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Genetic association analyses highlight biological pathways underlying mitral valve prolapse

Christian Dina[1,2,33], **Nabila Bouatia-Naji**[3,4,33], **Nathan Tucker**[5,33], **Francesca N. Delling**[6,7], **Katelynn Toomer**[8], **Ronen Durst**[9], **Maelle Perrocheau**[3,4], **Leticia Fernandez-Friera**[10,11], **Jorge Solis**[10,11], **PROMESA investigators**[12], **Thierry Le Tourneau**[1,2], **Ming-Huei Chen**[7,13], **Vincent Probst**[1,2], **Yohan Bosse**[14], **Philippe Pibarot**[14], **Diana Zelenika**[15], **Mark Lathrop**[15,16], **Serge Hercberg**[4,17,18,19,20], **Ronan Roussel**[20,21,22], **Emelia J. Benjamin**[6,7], **Fabrice Bonnet**[23,24], **LO Su Hao**[25], **Elena Dolmatova**[5], **Floriane Simonet**[1], **Simon Lecointe**[1,2], **Florence Kyndt**[1,2], **Richard Redon**[1,2], **Hervé Le Marec**[1,2], **Philippe Froguel**[26,27], **Patrick T. Ellinor**[5,28], **Ramachandran S. Vasan**[6], **Patrick Bruneval**[3,4,29], **Russell A. Norris**[8,35], **David J. Milan**[5,35], **Susan A. Slaugenhaupt**[30,35], **Robert A. Levine**[31,35], **Jean-Jacques Schott**[1,2,35], **Albert A. Hagege**[3,32,35], **MVP-France**[12], **Xavier Jeunemaitre**[3,4,33,35], and **Leducq Transatlantic MITRAL Network**[11]

[1]Inserm UMR1087, CNRS UMR 6291, Institut du Thorax, Nantes, France

[2]Centre Hospitalier Universitaire (CHU) Nantes, Université de Nantes, France

[3]Inserm UMR970 Paris Cardiovascular Research Center, Paris, France

[4]Paris Descartes University, Paris Sorbonne Cité, Paris, France

[5]Cardiovascular Research Center, Massachusetts General Hospital, Charlestown MA, USA

[6]Framingham Heart Study, Framingham, MA, USA

Correspondence should be addressed to: CD (christian.dina@univ-nantes.fr), N.B-N (nabila.bouatia-naji@inserm.fr) or X.J (xavier.jeunemaitre@inserm.fr).

[12]A full list of members and affiliations appears in the Supplementary Note.

[34]These authors contributed equally to this work.

[35]These authors directed equally this work.

Contributions

CD, NBN, and XJ organized and designed the genome-wide association study and the manuscript preparation. AAH, RAL, SAS, HLM, PP, JS and LFF organized and coordinated the network efforts and recruitment of patients. RD, VP, TLT, FK, PM and YB participated to the recruitment of patients and the interpretation of the echocardiographs. JJS and XJ coordinated the collection of patient samples. MP and SL managed the collection of patient samples and performed DNA extraction and genotyping. CD and NBN conceptualized the statistical analyses. CD supervised the statistical analyses. CD, MHC and FS performed the statistical analyses. DJM conceptualized and supervised the zebrafish experiments. NT performed zebrafish experiments interpreted the data and wrote parts of the manuscript. PTE and ED participated in zebrafish experiments and interpretation of data. RAN conceptualized and supervised the mouse studies and interpreted the data. KT performed immunohistochemistry staining of mouse organs. SHL generated the mice. FND, EB, DZ, ML, SH, RR, FB, R Redon, PF and RSV conducted the epidemiological studies in control cohorts and/or contributed samples to the GWAS and/or follow-up genotyping. PB supervised valve tissues DNA extraction. SAS contributed patient samples and organized follow-up genotyping and interpreted the GWAS and follow-up data. CD, NBN, and XJ wrote the manuscript. FND, RAN, DJL, SAS, RAL, JJS and AAH edited the manuscript and all authors approved its content.

Competing financial interests.

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[7]Department of Medicine (Cardiovascular Division), Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA, USA

[8]Department of Regenerative Medicine and Cell Biology, Cardiovascular Developmental Biology Center, Children's Research Institute, Medical University of South Carolina, Charleston, SC, USA

[9]Cardiology Department, Hadassah Hebrew University Medical Center, Jerusalem, Israel

[10]Hospital Universitario Montepríncipe. Universidad CEU San Pablo, Madrid, Spain

[11]Centro Nacional de Investigaciones Cardiovasculares, Carlos III (CNIC), Madrid, Spain

[13]Department of Neurology, Boston University School of Medicine, Boston, MA, USA

[14]Institut universitaire de cardiologie et de pneumologie de Québec, Laval University, Québec, Canada

[15]Centre national de génotypage, Evry, France

[16]Genome Quebec, Montreal, Canada

[17]Paris 13 University, Sorbonne Paris Cité, Bobigny, France

[18]Inserm U1153, INRA U1125, Nutritional Epidemiology Research Unit, Epidemiology and biostatistics Center, Bobigny, France

[19]AP-HP, Department of Public Health, Avicenne Hospital, Bobigny, France

[20]Paris Diderot University, Paris, France

[21]Inserm UMRS 1138, Centre de Recherche des Cordeliers, Paris, France

[22]AP-HP, Department of endocrinology, diabetes and nutrition, FIRE Department Hospital University, Bichat Hospital, Paris, France

[23]INSERM, CIC 0203, University Hospital of Pontchaillou, Rennes, FRANCE

[24]Department of Endocrinology, University Hospital, Rennes, FRANCE

[25]Department of Biochemistry and Molecular Medicine, School of Medicine, University of California, Davis CA, USA

[26]CNRS UMR 8199, Lille Pasteur Institute, Lille 2 University, European Genomic Institute for Diabetes (EGID), Lille, France

[27]Department of Genomics of Common Disease, School of Public Health, Imperial College London, London, United Kingdom

[28]Program in Medical and Population Genetics, Broad Institute, Cambridge, MA, USA

