

Whole-Genome Sequencing Analysis of *Chromobacterium piscinae* Strain ND17, a Quorum-Sensing Bacterium

 Kok-Gan Chan, Nina Yusrina Muhamad Yunos

Division of Genetics and Molecular Biology, Institute of Biological Sciences, Faculty of Science, University of Malaya, Kuala Lumpur, Malaysia

Here, we report the draft genome sequence of *Chromobacterium piscinae* strain ND17. This bacterium was isolated from a fresh water sample in Malaysia and exhibits quorum-sensing activity. This first draft genome of *C. piscinae* strain ND17 will pave the way to future studies of the quorum-sensing properties of this isolate.

Received 17 January 2016 Accepted 20 January 2016 Published 3 March 2016

Citation Chan K-G, Yunos NYM. 2016. Whole-genome sequencing analysis of *Chromobacterium piscinae* strain ND17, a quorum-sensing bacterium. *Genome Announc* 4(2):e00081-16. doi:10.1128/genomeA.00081-16.

Copyright © 2016 Chan and Yunos. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Kok-Gan Chan, kokgan@um.edu.my.

Bacteria communicate to control physiological activities such as virulence determinants, competence, symbiosis, antibiotic production, and biofilm formation (1). This bacterial cell-to-cell communication is known as “quorum sensing” (QS), a term first introduced by Fuqua and colleagues (2, 3). QS is achieved by generation and response of QS signaling molecules which refer to small, self-generated signal molecules (4). QS is a function that is population density dependent (5).

Chromobacterium piscinae is a Gram-negative, aerobic, rod-shaped, motile bacterium with violet pigmentation (6). In this study, we report the whole genome of *Chromobacterium piscinae* strain ND17. *C. piscinae* strain ND17 was isolated from a fresh water sample in Malaysia. Genomic DNA was extracted by using a MasterPure DNA purification kit (Epicentre, Inc., Madison, WI, USA). Next, a NanoDrop spectrophotometer (Thermo Scientific, Waltham, MA, USA) and Qubit 2.0 fluorometer (Life Technologies, Carlsbad, CA, USA) were used to determine the quality of the DNA extracted. An Illumina MiSeq sequencer (Illumina, Inc., San Diego, CA, USA) was used to sequence the bacterium genome. An average coverage of 38.5-fold was obtained for the draft genome of 4,089,562 bp in 223 contigs with an N_{50} of 43,813. The paired-end reads were trimmed and *de novo* assembled with CLC Genomic Workbench version 5.1 (CLC Bio, Denmark). Prodigal (version 2.60) was used for gene prediction (7) and Rapid Annotation Sub-system Technology (RAST) was used for gene annotation (8). tRNA was predicted with tRNAscan SE version 1.21 (9) and rRNA with RNAmmer (10).

The G+C content of the *C. piscinae* strain ND17 genome is 62.6%. The total number of predicted genes is 3,916, of which 3,506 are protein coding genes with predicted function numbers, equivalent to approximately 90% of the total number of predicted genes. A total of 81 tRNAs, two copies of 5S rRNA, and a copy of each 16S rRNA and 23S rRNA were predicted for strain ND17. Based on the annotation result, the *C. piscinae* strain ND17 genome is comprised of 89 genes responsible for virulence, disease, and defense, where most of the genes are connected with antibiotics and toxic compounds resistance. This is the first finding reported on QS of *C. piscinae*. We hope

this annotated genome of strain ND17 will be a valuable tool for better understanding the QS mechanism of *C. piscinae* strain ND17.

Nucleotide sequence accession numbers. This whole-genome-shotgun project has been deposited at DDBJ/EMBL/GenBank under accession no. [JTGE000000000](https://www.ncbi.nlm.nih.gov/nuccore/JTGE000000000). The version described in this paper is the first version, JTGE01000000.

FUNDING INFORMATION

This work was supported by the University of Malaya via High Impact Research Grants (UM.C/625/1/HIR/MOHE/CHAN/01, grant no. A-000001-50001, and UM-MOHE HIR Grant UM.C/625/1/HIR/MOHE/CHAN/14/1, grant no. H-50001-A000027), which have been awarded to Kok-Gan Chan.

REFERENCES

- Miller MB, Bassler BL. 2001. Quorum sensing in bacteria. *Annu Rev Microbiol* 55:165–199. [http://dx.doi.org/10.1146/annurev.micro.55.1.165](https://doi.org/10.1146/annurev.micro.55.1.165).
- Fuqua WC, Winans SC, Greenberg EP. 1994. Quorum sensing in bacteria: the LuxR-LuxI family of cell density-responsive transcriptional regulators. *J Bacteriol* 176:269–275.
- Yunos NYM, Tan WS, Koh CL, Sam CK, Mohamad NI, Tan PW, Adrian T-G-S, Yin WF, Chan KG. 2014. *Pseudomonas cremoricolorata* strain ND07 produces *N*-acyl homoserine lactones as quorum sensing molecules. *Sensors* 14:11595–11604.2 [http://dx.doi.org/10.3390/s140711595](https://doi.org/10.3390/s140711595).
- De Kievit TR, Iglewski BH. 2000. Bacterial quorum sensing in pathogenic relationships. *Infect Immun* 68:4839–4849. [http://dx.doi.org/10.1128/IAI.68.9.4839-4849.2000](https://doi.org/10.1128/IAI.68.9.4839-4849.2000).
- Liu YC, Chan KG, Chang CY. 2015. Modulation of host biology by *Pseudomonas aeruginosa* quorum sensing signal molecules: messengers or traitors. *Front Microbiol* 6:1226. [http://dx.doi.org/10.3389/fmicb.2015.01226](https://doi.org/10.3389/fmicb.2015.01226).
- Kämpfer P, Busse HJ, Scholz HC. 2009. *Chromobacterium piscinae* sp. nov. and *Chromobacterium pseudoviolaceum* sp. nov., from environmental samples. *Int J Syst Evol Microbiol* 59:2486–2490. [http://dx.doi.org/10.1099/ijs.0.008888-0](https://doi.org/10.1099/ijs.0.008888-0).
- Hyatt D, Chen GL, LoCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. [http://dx.doi.org/10.1186/1471-2105-11-119](https://doi.org/10.1186/1471-2105-11-119).
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formisano K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson

- R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotation using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
9. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 25:955–964. <http://dx.doi.org/10.1093/nar/25.5.0955>.
10. Lagesen K, Hallin P, Rødland EA, Staerfeldt H-H, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.