

Whole-Genome Sequences of Two *Campylobacter coli* Isolates from the Antimicrobial Resistance Monitoring Program in Colombia

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***Campylobacter coli*, along with *Campylobacter jejuni*, is a major agent of gastroenteritis and acute enterocolitis in humans. We report the whole-genome sequences of two multidrug-resistance *C. coli* strains, isolated from the Colombian poultry chain. The isolates contain a variety of antimicrobial resistance genes for aminoglycosides, lincosamides, fluoroquinolones, and tetracycline.**

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Campylobacter spp. are microaerobic, non-spore-forming, Gram-negative, and oxidase-positive members of the *Campylobacteraceae* family (1). *Campylobacter* spp. are zoonotic pathogens (2) and some of the main bacteria associated with human food-borne illness (3, 4). *Campylobacteriosis* is frequently associated with the consumption of undercooked poultry meat and the mishandling of the raw poultry products (5, 6). Different *Campylobacter* spp. are recognized as causing human gastroenteritis worldwide (7–9). *Campylobacter coli* is commonly isolated from swine and less so from poultry and humans (7, 9). Although *C. coli* accounts for fewer infections in humans than *Campylobacter jejuni*, its impact is considerable (10), taking into account the increased capability for antimicrobial resistance (11), where multidrug efflux pumps play an important role as mechanisms of antibiotic resistance (3). A small number of studies in Colombia have focused on understanding the epidemiology of *Campylobacter* spp. and their associated antimicrobial resistance. To correct this deficiency, the Colombian Integrated Program for Antimicrobial Resistance Surveillance (COIPARS) (12, 13) has been included in the Colombia-wide *Campylobacter* surveillance program, whose priority is to generate information for different governmental institutions, agricultural enterprises, and food animal production systems. The findings from our studies in Colombia have shown that *C. coli* and *C. jejuni* contamination of raw

poultry and poultry meat products present risk factors associated with acute illness in consumers of these products.

Here, we present the whole-genome sequences of two multidrug-resistant *C. coli* strains (M1483 and M1486), isolated from poultry meat collected from two retail stores in Bogotá, Colombia, as part of the COIPARS antimicrobial resistance monitoring program. Genomic DNA was isolated from overnight cultures using the PureLink Genomic DNA minikit (Invitrogen, Grand Island, NY, USA), and DNA libraries were prepared using SureSelect QXT sample preparation kit (Agilent, Santa Clara, CA, USA). The libraries were prepared according to the manufacturer's instructions and sequenced on an Illumina HiScanSQ instrument with 1 × 151-bp single reads, according to standard Illumina protocols. The *C. coli* M1483 and M1486 genomes were assembled using the reference-guided assembler ARGO, developed at NCBI, and the *de novo* assembler SPAdes (14). The genome sequence of strains M1483 and M1486 consisted of 45 and 55 contigs, yielding total sequences of 1,683,490 bp and 1,780,967 bp, respectively. The overall G+C content of the isolates was determined to be 32%. Sequences were annotated using the NCBI Prokaryotic Genome Automatic Annotation Pipeline (PGAAP) and have been deposited in GenBank. The results of the genome annotation presenting the number of genes, coding se-

TABLE 1 *Campylobacter coli* genome annotation statistics

Strain	NCBI BioSample	No. of genes	No. of CDSs ^a	No. of pseudogenes	No. of CRISPR arrays	No. of rRNAs	No. of tRNAs	No. of ncRNAs ^b	GenBank accession no.
M1483	SAMN04358093	1,782	1,739	55	0	3	37	3	LNQXL00000000
M1486	SAMN04358091	1,916	1,873	58	1	3	37	3	LNQXK00000000

^a CDSs, coding sequences.

^b ncRNAs, noncoding RNAs.

quences, pseudogenes, CRISPR arrays, rRNAs, tRNAs, and non-coding RNAs are summarized in Table 1.

A search for resistance-associated genes present in the isolates was performed using ResFinder version 2.1 (15) and enriched using RAST version 2.0 (16), both with default parameters. We found antimicrobial resistance genes for aminoglycosides (Aph 3'-III), lincosamides (*InuC*), fluoroquinolones (*gyrA* and *gyrB*), and tetracyclines (EF-G and TetO). Additionally, we found efflux pump genes (CmeA, CmeB, TolC, MATE, MFS, MacA, MacB, RND, AcrB, and OM) and CmeABC operon genes, both associated with increased multidrug resistance.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession numbers listed in Table 1. The versions described in this paper are the second versions.