[29]AP-HP, Department of Pathology Hôpital Européen Georges Pompidou, Paris, France

[30]Center for Human Genetic Research, Massachusetts General Hospital and Harvard Medical School, Boston, MA, USA

[31]Cardiac Ultrasound Laboratory, Massachusetts General Hospital and Harvard Medical School, Boston, MA, USA

[32]AP-HP, Department of Cardiology, Hôpital Européen Georges Pompidou, Paris, France

[33]AP-HP, Department of genetics, Hôpital Européen Georges Pompidou, Paris, France

Abstract

Non-syndromic mitral valve prolapse (MVP) is a common degenerative cardiac valvulopathy of unknown aetiology that predisposes to mitral regurgitation, heart failure and sudden death[1]. Previous family and pathophysiological studies suggest a complex pattern of inheritance[2–5]. We performed a meta-analysis of two genome-wide association studies in 1,442 cases and 2,439 controls. We identified and replicated in 1,422 cases and 6,779 controls six loci and provide functional evidence for candidate genes. We highlight *LMCD1* encoding a transcription factor[6], for which morpholino knockdown in zebrafish results in atrioventricular (AV) valve regurgitation. A similar zebrafish phenotype was obtained for tensin1 (*TNS1*), a focal adhesion protein involved in cytoskeleton organization. We also show the expression of tensin1 during valve morphogenesis and describe enlarged posterior mitral leaflets in *Tns1*$^{-/-}$ mice. This study identifies the first risk loci for MVP and suggests new mechanisms involved in mitral valve regurgitation, the most common indication for mitral valve repair[7].

The prevalence of non-syndromic MVP has been estimated as 2.4% in the general population[8]. Family aggregation[9,10], presence in rare connective tissue syndromes[11] as well as the identification of four linked loci[2–5] indicate genetic heterogeneity for MVP. Additional factors such as age- and sex-dependent penetrance, with possible association with myocardial structural and functional abnormalities, suggest additional genetic complexity[12]. We conducted an initial discovery meta-analysis on two independent French genome-wide association studies (GWAS) including 1,412 MVP cases and 2,439 controls (Supplementary Table 1), all of European ancestry, for ~4.8 million genotyped or imputed common (MAF > 0.1) single nucleotide polymorphisms (SNPs) (Supplementary Figure 1). Three loci showed genome-wide (GW) significant associations with MVP ($P < 5\times10^{-8}$) (Table 1). The strongest association (rs12465515; OR=1.33, $P=1.08\times10^{-8}$) was observed on Chr2q35 in a ~424 Kb gene-desert region where the nearest genes are *TNP1*, *IGFBP5* and *IGFBP2* (upstream) and *DIRC3* and *TNS1* (downstream) (Table 1). The two other GW-significant loci were at Chr17p13 (lead SNP rs216205, OR=1.35, $P=3.02 \times 10^{-8}$) in an intron of *SMG6* and at Chr22q12 near *MN1* and *PITPNB* (rs11705555 OR=1.34, $P=4.47 \times 10^{-8}$) (Table 1). We followed-up 23 loci with evidence of a suggestive association ($P<1\times10^{-5}$) in a first replication panel that included European Americans and European Spanish cases and controls (Set 1 and Set 2, Supplementary Figure 1).

We genotyped or imputed a total of 47 SNPs (23 loci). An intermediate meta-analysis including the discovery and the follow-up Sets 1 and 2 (N_{cases}= 2,312 and $N_{controls}$=8,296) identified a subset of 24 SNPs (15 loci) with significant associations with MVP (P<0.01), which were genotyped or imputed in two additional case-control studies from Canada and France (Sets 3 and Set 4, Supplementary Figure 1). In the global meta-analysis that included 2,864 cases and 9,218 controls, three additional loci associate with MVP at the genomic level (Table 1, Supplementary Table 2). The overall strongest association was observed on Chr3p13 for rs171408 that maps in an intron of *LMCD1* (OR=1.32, $P =1.29 \times 10^{-11}$, Table 1). Two additional signals were identified on Chr21q22 near *CBR1* and *SETD4* (rs62229266, OR=1.22, $P=1.18\times10^{-8}$) and on Chr14q24 near *SIPA1L1* and *PCNX*

(rs17767392, OR=1.23, $P = 2.27\times10^{-8}$) (Table 1). We also confirmed the three GW-significant signals identified in the discovery samples with lead SNPs rs12465515 near *IGFB55* and *TNS1* (OR= 1.25, $P = 3.11\times10^{-11}$), rs11705555 near *PITPNB* and *MN1* (OR=1.23, $P=1.39\times10^{-8}$) and rs216205 in *SMG6* (OR=1.24, $P=1.46\times10^{-8}$). Overall, we observed consistency in the direction of effects as well as nominal significant association in the follow-up meta-analysis and did not detect significant heterogeneity (P>0.05) among case control studies (Table 1).

Many patients in the general population with MVP show few clinical symptoms, if any[1]. Nonetheless, a substantial subset of patients are at risk of heart failure and cardiac death, and MVP is the most common cause of isolated mitral regurgitation requiring surgical repair[7]. To investigate if the confirmed MVP risks alleles could be more prevalent among the more severely affected patients who required valve repair or replacement, we analysed the 1,680 French patients who underwent surgical intervention and compared them to 3,259 French controls (Supplementary Table 3). We did not find a stronger effect of any MVP risk alleles, except a slight increase in the frequency of the risk allele of rs11705555 at the *PITPNB*/MN1 locus (OR=1.31, 95%CI (1.19–1.44), $P=1.88\times10^{-8}$). Overall, our findings support that MVP is under significant genetic control with susceptibility loci of relatively homogeneous effect sizes (OR from 1.22 to 1.33).

