

ORIGINAL MANUSCRIPT

Polymorphisms of the centrosomal gene (FGFR1OP) and lung cancer risk: a meta-analysis of 14 463 cases and 44 188 controls

Xiaozheng Kang^{1,2,3,†}, Hongliang Liu^{1,4,†}, Mark W. Onaitis^{1,2}, Zhensheng Liu^{1,4}, Kouros Owzar^{1,5}, Younghun Han⁶, Li Su^{7,8}, Yongyue Wei^{7,8}, Rayjean J. Hung⁹, Yonathan Brhane⁹, John McLaughlin¹⁰, Paul Brennan¹¹, Heike Bickeböllner¹², Albert Rosenberger¹², Richard S. Houlston¹³, Neil Caporaso¹⁴, Maria Teresa Landi¹⁴, Joachim Heinrich¹⁵, Angela Risch¹⁶, Xifeng Wu¹⁷, Yuanqing Ye¹⁷, David C. Christiani^{7,8}, Christopher I. Amos⁶ and Qingyi Wei^{1,4,*}; Transdisciplinary Research in Cancer of the Lung (TRICL) Research Team

¹Duke Cancer Institute and ²Division of Cardiovascular and Thoracic Surgery, Department of Surgery, Duke University Medical Center, 905 S. LaSalle Street, Durham, NC 27710, USA, ³Key Laboratory of Carcinogenesis and Translational Research (Ministry of Education), Department of Thoracic Surgery I, Peking University Cancer Hospital and Institute, Beijing 100142, China, ⁴Department of Medicine, Duke University School of Medicine, Durham, NC 27710, USA, ⁵Department of Biostatistics and Bioinformatics, Duke University Medical Center, Durham, NC 27710, USA, ⁶Community and Family Medicine, Geisel School of Medicine, Dartmouth College, Hanover, NH 03755, USA, ⁷Massachusetts General Hospital, Boston, MA 02114, USA, ⁸Department of Environmental Health, Harvard School of Public Health, Boston, MA 02115, USA, ⁹Lunenfeld-Tanenbaum Research Institute of Mount Sinai Hospital, Toronto, Ontario, Canada, ¹⁰Public Health Ontario, Toronto, Ontario M5T 3L9, Canada, ¹¹Genetic Epidemiology Group, International Agency for Research on Cancer (IARC), 69372 Lyon, France, ¹²Department of Genetic Epidemiology, University Medical Center, Georg-August-University Göttingen, 37073 Göttingen, Germany, ¹³Division of Genetics and Epidemiology, the Institute of Cancer Research, London SW7 3RP, UK, ¹⁴Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Bethesda, MD 20892, USA, ¹⁵Helmholtz Centre Munich, German Research Centre for Environmental Health, Institute of Epidemiology I, 85764 Neuherberg, Germany, ¹⁶Department of Molecular Biology, University of Salzburg, 5020 Salzburg, Austria and ¹⁷Department of Epidemiology, University of Texas MD Anderson Cancer Center, Houston, TX 77030, USA

†To whom correspondence should be addressed. Tel: +1 919 660 0562; Fax: +1 919 660 0178; Email: qingyi.wei@duke.edu

†These authors contributed equally to this work.

Abstract

Centrosome abnormalities are often observed in premalignant lesions and *in situ* tumors and have been associated with aneuploidy and tumor development. We investigated the associations of 9354 single-nucleotide polymorphisms (SNPs) in 106 centrosomal genes with lung cancer risk by first using the summary data from six published genome-wide association studies (GWASs) of the Transdisciplinary Research in Cancer of the Lung (TRICL) (12 160 cases and 16 838 controls) and then conducted *in silico* replication in two additional independent lung cancer GWASs of Harvard University (984 cases and 970 controls) and deCODE (1319 cases and 26 380 controls). A total of 44 significant SNPs with false discovery rate (FDR) ≤ 0.05 were mapped to one novel gene *FGFR1OP* and two previously reported genes (*TUBB* and *BRCA2*). After combined the results from TRICL with those from Harvard and deCODE, the most significant association ($P_{\text{combined}} = 8.032 \times 10^{-6}$) was with

Received: October 17, 2015; Revised: January 6, 2016; Accepted: January 25, 2016

© The Author 2016. Published by Oxford University Press. All rights reserved. For Permissions, please email: journals.permissions@oup.com.

