



## Identification and Characterization of FTY720 for the Treatment of Human African Trypanosomiasis

Amy J. Jones, Marcel Kaiser, b,c Vicky M. Avery

Eskitis Institute for Drug Discovery, Griffith University, Nathan, Brisbane, Queensland, Australia<sup>a</sup>; Swiss Tropical and Public Health Institute, Basel, Switzerland<sup>b</sup>; University of Basel, Basel, Switzerland<sup>c</sup>

The screening of a focused library identified FTY720 (Fingolimod; Gilenya) as a potent selective antitrypanosomal compound active against Trypanosoma brucei gambiense and T. brucei rhodesiense, the causative agents of human African trypanosomiasis (HAT). This is the first report of trypanocidal activity for FTY720, an oral drug registered for the treatment of relapsing multiple sclerosis, and the characterization of sphingolipids as a potential new class of compounds for HAT.

uman African trypanosomiasis (HAT), caused by the proto-Zoan parasites Trypanosoma brucei gambiense and T. brucei rhodesiense, is prevalent in 36 sub-Saharan countries (1). The disease is extremely debilitating, progressing through two stages: an early or hemolymphatic stage in which the parasites are confined to the blood and lymphatic systems, and a second or central nervous system (CNS) stage in which the parasites penetrate the choroid barrier and invade the CNS (2). Without treatment, the disease is nearly always fatal. The currently available drugs are stage and species specific, have poor safety profiles, and require complex protracted parenteral administration, which is impractical in the rural resource-poor regions where the disease occurs (3). New compounds are desperately needed for the treatment of HAT. Despite two new compounds currently in clinical development (4-6), the extremely high attrition rates in drug discovery mean it is paramount that novel molecules continue to be identified and developed for the treatment of HAT (7).

The screening of a library composed of 741 compounds with either FDA approval or that had reached the late stages of clinical development, was undertaken against bloodstream-form T. brucei *brucei* isolates, according to previously published protocols (8, 9). Active hits, i.e., those meeting the predefined selection criterion of >50% activity at both 20 and 2 µM, were simultaneously rescreened in a dose-response study against *T. brucei brucei* and the mammalian cell line HEK293, as previously described (9), to determine 50% inhibitory concentrations (IC<sub>50</sub>s) and selectivity indices (SI), respectively. Of the 31 compounds identified as active in the primary screening campaign, 4 compounds (Table 1) had  $IC_{50}$ s of  $<10 \mu M$  and an SI of >10, criteria defined by the Drugs for Neglected Diseases initiative (DNDi) for the selection of hit compounds for HAT (10).

FTY720, a drug registered by the U.S. FDA in 2010 as an oral treatment for relapsing multiple sclerosis (MS), exhibited promising selective trypanocidal activity against T. brucei brucei (Table 1) (11). In addition, FTY720 is trypanocidal and not trypanostatic, as exposure to the  $MIC_{90}$  for 32 h resulted in >80% of the trypanosomes being killed and eliminated, with viability determined by microscopic counting of the parasites, according to previously published protocols (9).

This is the first report of the trypanocidal activity of FTY720, a structural analogue of sphingosine that is phosphorylated in vivo by sphingosine kinase (SPHK) to FTY720P(S) and FTY720P(R) (11). To further characterize FTY720 and determine if it is the only sphingolipid compound with trypanocidal activity, a panel of commercially available sphingolipid compounds and SPHK inhibitors were evaluated against T. brucei brucei, T. brucei rhodesiense, and T. brucei gambiense (Table 2), according to published methodologies (6, 8).

FTY720 and its phosphorylated enantiomers FTY720P(S) and FTY20P(R) exhibited activity against all 3 T. brucei spp., with IC<sub>50</sub>s in the range of 0.072 to 1.98  $\mu$ M (Table 2). This is in contrast to what is observed in MS, in which only FTY720P(S) exerts a therapeutic effect through its agonistic activity at sphingosine 1 phosphate receptors (S1P), and FTY720P(R) and FTY720 display no or weak activity (12). The trypanocidal activities observed for FTY720 and both FTY720P enantiomers suggest that S1P receptors may not be the target of FTY720 in trypanosomes, a hypothesis that is further supported by the absence of any receptors bearing sequence homology to S1P receptors in trypanosomes. However, additional experiments are required to confirm this hypothesis.

