

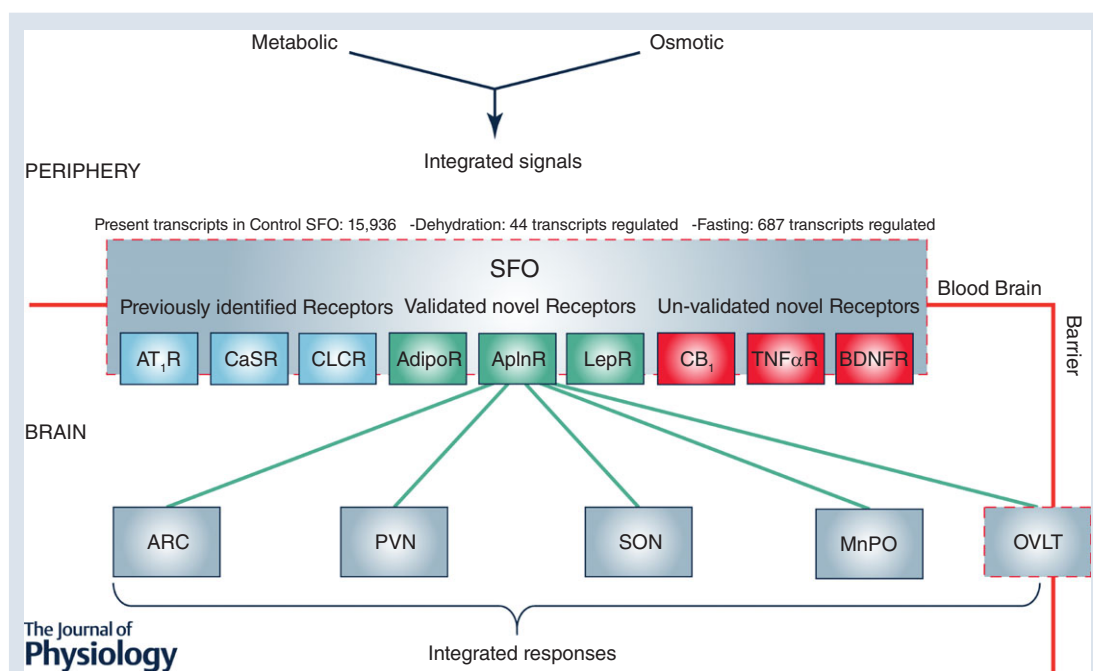
Physiological roles for the subfornical organ: a dynamic transcriptome shaped by autonomic state

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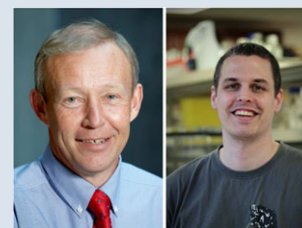
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Abstract The subfornical organ (SFO) is a circumventricular organ recognized for its ability to sense and integrate hydromineral and hormonal circulating fluid balance signals, information which is transmitted to central autonomic nuclei to which SFO neurons project. While the role of SFO was once synonymous with physiological responses to osmotic, volumetric and cardiovascular challenge, recent data suggest that SFO neurons also sense and integrate information

Alastair Ferguson received his PhD at the University of Calgary and after a postdoctoral fellowship at McGill University joined the Department of Physiology as an Assistant Professor at Queen's University in 1984. The research programme focuses on understanding the CNS pathways involved in regulation of the integrated autonomic nervous system. A variety of techniques are directed toward elucidating the role of critical autonomic nuclei and multiple neuroregulatory signalling molecules in the integration and regulation of the autonomic nervous system. **Charles Hindmarch** is a cardiovascular neuroscientist working at the University of Bristol as a Research Associate and at the University of Malaya as a Visiting Professor. The central theme of his research is to establish various models of hypertension exposed to a variety of physiological cues including dietary (salt/fat), age and activity (treadmill exercised). He employs high throughput transcriptomics such as RNAseq to profile both specific structures and single cells in the brain in order to establish appropriate gene targets for molecular, physiological and functional validation.