rs151606 within *FGFR1OP*. The rs151606 T>G was associated with an increased risk of lung cancer [odds ratio (OR) = 1.10, 95% confidence interval (95% CI) = 1.05–1.14]. Another significant tagSNP rs12212247 T>C ($P_{\text{combined}} = 9.589 \times 10^{-6}$) was associated with a decreased risk of lung cancer (OR = 0.93, 95% CI = 0.90–0.96). Further *in silico* functional analyzes revealed that rs151606 might affect transcriptional regulation and result in decreased *FGFR1OP* expression ($P_{\text{trend}} = 0.022$). The findings shed some new light on the role of centrosome abnormalities in the susceptibility to lung carcinogenesis.

Abbreviations

eQTL	expression quantitative trait loci
FDR	false discovery rate
GWASs	genome-wide association studies
LD	linkage disequilibrium
SNPs	single-nucleotide polymorphisms

Introduction

Lung cancer represents the number one cancer-related mortality worldwide, with more than 1.6 million cases diagnosed and 1.3 million deaths per year (1). In the United States, it is the primary cause of deaths from cancers in both men and women, leading to more deaths than from breast, colorectal and prostate cancers all combined. Although lung cancer is commonly considered a disease caused by environmental exposures, genetic factors also play a role in the etiology (2). Genomic regions at chromosomes 15q25.1 (3,4), 5p15.33 (4–6), 6p21.33 (4), 3q28 (4,7,8), 13q13.1 (8) and 22q12.1 (8) have been identified to be associated with lung cancer susceptibility by genome-wide association studies (GWASs) in populations of European descent.

Centrosomes are the pivotal regulating element of cell division and act as a reaction center where the cell cycle key regulating elements assemble and play an important role in cell polarization and ciliogenesis (9,10). Recent studies have revealed that deregulation of centrosome-related proteins induce centrosome abnormalities (11), which in turn result in tumor development through the induction of chromosome instability and aneuploidy (12,13). A growing body of evidence has linked centrosome abnormalities to the developments of solid tumors and hematopoietic malignancies (10), and the underlying mechanisms are now an active area of research (14–16).

Centrosome abnormalities and chromosome instability are also frequently observed in human lung cancer (17). Genes involved in centrosome dysregulation have emerged as the candidate targets to study the mechanism of initiation and progression of lung cancer. In this study, we hypothesized that genetic variants of centrosomal genes are associated with lung cancer risk. To test this hypothesis, we performed a meta-analysis of the association results of SNPs in centrosomal genes from eight lung cancer GWASs from Transdisciplinary Research in Cancer of the Lung (TRICL) and ILCCO, including 14 463 cases and 44 188 controls.

Materials and methods

Study populations

The study populations have been detailed previously (8,18). Briefly, this study included six previously reported lung cancer GWASs of from the TRICL, which was built upon the collaborative network of the International Lung Cancer Consortium (ILCCO) (8,18) including 12 160 lung cancer cases and 16 838 controls of European descent: the MD Anderson Cancer Center (MDACC) GWAS, the Institute of Cancer Research (ICR) GWAS, the National

Cancer Institute (NCI) GWAS, the International Agency for Research on Cancer (IARC) GWAS, Toronto study from Lunenfeld-Tanenbaum Research Institute study (Toronto) GWAS and the German Lung Cancer Study (GLC) GWAS, Germany. The additional datasets of another two independent GWASs of Caucasian populations were from Harvard Lung Cancer Study (19) (984 cases and 970 controls) and Icelandic Lung Cancer Study (deCODE) (1319 cases and 26 380 controls) (20) from the ILCCO. A written informed consent was obtained from each participant, and this study was approved by the institutional review boards for each of the participating institutions.

Genotyping platforms and quality controls

For these GWASs, genotyping was performed using Illumina HumanHap 317, 317+240S, 370Duo, 550, 610 or 1M arrays. Imputation was performed by using data from the 1000 Genomes Project (phase I integrated release 3, March 2012) (21) as reference and IMPUTE2 v2.1.1 (22), MaCH v1.0 (23) or minimac (version 2012.10.3) (24) software. Poorly imputed SNPs defined as an information score < 0.40 with IMPUTE2 or an $r^2 < 0.30$ with MaCH were excluded from the final analyses. Standard quality control on samples was performed on all scans, excluding individuals with low call rate (< 90%) and extremely high or low heterozygosity ($P < 1.0 \times 10^{-4}$), as well as all individuals evaluated to be of non-European ancestry (using the HapMap phase II CEU, JPT/CHB and YRI populations as a reference).