MVP-associated loci implicate four intergenic (*IGFBP5*/*TNS1*, *SETD4*/*CBR1*, *PITBNB*/*MN1*, and *PCNX*/*SIPA1L1*) and two intronic (*LMCD1* and *SMG6*) regions. From an initial list of 53 genes (±500Kb to ±1Mb of the lead SNP), we identified candidate genes at each locus based on proximity to sentinel SNP, expression level in the heart, presence of eQTL signal in publically available databases (GTEX), proximity to previously identified GWAS signals for cardiovascular traits and a biological link with mitral valve or general cardiac development (Supplementary Table 5 and Supplementary Methods for full details of gene prioritization strategy per locus).

We then investigated the expression pattern of candidate genes during valve development in mouse embryos by immunohistochemistry (IHC) at three time points that represent: *i*) completion of endothelial-to-mesenchymal transformation (EMT; E13.5), *ii*) valve sculpting and elongation (E17.5) and *iii*) achievement of the mature adult form (9 months old). Functional antibodies were only available for Tns1 and Igfbp5 on Chr2 and Pitpnb on Chr22. We also prioritized candidate genes in the MVP risk loci using morpholino knockdown (KD) based on the presence of clear zebrafish orthologs which, after filtering (Supplementary note), limited our analysis to eight genes at three loci: *igfbp2a*, *igfbp2b*, *igfbp5a*, *igfbp5b*, and *tns1* located at the chr2q35 locus; *lmcd1* at the chr3p13 locus and *smg6* and *sgsm2* at the chr17p13 locus.

On Chr2q35, rs12465515 lies within a large intergenic region with *TNS1 and IGFBP5* identified as the best two candidate genes (Supplementary Table 4). *TNS1* maps 750 kb downstream from the association signal at the chr2q35 locus (Figure 2A). Tensin1, coded by *TNS1*, localizes to focal adhesions and interacts with actin, as does Filamin A, whose genetic variants can cause a rare X-linked form of MVP[13]. Tensin1 interacts with cytoplasmic tails of integrins to anchor stress fibers and plays an important role in metastatic capacities of

cancer cells [14]. On the other hand, *IGFBP5*, encoding insulin-like growth factor-binding protein 5 (IGFBP5), is known to modulate muscle differentiation and mediate high glucose-induced pro-fibrotic effects in cardiac fibroblasts[15]. IGFBP5 was also demonstrated to modulate migration and adhesion of cancer cells and could potentially be at play in valve development and valvular interstitial cell integrity.

Only faint nonspecific staining was observed for *Igfbp5* in valves of developing and adult mice (Supplementary Figure 2). In contrast, murine IHC data showed a sustained expression for Tensin1 during valve morphogenesis, being stronger along the atrialis aspect of the forming leaflet (Figure 3A). We also found that Tensin1 expression is maintained during adulthood and localized in the endothelial and valvular interstitial cells (Figure 3A). Hematoxylin and eosin (H&E) histological staining in 9-month old Tensin1$^{-/-}$ mice showed enlarged posterior mitral leaflets compared to wild-type littermates (Figure 3B). In addition, valves from *tns1*−/− mouse showed evidence of myxomatous degeneration, indicated by increased proteoglycan content and loss of normal matrix stratification as indicate the accumulation of proteoglycan in the valves (Figure 3C). Preliminary echographic exploration of Tns1$^{-/-}$ mice (n=2) showed slight leaflet displacement (0.4 mm) compared to wildtype (0.1 mm) consistent with larger leaflets but no mitral regurgitation (Supplementary Figure 3), indicating subtle anomalies of the mitral valve that deserve future confirmation. In zebrafish experiments, a significant increase in AV regurgitation incidence was observed in *tns1* knockdown for both morpholinos (3- and 1.1-fold increase, respectively; P=0.02 and P=0.01) (Supplementary Video 2) but not in simultaneous knockdown of both *igfbp2* and *igfbp5* isoforms (Figure 2C). In situ hybridization identified high *tns1* expression throughout the developing heart, and knockdown diminished the aggregation of endothelial cells at the developing valve (Supplementary Figure 4). Further, although *notch1b* expression remained normally localized to the developing valve, the distribution of the valve development marker *bmp4* was highly disorganized (Supplementary Figure 5). Together, these results support Tensin 1 as the best candidate at the Chr2q35 locus for MVP pathogenesis.

The association signal at the Chr3p13 is intronic to *LMCD1*. Also named Dyxin, LMCD1 is a member of the LIM domain family of zinc finger proteins that act as co-regulators of transcription. It is highly expressed in mouse cardiac tissue and was demonstrated to be a direct repressor of GATA6, an important regulator of cardiac development[6]. Somatic mutations in *LMCD1* were described as potential oncogenic events in hepatocellular carcinoma metastasis by promoting cell migration[16]. The zebrafish knockdown of *lmcd1* results in significantly increased AV regurgitation for both morpholinos (4.7 and 1.2 fold increase in AV regurgitation; P=0.001 and P=0.009) (Figure 4C, Supplementary Video 3). In addition, morphological analysis of the developing myocardium revealed *lmcd1* morphants exhibited a moderate reduction in cardiac looping (Supplementary Video 3). However, although *lmcd1* is expressed throughout the heart, after *lmcd1* knockdown, expression patterns of valve development markers *notch1b* and *bmp4* displayed no abnormalities (Supplementary Figure 5) and no mis-localization of endothelial cell aggregation was observed (Supplementary Figure 4). Further investigation will be required to determine if the AV regurgitation is due to a primary valve defect or a more general defect in cardiac development. Previous *in vitro* and *in vivo* studies showed that Lmcd1/

Dyxin augment calcineurin[17]. Calcineurin signalling is required for AV endocardium EMT and subsequent valve morphogenesis in zebrafish[18]. rs355134, a highly correlated variant to the top SNP rs171408 (r^2=0.84 according to 1000 genomes data), is located in a predicted myocyte enhancer factor 2A (MEF2A) binding site, a key transcription factor in cardiac development[19]. We have previously shown that Mef2C regulated matrix production in mouse valves[20]. Our data extend the role of LMCD1 to valve development, and the putative implication of LMCD1 in matrix production regulated by MEF2A deserves future investigation.