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from circulating signals of metabolic status. Using microarrays, we have confirmed the expression of receptors already described in the SFO, and identified many novel transcripts expressed in this circumventricular organ including receptors for many of the critical circulating energy balance signals such as adiponectin, apelin, endocannabinoids, leptin, insulin and peptide YY. This transcriptome analysis also identified SFO transcripts, the expressions of which are significantly changed by either 72 h dehydration, or 48 h starvation, compared to fed and euhydrated controls. Expression and potential roles for many of these targets are yet to be confirmed and elucidated. Subsequent validation of data for adiponectin and leptin receptors confirmed that receptors for both are expressed in the SFO, that discrete populations of neurons in this tissue are functionally responsive to these adipokines, and that such responsiveness is regulated by physiological state. Thus, transcriptomic analysis offers great promise for understanding the integrative complexity of these physiological systems, especially with development of technologies allowing description of the entire transcriptome of single, carefully phenotyped, SFO neurons. These data will ultimately elucidate mechanisms through which these uniquely positioned neurons respond to and integrate complex circulating signals.

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Abstract figure legend This schematic diagram highlights the central role of the subfornical organ (SFO) as a CNS structure without the normal blood brain barrier which plays critical roles in sensing and integrating peripheral signals of body fluid and metabolic status which do not cross the normal blood brain barrier. It highlights the primary outputs of SFO neurons to other CNS autonomic control centres including the arcuate (ARC), paraventricular (PVN), supraoptic (SON), and median preoptic (MnPO) nuclei, as well as the organum vasculosum of the lamina terminalis (OVLT), through which these integrative SFO neurons influence autonomic outputs. This diagram also summarises data from transcriptomic analysis, highlighting the numbers of genes expressed in SFO, the numbers regulated by dehydration and food deprivation, as well as validated and yet to be validated targets.

Introduction

Cardiovascular disease and hypertension are associated with the development of obesity, insulin resistance and diabetes. These comorbidities represent critical components of 'metabolic syndrome', diagnosed in nearly 25% of the North American population in 2002 (Ford *et al.* 2002). Since the discovery of leptin in the early 1990s, a number of other neuroactive peptides and adipokines associated with metabolic syndrome, including adiponectin, amylin, angiotensin II, glucagon-like peptide-1 (GLP-1), oxytocin (OT), α -melanocyte stimulating hormone (α -MSH), peptide YY/neuropeptide Y (Buijs, 1990; Ferguson & Washburn, 1998; Cowley *et al.* 1999; Kadowaki *et al.* 2006), have been shown to act not only as peripheral hormones, but also as neural signalling molecules in critical autonomic control centres of the brain. Importantly, the majority of these neuroactive signalling molecules exert what are apparently diverse physiological effects on cardiovascular (blood pressure, heart rate, baroreflex sensitivity), metabolic (food intake, metabolic rate), immune and reproductive functions by acting in these autonomic control centres in the hypothalamus and medulla. Although such commonality suggests these CNS centres are potential sites at which pathological changes

may underlie all of the comorbidities associated with the metabolic syndrome, an understanding of the CNS circuitry through which this may occur has yet to emerge. An additional and intriguing part of such models is the inclusion of feedback control circuitry, through which these same molecules (e.g. glucose, adiponectin, amylin, angiotensin, cholecystokinin (CCK), GLP-1, insulin, leptin, peptide YY (PYY), ghrelin) act as circulating signals providing critical information regarding homeostatic status to the CNS, despite the fact that most of these messengers do not diffuse freely across the blood-brain barrier (BBB) (Abstract Figure). Peptide specific transporters, such as the leptin transporter, have been suggested to mediate blood to brain signalling (Banks & Kastin, 1987), but their roles are not well defined and for many important circulating signals they do not exist (Spranger *et al.* 2006; Price *et al.* 2007). Intriguingly, there are specific regions of the brain that lie outside of the BBB, collectively termed circumventricular organs (CVOs), and a well-established body of evidence has shown that these specialized structures do play critical roles in sensing and responding to circulating signals associated with fluid balance, cardiovascular, metabolic and immune function (McKinley *et al.* 2003; Hoyda *et al.* 2009; Sisó *et al.* 2010; Mimee *et al.* 2013) (Abstract Figure).

