

SCIENTIFIC REPORTS



OPEN

Enhanced Boron Tolerance in Plants Mediated by Bidirectional Transport Through Plasma Membrane Intrinsic Proteins

Received: 15 September 2015

Accepted: 28 January 2016

Published: 23 February 2016

Kareem A. Mosa^{1,2,3}, Kundan Kumar^{1,4}, Sudesh Chhikara¹, Craig Musante⁵, Jason C. White⁵ & Om Parkash Dhankher¹

High boron (B) concentration is toxic to plants that limit plant productivity. Recent studies have shown the involvement of the members of major intrinsic protein (MIP) family in controlling B transport. Here, we have provided experimental evidences showing the bidirectional transport activity of rice OsPIP1;3 and OsPIP2;6. Boron transport ability of OsPIP1;3 and OsPIP2;6 were displayed in yeast HD9 mutant strain ($\Delta fps1\Delta acr3\Delta ycf1$) as a result of increased B sensitivity, influx and accumulation by OsPIP1;3, and rapid efflux activity by OsPIP2;6. RT-PCR analysis showed strong upregulation of OsPIP1;3 and OsPIP2;6 transcripts in roots by B toxicity. Transgenic *Arabidopsis* lines overexpressing OsPIP1;3 and OsPIP2;6 exhibited enhanced tolerance to B toxicity. Furthermore, B concentration was significantly increased after 2 and 3 hours of tracer boron (¹⁰B) treatment. Interestingly, a rapid efflux of ¹⁰B from the roots of the transgenic plants was observed within 1 h of ¹⁰B treatment. Boron tolerance in OsPIP1;3 and OsPIP2;6 lines was inhibited by aquaporin inhibitors, silver nitrate and sodium azide. Our data proved that OsPIP1;3 and OsPIP2;6 are indeed involved in both influx and efflux of boron transport. Manipulation of these PIPs could be highly useful in improving B tolerance in crops grown in high B containing soils.

Boron (B) is an essential and immobile micronutrient required for all plant nutrition. The main functions of B relate to cell wall strength, nucleic acid synthesis, hormone responses, membrane function and cell cycle regulation^{1,2}. Higher concentration of B is toxic to plants and leads to nutritional disorder that eventually limits plant production in arid and semi-arid environment³. High concentrations of B may occur naturally in soil or in groundwater, or added to the soil from mining, fertilizers, or irrigation water⁴. Recent studies have implicated the involvement of efflux type B transporters and members of major intrinsic protein (MIP) family in controlling B toxicity tolerance⁵.

Major Intrinsic Protein (MIP) superfamily is highly conserved with members ranging in size from 23 to 31 kDa⁶. In higher plants, MIPs are divided into five main subfamilies based on their sequence similarities and localization: Plasma membrane Intrinsic Proteins (PIPs), Tonoplast membrane Intrinsic Proteins (TIPs), Nodulin 26-like Intrinsic membrane Proteins (NIPs) and the Small basic Intrinsic Proteins (SIPs)^{7,8}. Recently, uncharacterized X Intrinsic Proteins (XIPs) were identified in some plant and moss species^{9,10}. Among the MIP subfamilies, members of the PIP subfamily are the most studied. PIP subfamily is further divided into two groups: PIP1s and PIP2s. PIP1s isoforms have very low water channel activity¹¹, whereas, PIP2s isoforms have been shown to possess high water channel activity^{12,13}. In rice, MIP family is comprised of 11 PIPs, 10 TIPs, 10 NIPs and two SIPs members¹⁴.

Several researchers have identified a number of B efflux transporters in plants. The first efflux-type B transporter identified was AtBOR1 from *Arabidopsis thaliana*¹⁵. Overexpression of AtBOR1 conferred tolerance

¹Stockbridge School of Agriculture, University of Massachusetts, Amherst, MA 01003, USA. ²Department of Biotechnology, Faculty of Agriculture, Al-Azhar University, Cairo, Egypt. ³Department of Applied Biology, College of Sciences, University of Sharjah, P.O. Box 27272, Sharjah, United Arab Emirates. ⁴Department of Biological Sciences, Birla Institute of Technology & Science, K. K. Birla Goa Campus, Goa 403726, India. ⁵Department of Analytical Chemistry, The Connecticut Agricultural Experiment Station, New Haven, CT 06504-1106, USA. Correspondence and requests for materials should be addressed to O.P.D. (email: parkash@umass.edu)

to *Arabidopsis* under B deficient conditions and plays a key role in xylem loading¹⁶. BOR1 homolog in barley (*Hordeum vulgare*), HvBot1, has been identified as tolerance QTL encoding a putative transmembrane B transporter with similarity to bicarbonate transporter in animals¹⁷, enabled the barley plant to tolerate high levels of B¹⁸. Overexpression of an AtBOR1 paralog, AtBOR4, in transgenic *Arabidopsis* plants also increased their tolerance to high B levels¹⁹. Homologues of AtBOR1, *Hv-BOR2* and *Ta-BOR2* from barley and wheat, respectively, were cloned and positive correlations between mRNA levels of *BOR2* genes and tolerance of high B were described among different cultivars in both barley and wheat, supporting the role of BOR2 in tolerance of high B¹⁸. AtBOR2, encodes an efflux B transporter in *Arabidopsis* which is localized in plasma membrane, and is strongly expressed in lateral root caps and epidermis of elongation zones of roots and have role in crosslinking of rhamnolacturonan II and root elongation under boron limitation in *Arabidopsis*. *BOR2* and *BOR1* mutants had reduced root elongation under low B availability²⁰. In rice, OsBOR4, a boron efflux transporter, is required for normal pollen germination and/or pollen tube elongation, and homozygous mutants showed defects in pollen tube germination and/or elongation²¹.

Among MIPs, AtNIP5;1 was the first B transporter gene identified from *Arabidopsis* which is required for efficient uptake of B in *Arabidopsis* roots²². *Arabidopsis* NIP6;1 functions in xylem–phloem transport for preferential distribution of B into young growing tissues²³. In barley, HvNIP2;1, was proposed for B toxicity tolerance, and mediated by reduced expression of HvNIP2;1 to limit B uptake²⁴. A homolog of AtNIP5;1 was identified as MtNIP3 from *Medicago truncatula* and proposed to be associated with B tolerance²⁵. An MIP subfamily member, AtTIP5;1 has been shown to be responsible for providing tolerance to B toxicity in overexpressed transgenic *Arabidopsis* plants²⁶. PIPs subfamily has been demonstrated to have a role in B permeability. The expression of maize Zm-PIP1 in *Xenopus laevis* oocytes resulted in increased B permeability²⁷ and expression of Hv-PIP1;3 and Hv-PIP1;4 from barley increased the sensitivity of yeast cells to B²⁸. Recently we reported that members of rice PIP2 subgroup, OsPIP2;4 and OsPIP2;7, have been shown to be involved in mediating B permeability and provide tolerance in overexpressed *Arabidopsis* plants most likely by internal redistribution of toxic B²⁹.

In this present study, here we report the bidirectional transport activity of two rice PIPs, OsPIP1;3 and OsPIP2;6 as a representative members of PIP1 and PIP2 subgroups, respectively. Enhanced boron tolerance in the transgenic *Arabidopsis* plants overexpressing OsPIP1;3 and OsPIP2;6 is discussed.

Results

OsPIP1;3 Expression Enhances B Sensitivity and Increases B Content in Yeast. We tested the ability of OsPIP1;3 and OsPIP2;6, as a representative members of PIP1 and PIP2 subgroups, to functionally complement the function of aquaglyceroporin Fps1 in *S. cerevisiae*. Both *OsPIP1;3*, and *OsPIP2;6* genes were expressed in the HD9 yeast strain ($\Delta fps1\Delta acr3\Delta ycf1$) lacking the expression of the bidirectional aquaglyceroporin pump Fps1, arsenite export pump ACR3, and the vacuolar transporter YCF1³⁰, for complementation of the B sensitivity and mobilization. The preliminary results with full-length coding region of OsPIPs failed to show any B transport activity in yeast HD9 strain. There was no difference in the growth of HD9 strain ($\Delta fps1\Delta acr3\Delta ycf1$) expressing OsPIP genes and the empty vector controls (Fig. 1A). It has been reported that some of the plant aquaporins required a truncation of the N-terminal hydrophobic region in order to complement the function in yeast Fps1 mutant³¹. Therefore, we attempted to complement the B transport activity of OsPIP1;3, and OsPIP2;6 after truncating their N-terminal hydrophobic region. Yeast growth was strongly impaired in cells expressing truncated OsPIP1;3 compared to yeast transformed with the pYES3 empty vector control on plates containing medium supplemented with 10 mM and 20 mM of boric acid (Fig. 1B). However, there was no significant difference between pYES3 empty vector and yeast expressing OsPIP2;6.

Total B content in the yeast cells expressing the truncated OsPIP1;3 and OsPIP2;6 was analyzed. The yeast cells expressing OsPIP1;3 showed significantly higher B accumulation as compared with pYES3 vector control (Fig. 1C), whereas, yeast cells expressing OsPIP2;6 had no significant difference in the B accumulation level compared to yeast cells expressing pYES3 empty vector. This B accumulation data is in accordance with the B sensitivity phenotype in HD9 strain complemented with OsPIP1;3.

Short Duration Influx and Efflux of B in HD9 Yeast Strain Expressing OsPIP1;3 and OsPIP2;6.

The role of OsPIP1;3 and OsPIP2;6 in transporting B was evaluated by measuring short duration influx assay using tracer ¹⁰B. HD9 strain expressing the empty vector pYES3, OsPIP1;3, and OsPIP2;6 were grown to an OD₆₀₀ of 1. For influx, cells were exposed to tracer boron ¹⁰B enriched boric acid for 0, 15, 30 and 60 min and ¹⁰B contents were measured by ICP-MS. Expression of OsPIP1;3 led to a significant increase in ¹⁰B uptake after 15, 30 and 60 min of exposure. While, there was no significant difference between yeast cells expressing the empty vector pYES3 and OsPIP2;6 (Supplementary Fig S1A). This time dependent influx results corresponded well with the levels of B sensitivity observed when growing the respective transformants on B-containing medium as shown in Fig. 1A.