Amongst the remaining MVP loci, we detected expression for *Pitpnb*, candidate gene on Chr22q12, in valve endothelial and interstitial cells within the mouse mitral leaflets at each of the time points investigated (Supplementary Figure 6). At the Chr17p13 locus, knockdown of candidate genes included smg6 and sgsm2, neither of which led to a valvular phenotype in the zebrafish (Supplementary Figure 7). Additional candidate genes need to be explored at this locus, which has been associated with aortic root size[21] and coronary heart disease[19].

Despite the widespread prevalence, the molecular basis of MVP has largely been elusive. The first molecular pathways implicated in MVP arose from the observation of the disease in patients with Marfan or Ehlers-Danlos syndromes, findings that highlight the importance of extracellular matrix composition[22], the TGF-beta growth factor pathway[12,23] and valve cell proliferation and differentiation[24]. Several structural mechanisms have also been proposed, such as enlargement and flattening of the mitral annulus[25] that can impose additional stresses on genetically susceptible valves and chordae[26]. This first GWAS of non-syndromic MVP reveals several susceptibility loci supporting the concept that genetic variants affecting the expression of proteins during valve development can progressively affect mitral valve function into adult life, as was recently shown for Filamin-A[27]. In particular, we provide genetic and functional evidence that *TNS1* and *LMCD1* both implicated in cell proliferation and migration are contributing to mitral valve degeneration possibly during valve development, thus revealing new pathways as possible innovative therapeutic targets.

Online methods

Leducq Transatlantic MITRAL Network

The majority of patients were recruited as a major project of the *Leducq Mitral* Network, a transatlantic consortium investigating the physiopathology of mitral valve disease with basic and clinical investigators from 10 clinical and research centres. Six centres recruited MVP patients, MVP-Nantes and MVP-France for the initial GWAS effort, MVP-USA, Framingham Heart Study (FHS) and PROlapso Mitral en cEntros eSpAñoles (PROMESA) at the Centro Nacional de Investigaciones Cardiovasculares (CNIC) for initial replication and HEGP-Surgical Cases with QCCMRC data-sets in the last replication stage. Cases were compared with controls (Framingham Heart Study, PROMESA-CNIC and QCCMRC) or general population (D.E.S.I.R for initial GWAS and one replication stage), SU.VI.MAX (GWAS) (Supplementary Figure 1).

Cases recruitment criteria

We used consensus inclusion criteria of adult (≥ 18 years) patients with idiopathic MVP if they presented displacement into the left atrium of any part of the mitral valve leaflet(s) ≥ 2 mm beyond a line connecting the annular hinge points on the parasternal long-axis view of the left ventricle by two-dimensional (2D) echocardiography[1,2]. We also included patients with previous surgery for pure severe MR due to MVP supported by an operative report and written confirmation of the diagnosis by the surgeon (MVP-France, MVP-Nantes and Surgery Cases). All cases were validated by a local, experienced team of cardiologists on the basis of clinical and echocardiography records. Recruitments excluded patients with MVP associated with other heart disease (coronary artery disease with papillary muscle disruption, hypertrophic cardiomyopathy or rheumatic disease) or known syndromes (e.g. Marfan and Ehlers-Danlos). Local ethics committees approved all studies and all patients and controls provided written informed consent. Recruitment procedures of DNA collection are detailed per cohort in Supplementary Note.

Analytical methods

GWAS genotyping and quality control—Genotyping of the discovery cohorts was independently performed by different genetic platforms that included standard quality control measures of genotyping and data acquisition from diverse high-density genotyping arrays (Supplementary Table 1). We excluded participants with genotype call rate < 97% and individual heterozygosity (IHe) level < 10,000 (determined as outlier limit after visual inspection). We excluded SNPs with a minor allele frequency (MAF) <0.1, call rate <95%, monomorphic, and with an exact Hardy Weinberg Equilibrium (HWE) $p < 0.0001$ in controls and $p < 10^{-7}$ in demographically homogenous cases to exclude SNPs that show very large deviations.

Imputation—To complement directly genotyped SNPs we performed large-scale imputation in the four discovery cohorts. First, genotyped SNPs in cases and controls were phased using the SHAPE-IT (v1) program[3]. Then, the imputation of 4.8 million common SNPs (MAF>0.1 in 1000G Europeans, proper-info > 0.4) was carried out using IMPUTE v2[4] in ~7 Mb chunks. The reference panel used was Phase I integrated variant set release (v3), in NCBI build 37 (hg19). We used similar procedures to impute non-genotyped SNPs in the replication cohort FHS using MACH software (0.3 r2_hat)[5].

Direct genotyping in the replication sets—MGH cases from the follow-up Set1 and all cases and controls of Set2 were genotyped at the Massachusetts General Hospital PNGU Core Lab using the Sequenom iPLEX Gold® application and MassARRAY® system. Follow-up Sets 3 and 4 were genotyped at the LGC genomics company using the KASP® genotyping chemistry. We excluded 9 individuals that failed genotyping for all SNPs, and SNPs with call rate <0.90. No SNP deviated from HWE (P>0.05).

Demographic analyses—The ancestry of participants was assessed using a multi-dimensional scaling technique implemented in PLINK[6]. SNPs were selected for short-range linkage disequilibrium (LD) independence ($r^2 > 0.2$). Multi-dimensional scaling method was applied on the Identity-By-State matrix and we excluded outliers on the first two

components (Supplementary Figure 8) using an expectation-maximization (EM)-fitted Gaussian mixture clustering method implemented in the R package M-CLUST, assuming one cluster and noise (Supplementary Note).

Statistical Analyses

Genome-wide and replication association with MVP status—We applied a logistic regression (additive model) as implemented in SNPTEST[4] to test the association with MVP in the GWAS discovery adjusted for the five first principal components as covariates. We also used SNPTEST and/or logistic regression on allele dosage in replication sets when cases and/or controls were imputed for genotypes (FHS in Set 1 and D.E.S.I.R. 2 in Set 4, Supplementary Figure 1) and took into account for relatedness among (FHS). For directly genotyped cases control studies (Set 2 and Set 3) we used logistic regression as implemented in PLINK.