Circumventricular organs as sensors of circulating signals

The CVOs are specialized, structurally unique CNS nuclei that lack the normal BBB (Weindl, 1986; Gross, 1992). The area postrema (AP), subfornical organ (SFO), and organum vasculosum of the lamina terminalis (OVLT) are the only CVOs containing neuronal cell bodies, as opposed to nerve terminals. These three regions are classified as the 'sensory CVOs' in view of their roles as critical integrative centres where circulating peptides act to regulate the cardiovascular, immune and neuroendocrine systems (Weindl & Sofroniew, 1981; Ferguson & Bains, 1996; McKinley *et al.* 2003; Cottrell & Ferguson, 2004; Fry & Ferguson, 2007). While the AP was initially known for its ability to sense circulating toxins and trigger nausea and emesis (Hornby, 2001), this CVO also plays roles in the regulation of energy balance (Ritter *et al.* 1981; Contreras *et al.* 1982; Hyde & Miselis, 1983), and in central cardiovascular regulation (Brody, 1988; Ferguson & Smith, 1991; Osborn *et al.* 2000). Receptor localization (Sexton *et al.* 1994), and electrophysiological (Riediger *et al.* 2001) and lesion studies (Rowland & Richmond, 1999), collectively identified the AP as the primary CNS site at which circulating amylin acts to inhibit food intake. Other studies have shown effects of GLP-1 (Rowland *et al.* 1997), CCK (Edwards *et al.* 1986; Sun & Ferguson, 1997), adrenomedullin (Shan & Krukoff, 2000; Yang & Ferguson, 2003), orexin (Yang & Ferguson, 2002), adiponectin (Fry *et al.* 2006), PYY (Price *et al.* 2008) and ghrelin (Lawrence *et al.* 2002; Fry & Ferguson, 2009) on AP neurons. In contrast, the SFO was initially recognized for its pre-eminent role as the primary CNS site at which circulating angiotensin acted to stimulate drinking (Simpson & Routtenberg, 1975), increase blood pressure (Mangiapanne & Simpson, 1980), and modulate vasopressin secretion (Iovino & Steardo, 1984). However, this limited perspective of the physiological roles of the SFO has changed significantly over the past 5 years, in part a result of our ability to identify novel targets with transcriptomic technologies, and this evolution will be the focus of the remainder of this article.

Subfornical organ. The SFO is a forebrain midline structure located on the dorsal surface of the third ventricle below the ventral hippocampal commissure, and is primarily known for its well established roles in cardiovascular and neuroendocrine regulation (Ferguson & Bains, 1997; McKinley *et al.* 1998). Roles for the SFO in anorexia and emaciation (Trivedi *et al.* 2003) and immune function (Takahashi *et al.* 1997) have also been suggested. The SFO can be subdivided anatomically into 'core' and 'shell' zones with the primary efferent projections to the paraventricular nucleus (PVN), supraoptic nucleus (SON), median preoptic nucleus, OVLT

and arcuate nucleus (Miselis, 1981; Lind, 1986; Gruber *et al.* 1987), while those originating from the shell project primarily to the bed nucleus of the stria terminalis (McKinley *et al.* 2003). These neural outputs position this CVO to effectively communicate with all of the critical hypothalamic autonomic control centres, and thus play important roles in the regulation of a much greater spectrum of homeostatic functions. SFO neurons have for some time been known to sense circulating signals involved in fluid (osmolarity, sodium, calcium, relaxin, atrial natriuretic peptide), cardiovascular (angiotensin, endothelin, vasopressin (VP)), and immune (interleukin 1 β) regulation (Cottrell & Ferguson, 2004). Specific excitatory projections have been found to VP and oxytocin (OT) neurons in the SON and PVN, as well as to parvocellular areas of the PVN that in turn project either to the median eminence (corticotropin-releasing hormone (CRH), thyrotropin-releasing hormone (TRH) neurons), the medulla (OT, VP, TRH neurons), or the spinal cord (OT, VP neurons) (Ferguson & Bains, 1996). More recent single cell recordings have shown direct effects of metabolic signals such as calcitonin (Schmid *et al.* 1998), amylin (Riediger *et al.* 1999) and ghrelin (Pulman *et al.* 2006) on SFO neurons (Abstract Figure). Thus studies were slowly building a catalogue describing the SFO as a structure with neurons able to sense a variety of different signalling molecules present in the circulation, but progress was driven effectively by the analysis on one signalling molecule at a time, with the driving force originating in studies demonstrating receptors/binding sites for that molecule in this CVO.