To test whether OsPIP1;3 and OsPIP2;6 have B efflux activity, HD9 strains expressing the empty vector pYES3 and truncated versions of OsPIP1;3 and OsPIP2;6 were allowed to accumulate ¹⁰B till the end of log phase, harvested, washed, and then cells were re-suspended in B free media. ¹⁰B contents were analyzed by the method previously described after 15, 30 and 60 minutes. Efflux from yeast cells expressing OsPIP2;6 was faster than yeast cells expressing the empty vector pYES3 and OsPIP1;3 after 15 minutes of resuspension in B free media. However, there was no difference between the empty vector pYES3 and OsPIP2;6 after 30 and 60 minutes of resuspension in B free media. Whereas, at all time points, yeast cells expressing OsPIP1;3 showed less efflux activity than the empty vector pYES3 and OsPIP2;6 (Supplementary Fig. 1B).

Differential Regulation of OsPIP1;3 and OsPIP2;6 Under Boron Toxicity. To evaluate the differential expression of *OsPIP1;3* and *OsPIP2;6* mRNA transcripts in rice root and shoot tissues under B toxicity,

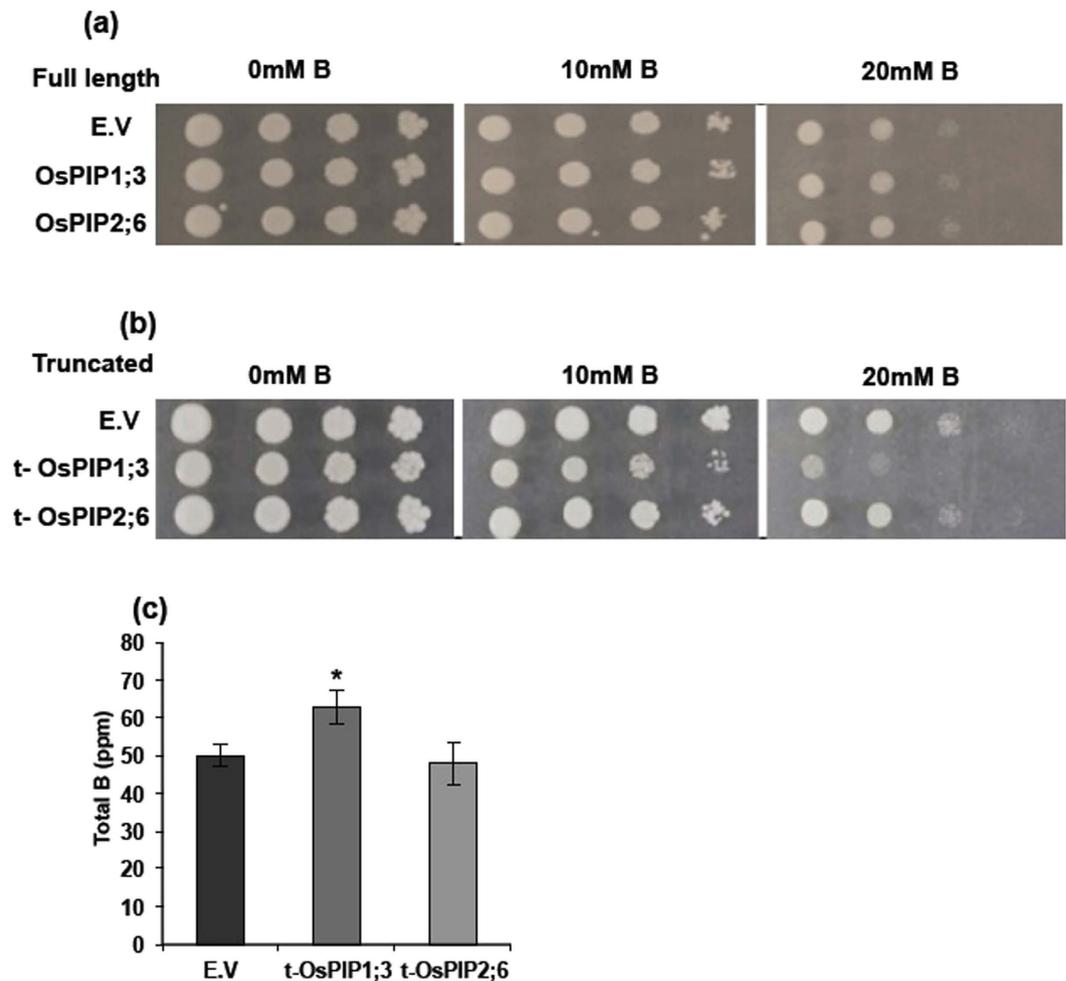


Figure 1. Functional complementation of *fps1* for B transport by OsPIP1;3 and OsPIP2;6 in *S. cerevisiae* HD9 strain ($\Delta fps1 \Delta acr3 \Delta ycf1$). Expression of full length (a) and truncated version (b) of OsPIP1;3 and OsPIP2;6 in HD9 yeast strain. The transformants were grown in liquid medium and 10-fold serial dilutions of the culture were spotted on plates with boric acid. Growth was recorded after 3–4 days at 30 °C. (c) Boron accumulation in yeast cells expressing pYES3 empty vector, t-OsPIP1;3, and t-OsPIP2;6. Data are means \pm S.D (n = 3), *P < 0.05.

we performed a quantitative RT-PCR (qRT-PCR) analysis. In shoots, transcript level of *OsPIP1;3* were down-regulated after 6, 12, and 24 hrs of B treatment, whereas, *OsPIP2;6* transcript levels showed contrasting pattern as it was upregulated after 6, 12 and 24 hrs of B toxicity (Fig. 2A). In roots, our results showed that the transcript levels of *OsPIP1;3* and *OsPIP2;6* were strongly upregulated up to 24 hrs of B treatment compared to the untreated controls (Fig. 2B). The transcript level for *OsPIP1;3* showed 15-fold increase at 24 hrs of B exposure, whereas, *OsPIP2;6* were induced by 30-fold at 24 hrs of B toxicity (Fig. 2B).

Enhanced Boron Toxicity Tolerance in Arabidopsis Plants Overexpressing OsPIP1;3 and OsPIP2;6. To investigate the function of OsPIP1;3 and OsPIP2;6 in boron permeability in plants, OsPIP1;3 was overexpressed in *A. thaliana* under *actin2* promoter-terminator cassette (*ACT2pt*) (Supplemental Fig. S2A).

Following transformation, several independent transgenic lines were screened for kanamycin resistance, and then three T₂ homozygous *Arabidopsis* lines overexpressing OsPIP1;3 (43, 51, and 60) were selected for further analysis. In addition, we used the three T₂ homozygous *Arabidopsis* lines overexpressing OsPIP2;6 (27, 33, and 40) that we generated in our previous study³² for further analysis. The overexpression of each transgene in the transgenic T₂ *Arabidopsis* plants was confirmed by semi-quantitative RT-PCR analysis (supplementary Fig. S2B).

To evaluate the phenotypic effect of B on the *Arabidopsis* transgenic lines overexpressing OsPIP1;3 and OsPIP2;6, seeds of transgenic lines and wild-type *Arabidopsis* were germinated on an 1/2x MS agar medium containing 0 or 2.5 mM of boric acid. On the control media without B, there were no differences in the phenotypes between the transgenic lines overexpressing OsPIP1;3 and OsPIP2;6, and wild-type control plants. However, on media supplemented with 2.5 mM of boric acid, the transgenic lines exhibited strong tolerance to B toxicity as compared with wild type plants (Figs 3 and 4). The growth of both shoots and roots of the transgenic lines were more vigorous than that of wild type plants on media containing B. The OsPIP1;3 and OsPIP2;6 transgenic plants had well developed green leaves compared to the wild types that had smaller and pale yellow leaves. Further,

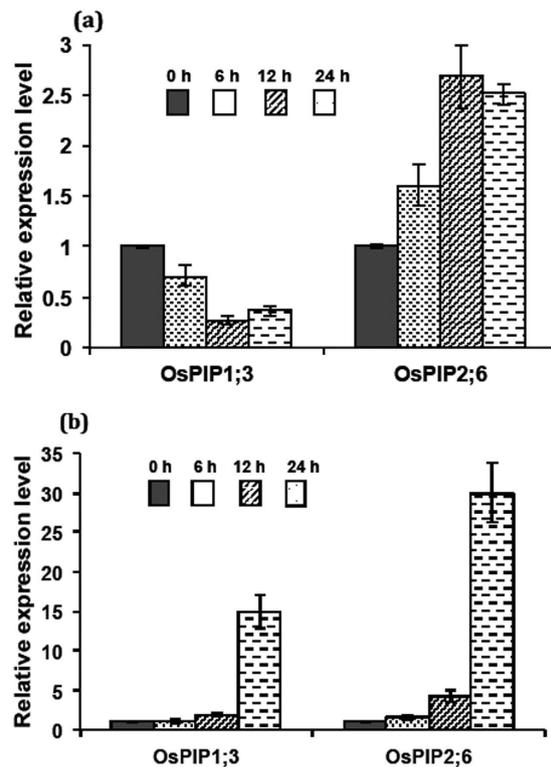


Figure 2. Relative expression of OsPIP1;3 and OsPIP2;6 genes in (a) shoots and (b) roots in response to boron exposure. Rice seedlings were exposed to 5 mM boric acid before harvesting shoots and roots tissues after 0, 6, 12 and 24 hrs of boron treatment. 18S rRNA was used for normalization of gene expression.