Genes and SNPs selection

The centrosomal genes were collected from the Molecular Signatures Database (<http://software.broadinstitute.org/gsea/msigdb>), which is a collection of annotated gene sets for use with gene set enrichment analysis. Overall, 106 centrosomal genes located on autosomal chromosomes were selected (Supplementary Table 1, available at *Carcinogenesis* Online). The genotyped or imputed common SNPs located within 2 kb up- and downstream of centrosomal genes were extracted from the GWAS datasets and the final meta-analysis included 9354 SNPs with genotyping call rate $\geq 90\%$, minor allele frequency $\geq 5\%$, and Hardy Weinberg Equilibrium exact $P \geq 10^{-4}$. As previously reported (8,25), SNPs from the 1000 Genomes project release Phase 1 integrated release version 3 and filtered with only SNPs showing imputation accuracy > 0.3 were retained. The detailed workflow is shown in Supplementary Figure 1, available at *Carcinogenesis* Online.

In silico functional validation

To predict the potential functions of risk-associated SNPs and their high-linkage disequilibrium (LD) ($r^2 \geq 0.60$) variants, we used three *in silico* tools: SNPinfo (26), RegulomeDB (27) and F-SNP (28). Expression quantitative trait loci (eQTL) analysis was carried out using lymphoblastoid cell data from 1000 Genomes Project (21) European subpopulation (EUR, 373 individuals) (phase I integrated release 3, March 2012).

Statistical methods

For each GWAS, ORs and associated 95% CIs were calculated by unconditional logistic regression using R (v2.6), Stata (v10, State College, Texas, USA) and PLINK (v1.06) (29) software. With the inverse variance method, meta-analysis under fixed and random-effects models was conducted on the results of log-additive model of 9354 SNPs. Cochran's Q statistic to test for heterogeneity and the I^2 statistic to quantify the proportion of the total variation due to heterogeneity were calculated (30). Fixed effects model was applied when there was no heterogeneity among GWASs (Q-test $P > 0.100$ and $I^2 < 50\%$); otherwise, random effects model was applied. The false discovery rate (FDR) procedure was employed to control for multiple testing (31) with $FDR \leq 0.050$. The correlation between SNPs and corresponding mRNA expression levels was estimated by using a linear regression model.

LocusZoom (32) was applied to generate regional association plots, using the 1000 Genomes European (EUR) reference data (phase I integrated release 3, March 2012) to compute LD. Haploview v4.2 (33) was employed to construct the LD plots. SNPs whose r^2 in linkage with any of the causal SNPs is less than 0.60 are also considered independent. All analyses were conducted with SAS (version 9.1.3; SAS Institute, Cary, NC, USA) unless specified otherwise.

Results

Meta-analysis of the main effects of SNPs in 106 centrosomal genes

The samples sizes of the used GWASs in this study are summarized in Table 1. In this study, we firstly carried out the analysis within the six TRICL GWASs, which included 12 160 lung cancer case subjects and 16 838 unaffected controls. After stringent quality control, we analyzed 9354 SNPs from 106 centrosomal genes (Figure 1). There are 843 SNPs and 96 SNPs with nominal $P < 0.050$ and < 0.001 under an additive (per allele) model, respectively; 44 SNPs remained significant at the FDR threshold ≤ 0.050 . These top SNPs are mapped to *TUBB*, *FGFR1OP* and *BRCA2*, respectively (Table 2). We removed SNPs that had a high pairwise LD with those in previously reported GWASs (4,8,34) (*TUBB* at 6p21.33 and *BRCA2* at 13q13.1), and only SNPs mapped to *FGFR1OP* were included for the additional analyses. The regional association plots demonstrated that SNP rs12212247

showed high LD ($r^2 \geq 0.60$) with other 31 SNPs in *FGFR1OP*, except for rs151606 (Figure 2; Supplementary Table 2, available at Carcinogenesis Online). Finally, two tagSNPs (rs151606 and rs12212247) in *FGFR1OP* were selected for the further analysis. As shown in Table 3, both SNPs had a high imputation quality in each dataset.

