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Physiological effects and mechanisms of action of endocrine disrupting chemicals that alter estrogen signaling

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INTRODUCTION

Numerous natural and synthetic chemicals have been identified that are reported to disrupt the normal functioning of the endocrine system and produce untoward effects in hormoneresponsive target tissues and organs in both humans and animals. These chemicals are broadly referred to as endocrine disrupting chemicals (EDCs) and consist of numerous physically and chemically distinct compounds. The physicochemical differences among EDCs are paralleled by unique biological effects and mechanisms of action within this class of compounds. The purpose of this review is to provide an overview of the physiological effects and unique mechanisms of action of four distinct EDCs that alter estrogen signaling. Two of these compounds, diethylstilbestrol (DES) and genistein (GEN), have been extensively studied and serve as classic examples of EDCs, while the other two compounds, methoxyacetic acid (MAA) and the essential oils of lavender and tea tree, have only recently been described as EDCs.

DIETHYLSTILBESTROL

Diethylstilbestrol (DES) is a nonsteroidal synthetic estrogen developed by Sir Charles Dodds and colleagues in 1938.¹ During the late 1940's physicians began using DES to treat pregnant women in an attempt to maintain normal placental steroid levels and to prevent miscarriages and premature births.² It was not until 1953 that a randomized controlled clinical trial evaluating the effectiveness of DES in preventing miscarriage and premature birth was reported.³ Results from this study showed that DES offered no protective effects with respect to miscarriage or premature birth. Despite the outcome of this clinical trial, DES continued to be prescribed to pregnant women until 1971, when it was reported that *in utero* DES exposure was associated with a rare reproductive tract cancer, termed vaginal

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clear cell adenocarcinoma, in approximately 0.1% of daughters of women who had taken the drug.^{4,5} As a result of this association the FDA advised against using DES in pregnant women in 1971. Subsequent studies have reported multiple adverse effects in males and females as a result of prenatal DES exposure that occur with greater frequency than clear cell adenocarcinoma. In females, these include anatomical malformations of the cervix, vagina, and uterus, decreased fertility, and fewer successful pregnancies.^{6–9} In contrast to females, no increased incidence of cancer has been reported in males prenatally exposed to DES. However, adverse effects have been observed in their reproductive tracts, including testicular hypoplasia, cryptorchidism, and epididymal cysts.^{6,7,10}

Epidemiological analyses continue for those men and women exposed to DES *in utero* as physicians and scientists strive to better understand the long-term effects in humans. These efforts have been accompanied by intensive laboratory-based research to elucidate the mechanisms by which DES elicits its carcinogenic and teratogenic effects and to better predict and prevent the untoward effects of *in utero* DES exposure in humans. Toward this end, a variety of rodent models have been developed to study the mechanism of action and effects of DES based primarily on different potentials as a chemical substance, pharmaceutical or hormonal agent. One well characterized model for DES exposure is the neonatal mouse model in which female and male pups are treated with DES (2µg/day) for the first five days of life and aged up to 18 months. DES treatment of female mice in this model produces a high incidence of reproductive tract tumors.^{11,12} The neonatal mouse model and other, previously utilized, rodent models were limited in that they could not elucidate the role of the estrogen receptor (ER) or DES as a hormonal substance in DES-induced toxicity.

The generation of mice lacking either estrogen receptor α (ER α) or estrogen receptor β (ER β), i.e. ER knockout (ERKO) mice, and their incorporation into the neonatal mouse model of DES exposure allowed for examination of the role of the ER subtypes in mediating the effects of DES *in vivo*.^{13,14} Wild type (WT) female mice treated neonatally with DES displayed characteristic DES-induced reproductive tract lesions (Table 1).¹⁵ Treatment resulted in atrophy, smooth muscle disorganization, hyalinization, squamous metaplasia of the glandular epithelium, and endometrial hyperplasia. In the vagina, persistent epithelial cornification and vaginal adenosis were observed in a small percentage of animals following DES treatment. Finally, DES induced progressive proliferative lesions of the epithelium in the oviduct. None of these reproductive tract lesions were observed in identically treated α ERKO female mice, indicating that DES toxicity is mediated via an ER α -dependent signaling pathway.^{15,16}

Similar experiments utilizing the neonatal mouse model of DES exposure were conducted with WT, α ERKO, and β ERKO male mice to determine if DES-induced effects in the male reproductive tract are ER-mediated. When WT males are exposed neonatally to DES, they exhibit significant decreases in seminal vesicle size at all ages, while no changes in seminal vesicle size are observed in identically treated α ERKO males and males treated with oil alone (Figure 1).¹⁷ With respect to the prostate gland which expresses clearly measureable amounts of both ER α and ER β , DES treatment of neonatal WT and β ERKO mice produced histological changes in the aged ventral and dorsolateral prostates indicative of the

phenotype of an estrogenized prostate gland, including increased stromal mass, epithelial hyperplasia and dysplasia, and interstitial lymphocyte infiltration.¹⁸ In addition, DES induced abnormal epithelial cell differentiation indicative of estrogen imprinting in the dorsolateral prostate of WT and β ERKO mice, including the presence of a continuous layer of basal cells lining the epithelial ducts and acini, luminal cell hyperplasia, and loss of expression of the functional differentiation marker dorsolateral prostate-2 protein in the dorsolateral prostate.¹⁷ None of these effects were observed in the prostate glands of identically treated α ERKO mice, indicating that a functional ER α is required for DES-mediated activity. Collectively, these data suggest that DES elicits its toxic effects in the male reproductive tract through an ER α -mediated mechanism.