For the GWAS meta-analysis, we applied the inverse normal strategy[7]. Because the number of controls greatly exceeds the number of cases in all studies, we used the effective sample size as advised in the METAL software [8]:$W = 4/(1/N_{cases}+1/N_{controls})$.

Regional association plots for Chr2q35, Chr3p13 and Chr17p13 were created using Locus Zoom[9].

Protein detection in mouse embryos and adult hearts

Standard histological and immunochemical procedures were used as previously described[10]. For all immunohistochemistry (IHC) experiments, 5-min antigen retrieval was performed with VectaStain and Pressure Cooker (Cuisinart). Antibodies used for immunological experiments were: Tensin1 (Novus), MF20 (Developmental Hybridoma Banks). Primary antibodies were used for IHC at a 1:100 dilution, Hoescht 33342 (nuclear stain) was used at a 1:10,000 dilution. Appropriate secondary antibodies were used for detection.

Histology and expression studies were performed on adult (9-month) wild-type (*Tensin1*$^{+/+}$) and knockout (*Tensin1*$^{-/-}$) hearts. For Histology: Adult (9-month) hearts were processed for hematoxylin and eosin stainings and immunohistochemistry (IHC) as previously described [11]. For all analyses male mice were used and N=3 for each genotype. Antibodies used for IHC were: Hyaluronan Binding Protein (HABP) to stain proteoglycans (1:100) (Callbiochem), collagen I (1:100) (MDbio), and Hoescht to stain nuclei (1:10,000) (Invitrogen).

Zebrafish experiments

Zebrafish experiments were performed in accordance with approved Institutional Animal Care and Use Committee (IACUC) protocols. TuAB zebrafish strains were reared according to standard techniques. Minimal effective doses of antisense morpholino oligonucleotides were injected at the single cell stage and compared to non-targeting morpholino injected controls. Nucleotides sequences are indicated in Supplementary Table 6. Embryos were scored for presence of AV regurgitation at 72 hours post-fertilization (hpf) using high speed videography. Semiquantitative PCR was used to demonstrate morpholino knockdown

efficacy (Supplementary Figure 9). In situ hybridizations for tissue specific expression of lmcd1, tns1, bmp4, and notch1b were performed as described[12]. In order to visualize the localization of the developing cardiac cushions, flk-EGFP reporter fish were microinjected with anti-tns1 or anti-lmcd1 morpholinos. After manual excision of the heart at 72hpf, hearts were counterstained with rhodamine labelled phalloidin and mounted using Vectashield. Confocal micrographs were acquired on a Zeiss 510 LSM with a 20X air lens. Final Images represent 2D projections of a z-series (ImageJ).

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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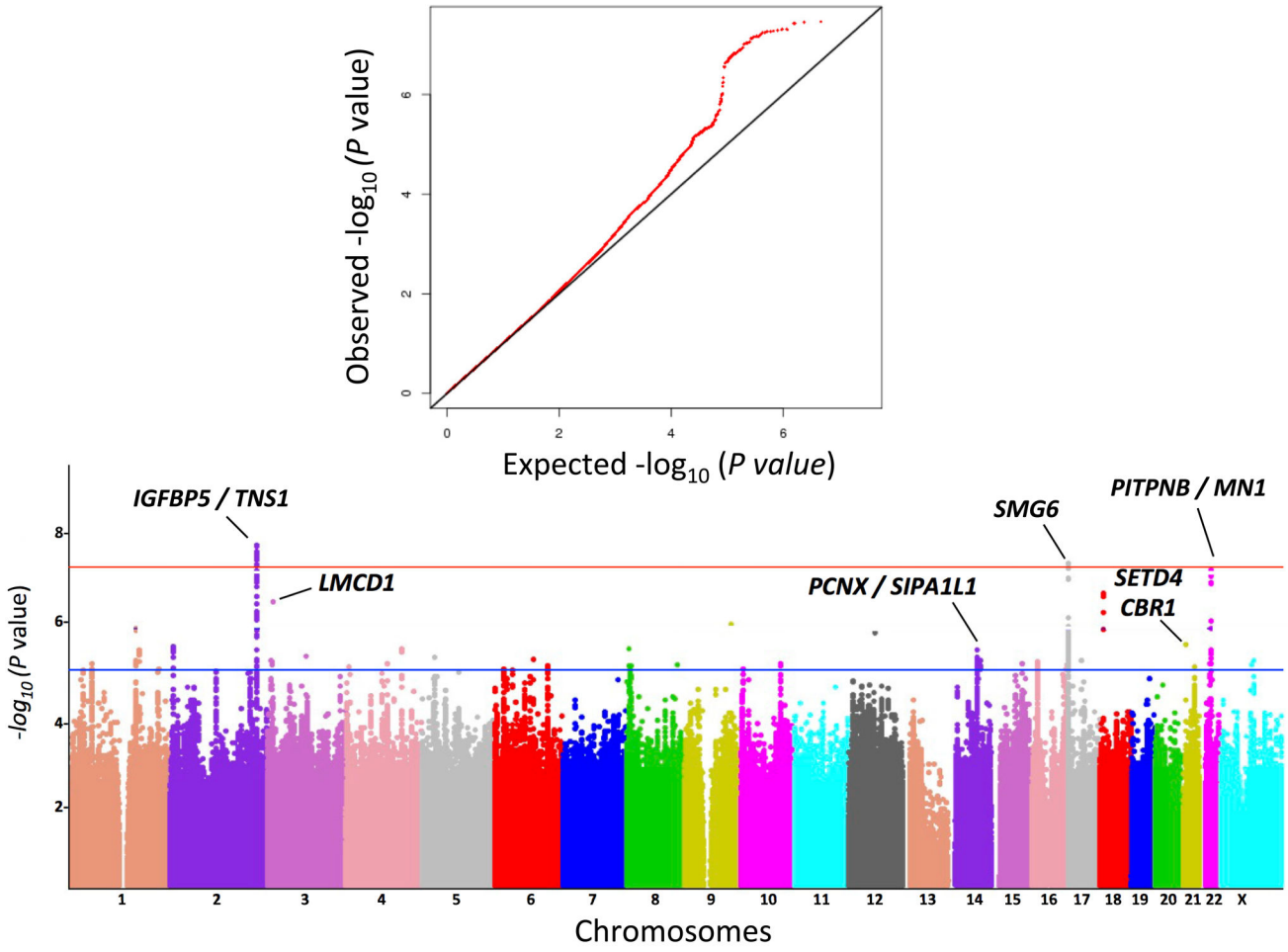

Figure 1. Quantile-quantile (A) and manhattan (B) plots representing the association of 4.8 million SNPs at the GWAS meta-analysis

Red line indicates genome-wide significant threshold ($P < 5 \times 10^{-8}$) and blue line the p-value thershold used for follow-up ($P < 10^{-5}$).