Transcriptomic analysis

The development of reliable tools for whole transcriptomic analysis offered an alternative approach where single studies could describe relative expression levels of the entire genome. We utilized this technology not only to catalogue the expression levels of all transcripts represented on the Affymetrix gene chip (>30,000), but also to assess changes in expression associated with the challenges of fluid (72 h) or food (48 h) deprivation in the SFO (Hindmarch *et al.* 2008). While these studies confirmed the expression of many previously identified receptors in the SFO (AT_{1A}, CaSR, ET_B, CLCR), they also identified for the first time receptors for many of the critical circulating energy balance signals, including adiponectin, apelin, endocannabinoids (CB₁), leptin, insulin and PYY, as well as peptides believed to play important roles in the regulation of feeding such as apelin, cholecystokinin (CCK) and brain derived neurotrophic factor (BDNF) (Fig. 1). Although these findings identify new targets and support the conclusion that the SFO plays important roles in monitoring these signals, and transmitting this

information to critical autonomic control centres in the hypothalamus, in reality they represent only the first step in a detailed systematic analysis as highlighted in three examples described below. Importantly, the original microarray data for these SFO experiments are held in the NCBI Gene Expression Omnibus, a functional genomics data repository for Microarray and Sequence data (<http://www.ncbi.nlm.nih.gov/geo>; Accession number: GSE12978), and are thus freely accessible to all, for additional analysis.

Validation: functional roles for SFO in energy balance?

The transcriptome of the SFO is dynamic and is modified by physiological state. Food deprivation (48 h) resulted in 687 transcripts regulated by greater than 2-fold while fluid deprivation resulted in just 44 transcripts (Hindmarch *et al.* 2008). These data suggested to us that in addition

to its well-established roles in the regulation of fluid balance and cardiovascular regulation the SFO may also play important roles in the regulation of energy homeostasis. We have subsequently demonstrated that electrical activation of SFO neurons stimulates food intake in satiated animals (Smith *et al.* 2010). We have also shown that while lesion of either AP or SFO in isolation does not influence long term food intake or body weight, lesion of both CVOs reduces both parameters (Baraboi *et al.* 2010*b*), suggesting complementary sensory functions of these CVOs. Such double lesions also result in reduced patterns of *c-fos* activation in PVN, the hypothalamic location of adrenal (CRH) and thyroid (TRH) control neurons, and in the nucleus of the tractus solitarius (NTS) in response to systemic GLP-1 and PYY receptor activation, confirming vital roles for these structures in sensing circulating signals (Baraboi *et al.* 2010*a,b*).