Having confirmed the necessary role of ER α in mediating the DES-induced phenotypic changes in the reproductive tracts of female and male mice, studies have focused on elucidating the mechanisms underlying these changes. One area of focus in this regard has been the examination of DES-induced changes in the expression of genes in the Hox and Wnt families that are known to be critical for the normal differentiation and organization of the reproductive tract. Several lines of evidence suggest that altered gene expression in the *Hox* and *Wnt* families may be central to the DES-induced effects on the reproductive tract. First, their expression in the paramesonephric duct is essential for the development and organization of the female reproductive tract.¹⁹ Second, null mouse models of the Hox and Wnt gene families exhibit reproductive tract phenotypes similar to those of DES-treated mice.^{20–23} Finally, DES treatment of neonatal mice reduces the expression of Hoxa9, Hoxa10, Hoxa11, and Wnt7a in the female reproductive tract.^{22,24,25} Based on this evidence, studies were conducted with aERKO mice in the neonatal mouse model of DES exposure to determine the role of ERa in mediating DES-induced decreases in Hox and Wnt gene expression. Results from these studies showed that the uteri of 5-day old WT mice neonatally exposed to DES had an approximately 50% reduction in Hoxa10 and Hoxa11 mRNA levels, and a greater than 80% reduction in Wnt7a mRNA levels when compared to their expression levels in the uteri of control animals.¹⁵ In the uteri of DES treated aERKO mice, the expression levels of Hoxa10, Hoxa11, and Wnt7a were similar to those measured in the uteri of control mice, indicating that ERa mediates the DES-induced reduction in expression of these genes (Figure 2). These results substantiate the central role that $ER\alpha$ plays in mediating the *in vivo* effects of DES and provide further support for the argument that DES-induced alterations in *Hox* and *Wnt* gene expression are at least partially responsible for the reproductive tract abnormalities observed in DES-treated mice.

The molecular mechanisms underlying DES-induced decreases in *Hox* and *Wnt* gene expression have yet to be fully characterized. One proposed mechanism put forth involved epigenetic modifications such as DNA methylation, since DES increases lactoferrin and *c*-*fos* gene expression in the mouse uterus through demethylation of the lactoferrin promoter and hypomethylation of exon 4 of the *c*-*fos* gene.^{26,27} Subsequent studies showed that DES had no effect on promoter methylation of the *Hoxa10* and *Hoxa11* genes, indicating that DES utilizes a different mechanism to decrease expression of these genes.²⁸ Subsequent studies examining the *Hoxa10* promoter identified a nonconsensus estrogen response element (ERE) that is differentially regulated by estradiol and DES.²⁹ Results from these

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studies showed that the maximal estradiol-induced luciferase activity was four times greater than that induced by DES in cells transfected with a luciferase reporter plasmid containing the *Hoxa10* nonconsensus ERE. These results suggest that the nonconsensus *Hoxa10* ERE is induced in a ligand-specific manner, which may be due to distinct conformational changes in the ER as a result of the unique ligand structures and/or the *Hoxa10* nonconsensus ERE. Furthermore, these results may provide the mechanistic explanation for decreased *Hoxa10* expression and the subsequent distinct phenotypes observed in the female reproductive tract following neonatal DES treatment. Whether a similar mechanism is responsible for DESinduced downregulation of *Hoxa11* and *Wnt7a* expression remains to be determined.

GENISTEIN

Members of the phytoestrogen family of nonsteroidal, plant-derived, compounds modulate ER signaling and therefore are potentially endocrine disrupting chemicals.³⁰ The potential clinical application of phytoestrogens for prevention of hormone-dependent cancers and for lowering cholesterol levels has prompted extensive research aimed at elucidating the physiological effects of this class of compounds. The isoflavone genistein is a member of the phytoestrogen family and is found at high concentrations in soy-based products. Both in vitro and in vivo studies have clearly demonstrated that genistein exhibits estrogenic activity and tyrosine kinase inhibitory activity; however, the potential therapeutic benefits of genistein are not clear due to conflicting study results.^{31–34} For example, genistein has been shown to reduce mammary cancer rates in rats in some studies, while in other studies it has been shown to increase mammary cancer rates in rats.^{35–37} Additional animal studies have linked genistein to diminished reproductive capacity^{38,39} and, in a neonatal mouse model, genistein has been shown to induce uterine adenocarcinomas.⁴⁰ The potential endocrine disrupting activity of genistein, coupled with the knowledge that humans are exposed to genistein through their diet, has stimulated interest in characterizing the biological effects of genistein on humans. While genistein has been shown to lower cholesterol levels in humans, little else is known regarding the beneficial and/or detrimental effects of human exposure to genistein.^{41,42} Exposure to genistein in infancy is of particular interest, as infants fed soy formula consume 6–9 mg/kg per day of isoflavones, approximately 65% of which is genistein, compared to 1 mg/kg per day for adults.⁴³ This genistein level is 6–11 times higher than the amount reported to produce hormonal effects that alter the menstrual cycle of adult women.⁴⁴ The presence of genistein in the human diet and the lack of knowledge regarding its definitive impact on human health have prompted investigations utilizing animal models to elucidate the mechanism of action of genistein and the biological consequences of exposure.