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REFERENCES

- Allos BM. 2001. *Campylobacter jejuni* infections: update on emerging issues and trends. Clin Infect Dis 32:1201–1206. <http://dx.doi.org/10.1086/319760>.
- Wieczorek K, Osek J. 2013. Antimicrobial resistance mechanisms among *Campylobacter*. BioMed Res Int 2013:340605. <http://dx.doi.org/10.1155/2013/340605>.
- Mavri A, Smole Možina S. 2013. Effects of efflux-pump inducers and genetic variation of the multidrug transporter *cmeB* in biocide resistance of *Campylobacter jejuni* and *Campylobacter coli*. J Med Microbiol 62:400–411. <http://dx.doi.org/10.1099/jmm.0.052316-0>.
- Read DS, Woodcock DJ, Strachan NJ, Forbes KJ, Colles FM, Maiden MC, Clifton-Hadley F, Ridley A, Vidal A, Rodgers J, Whiteley AS, Sheppard SK. 2013. Evidence for phenotypic plasticity among multihost *Campylobacter jejuni* and *C. coli* lineages, obtained using ribosomal multilocus sequence typing and Raman spectroscopy. Appl Environ Microbiol 79:965–973. <http://dx.doi.org/10.1128/AEM.02521-12>.
- Ma L, Wang Y, Shen J, Zhang Q, Wu C. 2014. Tracking *Campylobacter* contamination along a broiler chicken production chain from the farm level to retail in China. Int J Food Microbiol 181:77–84. <http://dx.doi.org/10.1016/j.ijfoodmicro.2014.04.023>.
- Marotta F, Garofolo G, Di Donato G, Aprea G, Platone I, Cianciavichia S, Alessiani A, Di Giannatale E. 2015. Population diversity of *Campylobacter jejuni* in poultry and its dynamic of contamination in chicken meat. BioMed Res Int 2015:859845. <http://dx.doi.org/10.1155/2015/859845>.
- Sheppard SK, Didelot X, Jolley KA, Darling AE, Pascoe B, Meric G, Kelly DJ, Cody A, Colles FM, Strachan NJ, Ogden ID, Forbes K, French NP, Carter P, Miller WG, McCarthy ND, Owen R, Littrup E, Egholm M, Affourtit JP, Bentley SD, Parkhill J, Maiden MC, Falush D. 2013. Progressive genome-wide introgression in agricultural *Campylobacter coli*. Mol Ecol 22:1051–1064. <http://dx.doi.org/10.1111/mec.12162>.
- Skarp CPA, Hänninen ML, Rautelin HIK. 2015. Campylobacteriosis: the role of poultry meat. Clin Microbiol Infect [Epub ahead of print.] <http://dx.doi.org/10.1016/j.cmi.2015.11.019>.
- Zautner AE, Goldschmidt AM, Thürmer A, Schuldes J, Bader O, Lugert R, Groß U, Stingl K, Salinas G, Lingner T. 2015. SMRT sequencing of the *Campylobacter coli* BfR-CA-9557 genome sequence reveals unique methylation motifs. BMC Genomics 16:1088. <http://dx.doi.org/10.1186/s12864-015-2317-3>.
- Richards VP, Lefébure T, Pavinski Bitar PD, Stanhope MJ. 2013. Comparative characterization of the virulence gene clusters (lipooligosaccharide [LOS] and capsular polysaccharide [CPS]) for *Campylobacter coli*, *Campylobacter jejuni* subsp. *jejuni* and related *Campylobacter* species. Infect Genet Evol 14:200–213. <http://dx.doi.org/10.1016/j.meegid.2012.12.010>.
- Bakhshi B, Naseri A, Alebouyeh M. 2016. Comparison of antimicrobial susceptibility of *Campylobacter* strains isolated from food samples and patients with diarrhea. Iran Biomed J 20:91–96.
- Donado-Godoy P, Bernal JF, Rodríguez F, Gomez Y, Agarwala R, Landsman D, Mariño-Ramírez L. 2015. Genome sequences of multidrug-resistant *Salmonella enterica* serovar paratyphi B (dT+) and Heidelberg Strains from the Colombian poultry chain. Genome Announc 3(5):e01265-15. <http://dx.doi.org/10.1128/genomeA.01265-15>.
- Donado-Godoy P, Castellanos R, León M, Arevalo A, Clavijo V, Bernal J, León D, Tafur MA, Byrne BA, Smith WA, Perez-Gutierrez E. 2015. The Establishment of the Colombian Integrated Program for Antimicrobial Resistance Surveillance (COIPARS): A Pilot Project on Poultry Farms, Slaughterhouses and Retail market. Zoonoses Public Health 62(suppl 1):58–69. <http://dx.doi.org/10.1111/zph.12192>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. <http://dx.doi.org/10.1093/jac/dks261>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formosa K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.