OsPIP1;3 and OsPIP2;6 transgenic plants demonstrated well-branched and longer roots compared to wild type plants (Figs 3A and 4A). The fresh shoot weight of OsPIP1;3 and OsPIP2;6 transgenic lines was significantly higher as compared to wild type plants on media supplemented with 2.5 mM B (Figs 3B and 4B). The root length of OsPIP1;3 and OsPIP2;6 overexpression lines were more than two-fold longer than that of the wild type plants on medium containing 2.5 mM B (Figs 3C and 4C).

Boron Accumulation in Transgenic Arabidopsis Overexpressing Rice PIP1;3 and PIP2;6. To test whether OsPIP1;3 and OsPIP2;6 are involved in B transport and uptake, wild type and transgenic *Arabidopsis* lines overexpressing OsPIP1;3 and OsPIP2;6 were grown hydroponically for three weeks and then treated with 2.5 mM boric acid for four days. Total B from shoots and roots of wild type and transgenic *Arabidopsis* were measured separately by ICP-MS. In shoots, overexpression of OsPIP1;3 and OsPIP2;6 showed no significant difference in B accumulation compared to the wild type (Fig. 5A,B). Similarly, there were no significant differences in B accumulation levels between the transgenic lines overexpressing OsPIP1;3 and OsPIP2;6 root tissues compared to wild type plant roots (Fig. 5C,D).

To confirm these results, another long-term uptake assays were conducted using a stable isotope of B (^{10}B). Regular Boric acid composition is $^{11}\text{B} : ^{10}\text{B} = 81.0 : 19.0$. Wild type and transgenic *Arabidopsis* lines overexpressing OsPIP1;3 and OsPIP2;6 were grown on 1/2x MS for three weeks and then treated with 2.5 mM ^{10}B enriched boric acid for four days. Accumulation of ^{10}B in shoots and roots of wild type and transgenic plants were measured separately by ICP-MS. Similar to previous results for boric acid, there was no significant difference in ^{10}B accumulation levels between the *Arabidopsis* wild type plants and the transgenic lines overexpressing OsPIP1;3 and OsPIP2;6 in both shoot and root tissues (Supplemental Fig. S3).

OsPIP1;3 and OsPIP2;6 Exhibited Bidirectional Transport Activity in Transgenic Arabidopsis Plants. In order to check if OsPIP1;3 and OsPIP2;6 are involved in the influx of B, a short-term B uptake (influx) using a stable isotope ^{10}B was performed. Wild type and transgenic *Arabidopsis* plants overexpressing OsPIP1;3 and OsPIP2;6 were first grown on 1/2x MS containing required amount of boric acid ($^{11}\text{B} : ^{10}\text{B} = 81.0 : 19.0$) for 3 weeks, and then plants were exposed to media containing 5 mM ^{10}B enriched boric acid and the amount of ^{10}B taken up by roots and shoots were determined at specified time points. In shoots, OsPIP1;3 and OsPIP2;6 overexpressed plants exhibited greater ^{10}B uptake than wild type plants after 1, 2, and 3 hrs (Fig. 6A). Similarly, a rapid influx of ^{10}B across the roots of OsPIP1;3 and OsPIP2;6 transgenic plants after 1, 2 and 3 hrs of exposure was observed (Fig. 6B).

To determine whether OsPIP1;3 and OsPIP2;6 are involved in the efflux of B, a short-term B efflux assay using a stable isotope ^{10}B was conducted. *Arabidopsis* wild type and transgenic plants overexpressing OsPIP1;3 and OsPIP2;6 were first exposed to 5 mM ^{10}B enriched boric acid in 1/2x MS liquid medium for 3 hrs and then

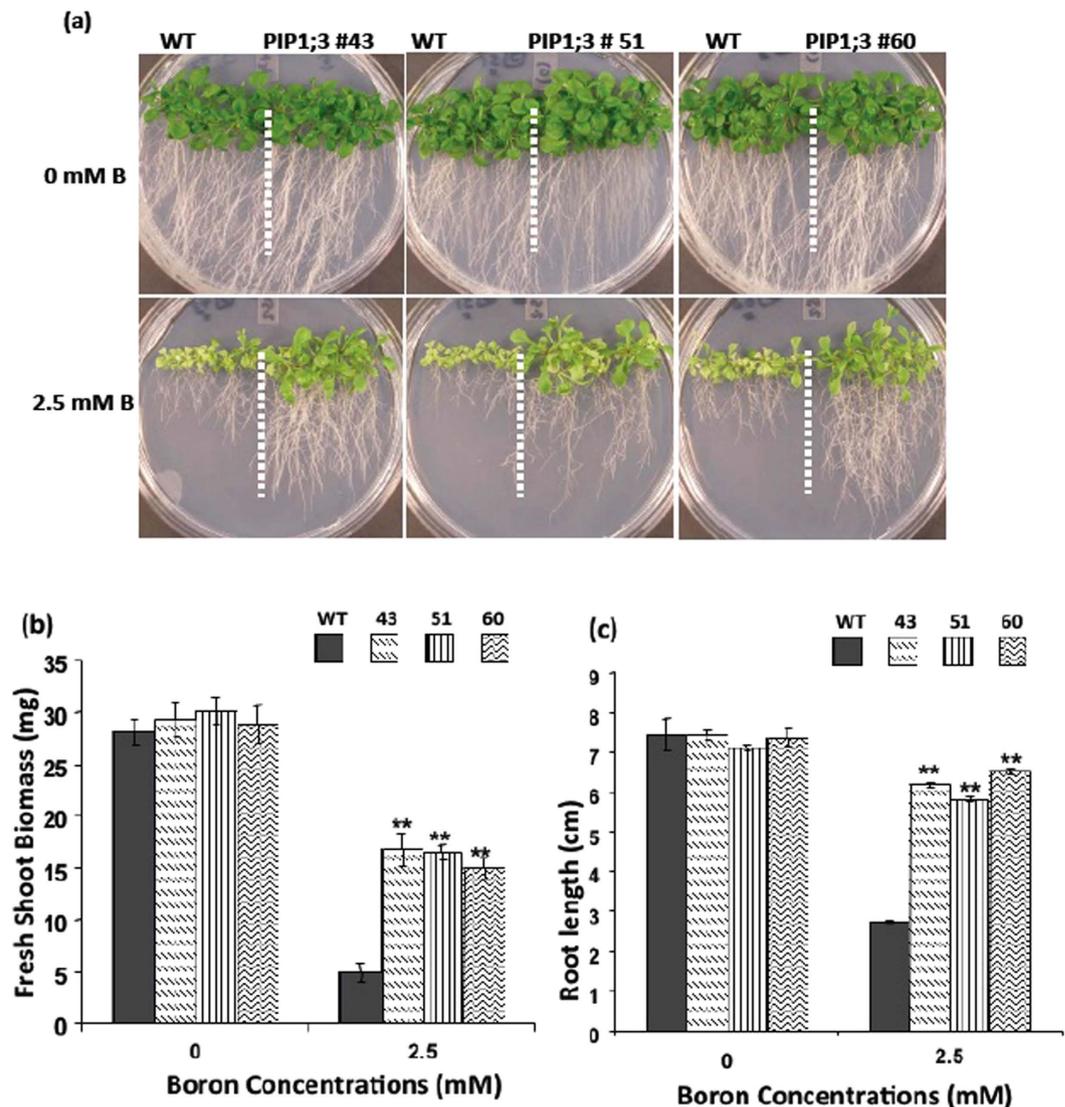


Figure 3. Boron resistance phenotype of transgenic *Arabidopsis* overexpressing OsPIP1;3. (a) Boron tolerance phenotype, (b) Fresh shoot weight and, (c) root length of the *Arabidopsis* transgenic lines 43, 51 and 60 as compared with wild type (WT) on 0 and 2.5 mM boric acid. The values are presented as an average of four replicates of 40 plants. The asterisks represent the significant difference in biomass accumulation and root length compared with wild type (WT) plants, *P < 0.05, **P < 0.01.

incubated in 1/2x MS liquid medium without any B up to 3 hrs. Concentration of ^{10}B in roots was measured at each time point (Fig. 6C). A rapid efflux of more than half of ^{10}B was observed within the first hour after transferring to 1/2x MS media without B in roots of the transgenic *Arabidopsis* expressing OsPIP1;3 or OsPIP2;6. However, after 2 and 3 hours period there was no significant difference in ^{10}B concentration in roots of the transgenic and wild types control plants (Fig. 6C).

Effect of Aquaporin Inhibitors on Boron Tolerance in OsPIP1;3 and OsPIP2;6 Transgenic Plants.