FTY720 was not the only sphingolipid derivative with trypanocidal activity. D-erythro-sphinganine, N-hexanoyl-D-sphingosine, and sphingosine all displayed trypanocidal activity against T. brucei, with IC<sub>50</sub>s ranging from 0.02 to 4.25 μM (Table 2). As in eukaryotic cells, sphingolipids form an essential component of T. brucei membranes, and the de novo synthesis of sphingolipids is essential for parasite viability (13). Sphingolipid biosynthesis is also unique in *T. brucei*, in contrast to the kinetoplastid parasites *Leishmania* (14) and *Trypanosoma cruzi* (15), which synthesize the higher-order sphingolipid, inositol phosphorylceramide (IPC); IPC is not detected in bloodstream *T. brucei* parasites (13, 16). In bloodstream T. brucei, the predominant higher-order sphingolipids are sphingomyelin, which is normally found in mammals, and ethanolamine phosphorylceramide (EPC) (17). The unique sph-

Received 30 August 2015 Returned for modification 30 September 2015 Accepted 4 December 2015

Accepted manuscript posted online 14 December 2015

Citation Jones AJ, Kaiser M, Avery VM. 2016. Identification and characterization of FTY720 for the treatment of human African trypanosomiasis. Antimicrob Agents Chemother 60:1859-1861. doi:10.1128/AAC.02116-15.

Address correspondence to Amy J. Jones, a.jones@griffith.edu.au.

Copyright © 2016, American Society for Microbiology. All Rights Reserved.

TABLE 1 IC<sub>50</sub>s of active compounds from the compound library against *T. brucei brucei* and HEK293 that met the hit selection criteria of IC<sub>50</sub> of <10  $\mu$ M and SI of >10

	$IC_{50}$ (mean $\pm$ SD) ( $\mu$ M) for <sup>a</sup> :				
Compound	T. brucei brucei	HEK293	$\mathrm{SI}^b$	Indication	Mechanism of action <sup>c</sup>
Nitidine chloride	$0.55 \pm 0.05$	$6.32 \pm 0.76$	11.39	Antimicrobial/antiviral/anti-inflammatory/analgesic	Inhibits topoisomerase I/II
Ethacridine lactate	$0.73 \pm 0.14$	ND	10.68	Topical antiseptic/abortifacient	Intercalates DNA/induces myometrial contraction
FTY720	$0.59 \pm 0.00$	ND	16.78	Multiple sclerosis	S1P receptors
Geldanamycin	$0.02 \pm 0.00$	$0.44 \pm 0.19$	100.85	Anticancer	Inhibits HSP-90
Pentamidine	$0.0017 \pm 0.0008$	ND	>394	Antibacterial/antiparasitic	DNA, RNA, phospholipids, and protein synthesis inhibitor
Diminazene aceturate	$0.046 \pm 0.02$	ND	>837	Antiparasitic	Binds to DNA
Puromycin	$0.042 \pm 0.01$	$0.71 \pm 0.04$	17	Antibacterial	Inhibits protein synthesis

a Results from two independent experiments. ND, no IC50 could be determined due to the compound not reaching a plateau of activity in the HEK293 assay.

ingolipid biosynthesis pathway in T. brucei has led to the pathway being proposed as a potential drug target in the parasite, and sphingosine has been shown to inhibit T. brucei brucei, with an IC<sub>50</sub> of 0.82 µM, which is similar to the value obtained in the present study (18). In addition to sphingolipid derivatives, a number of SPHK inhibitors also displayed activity against T. brucei, with IC<sub>50</sub>s of 0.02 to 10.79 μM (Table 2). CAY10621, PF543, and SKI 2 are selective inhibitors of SPHK 1, with IC<sub>50</sub>s of 3.3, 0.002, and 0.5 μM, respectively (19–21). However, there was no correlation between SPHK and antitrypanosomal activity. The SPHK inhibitor di-methyl sphingosine inhibited T. brucei brucei, with an IC<sub>50</sub> of 1.63 µM, which is in agreement with the value previously reported by Pasternack et al. (22). Although further studies are required to confirm if the SPHK inhibitors identified in this study are targeting TbSPHK, the enzyme was previously validated as a drug target in procyclic T. brucei brucei parasites (22). Unlike mammals that possess two SPHK isoforms, designated SPHK 1 and SPHK 2, only one orthologue, TbSPHK, has been identified to date in T. brucei brucei (23, 24). TbSPHK is constitutively expressed in both procyclic and bloodstream T. brucei brucei parasites, and its depletion

in procyclics results in impaired growth and abnormal organelle positioning (22).