To elucidate the mechanistic aspects of genistein signaling, estrogen receptor binding assays have been performed to determine if genistein directly binds to ER. Results from these analyses show that genistein has a 20- to 30-fold higher affinity for ER β than it does for ER α .^{45,46} However, *in vitro* assays demonstrate that genistein has only a slight preference for transactivation of gene expression through ER β when compared to ER α , indicating that genistein is able to signal through both ER α and ER β .⁴⁶ To clarify the role of the ER subtypes in genistein signaling *in vivo*, neonatal mouse models have been utilized to examine its effects on the uterus, which predominantly expresses ER α . In this model,

genistein elicited a classic uterotropic response including increased uterine wet weight, epithelial cell height, gland number, and lactoferrin expression.⁴⁷ Given the predominant expression of ER α in the mouse uterus, these results indicate that genistein can effectively signal through ER α *in vivo*. Further support for the role of ER α in genistein signaling is derived from studies showing that genistein activates the IGF-1 signaling pathway in the mouse uterus via an ER α -dependent mechanism.⁴⁸ Collectively, these data demonstrate that ER α mediates genistein signaling in the mouse uterus. Further studies incorporating the α ERKO mice complement the above observations by providing direct evidence that ER α is necessary for genistein significantly increased uterine wet weights in ovariectomized WT mice, while neither compound increased uterine wet weights in α ERKO mice (Figure 3, unpublished data).

The mouse ovary, in contrast to the uterus which predominantly expresses $ER\alpha$, expresses both ER α and ER β and provides a suitable model to determine the role of ER β in mediating genistein signaling. As a result, in vivo studies have been performed to examine the effect of genistein on the mouse ovary. Results from these studies demonstrated that genistein induced a dose-dependent increase in multi-oocyte follicles in WT CD-1 mice treated for 5 days with genistein when compared to vehicle controls (Table 2).⁴⁹ Given the established tyrosine kinase inhibitory activity of genistein, the nonestrogenic tyrosine kinase inhibitor lavendustin A was incorporated into these experiments to confirm that the results were due solely to the associated estrogenic activity of genistein. No multi-oocyte follicles were observed in mice treated with two different concentrations of lavendustin A, indicating that the genistein-induced increases in multi-oocyte follicles were due to its estrogenic activity and not to tyrosine kinase inhibition. Further experiments were conducted in C57BL/6, α ERKO, and β ERKO mice to determine if genistein signaling in the ovary is mediated by ER α , ER β or a non-receptor-mediated mechanism. Results from these studies showed that genistein induced a concentration-dependent increase in multi-oocyte follicles in C57BL/6 and aERKO mice, while the incidence of multi-oocyte follicles in BERKO mice was significantly lower.⁴⁹ These results indicate that ER β is necessary for the induction of multioocyte follicles in genistein-treated neonatal mice.

Taken together, these *in vivo* data show that genistein utilizes both ER α - and ER β -mediated mechanisms depending on the specific tissue to elicit estrogenic effects on the female mouse reproductive tract. While these models have proven useful in uncovering the effects and mechanism of action of genistein on the mouse reproductive tract, extrapolation of these results to humans has been hampered by conflicting data obtained from epidemiological studies. Therefore, continued efforts to elucidate the biological effects and mechanism of action of genistein are needed, as are further epidemiological studies, to obtain a clearer picture of the beneficial and detrimental effects of genistein exposure in humans.

METHOXYACETIC ACID

Methoxyacetic acid (MAA) is the major metabolite of the industrial solvent ethylene glycol monomethyl ether (EGME), used in paints, varnishes, dyes, and fuel additives. Occupational exposure to EGME, and consequently to its major metabolite MAA, have been associated

with reproductive toxicity. In women, exposure to ethylene glycol ethers has been associated with increased risk of spontaneous abortion and subfertility, while it is associated with decreased sperm counts in exposed males.^{50–56} Laboratory studies have demonstrated that EGME targets the ovarian luteal cell, suppresses cyclicity, and inhibits ovulation in female rats, while in male rats it reduces testicular size and fertility.^{57,58} Subsequent studies have established that MAA is responsible for the reproductive toxicities associated with EGME exposure.^{53,57}

MAA is a member of the short-chain fatty acid family, which includes the antiepileptic drug valproic acid (VPA) and the intestinal bacterial product sodium butyrate (NaB). Interestingly, VPA has also been associated with reproductive toxicities, including menstrual abnormalities and polycystic ovaries.^{59–61} One shared feature of MAA, VPA, and NaB is their ability to inhibit histone deacetylases and alter gene expression via histone hyperacetylation.^{62–66} However, what role this effect plays in the reproductive toxicities associated with MAA and VPA is unknown.