Of these two SNPs, rs151606 A>T was shown to be associated with an increased risk of lung cancer (OR = 1.12, 95% CI = 1.07–1.17, $P = 1.177 \times 10^{-6}$), while rs12212247 T>C was associated with a decreased risk (OR = 0.93, 95% CI = 0.90–0.97, $P = 8.926 \times 10^{-5}$). There was no heterogeneity observed for these risk estimates in these six GWASs (Figure 3).

We sought to expand these findings in another two independent lung cancer GWASs from Harvard University (984 cases and 970 controls) and deCODE (1319 cases and 26 380 controls). When the two studies were combined, rs12212247 and its 31 high LD SNPs within *FGFR1OP* were found with the same effect direction with a nominal $P = 0.039$. In addition, the protective effect of SNP rs151606 was insignificant (Table 3; Supplementary Table 3, available at Carcinogenesis Online). However, when we combined the summary results of all the eight studies, we observed a significant evidence for the association of rs151606 and lung cancer risk (OR = 1.10, 95% CI = 1.05–1.14, $P_{\text{combined}} = 8.032 \times 10^{-6}$) and rs12212247 (OR = 0.93, 95% CI = 0.90–0.96, $P_{\text{combined}} = 9.589 \times 10^{-6}$) (Table 3; Figure 3),

Table 1. Characteristics of the study populations included in the participating genome-wide association studies

Variable	ICR ^a		MDACC ^b		IARC ^c		NCI ^d		Toronto ^e		GLC ^f		Harvard ^g		deCODE ^h	
	Case	Control	Case	Control	Case	Control	Case	Control	Case	Control	Case	Control	Case	Control	Case	Control
Overall	1952	5200	1150	1134	2533	3791	5713	5736	331	499	481	478	984	970	1319	26380
AD	465	5200	619	1134	517	2824	1841	5736	90	499	186	478	597	970	547	26380
SQ	611	5200	306	1134	911	2968	1447	5736	50	499	97	478	216	970	259	26380

AD, adenocarcinoma; SQ, squamous cell carcinoma.

^aICR: the Institute of Cancer Research Genome-wide Association Study, UK.

^bMDACC: the MD Anderson Cancer Center Genome-wide Association Study, USA.

^cIARC: the International Agency for Research on Cancer Genome-wide Association Study, France.

^dNCI: the National Cancer Institute Genome-wide Association Study, USA.

^eToronto: the Lunenfeld-Tanenbaum Research Institute Genome-wide Association Study, Toronto, Canada.

^fGLC: German Lung Cancer Study, Germany.

^gHarvard: Harvard Lung Cancer Study, USA.

^hdeCODE: Icelandic Lung Cancer Study, Iceland.

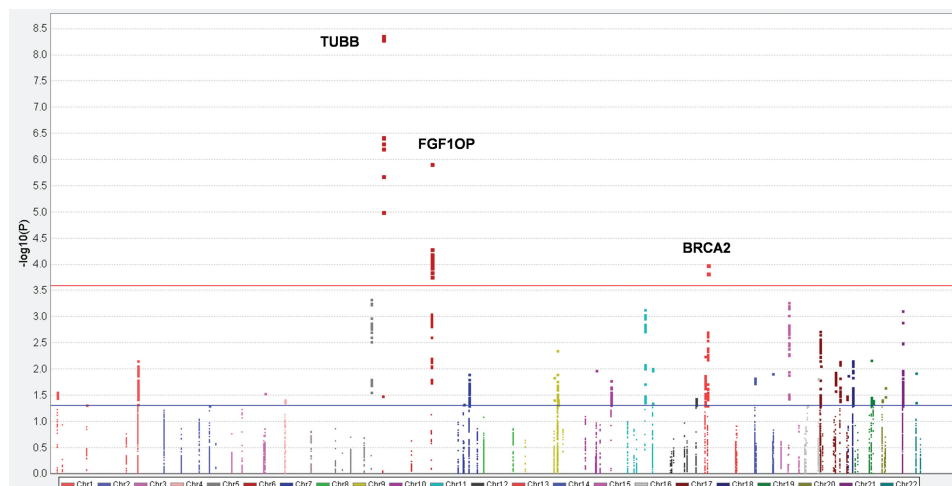


Figure 1. Manhattan plot of genome-wide association results. SNPs are plotted on the x-axis according to their positions on each chromosome. On the y-axis, the association P values with lung cancer risk are shown (as $-\log_{10} P$ values). Horizontal red line represents FDR threshold 0.05. Horizontal blue line represents nominal P values of 0.05.

given only 9354 SNPs were tested for their associations with lung cancer risk in the present study.