The potent selective antitrypanosomal activity exhibited by the sphingolipids in this study led to FTY720 being evaluated in an *in vivo* acute *T. brucei rhodesiense* murine model, according to previously published protocols (6). Disappointingly, FTY720 exhibited no *in vivo* trypanocidal activity following oral or intravenous administration (data not shown). The possible reasons for a lack of translation from *in vitro* to *in vivo* activity are complex and multifold. Potential reasons include the *in vivo* phosphorylation of FTY720 to FTY720P(S), sequestration of the drug in tissues, or significant protein binding resulting in insufficient free drug at trypanocidal concentrations (25). However, extensive studies will be required to determine the exact reasons that *in vivo* trypanocidal activity was not observed for FTY720.

This study is the first report of the antitrypanosomal activity of FTY720. Despite a lack of *in vivo* activity, the results presented in this paper further highlight the potential of sphingolipid derivatives and SPHK inhibitors as therapeutics for HAT; thus, further investigation into these chemical classes is warranted.

TABLE 2 In vitro trypanocidal activities of FTY720, sphingolipids, and SPHKI against T. brucei subspecies

	$IC_{50}$ (mean $\pm$ SD) ( $\mu$ M) for <sup>a</sup> :						
Compound	T. brucei brucei	T. brucei rhodesiense	T. brucei gambiense STIB930	T. brucei gambiense K048	SI HEK293 <sup>b</sup>		
FTY720	$0.59 \pm 0.01$	$0.20 \pm 0.01$	$0.01 \pm 0.01$	$0.072 \pm 0.08$	16.8		
FTY720P(S)	$1.21 \pm 0.07$	$0.71 \pm 0.09$	$0.03 \pm 0.01$	$0.40 \pm 0.13$	14.8		
FTY720P(R)	$1.98 \pm 0.26$	$1.06 \pm 0.21$	$0.11 \pm 0.05$	$0.67 \pm 0.05$	9		
Sphingosine	$1.26 \pm 0.35$	$0.63 \pm 0.09$	$0.54 \pm 0.02$	$0.53 \pm 0.08$	31.4		
D-Erythro-sphinganine	$1.71 \pm 0.05$	$0.99 \pm 0.04$	$0.86 \pm 0.04$	$0.72 \pm 0.08$	6		
D-Methyl-sphingosine	$1.63 \pm 0.12$	$0.58 \pm 0.11$	$0.02 \pm 0.02$	$0.18 \pm 0.01$	12.2		
N-hexanoyl-D-sphingosine	$2.89 \pm 0.83$	$3.03 \pm 0.17$	$6.17 \pm 0.91$	$4.25 \pm 0.43$	1.4		
CAY10621	$0.48 \pm 0.36$	$0.19 \pm 0.03$	$0.06 \pm 0.06$	$0.09 \pm 0.14$	20.8		
PF543	$10.79 \pm 3.07$	$3.10 \pm 1.25$	$0.51 \pm 0.14$	$2.13 \pm 0.16$	1.8		
SKI2	$1.95 \pm 0.92$	$3.73 \pm 0.09$	$0.20 \pm 0.03$	$0.34 \pm 0.27$	20.3		
Pentamidine	$0.0027 \pm 0.0007$		$0.0007 \pm 0.0002$	$0.051 \pm 0.016$	260.8		
Diminazene aceturate	$0.027 \pm 0.001$				957		
Puromycin	$0.038 \pm 0.002$				12		
Melarsoprol		$0.007 \pm 0.003$	$0.0006 \pm 0.002$	$0.014 \pm 0.002$			

<sup>&</sup>lt;sup>a</sup> Results from three independent experiments.

b SI, selectivity index.

<sup>&</sup>lt;sup>c</sup> S1P, sphingosine 1 phosphate; HSP, heat shock protein.

<sup>&</sup>lt;sup>b</sup> Selectivity index (SI) compared to HEK293 cells.

## **ACKNOWLEDGMENTS**

We thank G. Stevenson for his contributions to this study, particularly the medicinal chemistry advice, and M. Jud, S. Keller, and G. Riccio (Swiss TPH) for assistance with in vitro and in vivo tests.