These above-described findings prompted further investigation into the effect of two different aquaporin channel inhibitor/blocker, sodium azide (NaN_3) and silver nitrate (AgNO_3), on the phenotype of the *Arabidopsis* plants overexpressing OsPIPs. Seeds of *Arabidopsis* wild type control and three independent transgenic lines were grown for three weeks on 1/2x MS medium containing 0, 3 mM B, $100\ \mu\text{M}$ NaN_3 , $50\ \mu\text{M}$ AgNO_3 , 3 mM B + $100\ \mu\text{M}$ NaN_3 , and 3 mM B + $50\ \mu\text{M}$ AgNO_3 . There was no difference between the wild type plants and transgenic lines overexpressing OsPIP1;3 and OsPIP2;6 grown on the control plates without B or NaN_3 or AgNO_3 (Figs 7 and 8). However, as expected, the transgenic lines of OsPIP1;3 and OsPIP2;6 grown on 3 mM B were more tolerant than the wild type plants. Interestingly, when plants were grown on media containing 3 mM B + $100\ \mu\text{M}$ NaN_3 , tolerance to B was abolished and there were no phenotypic differences between transgenic OsPIP1;3, OsPIP2;6 lines and wild-type control plants. Similarly, the addition of $50\ \mu\text{M}$ AgNO_3 to the 3 mM B media decreased the tolerance phenotype of the transgenic lines overexpressing OsPIP1;3 and OsPIP2;6 (Figs 7 and 8). The fresh shoot biomass of the OsPIP1;3 and OsPIP2;6 transgenic lines had 2 to 3-fold more the biomass

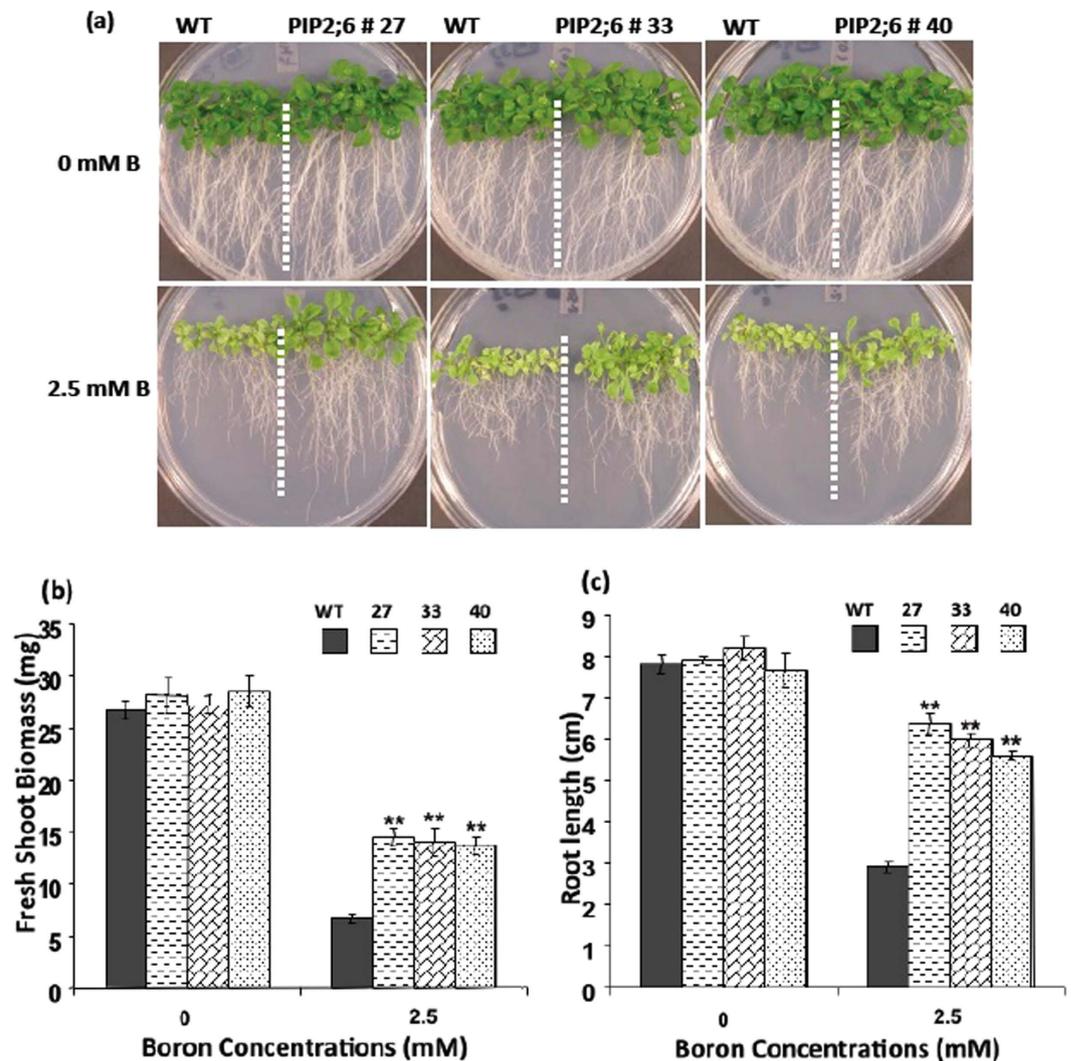


Figure 4. Boron resistance phenotype of transgenic *Arabidopsis* overexpressing *OsPIP2;6*. (a) Boron tolerance phenotype, (b) Fresh shoot weight and, (c) root length of the *Arabidopsis* transgenic lines 27, 33 and 40 as compared with wild type (WT) on 0 and 2.5 mM boric acid. The values are presented as an average of four replicates of 40 plants. The asterisks represent the significant difference in biomass accumulation and root length compared with wild type (WT) plants, * $P < 0.05$, ** $P < 0.01$.

of wild type controls when grown on 3 mM B (Supplementary Fig. S4). Whereas, the biomass of the *OsPIP1;3* and *OsPIP2;6* transgenic lines and wild type plants was almost same when plants were grown on 3 mM B + 100 μ M NaN_3 or 3 mM B + 50 μ M AgNO_3 (Supplementary Fig. S4). This proves the specific action of the inhibition of PIPs by sodium azide and silver nitrate and the tolerance to B in the transgenic lines is as a result of the activity of the *OsPIP1;3* and *OsPIP2;6*.

Discussion

Aquaporins are shown to play a pivotal role in metalloids transport in various organisms and our previous reports potentiate the role of members of rice PIPs, a subfamily of aquaporins in arsenite (AsIII) and B transport in plants^{32,29}. In current study, we have provided experimental evidences showing that *OsPIP1;3* and *OsPIP2;6* are involved in both influx and efflux of boron transport. Regulation analysis of *OsPIP1;3* and *OsPIP2;6* transcripts showed that both genes exhibited a strong upregulation in roots under B toxicity. The reason for slight decreases in *OsPIP1;3* transcript levels in shoots compared to *OsPIP2;6* is not known. However, given to the very high levels of transcripts of both *OsPIP1;3* and *OsPIP2;6* in roots (15- to 30-fold higher), this difference (2-fold decrease) in shoot is relatively minor. Further, this difference in the regulation of both genes in shoots might be due their difference in tissue-specificity or functional variability, which need further studies to clarify. It has previously been reported that expression of *HvPIP1;3* boron transporter was significantly increased upon exposure to 5 mM of B in barley²⁸. Similarly, the expression of *OsPIP2;4* and *OsPIP2;7* were strongly induced in roots by toxic B treatment, that favors the current evidences²⁹.

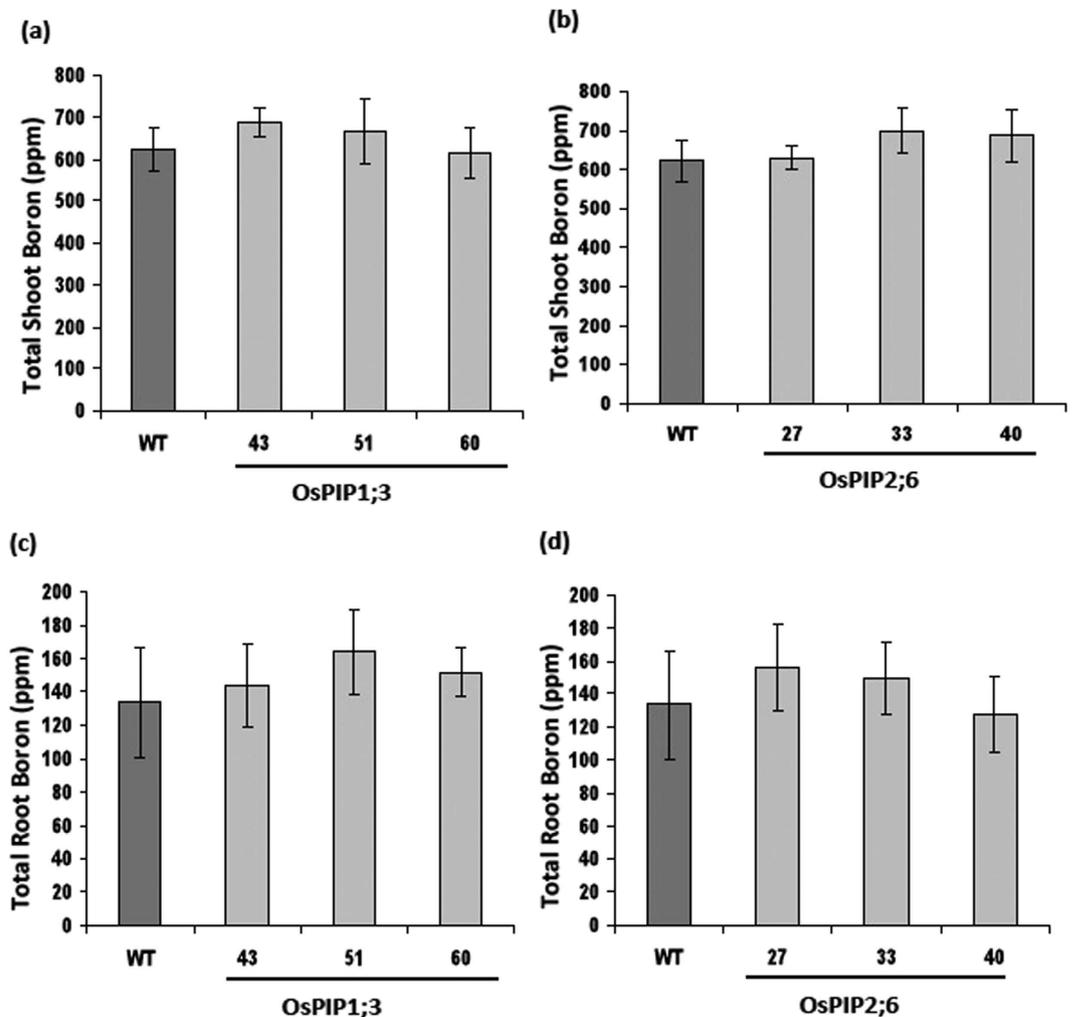


Figure 5. Analysis of total B accumulation in the transgenic *Arabidopsis* lines expressing OsPIP1;3 and OsPIP2;6. Total shoot B (a) in the transgenic *Arabidopsis* lines 43, 51, and 60 overexpressing OsPIP1;3 and (b) in the transgenic *Arabidopsis* lines 27, 33 and 40 overexpressing OsPIP2;6 in comparison with wild type (WT). Total root B (c) in the transgenic *Arabidopsis* lines 43, 51 and 60 overexpressing OsPIP1;3 and (d) in the transgenic *Arabidopsis* lines 27, 33 and 40 overexpressing OsPIP2;6 in comparison with wild type (WT). The average and standard error (SE) values are shown for four replicates of 25 plants each for WT and transgenic *Arabidopsis* lines.