Stratified analyses

Like many other cancers, lung cancer histologic subtypes can have dramatically different clinical behaviors. In the further analyses stratified by histology, only adenocarcinoma (AD) and squamous cell carcinoma (SQ) were included. SNP rs151606 was associated only with the risk of SQ (OR = 1.09, 95% CI = 1.02–1.16, $P_{\text{combined}} = 0.011$), but not with AD (OR = 1.06, 95% CI = 1.00–1.13, $P_{\text{combined}} = 0.058$); the strongest effect of rs12212247 was detected

in AD (OR = 0.94, 95%CI = 0.89–0.98, $P_{\text{combined}} = 0.007$), and then in SQ (OR = 0.95, 95%CI = 0.90–1.00, $P_{\text{combined}} = 0.043$) (Figure 3).

In silico functional validation

In silico eQTL analysis was performed by using the lymphoblastoid cell lines from 373 individuals of European descent. The rs151606 genotypes demonstrated significant association with decreased mRNA expression of *FGFR1OP* in both additive ($P_{\text{trend}} = 0.022$) and recessive models ($P_{\text{trend}} = 0.031$) (Figure 4). According to the prediction results of the online tool F-SNP, SNP rs151606 might be associated with mRNA expression by

Table 2. Associations between SNPs in centrosomal genes and lung cancer risk with FDR $P \leq 0.050$