The reproductive toxicities associated with MAA exposure in humans and animals coupled with the critical role of estrogen signaling in normal male and female reproductive function suggest that MAA may disrupt normal estrogen signaling. In addition, the aforementioned reproductive toxicities are similar to the reproductive phenotypes observed in aERKO and aromatase knockout mice, further suggesting that MAA may disrupt estrogen signaling. Therefore, studies have been conducted to characterize the effects of MAA on estrogen signaling *in vitro* and *in vivo*. MAA has been shown to enhance exogenous ER signaling *in* vitro by transiently transfecting an ER expression vector and a luciferase reporter into HeLa cells and treating them with estradiol in the presence and absence of MAA.⁶² These results have been confirmed in our laboratory by transfecting ER-negative HeLa cells with an expression vector for either ER α or ER β and the estrogen-inducible 3x-ERE-TATA-Luc firefly luciferase reporter plasmid. As shown in Figure 4A and 4B, MAA potentiates the activity of estradiol in HeLa cells transfected with the luciferase reporter and either ERa or ER^{β.67} These data are incongruent with the reproductive toxicities observed following MAA exposure; therefore, additional experiments were performed to examine the effect of MAA on endogenous ER signaling. Interestingly, when ER-positive MCF-7 cells were transfected with the 3x-ERE-TATA-Luc reporter plasmid, but no ER expression vector, and treated identically to the HeLa cells, no potentiation of E2-induced luciferase activity was observed (data not shown).⁶⁷ However, when an ER expression vector was cotransfected into the MCF-7 cells, MAA was able to potentiate the estrogen-stimulated luciferase activity. These results demonstrate that MAA differentially affects exogenous and endogenous ER signaling. Further experiments showed that MAA dose-dependently increases exogenous ERa protein expression levels in HeLa cells in the presence or absence of estradiol, which is due to activation of the CMV promoter contained in the expression vectors for both ERa and ER β (Figure 4C, 4D).⁶⁷ In contrast to its effects on the exogenous ER, MAA decreases endogenous ERa protein and mRNA expression levels by approximately 50% in MCF-7 cells treated with 5mM MAA for 24h when compared to vehicle controls (Figure 5A and 5B).⁶⁷ A similar effect was observed in the uteri of ovariectomized C57BL/6 mice treated with 400mg/kg MAA for 2.5h, as ERa mRNA levels decreased by approximately 30%

following MAA treatment when compared to vehicle controls.⁶⁷ Taken together, these data illustrate that MAA has similar effects on endogenous ERa expression *in vitro* and *in vivo*.

Subsequent to these studies, gene expression experiments were performed to determine if the observed decrease in ER α expression correlated with alterations in estrogen signaling. In MCF-7 cells, pre-treatment with 5mM MAA followed by 1nM estradiol attenuated the expression of the estrogen-regulated genes pS2, MYC, SPUVE, GREB1, and MCM3 when compared to treatment with 1nM estradiol alone (Figure 6).⁶⁷ MAA had a similar effect on estrogen-modulated gene expression *in vivo*, as ovariectomized C57BL/6 mice treated with 400mg/kg MAA and 1µg/kg estradiol had decreased uterine expression of the estrogen-inducible genes *Greb1*, *Inhbb*, and *Fos* when compared to uterine expression in mice treated with estradiol alone.⁶⁷ These studies show that MAA attenuates estradiol-mediated signaling *in vitro* and *in vivo* and illustrate the functional consequences of MAA-induced decreases in ER α expression. The MAA-mediated attenuation of estrogen-inducible gene expression is likely due to the observed decrease of ER expression following MAA treatment, as binding experiments have demonstrated that MAA does not compete with estradiol for binding to the ER.

Collectively, these data confirm that MAA disrupts ER signaling through a mechanism that involves decreased endogenous ERa expression. These results are consistent with the reproductive toxicities associated with EGME exposure and provide some insight into the mechanism of action of MAA with regard to reproductive toxicity, which, based on these results, include disrupted ER signaling as a result of decreased ER expression. These results also provide a framework for future studies to examine the impact of MAA on ER expression and signaling in other parts of the reproductive tract and to further pursue the mechanism(s) of action underlying MAA-induced reproductive toxicity.

LAVENDER OIL AND TEA TREE OIL

Lavender oil and tea tree oil are essential oils frequently used in over-the-counter health and beauty products, including topically applied lotions, gels, and creams, to impart pleasant aromas and for a variety of purported health benefits. Consequently, topical exposure to essential oils is widespread; however, little is known regarding the acute and long-term effects of such exposure in humans. Clinical evidence suggesting that lavender oil and tea tree oil may possess endocrine disrupting activity first surfaced with the report of three patients presenting with idiopathic male prepubertal gynecomastia, all of whom had topically applied products containing either lavender oil or a combination of lavender and tea tree oil.⁶⁸ Gynecomastia is clinically characterized by enlarged breast tissue and is the result of disrupted estrogen and androgen signaling which lead to increased or unopposed estrogen action on breast tissue.⁶⁹ Although pubertal male gynecomastia is common, an underlying cause is not identified in most cases (idiopathic). Hence pubertal gynecomastial does not usually require investigation. Pre-pubertal gynecomastia, nevertheless, is rare and must be investigated. This is the reason why the three boys were thoroughly investigated for the gynecomastia.^{70,71} These three male prepubertal patients with gynecomastia had normal low serum levels of endogenous sex steroids and no apparent underlying pathologies. One common feature among the three boys was that each of them had topically applied over-the-

counter products containing lavender oil, and in one case a product contained both lavender oil and tea tree oil. In all three boys, the onset of gynecomastia coincided with the topical application of these products and resolved with the discontinuation of their use, suggesting that lavender oil, and perhaps tea tree oil, may possess endocrine disrupting activity that leads to the development of gynecomastia.