The ability of OsPIP1;3 and OsPIP2;6 to transport B was examined using a yeast complementation assay with HD9 strain defective in Fps1, ACR3, and Ycf1³⁰. Preliminary results with full-length genes failed to show any B transport activity in yeast strain HD9. It has been reported that some members of plant PIPs require partial truncation of the hydrophobic N-terminal domains in order to be functionally expressed in yeast^{28,29}. Therefore, in the current study, we attempted to complement the B transport activity of OsPIP1;3 and OsPIP2;6 after truncating their N-terminal hydrophobic region. The B transport ability of OsPIP1;3 could be demonstrated as a result of increased B sensitivity and increased B influx and accumulation in yeast cells. Whereas, there was no difference in the growth of HD9 strain expressing OsPIP2;6 and the empty vector control. However, our observation should not be directly taken as the inability of OsPIP2;6 to transport boron as it showed strong efflux activity of B to external media within 15 minutes. This rapid efflux explains the B-insensitive phenotype and lack of B accumulation in yeast strain. Similarly, earlier studies also showed that expression of OsNIP1;1 did not affect the growth of $\Delta fps1 \Delta acr3 \Delta ycf1$ mutant yeast on AsIII containing medium³¹. However, OsNIP1;1 was able to mediate AsIII transport when expressed in *Xenopus* oocytes³³. In addition, heterologous expression of NIP1;2, NIP5;1, NIP6;1, NIP7;1 in yeast demonstrated their ability to transport AsIII^{31,34}, although AsIII tolerance was not observed in the T-DNA lines of *nip1;2*, *nip5;1*, *nip6;1*, and *nip7;1*³⁵. Therefore, these inconsistent results may rely on the respective expression assays and experimental conditions. Likewise, considering the fact that the failure of complementation of OsPIP2;6 in HD9 strain, our study also demonstrated the B transport ability of OsPIP2;6 in the overexpressed *Arabidopsis* transgenic plants. Further, we stressed that yeast cells are different than plant cells in terms of their ability to tolerate B toxicity. Optimization of yeast and *Arabidopsis* growth on various concentrations of boric

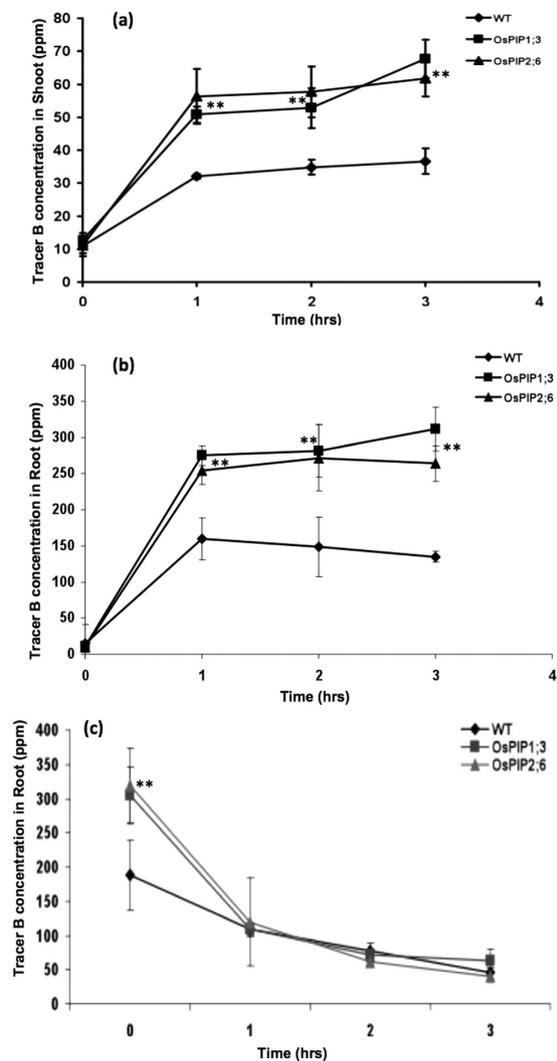


Figure 6. Influx and efflux of boron in the transgenic *Arabidopsis* lines expressing OsPIP1;3 and OsPIP2;6. Concentration of tracer boron ^{10}B in shoot (a) and roots (b) of transgenic *Arabidopsis* expressing OsPIP1;3 and OsPIP2;6 in comparison with wild type (WT) during short-term exposure to 2.5 mM ^{10}B enriched boric acid for 1, 2, and 3 hrs. (c) Concentration of tracer boron ^{10}B in roots of transgenic *Arabidopsis* expressing OsPIP1;3 and OsPIP2;6 in comparison with wild type (WT) during short-term efflux of 2.5 mM ^{10}B enriched boric acid for 1, 2, and 3 hrs. The average and standard error (SE) values are presented for three replicates of 25 plants each for WT and transgenic *Arabidopsis* lines. The asterisks represent the significant difference in influx and efflux of B compared with wild type (WT) plants, *P < 0.05, **P < 0.01.

acids showed clearly that yeast can tolerate much higher (up to 20 mM) concentration of B than *Arabidopsis* (up to 3 mM).

When OsPIP1;3 and OsPIP2;6 were overexpressed in *Arabidopsis* to clarify their physiological function *in planta*, the transgenic plants exhibited enhanced tolerance to B toxicity. Similar results were previously observed for other aquaporins members. Transgenic *Arabidopsis* plants overexpressing AtTIP5;1 conferred tolerance to B toxicity²⁶. Overexpression of OsPIP2;4 and OsPIP2;7 in *Arabidopsis* also resulted in an increased tolerance to toxic B levels²⁹. It is also intriguing that total B concentration was not affected when the transgenic *Arabidopsis* plants overexpressing OsPIP1;3 and OsPIP2;6 treated with boric acid for four days while the B concentration was significantly increased after 2 and 3 hours of ^{10}B treatment. Interestingly, a rapid efflux of ^{10}B from the roots of the OsPIP1;3 and OsPIP2;6 overexpressed transgenic *Arabidopsis* plants was observed within 1 h of ^{10}B treatment. These results are in accordance with our recent report demonstrated the involvement OsPIP2;4 and OsPIP2;7 in both influx and efflux of B from roots and shoot tissues²⁹, which is also consistent with the well-known fact of the metalloids bidirectional transport properties of plant aquaporins³¹. Therefore, tolerance to toxic B in the OsPIP1;3 and OsPIP2;6 overexpressed transgenic *Arabidopsis* plants presumed to be as a result of the rapid efflux of the excess B from root tissues to external medium.

Further, it is interesting to note that this B tolerance phenotype in OsPIP1;3 and OsPIP2;6 overexpressed lines caused by efflux mechanism was inhibited by two aquaporin inhibitors; silver nitrate and sodium azide. It has been reported that silver nitrate and sodium azide can block the aquaporins channels in plants^{36,37}. Arsenite

