic switch. *PLoS Genet* 4(10):e1000221.
37. Fisher AL, Caudy M (1998) Groucho proteins: Transcriptional corepressors for specific subsets of DNA-binding transcription factors in vertebrates and invertebrates. *Genes Dev* 12(13):1931–1940.
38. Roy SG, Hansen IA, Raikhel AS (2007) Effect of insulin and 20-hydroxyecdysone in the fat body of the yellow fever mosquito, *Aedes aegypti*. *Insect Biochem Mol Biol* 37(12):1317–1326.
39. Powell S, et al. (2012) eggNOG v3.0: Orthologous groups covering 1133 organisms at 41 different taxonomic ranges. *Nucleic Acids Res* 40(Database issue):D284–D289.
40. Hansen IA, Attardo GM, Roy SG, Raikhel AS (2005) Target of rapamycin-dependent activation of S6 kinase is a central step in the transduction of nutritional signals during egg development in a mosquito. *J Biol Chem* 280(21):20565–20572.
41. Grant CE, Bailey TL, Noble WS (2011) FIMO: Scanning for occurrences of a given motif. *Bioinformatics* 27(7):1017–1018.
42. Mane-Padros D, Cruz J, Cheng A, Raikhel AS (2012) A critical role of the nuclear receptor HR3 in regulation of gonadotrophic cycles of the mosquito *Aedes aegypti*. *PLoS One* 7(9):e45019.