These clinical case reports, coupled with the link between gynecomastia and disrupted sex steroid signaling, prompted studies to characterize the effects of lavender oil and tea tree oil on estrogen and androgen signaling. Estrogenic activity was assessed by conducting doseresponse experiments in MCF-7 cells transfected with the estrogen-inducible 3x-ERE-TATA-Luc reporter plasmid. Both lavender oil and tea tree oil induced ERE-dependent luciferase activity in a dose-dependent manner, with maximal activity observed at 0.025% vol/vol for each oil that corresponded to approximately 50% of the activity induced by 1nM estradiol (Figure 7).⁶⁸ This effect was attenuated in the presence of the pure ER antagonist fulvestrant, indicating that the estrogenic activity of the oils is ER-dependent. Further studies showed that both lavender oil and tea tree oil modulate the endogenous expression of the estrogen-responsive genes MYC, CTSD, and IGFBP-3 in MCF-7 cells in a manner similar to that observed with 1nM estradiol with respect to both the magnitude and timing of the response (Figure 8).⁶⁸ These responses were ER-dependent as well, as they were attenuated in the presence of fulvestrant. The estrogenic activity of tea tree oil has been independently confirmed in a separate study that examined the effect of tea tree oil in MCF-7 proliferation assays.⁷² Results from these studies show that tea tree oil induced a significant dose-dependent estrogenic response in vitro, with the maximal proliferative response occurring at a concentration of 0.0125% that corresponded to 34% of the maximal estradiol response in the assays.

Since altered androgen signaling can also contribute to the development of gynecomastia, studies were conducted to characterize the effect of lavender oil and tea tree oil on androgen signaling. The MDA-kb2 breast cancer cell line, which expresses AR and has been stably transfected with an androgen-inducible mouse mammary tumor virus promoter, was incorporated into these experiments to evaluate the androgenic activity of both oils.⁷³ While increased luciferase activity was observed following treatment of MDA-kb2 cells with the AR agonist dihydrotestosterone (DHT), no increase in luciferase activity was observed following exposure to lavender oil or tea tree oil regardless of concentration, indicating that the oils do not possess androgenic activity.⁶⁸ However, both oils elicited a concentrationdependent inhibition of DHT-induced luciferase activity, indicating that both oils possess antiandrogenic activity (Figure 9).⁶⁸ This antiandrogenic effect extended to the DHTinduced expression of the androgen-responsive endogenous genes CYP4F8, C1orf116, UGT2B28, and SEC14L2 in MDA-kb2 cells treated with 0.1nM DHT and either lavender oil or tea tree oil (Figure 10).⁶⁸ Expression levels of AR mRNA and protein were not altered by either oil, indicating that the observed antiandrogenic effect of the oils is not due to downregulation of AR expression.

Taken together, these data illustrate that both lavender and tea tree oil possess estrogenic and antiandrogenic properties, and this activity may underlie the onset of prepubertal gynecomastia in the three patients described earlier with idiopathic prepubertal

gynecomastia. The endocrine disrupting activity of these oils may be shared with other essential oils as well, as estrogenic and antiandrogenic properties have been reported for other essential oils and some of their monoterpene constituents.^{74–79} In light of these data and the widespread use of essential oils in over-the-counter health and beauty products, additional studies should be conducted to more fully characterize their endocrine disrupting properties. One prominent challenge with respect to such studies is that essential oils contain numerous components, most of which are monoterpenes. Therefore, identifying which component or components of the oils possess the endocrine disrupting activity is difficult. However, identification of these components would greatly facilitate identification of other oils that may possess similar endocrine disrupting activity, as their presence in an essential oil would suggest potential endocrine disrupting activity.

CONCLUSIONS

In this review we have provided examples of the activity and actions of several different EDC type compounds of varying chemical structure, properties, and sources. Findings from *in vitro*, cell based assays, and/or experimental animal studies have provided evidence of their hormonal biological actions involving receptor mediated activities as a mechanism for their toxicities. Additionally, an example is given of a unique EDC clinical observation regarding the properties of some essential oils. As expected, further observations of other types of toxicities and development of additional experimental models and systems will be required for the evaluation of the mechanisms involved. Finally, the impact that these exposures may have on the human population will require appreciation of a potential chemical effects from certain exposure as well as broader epidemiological assessments in support of the experimental findings.

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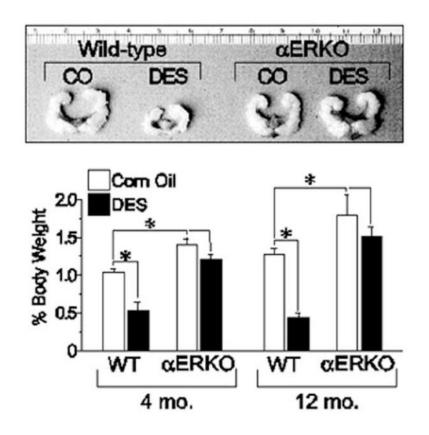


Figure 1.