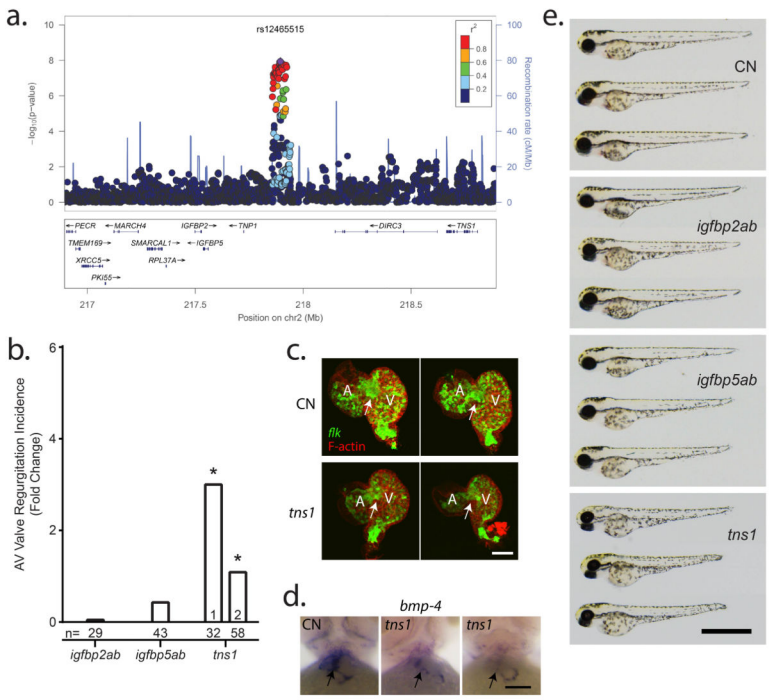

Figure 2. Cardiac regurgitation in zebrafish morpholino knockdown for candidate genes on Chr2q35

a) Genomic context of the association signal observed in the GWAS meta-analysis. The regional association plot was generated using locus zoom and displays surrounding genes, with TNS1, IGFBP2 and IGFBP5 identified as best potential candidates at this locus. **b) Mitral regurgitation observed at 72 hours post fertilization (hpf) in zebrafish embryos after morpholino mediated knockdown**. All results are presented as fold change compared to clutchmate controls. n=number of biological replicates per morpholino. (*) indicates $p<0.05$. **c) 2-dimensional projections of z-series image stacks taken on excised control (CN) and *tns1* knockdown zebrafish hearts**. Green denotes EGFP expression, a marker of endothelium under the control of the *flk* promoter. Red staining indicates the distribution of F-actin, which is highly expressed in the functional myocardium. Scale bar represents 50μm. **d) anti-*bmp-4* probe labels the valve and surrounding myocardium in CN and *tns1* knockdown embryos**. Scale bar represents 50μm. **e) Brightfield micrographs displaying gross morphology of 72hpf zebrafish morphants**. Scale bar represents 1mm. Body axis length of morpholino-injected fish is slightly reduced compared to wild-types.