SNP	Gene	Chr	Position (hg19)	Allele ^a	EAF	Q ^b	I ²	Effects ^c	OR (95%CI)	P	FDR
rs115897636	TUBB	6	30689001	T/A	0.12	0.869	0	+++++	1.17 (1.11–1.24)	4.23E–09	<.0001
rs114884831	TUBB	6	30687614	G/C	0.11	0.725	0	+++++	1.18 (1.12–1.24)	4.98E–09	<.0001
rs115605279	TUBB	6	30688575	G/C	0.11	0.724	0	+++++	1.18 (1.12–1.24)	5.05E–09	<.0001
rs116812176	TUBB	6	30693816	A/G	0.20	0.456	0	+++++	1.12 (1.07–1.17)	3.60E–07	0.001
rs114380302	TUBB	6	30692965	G/A	0.20	0.448	0	+++++	1.12 (1.07–1.17)	3.78E–07	0.001
rs116338781	TUBB	6	30688427	G/T	0.20	0.453	0	+++++	1.12 (1.07–1.17)	4.88E–07	0.001
rs114460911	TUBB	6	30693633	A/G	0.20	0.522	0	+++++	1.11 (1.07–1.16)	6.09E–07	0.001
rs151606	FGFR1OP	6	167430482	A/T	0.34	0.552	0	+++++	1.12 (1.07–1.17)	1.18E–06	0.001
rs114075700	TUBB	6	30694374	T/C	0.18	0.485	0	+++++	1.11 (1.07–1.17)	2.01E–06	0.002
rs115969492	TUBB	6	30693121	A/G	0.33	0.780	0	+++++	1.10 (1.05–1.15)	9.78E–06	0.009
rs239935	FGFR1OP	6	167411788	A/G	0.48	0.309	16	+++++	1.07 (1.04–1.11)	5.03E–05	0.025
rs400837	FGFR1OP	6	167411008	C/T	0.47	0.338	12	+++++	1.07 (1.04–1.11)	6.28E–05	0.025
rs424185	FGFR1OP	6	167410907	T/C	0.47	0.340	12	+++++	1.07 (1.04–1.11)	6.78E–05	0.025
rs376097	FGFR1OP	6	167410878	G/A	0.47	0.340	12	+++++	1.07 (1.04–1.11)	6.78E–05	0.025
rs239934	FGFR1OP	6	167412048	A/G	0.49	0.379	6	+++++	1.07 (1.04–1.11)	7.24E–05	0.025
rs1322077	FGFR1OP	6	167424293	T/C	0.46	0.334	13	-----	0.93 (0.90–0.97)	7.35E–05	0.025
rs9457251	FGFR1OP	6	167426707	T/C	0.46	0.320	15	-----	0.93 (0.90–0.97)	7.54E–05	0.025
rs9457249	FGFR1OP	6	167426438	A/G	0.46	0.320	15	-----	0.93 (0.90–0.97)	7.58E–05	0.025
rs9459836	FGFR1OP	6	167419105	G/A	0.46	0.321	15	-----	0.93 (0.90–0.97)	8.56E–05	0.025
rs13195812	FGFR1OP	6	167443902	C/T	0.45	0.317	15	-----	0.93 (0.90–0.97)	8.59E–05	0.025
rs9459849	FGFR1OP	6	167444160	G/T	0.45	0.313	16	-----	0.93 (0.90–0.97)	8.69E–05	0.025
rs9459841	FGFR1OP	6	167431949	G/A	0.45	0.315	15	-----	0.93 (0.90–0.97)	8.77E–05	0.025
rs9459840	FGFR1OP	6	167431910	G/A	0.45	0.315	15	-----	0.93 (0.90–0.97)	8.77E–05	0.025
rs4710171	FGFR1OP	6	167430186	A/G	0.45	0.315	15	-----	0.93 (0.90–0.97)	8.79E–05	0.025
rs1060404	FGFR1OP	6	167429467	A/G	0.45	0.315	15	-----	0.93 (0.90–0.97)	8.80E–05	0.025
rs12212247	FGFR1OP	6	167413539	T/C	0.46	0.329	13	-----	0.93 (0.90–0.97)	8.93E–05	0.025
rs2237272	FGFR1OP	6	167443443	C/T	0.45	0.313	16	-----	0.93 (0.90–0.97)	9.17E–05	0.025
rs6904946	FGFR1OP	6	167433948	T/C	0.45	0.310	16	-----	0.93 (0.90–0.97)	9.19E–05	0.025
rs10484531	FGFR1OP	6	167454434	G/A	0.45	0.311	16	-----	0.93 (0.90–0.97)	9.21E–05	0.025
rs9457256	FGFR1OP	6	167436461	T/C	0.45	0.307	17	-----	0.93 (0.90–0.97)	9.26E–05	0.025
rs3752520	FGFR1OP	6	167436159	T/C	0.45	0.307	17	-----	0.93 (0.90–0.97)	9.26E–05	0.025
rs9457252	FGFR1OP	6	167433925	G/A	0.45	0.307	17	-----	0.93 (0.90–0.97)	9.27E–05	0.025
rs6929466	FGFR1OP	6	167422922	C/T	0.45	0.302	17	-----	0.93 (0.90–0.97)	9.53E–05	0.025
rs9295385	FGFR1OP	6	167448181	A/G	0.45	0.310	16	-----	0.93 (0.90–0.97)	9.63E–05	0.025
rs10455982	FGFR1OP	6	167448728	C/T	0.45	0.307	17	-----	0.93 (0.90–0.97)	9.74E–05	0.025
rs10946204	FGFR1OP	6	167451129	T/C	0.45	0.308	16	-----	0.93 (0.90–0.97)	9.75E–05	0.025
rs10946203	FGFR1OP	6	167446735	A/C	0.45	0.297	18	-----	0.93 (0.90–0.97)	9.88E–05	0.025
rs9534269	BRCA2	13	32939286	T/G	0.26	0.252	24	-----+	0.93 (0.89–0.96)	1.03E–04	0.025
rs6900701	FGFR1OP	6	167434112	A/G	0.45	0.331	13	-----	0.93 (0.90–0.97)	1.09E–04	0.026
rs2237276	FGFR1OP	6	167442115	C/T	0.45	0.299	18	-----	0.93 (0.90–0.97)	1.14E–04	0.026
rs9457254	FGFR1OP	6	167434135	G/A	0.45	0.330	13	-----	0.93 (0.90–0.97)	1.14E–04	0.026
rs9457259	FGFR1OP	6	167444281	C/T	0.45	0.323	14	-----	0.93 (0.90–0.97)	1.38E–04	0.031
rs9943876	BRCA2	13	32927894	C/T	0.30	0.438	0	-----+	0.93 (0.90–0.97)	1.48E–04	0.032
rs2237275	FGFR1OP	6	167442994	C/A	0.44	0.308	16	-----	0.93 (0.90–0.97)	1.70E–04	0.036

Chr, chromosome; EAF, effect allele frequency; FDR, false discovery rate; OR, odds ratio; SNP, single nucleotide polymorphism.