Effect of neonatal DES exposure on seminal vesicle weight in adult WT and αERKO mice. *Top*, seminal vesicles from 6-month old WT and αERKO mice following neonatal exposure to either vehicle (corn oil, CO) or DES (scale is in centimeters). *Bottom*, quantitative analysis of seminal vesicle weights from WT and *α*ERKO mice at 4 and 12 months of age following neonatal exposure to CO or DES. *, P<0.05. (Modified from Prins, G.S., Birch, L., Couse, J.F., Choi, I., Katzenellenbogen, B., and Korach, K.S., 2001 Cancer Res., 61: 6089-6097.)

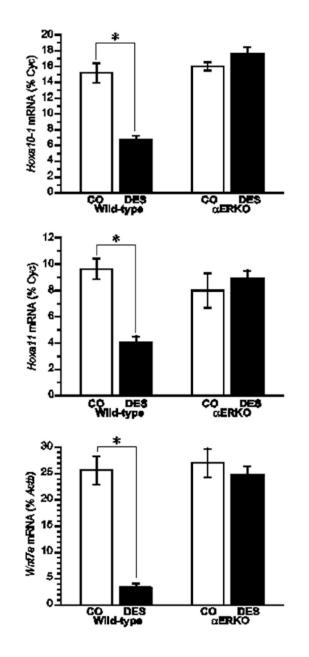


Figure 2.

Effect of neonatal DES exposure on uterine expression of *Hoxa10*, *Hoxa11*, and *Wnt7a* in WT and αERKO mice. *Top and middle*, Quantitative analyses of ribonuclease protection assays (*Hoxa10* and *Hoxa11*) (+/– SEM) showing average percentage of cyclophilin (normalization mRNA) for each treatment group. *Bottom*, Quantitative analysis of semiquantitative RT-PCR showing the average *Wnt7a* levels as a percentage of *Actb* (+/– SEM) for each treatment group. *, P<0.01. (Modified from Couse, J.F., Dixon, D., Yates, M., Moore, A.B., Ma, L., Maas, R., and Korach, K.S., 2001 Dev. Biol., 238: 224-238.).

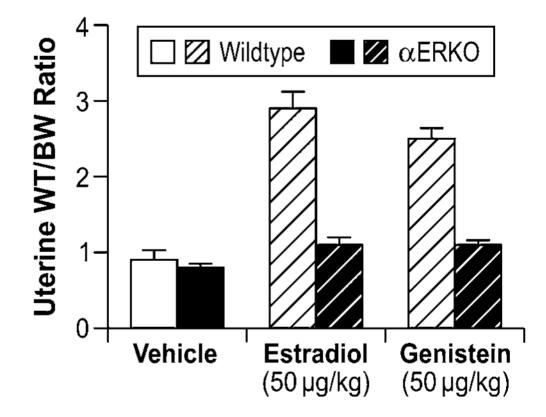


Figure 3.

Role of ER α in modulating genistein-induced uterotropic effects. Ovariectomized WT and α ERKO mice (n=4 mice/treatment group) were treated daily with subcutaneous injections of corn oil (vehicle), 10µg/kg estradiol, or 50µg/kg genistein for three days. On the fourth day whole body weights were measured and uteri were collected and weighed. The data represent uterine wet weight with respect to whole body weight. (Unpublished data from Lindzey, J. and Korach, K.S.).

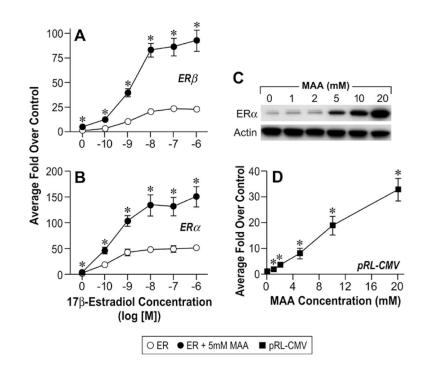


Figure 4.

MAA activates the CMV promoter in vitro. A. HeLa cells were transiently transfected with a human ERβ expression vector, the estrogen-responsive 3X-ERE-TATA-Luc reporter plasmid, and the CMV-\beta-gal reporter plasmid and treated for 18h with either vehicle, increasing concentrations of E2, 5mM MAA, or increasing concentrations of E2 plus 5mM MAA. The histogram represents the average fold over control of duplicate samples from three independent experiments. The error bars represent the standard error of the mean (SEM), *p<0.05 comparing identical treatments +/- 5mM MAA. B. HeLa cells were transiently transfected with a human ERa expression vector, 3X-ERE-TATA-Luc and CMV- β -gal and treated identically to the cells in Figure 1A. The data shown represent the average fold over control of duplicate samples from three independent experiments and the error bars indicated the SEM, p<0.05 comparing identical treatments +/-5mM MAA. C. HeLa cells were transiently transfected with a human ERa expression vector and treated with either vehicle or increasing concentrations of MAA for 18h. Total protein was isolated from the cells and ERa protein expression was analyzed by Western blot. The data shown are representative of three independent experiments. D. HeLa cells were transiently transfected with the pRL-CMV reporter plasmid and treated with either vehicle or increasing concentrations of MAA for 18h. The data shown represent the average fold over control of duplicate samples from three independent experiments. The error bars represent the SEM, *p<0.01 comparing treatment with MAA to vehicle control. (Modified from Henley, D.V., Mueller, S., and Korach, K.S. In Press Environ Health Persp.).