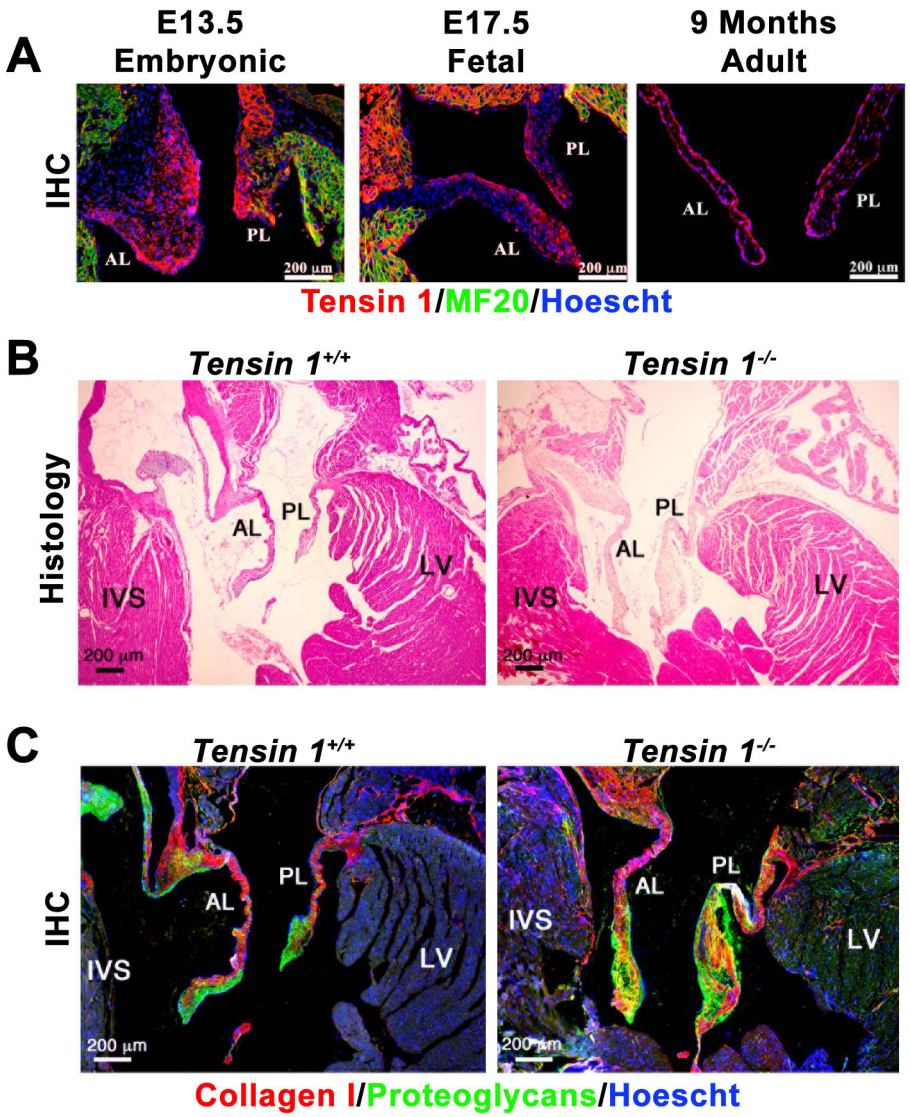

Figure 3. Murine Tensin 1 expression during developing valves and knockout phenotype at 9 months

A) Tensin1 expression in the mouse developing heart. IHC was performed for Tensin1 (red) at E13.5 (complete epithelial mesenchymal transformation), E17.5 (valve sculpting and elongation) and 9 months of age. MF20 (green) labels myocytes, Hoescht (Blue) labels nuclei. (**B) Tensin1 knockout mice exhibit enlarged mitral leaflets.** Hematoxylin and Eosin (H&E) histological staining was performed on Wild-type (Tensin$^{+/+}$) and Tensin knockout (Tensin1$^{-/-}$) mice. Scale bars are denoted. **(C) Tensin1 knockout mice exhibit myxomatous mitral leaflets.** Immunohistochemistry (IHC) for collagen (red), proteoglycans (green) show failure of normal matrix stratification and expansion of proteoglycan expression in the tensin1−/− mitral leaflets indicative of a myxomatous phenotype. AL= Anterior Leaflet, PL= Posterior Leaflet, LV=Left Ventricle, IVS=interventricular septum. Scale bars are denoted.

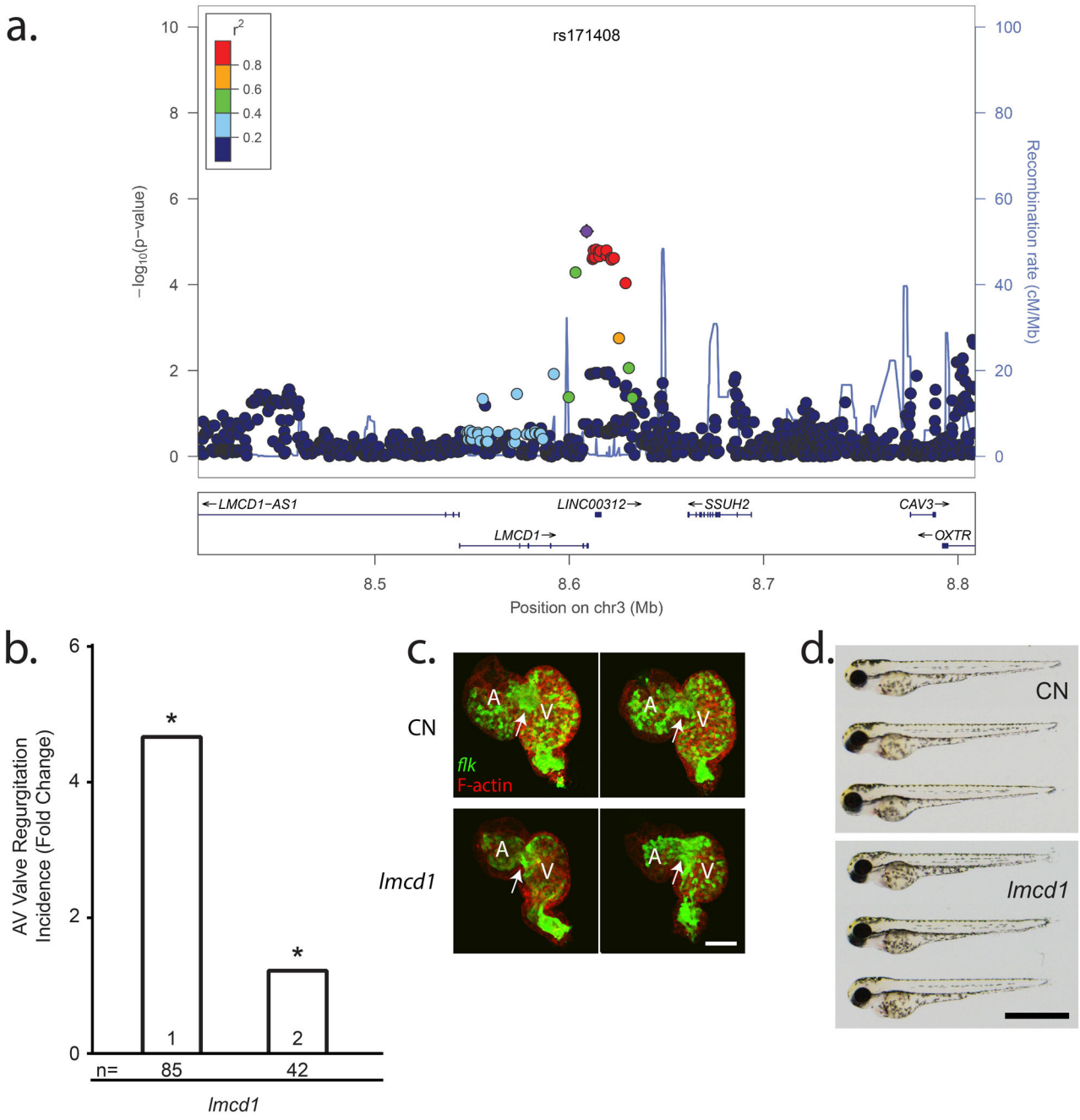

Figure 4. Cardiac regurgitation in zebrafish morpholino knockdown for Lmcd1 on Chr3p13

a) Genomic context of the association signal observed in the GWAS meta-analysis. The regional association plot was generated using locus zoom and displays surrounding genes, with *LMCD1* identified as best potential candidate as the signal is intronic to *LMCD1*. **b) Mitral regurgitation observed at 72 hours post fertilization (hpf) in zebrafish embryos after morpholino mediated knockdown**. All results are presented as fold change compared to clutchmate controls. n=number of biological replicates per morpholino. (*) indicates $p<0.05$.

