



## Draft Genome Sequence of Goose Dicistrovirus

## Alexander L. Greninger, Keith R. Jerome

Department of Laboratory Medicine, University of Washington, Seattle, Washington, USA

We report the draft genome sequence of goose dicistrovirus assembled from the filtered feces of a Canadian goose from South Lake Union in Seattle, Washington. The 9.1-kb dicistronic RNA virus falls within the family *Dicistroviridae*; however, it shares <33% translated amino acid sequence within the nonstructural open reading frame (ORF) from aparavirus or cripavirus.

Received 14 January 2016 Accepted 15 January 2016 Published 3 March 2016

Citation Greninger AL, Jerome KR. 2016. Draft genome sequence of goose dicistrovirus. Genome Announc 4(2):e00068-16. doi:10.1128/genomeA.00068-16. Copyright © 2016 Greninger and Jerome. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license. Address correspondence to Alexander L. Greninger, gerbix@gmail.com.

Canadian geese (*Branta canadensis*) are well known for their prolific production of feces, depositing up to one dropping per minute and up to 1.5 lb per day (1). This fact combined with their growing populations have raised concerns over the relation of the goose population to public health. Goose feces are known to contain a number of pathogens that infect humans and affect surface waters (2–4). Exploration of the viruses of Canadian geese has led to the finding of Newcastle disease virus, influenza virus, and bornavirus (1, 5, 6).

In this study, we sequenced filtrate of a singular goose dropping from Lake Union Park in Seattle, Washington. One gram of goose feces was mixed 1:10 (weight/volume) in phosphatebuffered saline (PBS), vortexed, and syringe-filtered through a  $0.8-\mu$ m filter (Millipore). RNA from 1 mL of filtrate was extracted using the Zymo RNA miniprep kit using on-column DNase treatment. Sequencing library preparation from cDNA was performed using Nextera XT with 15 cycles of PCR amplification as described previously (7, 8). A total of 3.77 million paired-end 2- × 300-bp sequencing reads were *de novo* assembled using Abyss (9).

A 9,130-nucleotide contig was assembled that aligned by blastx to members of the *Dicistroviridae* family. The contig had  $55 \times$ coverage and contained two large open reading frames (ORFs) of 5,481 and 2,577 nucleotides, along with a 352-nucleotide intergenic region. The first ORF contained conserved domains for RNA helicase, C3G peptidase, and RNA-dependent RNA polymerase, while the second ORF contained conserved domains for picornavirus and cricket paralysis virus (CRPV) capsid proteins. The putative translated nonstructural protein gene sequence best aligned with 33% amino acid identity to cricket paralysis virus (AKA63263) while the capsid protein best aligned with 35% amino acid identity to Drosophila C virus (NP\_044946). The translated nonstructural gene was also 32% identical by amino acid to the acute bee paralysis virus polymerase protein, indicating substantial divergence from both cripavirus and asparaviruses of the Discistroviridae family.

The genome likely encodes an insect virus that is only found associated with goose feces, rather than a bona fide goose virus. Of note, a 4,177-nucleotide contig was also assembled that aligned with 96% identity by nucleotide to subterranean clover mottle virus isolate MJ (AY376453) (10). Further study will be required to decipher the true host of goose dicistrovirus, as well as charac-

terize unique coding aspects of the dicistrovirus such as the intergenic region internal ribosome binding sequence.

Nucleotide sequence accession number. The GenBank accession number for goose dicistrovirus is KT873140.

## REFERENCES

- 1. Dieter RA, Dieter RS, Dieter RA, Gulliver G. 2001. Zoonotic diseases: health aspects of Canadian geese. Int J Circumpolar Health 60:676–684.
- Graczyk TK, Fayer R, Trout JM, Lewis EJ, Farley CA, Sulaiman I, Lal AA. 1998. Giardia sp. cysts and infectious *Cryptosporidium parvum* oocysts in the feces of migratory Canada geese (*Branta canadensis*). Appl Environ Microbiol 64:2736–2738.
- 3. Hussong D, Damaré JM, Limpert RJ, Sladen WJ, Weiner RM, Colwell RR. 1979. Microbial impact of Canada geese (*Branta canadensis*) and whistling swans (*Cygnus columbianus columbianus*) on aquatic ecosystems. Appl Environ Microbiol 37:14–20.
- Alderisio KA, DeLuca N. 1999. Seasonal enumeration of fecal coliform bacteria from the feces of ring-billed gulls (*Larus delawarensis*) and Canada geese (*Branta canadensis*). Appl Environ Microbiol 65:5628–5630.
- Guo J, Baroch J, Randall A, Tizard I. 2013. Complete genome sequence of an avian bornavirus isolated from a healthy Canadian goose (*Branta canadensis*). Genome Announc 1(5):e00839-13. http://dx.doi.org/ 10.1128/genomeA.00839-13.
- Pasick J, Berhane Y, Embury-Hyatt C, Copps J, Kehler H, Handel K, Babiuk S, Hooper-McGrevy K, Li Y, Mai Le Q, Lien Phuong S. 2007. Susceptibility of Canada geese (*Branta canadensis*) to highly pathogenic avian influenza virus (H5N1). Emerg Infect Dis 13:1821–1827. http:// dx.doi.org/10.3201/eid1312.070502.
- Greninger AL, Naccache SN, Messacar K, Clayton A, Yu G, Somasekar S, Federman S, Stryke D, Anderson C, Yagi S, Messenger S, Wadford D, Xia D, Watt JP, Van Haren K, Dominguez SR, Glaser C, Aldrovandi G, Chiu CY. 2015. A novel outbreak enterovirus D68 strain associated with acute flaccid myelitis cases in the USA (2012–14): a retrospective cohort study. Lancet Infect Dis 15:671–682. http://dx.doi.org/10.1016/S1473 -3099(15)70093-9.
- Greninger AL, Messacar K, Dunnebacke T, Naccache SN, Federman S, Bouquet J, Mirsky D, Nomura Y, Yagi S, Glaser C, Vollmer M, Press CA, Klenschmidt-DeMasters BK, Dominguez SR, Chiu CY. 2015. Clinical metagenomic identification of *Balamuthia mandrillaris* encephalitis and assembly of the draft genome: the continuing case for reference genome sequencing. Genome Med 7:113. http://dx.doi.org/10.1186/s13073 -015-0235-2.
- Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJM, Birol I. 2009. ABySS: a parallel assembler for short read sequence data. Genome Res 19:1117–1123. http://dx.doi.org/10.1101/gr.089532.108.
- Dwyer GI, Njeru R, Williamson S, Fosu-Nyarko J, Hopkins R, Jones RA, Waterhouse PM, Jones MGK. 2003. The complete nucleotide sequence of subterranean clover mottle virus. Arch Virol 148:2237–2247. http://dx.doi.org/10.1007/s00705-003-0144-3.