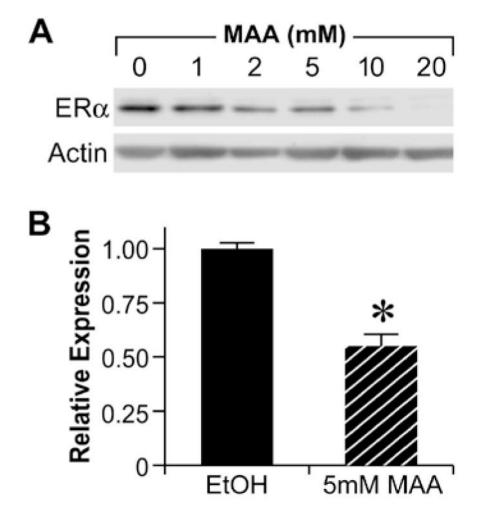


Figure 5.

Methoxyacetic Acid (MAA) decreases endogenous ER α expression.. A. MCF-7 cells were treated for 24h with either vehicle or increasing concentrations of MAA. Total protein was isolated from each sample and analyzed for ER α protein expression by Western blot. The data shown are representative of results from three independent experiments. B. MCF-7 cells were treated for 24h with either vehicle or 5mM MAA. Total RNA was isolated from each sample and steady state mRNA levels of ER α were measured by real-time PCR. The data shown represent the average fold over control obtained from duplicate samples in four independent experiments, and the error bars indicate the SEM, *p<0.01 comparing treatment with 5mM MAA to vehicle control. (Modified from Henley, D.V., Mueller, S., and Korach, K.S. In Press Environ Health Persp.).

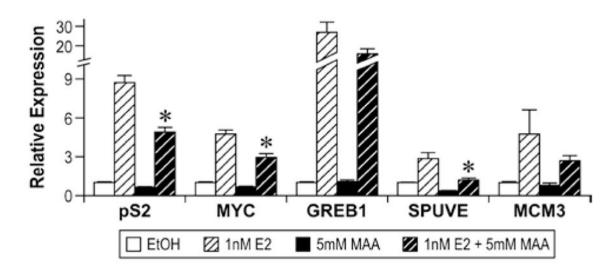


Figure 6.

Methoxyacetic Acid (MAA) disrupts estrogen-mediated endogenous gene expression. MCF-7 cells were pretreated with either vehicle or 5mM MAA for 2h and then treated for 18h with either vehicle or 1nM E2. Total RNA was isolated from the samples and the expression of endogenous estrogen-responsive genes was analyzed by real-time PCR. The data shown represent the average fold over control of duplicate samples from at least three independent experiments. Error bars represent the SEM, *p<0.01 comparing treatment with 1nM E2 + 5mM MAA to treatment with 1nM E2 alone. The error bars represent SEM. (Modified from Henley, D.V., Mueller, S., and Korach, K.S. In Press Environ Health Persp.).

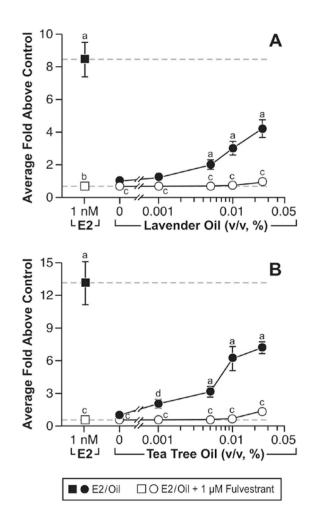


Figure 7.

In vitro estrogenic activity of lavender oil and tea tree oil in reporter gene assays. MCF-7 cells were transiently transfected with the estrogen-inducible 3x-ERE-luciferase reporter plasmid and treated with increasing concentrations of either lavender oil or tea tree oil for 18 hours in the presence and absence of fulvestrant, an ER antagonist. Estradiol treatment (1 nM) was included as a positive control. Data are plotted as fold-luciferase activity over vehicle control. (Modified from Henley, D.V., Lipson, N., Korach, K.S., and Bloch, C.A., *NEJM*, 356, 481, 2007).

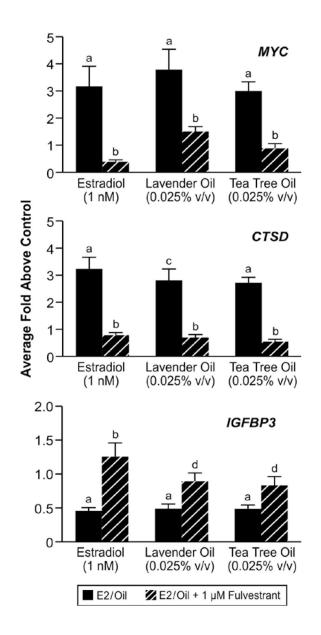


Figure 8.

In vitro effects of lavender oil and tea tree oil on the expression of estrogen-regulated endogenous genes. MCF-7 cells were treated with 1 nM estradiol, 0.025% lavender oil, or 0.025% tea tree oil in the presence or absence of fulvestrant and real-time PCR was performed to measure changes in the expression of the estrogen-inducible genes MYC, IGFBP3, and CTSD. Data shown are normalized and relative to a vehicle control. (Modified from Henley, D.V., Lipson, N., Korach, K.S., and Bloch, C.A., *NEJM*, 356, 481, 2007).