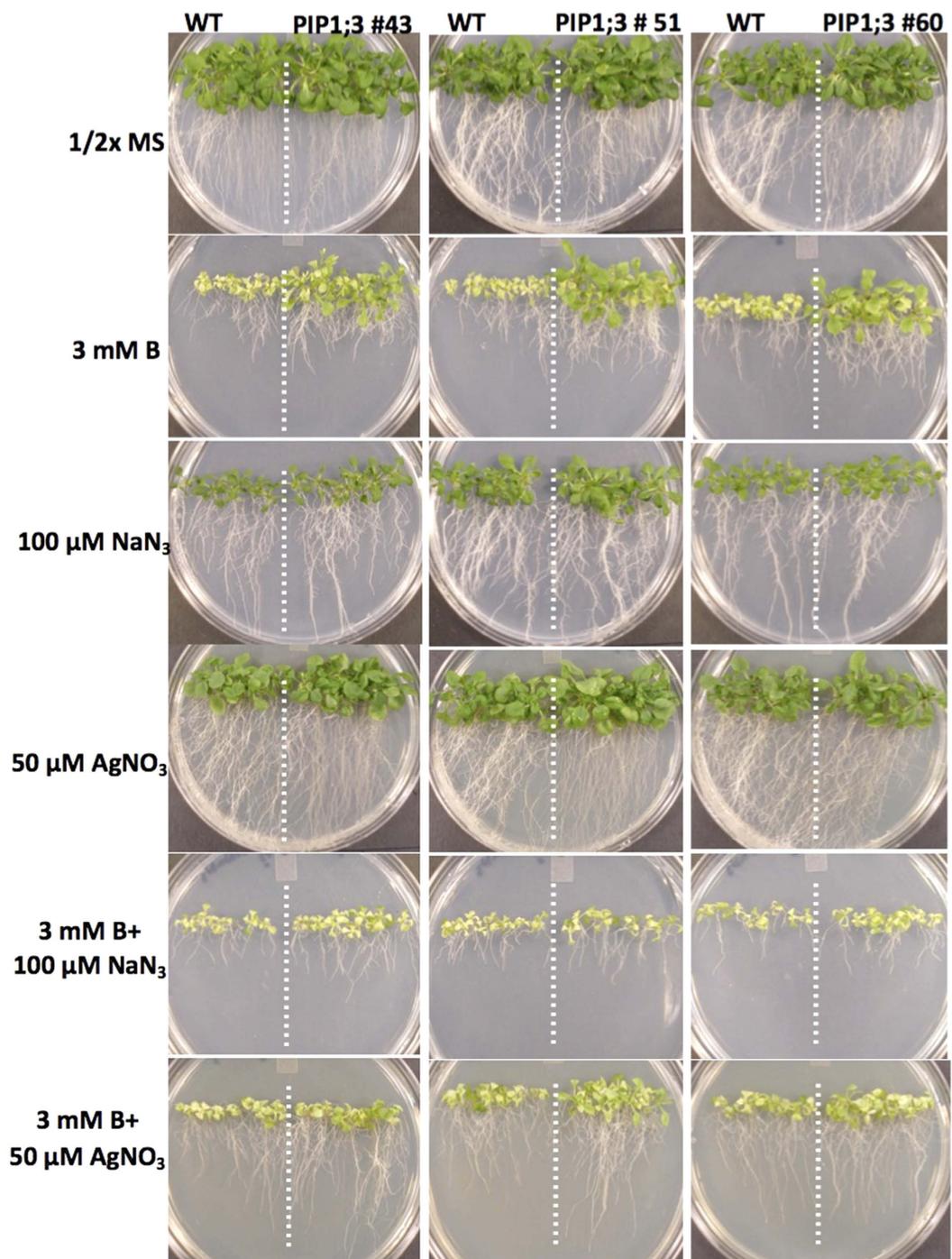


Figure 7. Phenotype of *Arabidopsis* expressing OsPIP1;3 on boron containing media and boron transporter inhibitors. *Arabidopsis* transgenic lines 43, 51 and 60 expressing OsPIP1;3 as compared with wild type (WT) on 0, 3 mM B, 100 μ M sodium azide, 50 μ M silver nitrate, 3 mM B + 100 μ M sodium azide, and 3 mM B + 50 μ M silver nitrate.

uptake in roots and fronds of *P. Vittata* was reduced by 64% and 58% in response to silver nitrate treatment as a possible result of inhibiting aquaporin transporters³⁸. Boron transport via PIP aquaporins in barley roots was reduced by 40–50% in response to metabolic inhibition by sodium azide²⁸. In this study, we demonstrated phenotypic evidence showing the inhibition of aquaporin gating by silver nitrate and sodium azide. Adding these two inhibitors to the media containing toxic B, abolished the tolerance phenotype of the transgenic *Arabidopsis* plants overexpressing OsPIP1;3 and OsPIP2;6 as a result of blocking the transport (both influx and efflux) mechanism of PIPs.

In conclusion, the findings presented here suggesting a bidirectional B transport activity of OsPIP1;3 and OsPIP2;6 and their differential regulation in roots and shoots exposed to high B. These results are in agreement

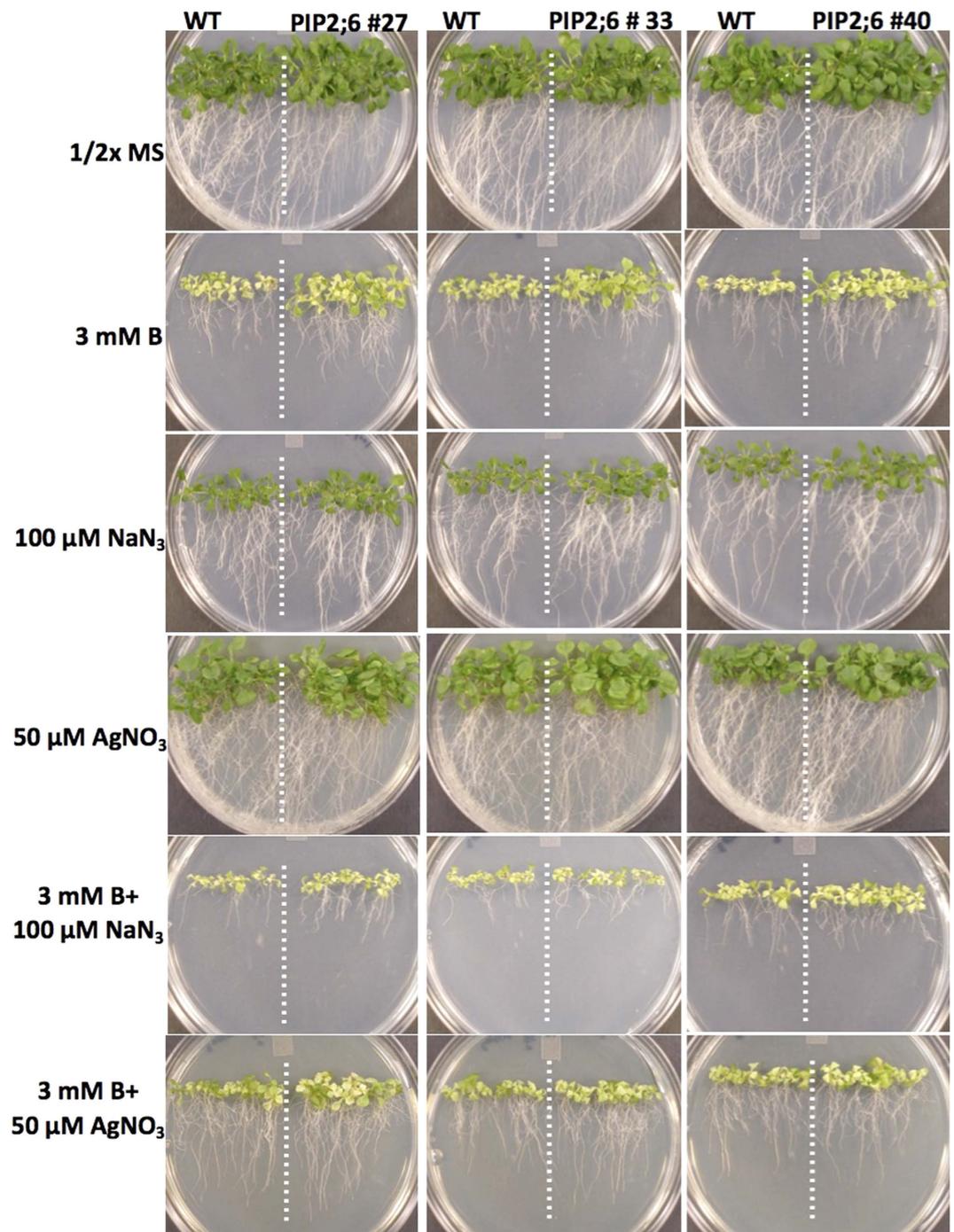


Figure 8. Phenotype of *Arabidopsis* expressing OsPIP2;6 on boron containing media and boron transporter inhibitors. *Arabidopsis* transgenic lines 27, 33 and 40 expressing OsPIP2;6 as compared with wild type (WT) on 0, 3 mM B, 100 μ M sodium azide, 50 μ M silver nitrate, 3 mM B + 100 μ M sodium azide, and 3 mM B + 50 μ M silver nitrate.

with our previous findings for OsPIP2;4 and OsPIP2;7²⁹, supporting our hypothesis that PIPs are involved in providing B tolerance by maintaining the B homeostasis in plant cells via their bidirectional transport activity under elevated B stress conditions. Similarly, we have previously shown the overexpression of OsPIP2;4, OsPIP2;6 and OsPIP2;7 showed strong tolerance to AsIII via similar bidirectional transport activity and were differentially regulated by AsIII³². Based on these findings, we drew a working model (Fig. 9) showing a proposed mechanism along with conclusion that for OsPIP1;3, OsPIP2;4, OsPIP2;6 and OsPIP2;7, contribute largely to the B and AsIII transport and tolerance mechanism in plants. OsPIP1;3, OsPIP2;4, OsPIP2;6, and OsPIP2;7 proteins localized at the plasma membrane are involved in the bidirectional transport of AsIII and B, and most likely other metalloids as well, which need to be tested in the future studies. More studies are needed to investigate the other possible

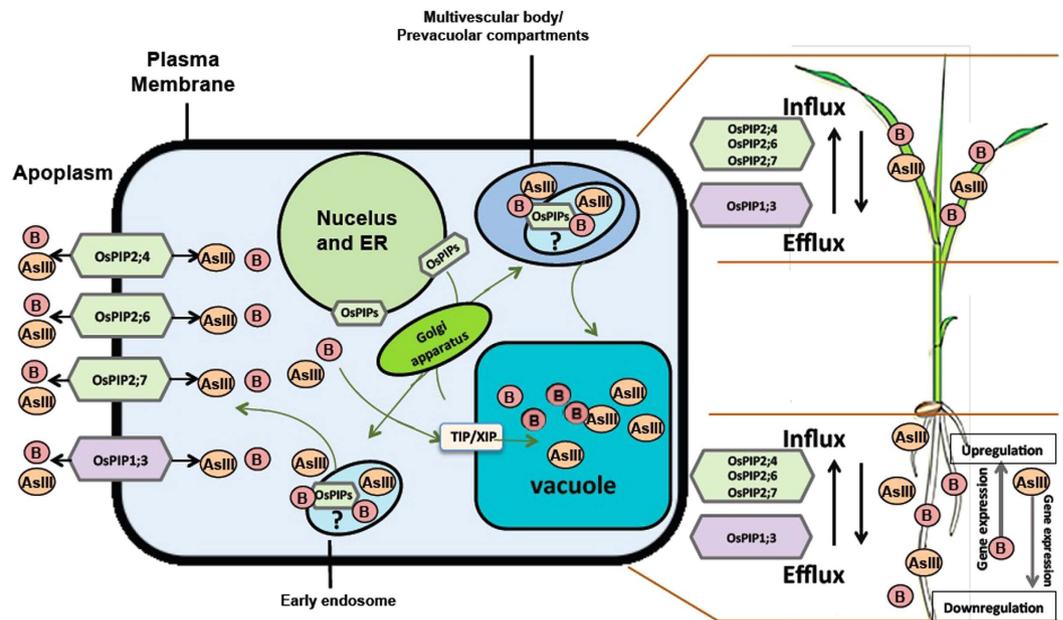


Figure 9. Schematic diagram indicating boron and arsenite (AsIII) transport mechanisms through OsPIPs and their differential regulation by boron and AsIII. The bidirectional transport (influx and efflux) through OsPIP1;3, OsPIP2;4, OsPIP2;6 and OsPIP2;7 proteins localized at the plasma membrane have been demonstrated in the diagram. Other possible unconfirmed mechanisms (marked with '?') is represented here, PIPs might be involved in the internal re-distribution of metalloids by transporting it from the endoplasmic reticulum (ER) via the Golgi apparatus to the plasma membrane. Also, metalloids could be transported through early endosome to the multivesicular body/prevacuolar compartments before it's finally placed into vacuole. AsIII and B may be transported to vacuole by uncharacterized X Intrinsic Proteins (XIPs) and/or the Tonoplast membrane Intrinsic Proteins (TIPs).

mechanisms marked with '?' in Fig. 9. PIPs could transport metalloids through early endosome to vacuole or to the plasma membrane, supporting the hypothesis that it might be involved in the internal re-distribution of metalloids (Fig. 9). Further characterization of these and other PIPs is required to clarify the functional significance of each member in boron transport in plants. Manipulation of these PIPs in crops could prove highly useful in improving B tolerance in crops grown in high B containing soils.