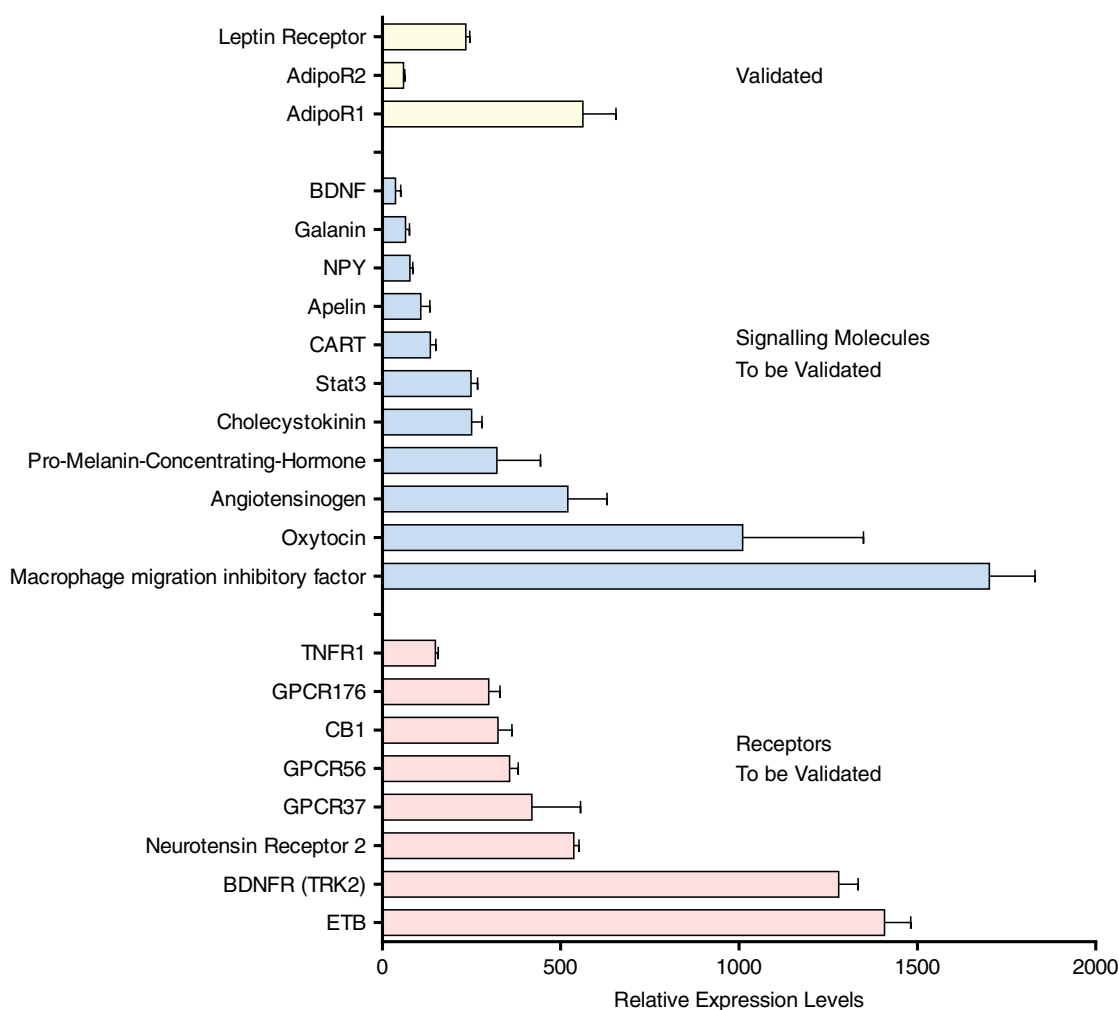


Figure 1. Targets in SFO identified by microarray analysis

This histogram shows data from our microarray analysis (Hindmarch *et al.* 2008) highlighting the relative expression levels of transcripts from our microarray analysis, the validation of which we describe in this paper (yellow). In addition we show here some novel target transcripts identified in our array study which are known to be potential signalling molecules (blue), or receptors (pink) associated with the regulation of energy balance.

Validation: adiponectin actions in the SFO – dynamic changes with food deprivation? Our array studies not only identified adiponectin receptors in the SFO for the first time, but in addition suggested that the expression level of adipoR2 was in fact changed by food deprivation. In this case our initial approach to validation began with molecular confirmation that adipoR1 and R2 were expressed in SFO using standard PCR (Fig. 2A), and selectively altered levels of AdipoR2 expression following food deprivation were then confirmed using quantitative RT-PCR (Fig. 2B). Patch clamp recordings confirmed functional roles for these receptors showing that adiponectin hyperpolarized 35% and depolarized 22% of SFO neurons as shown in Fig. 2C (Alim *et al.* 2010). Intriguingly similar recordings showing that adiponectin depolarized 77% (compared to only 22% in controls) of SFO neurons from food deprived animals while no cells hyperpolarized (Alim *et al.* 2010) (Fig. 2D), provided direct evidence of functional consequences

associated with these transcriptome changes induced by physiological state.