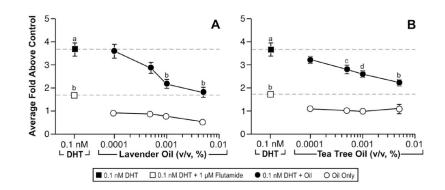


Figure 9.

In vitro antiandrogenic activity of lavender oil and tea tree oil in reporter gene assays. MDA-kb2 cells stably transfected with the MMTV-luciferase reporter plasmid were treated for 24 hours with increasing concentrations of lavender oil or tea tree oil in the presence or absence of 0.1 nM dihydrotestosterone (DHT). The androgen receptor antagonist flutamide (1 μ M) was included as a control antagonist for the androgen receptor. Luciferase activity was measured and data plotted as the average fold above control. (Modified from Henley, D.V., Lipson, N., Korach, K.S., and Bloch, C.A., *NEJM*, 356, 483, 2007).

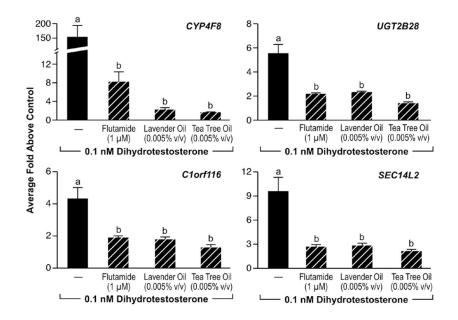


Figure 10.

In vitro effects of lavender oil and tea tree oil on androgen-stimulated endogenous gene expression. MDA-kb2 cells were treated with 0.1 nM dihydrotestosterone for 24 hours in the presence or absence of 0.0005% lavender oil, 0.0005% tea tree oil, or 1µM flutamide and real-time PCR was performed to measure changes in the expression of androgen-inducible genes. Data shown are normalized and relative to a vehicle control. (Modified from Henley, D.V., Lipson, N., Korach, K.S., and Bloch, C.A., *NEJM*, 356, 483, 2007).

Table 1

Incidence of pathology observed in wild-type and aERKO female mice following neonatal DES exposure.

Tissue/Lesion 4–5 8–9 12 Uterus	12-13 18-23	3-4-5	2 2 0	11 12	01
Uterus				C1-71	C7-QI
Atrophy 0/12 22/26 12	12/25 13/16	6/0 9	9 0/14	0/15	6/0
Disorg. Smooth Muscle 10/10 26/26 21	21/25 10/16	6/0 9	9 0/14	0/15	6/0
Hyalinization 10/10 25/26 23	23/25 12/16	6/0 9	9 0/14	0/15	6/0
Squamous metaplasia 3/10 9/26 15	15/25 2/16	6/0 9	9 0/14	0/15	6/0
Endometrial hyperplasia $0/12$ $2/26^{a}$ $10/3$	10/25 ^b 7/16 ^c	5 <i>c</i> 0/9	9 0/14	0/15	6/0
Vagina					
Persistent cornification 10/10 26/26 25	25/25 16/16	6/0 9	9 0/14	0/15	6/0
No cornification 0/10 0/26 0	0/25 0/16	6 8/8	e 13/13e	15/15 ^e	<i>ə</i> 6/6
Adenosis 0/12 2/26 (0/25 0/16	6/0 9	9 0/14	0/15	6/0
PPL of the Oviduct 11/12 23/23 21	21/24 14/14	4 0/9	9 0/14	0/15	6/0

Unless otherwise noted there was a 0% incidence in corn-oil treated animals of the same genotype. PPL = progressive proliferative lesion. Adapted from source: Couse J.F., Dixon D., Yates M., Moore A.B., Ma L., Maas R., and Korach K.S., *Dev. Biol.*, 238, 224–238, 2001. Source: Henley D.V., and Korach K.S., *Endocrinology*, 147(6) (Supplement), S26, 2006.

 e exhibited in 100% of corn oil exposed mice of same genotype and age.

 e exhibited in 1 of 12 corn oil exposed wild-type mice;

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Incidence of multi-oocyte follicles following treatment with genistein or Lavendustin A

		Genistein (µg/pup/day)	pn/dnd/Яn)	y)	T III anno 1	(innidud/Sul to intennity int
Genotype	Vehicle	1	10	100	1	10
Wild-type CD-1	0/8 (0)	0/8 (0) 1/8 (2) 2/8 (4)	2/8 (4)	6/8 (8)	0/8 (0)	0/8 (0)
Wild-type C57BL/6 1/11 (1) 1/11 (1) 9/11 (3)	1/11 (1)	1/11 (1)	9/11 (3)	11/11 (10)	pu	pu
αERKO	1/3 (1)	2/4 (1)	4/6 (4)	pu	pu	pu
ßERKO	1/2 (1)	0/4 (0)	1/2 (1) 0/4 (0) 0/5 (0)	1/3 (2)	nd	pu

that treatment group. nd, not determined. Adapted from source: Jefferson, W.N., Couse, J.F., Padilla-Banks, E., Korach, K.S., and Newbold, R.R., *Biol. Reprod.*, 67, 1285–1296, 2002. Source: Henley, D.V., and Korach, K.S., Endocrinology, 147(6) (Supplement), S27, 2006.