Methods

Plant Material and Boron Treatment. Rice (*Oryza sativa* cv. Nipponbare) seeds were germinated for two weeks in vermiculite under the green house conditions of 16/8 hours light/dark cycle, respectively, at 28 °C. Two weeks old rice plants were transferred to hydroponic solution with Hoagland's nutrients and kept to acclimatize for one week before the treatment. Five mM boric acid were added to hydroponic solution before harvesting shoots and roots tissues at 0, 6, 12 and 24 hrs after B treatments. Harvested tissues were frozen into liquid nitrogen, and stored at -80 °C until further use.

RNA Extraction and RT-PCR Analysis. Total RNA from shoot and root tissues were isolated using RNeasy plant mini kit (Qiagen) following the manufacturer's instructions. Five µg of total RNA was used for reverse transcription using the ThermoScript™ RT-PCR System (Invitrogen) for first-strand cDNA synthesis following the manufacturer's instructions. Gene-specific primers used for the quantitative real time PCR (qRT-PCR) analysis were designed using the PrimerQuest (Integrated DNA Technologies). The sequences of the qRT-PCR primers are as follows: OsPIP1;3 forward primer: CTGGTGATCGATGAAGCTAG; reverse primer: ACACAAGTACCATTCTCACAC; and OsPIP2;6 forward primer: GCCAGGTGCATGATTTGTT; reverse primer: GCCGAAGCAGTTTGTATCTC. qRT-PCR was performed following the instruction for Mastercycler® ep realplex (Eppendorf AG, Hamburg, Germany) with Absolute Blue qPCR SYBR Green Mix (Thermo Fisher Scientific, Surrey, UK). Relative expression level was calculated using $2^{-\Delta\Delta CT}$ method³⁹. Rice 18S ribosomal RNA (rRNA) was used as housekeeping genes for normalization. For semi-quantitative RT-PCR, Taq Polymerase (TaKaRa) was used according to the manufacturer's instruction. The PCR amplification conditions were: 95 °C for 2 min; 95 °C for 45 s, 58 °C for 45 s, 72 °C for 45 s; final extension at 72 °C for 7 min.

Yeast Complementation Assays. Full-length or truncated *OsPIP1;3* and *OsPIP2;6* gene sequences were amplified by PCR. For N-terminal truncation of OsPIPs, first 30–40 amino acids, were deleted by designing forward primer lacking these N-terminal sequences as previously described²⁹. Amplified products were cloned into the pGEM-T easy vector (Promega, UK) using manufacturer's instructions. The sequences were verified by DNA sequencing and then were cloned into *EcoRI* and *XhoI* sites of pYES3 vector for subsequent transformation into

HD9 (Δ fps1 Δ acr3 Δ ycf1) yeast (*Saccharomyces cerevisiae*) strain³⁰. The transformants were selected onto minimal SD medium minus tryptophan and confirmed by PCR. Yeast culture of HD9 expressing empty vector pYES3 or OsPIP1;3 and OsPIP2;6 were grown in liquid SD medium with either 2% glucose or 2% galactose and the appropriate supplements. Yeast phenotypes assays were performed as previously described by Kumar *et al.* (2013)²⁹. Briefly, the cultures were diluted to an OD₆₀₀ of 1.0. Cells with 10-fold dilutions series were spotted onto SD + Gal plates and SD + glu plates (as a control) with various concentrations of boric acid as indicated. The indicated concentration of boric acids were selected based on optimization of yeast strain growth conditions. Yeast growth on the plates was observed after 3–4 days of incubation at 30 °C.

Boron Transport Assays in Yeast. HD9 strain transformed with OsPIP1;3 and OsPIP2;6 and empty vector pYES3 were grown at 30 °C in SD minus tryptophane medium supplemented with 2% galactose to an OD₆₀₀ of 1.0. For B uptake assay, cells were supplied with 10 mM boric acid and allowed to grow overnight. Yeast cells were washed with Tris-HCl buffer containing 0.5% NaCl and then quickly washed by deionized water and dried at 70 °C, which were then digested with concentrated HNO₃ and H₂O₂ for total B was analysis by ICP-MS (Perkin Elmer). For B influx assay, cells were exposed to 10 mM ¹⁰B enriched boric acid (99%, Cambridge Isotope Laboratories, Andover, MA, USA) for 0, 15, 30 and 60 min. Cells were harvested and washed once with chilled 25 mM phosphate buffer pH 6.0 containing 0.5% NaCl, then by 10 mM Tris HCl followed by deionized water and dried at 70 °C. Total B contents were estimated by ICP-MS (Perkin Elmer) after digestion with concentrated HNO₃ and H₂O₂. For B efflux assay, cells were allowed to accumulate B till the end of log phase in 10 mM ¹⁰B enriched boric acid containing SD (-tryptophan) media. Cells were washed in chilled phosphate buffer as described above and re-suspended in the media without B. Aliquot of cells were withdrawn at 0, 20, 40 and 60 minute and the B content were analyzed by the method previously described. Total B contents at 20, 40 and 60 min were plotted as a fraction of B content at time 0, which is the maximum amount of B accumulated.

Overexpression of OsPIP1;3 and OsPIP2;6 in Arabidopsis and Characterization of Transgenic Plants. Full-length fragments of *OsPIP1;3* gene was amplified by PCR using the following primers; Forward: TACGTCGAATTCTCCATGGAGGGGAAGGAGGA; Reverse: TAGCTGCTCGAGTTAGTCCCGGCTCTTGAA, and then cloned into the pGEMT easy vector (Promega, UK) using manufacturer's instructions. The *NcoI/XhoI* fragment of *OsPIP1;3* was sub-cloned under the constitutively expressed *actin2* gene promoter-terminator expression cassette, *ACT2pt*⁴⁰, and cloned into the binary vector pBIN19 using the *KpnI/SacI* restriction sites. Plasmids were introduced into the *Agrobacterium tumefaciens* strain C58 using the heat shock method.

Arabidopsis plants were transformed by vacuum infiltration as described previously⁴¹. Transgenic plants were selected on kanamycin and the resulting T₀ seedlings of *Arabidopsis* expressing OsPIP1;3 were grown in soil and independent T₁ transgenic lines were obtained. Homozygous T₂ lines were generated by growing the T₁ transgenic lines showing 3:1 (resistant: sensitive) Mendelian segregation ratio. Transcript analysis was performed on the T₂ transgenic *Arabidopsis* lines by a semi-quantitative RT-PCR as described above. For studying the role of OsPIP2;6 in response to B treatment, we used the three T₂ homozygous *Arabidopsis* lines overexpressing OsPIP2;6 (27, 33, and 40) that were generated in our previous study³². For B phenotypic analysis, seeds of the wild type and transgenic overexpression lines were germinated and grown on 1/2x MS agar plates containing 0 or 2.5 mM boric acid for three weeks. Shoot biomass and root lengths were determined. For aquaporin inhibitors treatment, 100 μM NaN₃ (sodium azide) and 50 μM AgNO₃ (silver nitrate) were used with or without boric acid.

Boron Transport Assays in Arabidopsis. Wild type and transgenic *Arabidopsis* seeds were grown on a nylon mesh placed on 1/2x MS plates for two weeks with 16/8 hrs light/dark cycles at 22 °C. The nylon mesh along with seedlings was then transferred, placed on a support into the magenta boxes containing 1/2x MS liquid media and plants were grown for one-week for acclimatization. For long-term B accumulation assay, the 1/2x MS liquid medium was replaced with new medium containing 2.5 mM boric acid and plants were grown for another four days. Shoot and root tissues were harvested, washed with deionized water, and dried at 70 °C for 48 hrs. Dried tissues were digested in concentrated nitric acid and hydrogen peroxide and then samples were analyzed by ICP-MS. For B influx assay, 5 mM ¹⁰B-enriched boric acid (99%, Cambridge Isotope Laboratories, Andover, MA, USA) was used. Boric acid concentration was selected based on the optimization of Arabidopsis plant growths on wide range of concentrations. The shoot and root tissues were harvested separately after 1, 2, and 3 hrs of ¹⁰B treatment. ¹⁰B in roots and shoots was measured by ICP-MS. For B efflux assay, plants were treated with 5 mM ¹⁰B-enriched boric acid for 3 hrs and washed quickly with deionized water and then transferred to 1/2x MS liquid medium without B. Roots and shoots were harvested after 0, 1, 2, and 3 hrs of transferring to B free media. ¹⁰B was analyzed in roots, shoots, and efflux media by ICP-MS

Statistical Analysis. One-way ANOVA followed by least significant difference (LSD) multiple comparison test ($p < 0.01$ and $p < 0.05$) was used to determine all differences of statistical significance among treatments. Standard error of the mean were calculated and represented in all figures. Elemental content is expressed on a dry weight basis.