Validation: leptin actions in SFO – dynamic changes in obesity? The leptin receptor (ObRb) was also reported as present in SFO by our microarray analysis, although the extensive literature on CNS distribution of leptin receptors had not identified expression in this CVO. Our follow-up validation in this instance included PCR of SFO tissue with primer sets directed toward different regions of the leptin receptor which confirmed our array work (Fig. 3A). Antibodies against the leptin receptors (despite concerns regarding specificity) also confirmed expression, as did the induction of p-Stat3 by leptin (Smith *et al.* 2009). Functional roles for ObRb in SFO were confirmed both by electrophysiology showing depolarizing (39%) (the same neurons that were depolarized by amylin) and hyperpolarizing (25%) effects of leptin on SFO neurons

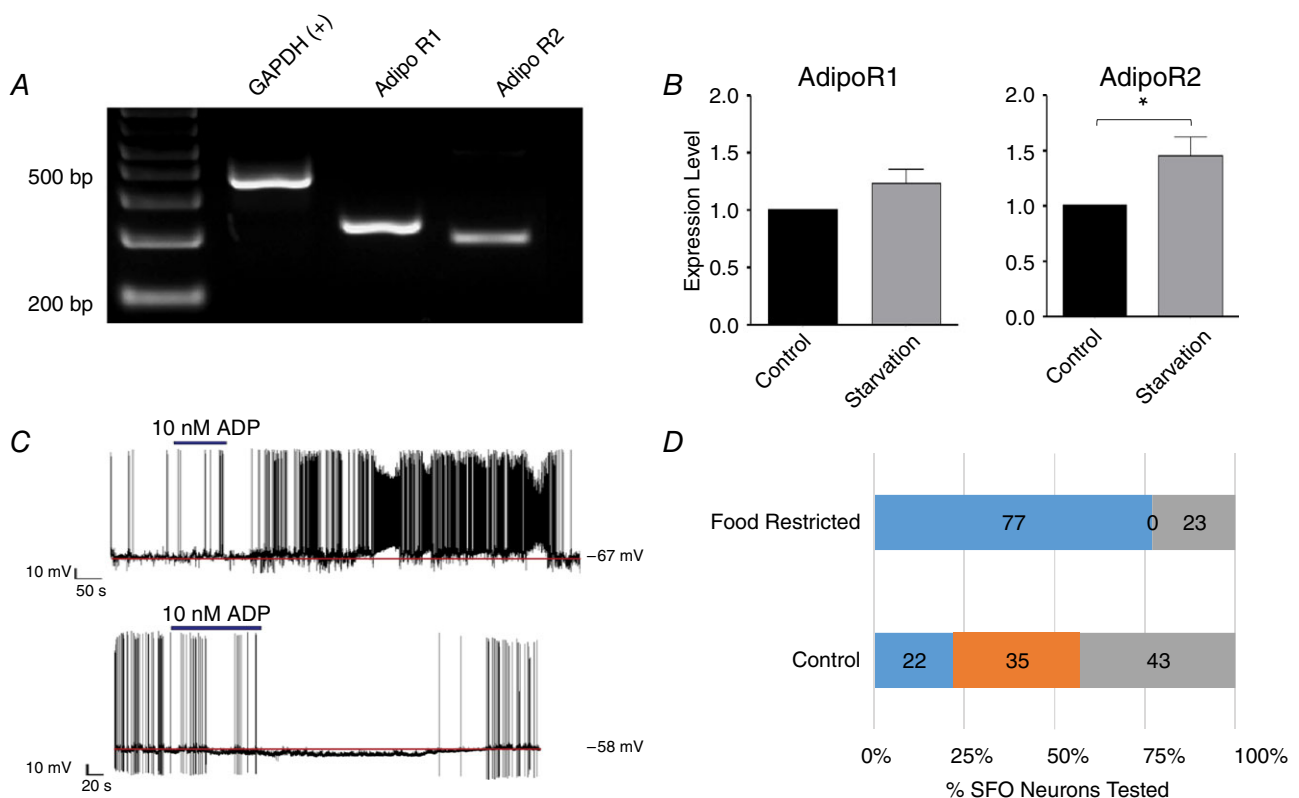


Figure 2. The process of adiponectin receptor validation (Alim *et al.* 2010) following our identification of these receptors in SFO