This work was supported by NHMRC project grant APP1067728 awarded to Vicky M. Avery. The funders had no role in the study design, data collection and interpretation, or the decision to submit the work for publication.

## **FUNDING INFORMATION**

Department of Health | National Health and Medical Research Council (NHMRC) provided funding to Vicky M. Avery under grant number APP1067728.

## **REFERENCES**

- 1. Franco JR, Simarro PP, Diarra A, Jannin JG. 2014. Epidemiology of human African trypanosomiasis. Clin Epidemiol 6:257-275. http://dx.doi .org/10.2147/CLEP.S39728.
- World Health Organization. 2013. Control and surveillance of human African trypanosomiasis: report of a WHO expert committee. World Health Organization, Geneva, Switzerland. http://apps.who.int/iris /bitstream/10665/95732/1/9789241209847\_eng.pdf.
- 3. Brun R, Blum J, Chappuis F, Burri C. 2010. Human African trypanosomiasis. Lancet 375:148-159. http://dx.doi.org/10.1016/S0140-6736 (09)60829-1.
- Torreele E, Bourdin Trunz B, Tweats D, Kaiser M, Brun R, Mazué G, Bray MA, Pécoul B. 2010. Fexinidazole-a new oral nitroimidazole drug candidate entering clinical development for the treatment of sleeping sickness. PLoS Negl Trop Dis 4:e923. http://dx.doi.org/10.1371/journal.pntd
- 5. Jacobs RT, Nare B, Wring SA, Orr MD, Chen D, Sligar JM, Jenks MX, Noe RA, Bowling TS, Mercer LT, Rewerts C, Gaukel E, Owens J, Parham R, Randolph R, Beaudet B, Bacchi CJ, Yarlett N, Plattner JJ, Freund Y, Ding C, Akama T, Zhang YK, Brun R, Kaiser M, Scandale I, Don R. 2011. SCYX-7158, an orally-active benzoxaborole for the treatment of stage 2 human African trypanosomiasis. PLoS Negl Trop Dis 5:e1151. http://dx.doi.org/10.1371/journal.pntd.0001151.
- 6. Kaiser M, Bray MA, Cal M, Bourdin Trunz B, Torreele E, Brun R. 2011. Antitrypanosomal activity of fexinidazole, a new oral nitroimidazole drug candidate for treatment of sleeping sickness. Antimicrob Agents Chemother 55:5602-5608. http://dx.doi.org/10.1128/AAC.00246-11.
- 7. Kola I, Landis J. 2004. Can the pharmaceutical industry reduce attrition rates? Nat Rev Drug Discov 3:711-715. http://dx.doi.org/10.1038 /nrd1470.
- 8. Sykes ML, Avery VM. 2009. Development of an Alamar Blue viability assay in 384-well format for high throughput whole cell screening of Trypanosoma brucei brucei bloodstream form strain 427. Am J Trop Med Hyg 81:665-674. http://dx.doi.org/10.4269/ajtmh.2009.09-0015.
- 9. Sykes ML, Baell JB, Kaiser M, Chatelain E, Moawad SR, Ganame D, Ioset JR, Avery VM. 2012. Identification of compounds with antiproliferative activity against Trypanosoma brucei brucei strain 427 by a whole cell viability based HTS campaign. PLoS Negl Trop Dis 6:e1896. http://dx.doi.org/10.1371/journal.pntd.0001896.
- 10. Don R, Ioset JR. 2014. Screening strategies to identify new chemical diversity for drug development to treat kinetoplastid infections. Parasitology 141:140–146. http://dx.doi.org/10.1017/S003118201300142X.
- 11. Brinkmann V, Billich A, Baumruker T, Heining P, Schmouder R, Francis G, Aradhye S, Burtin P. 2010. Fingolimod (FTY720): discovery and development of an oral drug to treat multiple sclerosis. Nat Rev Drug Discov 9:883-897. http://dx.doi.org/10.1038/nrd3248.