c) 2-dimensional projections of z-series image stacks taken on excised 72hpf control (CN) and *lmcd1* knockdown zebrafish hearts. Green denotes EGFP expression, a marker of endothelium under the control of the *flk* promoter. Red staining indicates the distribution of F-actin, which is highly expressed in the functional myocardium. Scale bar represents 50μm. **d) Brightfield micrographs displaying gross morphology of 72hpf embryos following *lmcd1* knockdown**. Scale bar represents 1mm. CN=control morpholino injected embryos. No detectable morphological difference is observed between morpholino-injected fish and wild-types.

Table 1

Genome-wide significant associations of single nucleotide polymorphisms with mitral valve prolapse

OR: odds ratio; 95%CI: 95% confidence interval; CC: case control. Locus designed for nearest or the best candidate gene. RA: risk allele. Alleles are indexed to the forward strand of NCBI Build 37. P-values reported are two-sided and based on an inverse-variance weighted meta-analysis model (fixed effects). P-value for heterogeneity corresponds to Cochran Q statistics.

					Discovery GWAS 1,442 cases vs. 2,439 controls			Follow-up 1,422 cases vs. 6,779 controls					Combined 2,864 Cases vs. 9,218 Controls	
					MVP-Fr. vs SU.VI.MAX (953/1,566)	MVP-N vs. D.E.S.I.R 1 (489/873)	Discovery meta-analysis	MGH+FHS cases vs. FHS (699/5,575)	CNIC CC Study (171/282)	Canada CC Study (102/102)	Surgery vs.DESIR2 (450/820)	Follow-up meta-analysis		
Chr	SNP	Locus	RA	Freq.	OR (95%CI) P-value	OR (95%CI) P-value	OR (95%CI) P-value	OR (95%CI) P-value	OR (95%CI) P-value	OR (95%CI) P-value	OR (95%CI) P-value	OR (95%CI) P-value	OR (95%CI) P-value	Hetero-geneity P-value
2	**rs12465515**	**IGFBP5/TNS1**	C	0.34	1.39 (1.23–1.56) 8.0×10^{-8}	1.22 (1.03–1.45) 0.019	1.33 (1.2–1.47) 1.2×10^{-8}	1.28 (1.14–1.47) 1.2×10^{-4}	1.11 (0.84–1.47) 0.451	0.87 (0.57–1.33) 0.521	1.12 (0.95–1.33) 0.158	1.19 (1.09–1.32) 1.6×10^{-4}	**1.25 (1.18–1.33)** **3.1×10^{-11}**	0.145
3	**rs171408**	**LMCD1**	G	0.22	1.41 (1.22–1.61) 3.0×10^{-6}	1.14 (0.93–1.39) 0.188	1.3 (1.16–1.47) 5.4×10^{-6}	1.19 (1.02–1.37) 0.023	1.64 (1.19–2.27) 0.002	1.18 (0.76–1.82) 0.464	1.52 (1.25–1.85) 3.6×10^{-5}	1.32 (1.19–1.47) 3.7×10^{-7}	**1.32 (1.22–1.43)** **1.3×10^{-11}**	0.108
14	**rs17767392**	**PCNX/SIPA1L1**	T	0.25	1.33 (1.16–1.52) 2.0×10^{-5}	1.12 (0.93–1.35) 0.224	1.26 (1.13–1.4) 3.1×10^{-5}	1.19 (1.04–1.37) 0.009	1.04 (0.76–1.41) 0.814	1.2 (0.77–1.89) 0.424	1.32 (1.09–1.56) 0.004	1.20 (1.10–1.33) 1.7×10^{-4}	**1.23 (1.15–1.32)** **2.3×10^{-8}**	0.522
17	**rs216205**	**SMG6**	T	0.74	1.41 (1.24–1.61) 3.0×10^{-7}	1.25 (1.04–1.5) 0.018	1.35 (1.22–1.5) 3.0×10^{-8}	1.15 (1.00–1.33) 0.044	1.06 (0.79–1.43) 0.697	1.22 (0.80–1.86) 0.364	1.15 (0.95–1.38) 0.141	1.15 (1.03–1.27) 0.009	**1.24 (1.15–1.33)** **1.5×10^{-8}**	0.273
21	**rs62229266**	**SETD4/CBR1**	T	0.36	1.19 (1.05–1.35) 0.005	1.37 (1.16–1.64) 1.7×10^{-4}	1.25 (1.13–1.38) 8.2×10^{-6}	1.35 (1.18–1.54) 2.3×10^{-5}	1.15 (0.88–1.52) 0.303	0.86 (0.57–1.30) 0.469	1.04 (0.88–1.23) 0.605	1.19 (1.08–1.3) 2.6×10^{-4}	**1.22 (1.14–1.30)** **1.2×10^{-8}**	0.066
22	**rs11705555**	**PITPNB/MN1**	C	0.26	1.32 (1.16–1.49) 2.8×10^{-5}	1.37 (1.15–1.64) 4.0×10^{-4}	1.34 (1.21–1.49) 4.5×10^{-8}	1.16 (1.01–1.35) 0.032	1.09 (0.81–1.45) 0.594	0.91 (0.58–1.41) 0.657	1.19 (0.99–1.41) 0.059	1.15 (1.04–1.27) 0.007	**1.23 (1.15–1.33)** **1.4×10^{-8}**	0.313