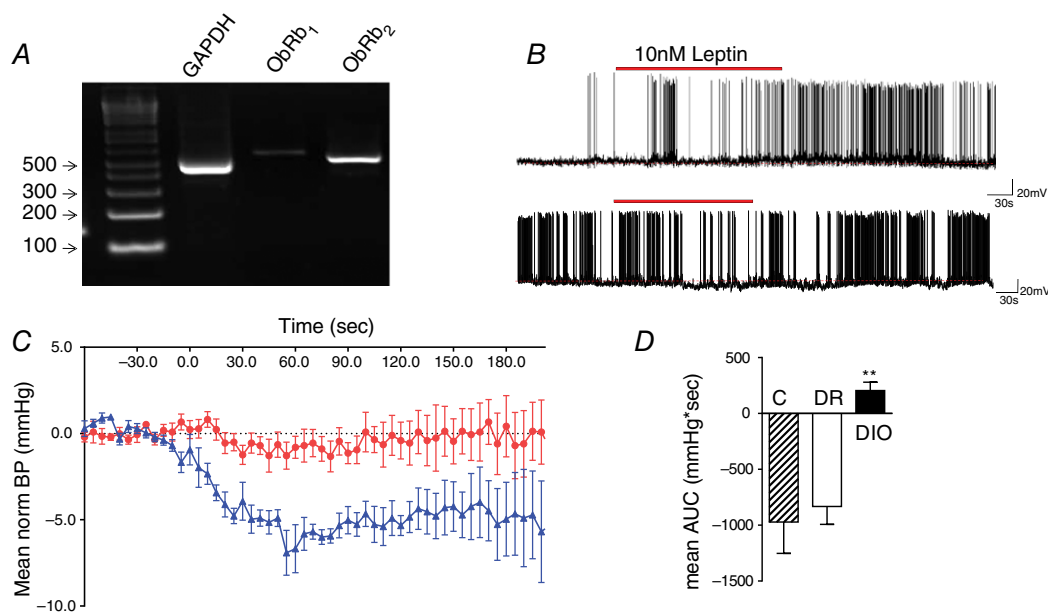
A shows an agarose gel from RT-PCR analysis of whole tissue from SFO using primer sets for GAPDH, AdipoR1 and AdipoR2, all of which were seen to be clearly expressed in the SFO. B, the histograms show qRT-PCR data confirming microarray analysis indicating that while AdipoR2 was significantly increased by 48 h of food restriction (*), AdipoR1 was not. C, patch clamp recordings from SFO neurons also validate functional roles for these receptors in that proportions of these cells are either depolarized or hyperpolarized by bath administration of adiponectin (ADP, 10 nM, indicated by the bar). D, the histogram summarizes proportions of dissociated SFO neurons from control and food deprived (48 h) groups showing depolarizations (blue bars) or hyperpolarizations (orange bars) in response to adiponectin and illustrate a large shift toward depolarizing effects in the latter group.

(Smith *et al.* 2009) (Fig. 3B), and by microinjection studies showing that direct administration into SFO caused changes in blood pressure (Fig. 3C), effects that are no longer observed in the diet induced obesity (DIO) obese phenotype (aged matched chow fed and diet resistant animals retain responsiveness to leptin as illustrated in Fig. 3D (Smith & Ferguson, 2012).

Validation: untapped targets? Additional SFO genes identified by our array studies which are associated with CNS regulation of energy balance, and have yet to be validated and pursued, include receptors for endocannabinoids (CB₁), BDNF (BDNFR) and tumour necrosis factor α (TNFR1) (Abstract Figure), and signalling molecules such as cocaine and amphetamine related transcript (CART), OT, pro-melanin-concentrating hormone (PMCH) and signal transducer and activator of transcription 3 (STAT3) (Hindmarch *et al.* 2008). Finally, we have identified true glucose sensing neurons in the SFO (Medeiros *et al.* 2012), and have preliminary data showing that acute (<24 h) changes in glucose concentration modify the responsiveness of SFO neurons to CCK. These findings highlight the role of physiological state in modifying

the sensory abilities of SFO in the regulation of energy balance. Thus, our initial array interrogation of the SFO has identified this CVO as a region of the CNS which performs critical roles in continually monitoring circulating metabolic, cardiovascular and immune signalling molecules. Intriguingly, the SFO, through its efferent connections to hypothalamic autonomic control centres, may then coordinate the integrated regulation of metabolic, cardiovascular, immune and neuroendocrine outputs.