- 12. Albert R, Hinterding K, Brinkmann V, Guerini D, Muller-Hartwieg C, Knecht H, Simeon C, Streiff M, Wagner T, Welzenbach K, Zecri F, Zollinger M, Cooke N, Francotte E. 2005. Novel immunomodulator FTY720 is phosphorylated in rats and humans to form a single stereoisomer. Identification, chemical proof, and biological characterization of the biologically active species and its enantiomer. J Med Chem 48:5373-5377.
- 13. Sutterwala SS, Creswell CH, Sanyal S, Menon AK, Bangs JD. 2007. De novo sphingolipid synthesis is essential for viability, but not for transport of glycosylphosphatidylinositol-anchored proteins, in African trypanosomes. Eukaryot Cell 6:454-464. http://dx.doi.org/10.1128/EC.00283-06.
- 14. Kaneshiro ES, Jayasimhulu K, Lester RL. 1986. Characterization of inositol lipids from Leishmania donovani promastigotes: identification of an inositol sphingophospholipid. J Lipid Res 27:1294-1303.
- 15. Salto ML, Bertello LE, Vieira M, Docampo R, Moreno SN, de Lederkremer RM. 2003. Formation and remodeling of inositolphosphoceramide during differentiation of Trypanosoma cruzi from trypomastigote to amastigote. Eukaryot Cell 2:756-768. http://dx.doi.org/10.1128/EC.2.4 .756-768,2003
- 16. Patnaik PK, Field MC, Menon AK, Cross GA, Yee MC, Bütikofer P. 1993. Molecular species analysis of phospholipids from Trypanosoma brucei bloodstream and procyclic forms. Mol Biochem Parasitol 58:97-105. http://dx.doi.org/10.1016/0166-6851(93)90094-E.
- 17. Sutterwala SS, Hsu FF, Sevova ES, Schwartz KJ, Zhang K, Key P, Turk J, Beverley SM, Bangs JD. 2008. Developmentally regulated sphingolipid synthesis in African trypanosomes. Mol Microbiol 70:281-296. http://dx .doi.org/10.1111/j.1365-2958.2008.06393.x.
- 18. Jones DC, Hallyburton I, Stojanovski L, Read KD, Frearson JA, Fairlamb AH. 2010. Identification of a κ-opioid agonist as a potent and selective lead for drug development against human African trypanosomiasis. Biochem Pharmacol 80:1478-1486. http://dx.doi.org/10.1016/j.bcp.2010
- 19. Wong L, Tan SS, Lam Y, Melendez AJ. 2009. Synthesis and evaluation of sphingosine analogues as inhibitors of sphingosine kinases. J Med Chem 52:3618-3626. http://dx.doi.org/10.1021/jm900121d.
- Schnute ME, McReynolds MD, Kasten T, Yates M, Jerome G, Rains JW, Hall T, Chrencik J, Kraus M, Cronin CN, Saabye M, Highkin MK, Broadus R, Ogawa S, Cukyne K, Zawadzke LE, Peterkin V, Iyanar K, Scholten JA, Wendling J, Fujiwara H, Nemirovskiy O, Wittwer AJ, Nagiec MM. 2012. Modulation of cellular S1P levels with a novel, potent and specific inhibitor of sphingosine kinase-1. Biochem J 444:79-88. http: //dx.doi.org/10.1042/BJ20111929.
- 21. French KJ, Schrecengost RS, Lee BD, Zhuang Y, Smith SN, Eberly JL, Yun JK, Smith CD. 2003. Discovery and evaluation of inhibitors of human sphingosine kinase. Cancer Res 63:5962-5969.
- 22. Pasternack DA, Sharma AI, Olson CL, Epting CL, Engman DM. 2015. Sphingosine kinase regulates microtubule dynamics and organelle positioning necessary for proper G<sub>1</sub>/S cell cycle transmission in Trypanosoma brucei. mBio 6(5):e01291-15. http://dx.doi.org/10.1128/mBio.01291-15.
- 23. Liu H, Sugiura M, Nava VE, Edsall LC, Kono K, Poulton S, Milstien S, Kohama T, Spiegel S. 2000. Molecular cloning and functional characterization of a novel mammalian sphingosine kinase type 2 isoform. J Biol Chem 275:19513–19520. http://dx.doi.org/10.1074/jbc.M002759200.
- 24. Smith TK, Bütikofer P. 2010. Lipid metabolism in Trypanosoma brucei. Mol Biochem Parasitol 172:66-79. http://dx.doi.org/10.1016/ j.molbiopara.2010.04.001.
- 25. Center for Drug Evaluation and Research. 2009. Pharmacology review(s). Application no. 22-527. Center for Drug Evaluation and Research, U.S. Food and Drug Administration, Silver Spring, MD. http:// www.accessdata.fda.gov/drugsatfda\_docs/nda/2010/022527Orig1s000 pharmr.pdf.