References

1. Brown, P. H. *et al.* Boron in plant biology. *Plant biology* **4**(2), 205–223 (2002).
2. Reguera, M., Espí, A., Bolaños, L., Bonilla, I. & Redondo-Nieto, M. Endoreduplication before cell differentiation fails in boron-deficient legume nodules. Is boron involved in signalling during cell cycle regulation? *New Phytologist* **183**(1), 8–12 (2009).
3. Pennisi, M., Bianchini, G., Muti, A., Kloppmann, W. & Gionfanti, R. Behaviour of boron and strontium isotopes in groundwater-aquifer interactions in the Cornia Plain (Tuscany, Italy). *Applied Geochemistry* **21**(7), 1169–1183 (2006).

4. Nable, R. O., Bañuelos, G. S. & Paull, J. G. Boron toxicity. *Plant and Soil* **193**(1–2), 181–198 (1997).
5. Reid, R. Understanding the boron transport network in plants. *Plant and Soil* **385**(1–2), 1–13 (2014).
6. Gomes, D. *et al.* Aquaporins are multifunctional water and solute transporters highly divergent in living organisms. *Biochimica et Biophysica Acta* **1788**(6), 1213–1228 (2009).
7. Johanson, U. *et al.* The complete set of genes encoding major intrinsic proteins in Arabidopsis provides a framework for a new nomenclature for major intrinsic proteins in plants. *Plant physiology* **126**(4), 1358–1369 (2001).
8. Maurel, C., Verdoucq, L., Luu, D. T. & Santoni, V. Plant aquaporins: membrane channels with multiple integrated functions. *Annu. Rev. Plant Biol.* **59**, 595–624 (2008).
9. Danielson, J. Å. & Johanson, U. Unexpected complexity of the aquaporin gene family in the moss *Physcomitrella patens*. *BMC Plant Biology* **8**(1), 45 (2008).
10. Bienert, G. P., Bienert, M. D., Jahn, T. P., Boutry, M. & Chaumont, F. Solanaceae XIPs are plasma membrane aquaporins that facilitate the transport of many uncharged substrates. *The Plant Journal* **66**(2), 306–317 (2011).
11. Chaumont, F., Barrieu, F., Jung, R. & Chrispeels, M. J. Plasma membrane intrinsic proteins from maize cluster in two sequence subgroups with differential aquaporin activity. *Plant Physiology* **122**(4), 1025–1034 (2000).
12. Kammerloher, W., Fischer, U., Piechotka, G. P. & Schäffner, A. R. Water channels in the plant plasma membrane cloned by immunoselection from a mammalian expression system. *The Plant Journal* **6**(2), 187–199 (1994).
13. Weig, A., Deswarte, C. & Chrispeels, M. J. The major intrinsic protein family of Arabidopsis has 23 members that form three distinct groups with functional aquaporins in each group. *Plant Physiology* **114**(4), 1347–1357 (1997).
14. Sakurai, J., Ishikawa, F., Yamaguchi, T., Uemura, M. & Maeshima, M. Identification of 33 rice aquaporin genes and analysis of their expression and function. *Plant and Cell Physiology* **46**(9), 1568–1577 (2005).
15. Takano, J. *et al.* Arabidopsis boron transporter for xylem loading. *Nature* **420**(6913), 337–340 (2002).
16. Miwa, K., Takano, J. & Fujiwara, T. Improvement of seed yields under boron-limiting conditions through overexpression of BOR1, a boron transporter for xylem loading, in Arabidopsis thaliana. *The Plant Journal* **46**(6), 1084–1091 (2006).
17. Sutton, T. *et al.* Boron-toxicity tolerance in barley arising from efflux transporter amplification. *Science* **318**(5855), 1446–1449 (2007).
18. Reid, R. Identification of boron transporter genes likely to be responsible for tolerance to boron toxicity in wheat and barley. *Plant and cell physiology* **48**(12), 1673–1678 (2007).
19. Miwa, K. *et al.* Plants tolerant of high boron levels. *Science* **318**(5855), 1417–1417 (2007).
20. Miwa, K. *et al.* Roles of BOR2, a boron exporter, in cross linking of rhamnogalacturonan II and root elongation under boron limitation in Arabidopsis. *Plant physiology* **163**(4), 1699–1709 (2013).
21. Tanaka, N. *et al.* Roles of pollen-specific boron efflux transporter, OsBOR4, in the rice fertilization process. *Plant and Cell Physiology* **54**(13), 1366–1376 (2013).
22. Takano, J. *et al.* The Arabidopsis major intrinsic protein NIP5; 1 is essential for efficient boron uptake and plant development under boron limitation. *The Plant Cell* **18**(6), 1498–1509 (2006).
23. Tanaka, M., Wallace, I. S., Takano, J., Roberts, D. M. & Fujiwara, T. NIP6; 1 is a boric acid channel for preferential transport of boron to growing shoot tissues in Arabidopsis. *The Plant Cell* **20**(10), 2860–2875 (2008).
24. Schnurbusch, T. *et al.* Boron toxicity tolerance in barley through reduced expression of the multifunctional aquaporin HvNIP2; 1. *Plant Physiology* **153**(4), 1706–1715 (2010).
25. Bogacki, P., Peck, D. M., Nair, R. M., Howie, J. & Oldach, K. H. Genetic analysis of tolerance to Boron toxicity in the legume *Medicago truncatula*. *BMC plant biology* **13**(1), 54 (2013).
26. Pang, Y. *et al.* Overexpression of the tonoplast aquaporin AtTIP5; 1 conferred tolerance to boron toxicity in Arabidopsis. *Journal of Genetics and Genomics* **37**(6), 389–397 (2010).
27. Dordas, C. & Brown, P. H. Evidence for channel mediated transport of boric acid in squash (*Cucurbita pepo*). *Plant and Soil* **235**(1), 95–103 (2001).
28. Fitzpatrick, K. L. & Reid, R. J. The involvement of aquaglyceroporins in transport of boron in barley roots. *Plant, cell & environment* **32**(10), 1357–1365 (2009).
29. Kumar, K. *et al.* Two rice plasma membrane intrinsic proteins, OsPIP2; 4 and OsPIP2; 7, are involved in transport and providing tolerance to boron toxicity. *Planta* **239**(1), 187–198 (2014).
30. Liu, Z. *et al.* Arsenite transport by mammalian aquaglyceroporins AQP7 and AQP9. *Proc natl acad Sci* **99**(9), 6053–6058 (2002).
31. Bienert, G. P. *et al.* A subgroup of plant aquaporins facilitate the bi-directional diffusion of As (OH) 3 and Sb (OH) 3 across membranes. *BMC Biology* **6**(1), 26 (2008).
32. Mosa, K. A. *et al.* Members of rice plasma membrane intrinsic proteins subfamily are involved in arsenite permeability and tolerance in plants. *Transgenic research* **21**(6), 1265–1277 (2012).
33. Ma, J. F. *et al.* Transporters of arsenite in rice and their role in arsenic accumulation in rice grain. *Proc natl acad Sci* **105**(29), 9931–9935 (2008).
34. Isayenkov, S. V. & Maathuis, F. J. The Arabidopsis thaliana aquaglyceroporin AtNIP7; 1 is a pathway for arsenite uptake. *FEBS Letters* **582**(11), 1625–1628 (2008).
35. Kamiya, T. *et al.* NIP1; 1, an aquaporin homolog, determines the arsenite sensitivity of Arabidopsis thaliana. *Journal of Biological Chemistry* **284**(4), 2114–2120 (2009).
36. Niemietz, C. M. & Tyerman, S. D. New potent inhibitors of aquaporins: silver and gold compounds inhibit aquaporins of plant and human origin. *FEBS Letters* **531**(3), 443–447 (2002).
37. Postaire, O. *et al.* A PIP1 aquaporin contributes to hydrostatic pressure-induced water transport in both the root and rosette of Arabidopsis. *Plant Physiology* **152**(3), 1418–1430 (2010).
38. Mathews, S., Rathinasabapathi, B. & Ma, L. Q. Uptake and translocation of arsenite by *Pteris vittata* L.: effects of glycerol, antimonic acid and silver. *Environmental Pollution* **159**(12), 3490–3495 (2011).
39. Livak, K. J. & Schmittgen, T. D. Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ method. *Methods* **25**(4), 402–408 (2001).
40. Dhankher, O. P. *et al.* Engineering tolerance and hyperaccumulation of arsenic in plants by combining arsenate reductase and γ -glutamylcysteine synthetase expression. *Nat Biotech.* **20**(11), 1140–1145 (2002).
41. Bechtold, N. & Pelletier, G. In planta *Agrobacterium*-mediated transformation of adult *Arabidopsis thaliana* plants by vacuum infiltration. In *Arabidopsis protocols* (pp. 259–266) Humana Press (1998).

Acknowledgements

Authors acknowledge the funding support for this work a by grant #S1899000000001 from the Ministry of Higher Education and Scientific Research in Egypt through the Egyptian Cultural and Educational Bureau, Washington, DC to OPD and KAM (GM: 714) and by a partial funding by USDA—NIFA Hatch program (MAS 00475) to OPD. KAM also would like to thank University of Sharjah, UAE for supporting KAM while writing this manuscript.

Author Contributions

K.A.M. and O.P.D. conceived and designed the experiments; K.A.M. and K.K. performed the experiments and analyzed the data; S.C. assisted in the cloning of genes; J.C.W. and C.M. performed B analysis by ICP-MS; and K.A.M. and O.P.D. wrote the manuscript.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Mosa, K. A. *et al.* Enhanced Boron Tolerance in Plants Mediated by Bidirectional Transport Through Plasma Membrane Intrinsic Proteins. *Sci. Rep.* **6**, 21640; doi: 10.1038/srep21640 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>