One-cell-at-a-time. We have presented here data that firstly profile the transcriptome of the control, dehydrated and fasted SFO, and identified regulated targets that have been validated by additional studies. That said, it needs to be emphasized that such profiles of tissue composed of entire nuclei are based on the average expression of the entire population of discrete SFO neurons rather than being representative of the component subpopulations of cells of this tissue. Thus, describing the properties of these single neurons will also be critical to understanding the functional physiological roles of the SFO. We have already described that significant proportions (>25%) of SFO neurons respond to signals such as angiotensin



(>60%), ghrelin (>25%), leptin (>60%), or adiponectin (>50%) and this suggests that each neuron has specific receptors (i.e. sensors) for a number of these molecules. Further, studies have confirmed that single SFO neurons can sense multiple signals (Anderson *et al.* 2001; Pulman *et al.* 2006; Smith *et al.* 2009). There is also overwhelming evidence that expression of a transcript can differ from cell to cell (Xi *et al.* 1999; Yamashita *et al.* 2002; Pulman *et al.* 2006; Hoyda *et al.* 2007) but until recently it has been technically challenging to describe the entire transcriptome of a single neuron. Technological advances have now paved the way for single-cell profiling of the SFO to become a realistic ambition. Regardless of the technology used (Lee *et al.* 2013) Next Generation Sequencing (NGS) of the transcriptome (RNAseq) involves the preparation and subsequent sequencing of a library of short sequences that represent both coding and non-coding transcripts within a particular tissue or cell. The read-data that result from these experiments can be aligned back to the appropriate genome in order to identify the relative abundance of each transcript. Single cell transcriptomics is already changing the way we are able to describe populations of cells; while previous efforts to phenotype populations of individual neurons relied upon physiological profiling (electrophysiological, molecular), libraries of substantive numbers of individual cell transcriptomes can now be sequenced. This approach is not without its limitations since one must either destroy the cell (via patch) or dissociate and sort the cells in order to capture the nuclear material for sequencing. Single cell transcriptomic analysis is, however, moving within reach, and the recent identification of 47 molecularly distinct subclasses following a molecular census of 3005 single cell transcriptomes from somatosensory S1 cortex and hippocampus CA1 in mice (Zeisel *et al.* 2015) substantiates the potential value of this approach. In the SFO or other neuronal structures, we expect that the profiling of the control SFO transcriptome will reveal similar intra-cell diversity as found in the S1 and CA1 cells. We anticipate that it will be possible to capture accurate signatures for those SFO neurons that respond to circulating signals such as angiotensin, leptin or adiponectin, or core compared to shell neurons, or interneurons compared to projection neurons, or neurons that project to one output nucleus compared to another. Such classification will then open the door to examination of how these individual sub-populations of SFO neurons are differentially regulated by specific physiological challenges.

Concluding remarks

In conclusion, we have described here an emerging literature supporting the idea that the SFO plays important roles in the regulation of energy balance in addition to its

well established roles in the control of fluid balance, the cardiovascular system and immune regulation (Abstract Figure). We have described the role that transcriptomic profiling of the whole SFO has played in the development of these ideas. Transcriptome-wide data analysis continues to tell us that changes in transcript expression associated with physiological challenges are complex, integrated and plastic. Many potential targets remain as subjects for future validation studies. Electrophysiological and molecular data also tell us that there is great diversity within a population of cells even within the same tissue. In order to properly describe the roles of neuronal tissue like the SFO in biological phenomena such as energy homeostasis it is important that we begin to describe the individual cells that make up these structures.

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Additional information

Competing interests

None declared.

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