^aReference allele/effect allele.

^bFixed effect models were used when no heterogeneity was found between studies (Q test $P > 0.10$ and $I^2 < 50.0\%$); otherwise, random effect models were used.

^c‘+’ means positive association and ‘-’ means negative association.

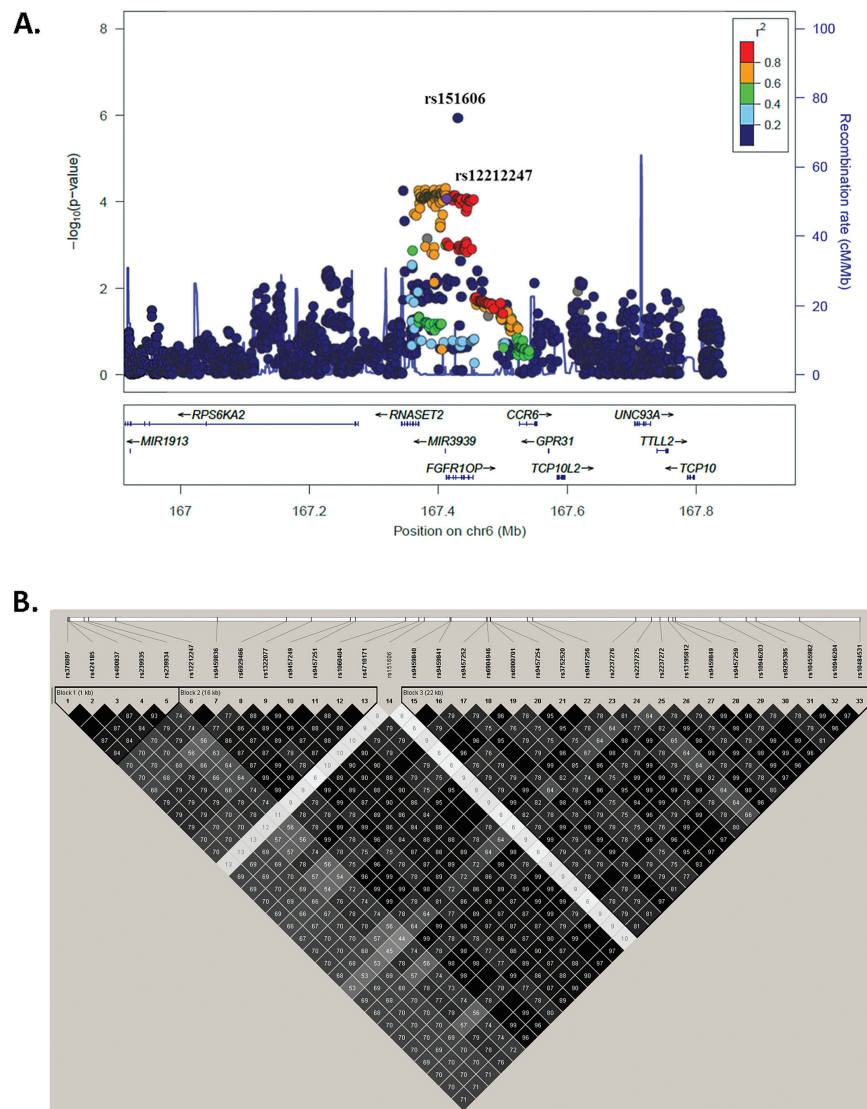


Figure 2. Regional association plots. Data points are colored according to their level of LD of the each SNP with the tagging SNPs. (A) SNPs in the region 500 kb up- or downstream of the marker SNP and (B) SNPs with $FDR \leq 0.050$. In A, the left-hand y-axis shows the association P value of individual SNPs, which is plotted as $-\log_{10}(P)$ against chromosomal base pair position; the right-hand y-axis shows the recombination rate estimated from the hg19/1000 Genomes European population.

influencing the binding activity of transcriptional factors (i.e. HNF-3b, XFD-2). Although rs12212247 might have an effect on the activity of transcription factor binding site (TFBS) as predicted by SNPinfo and RegulonDB score of 2b (Supplementary Table 4, available at Carcinogenesis Online), which indicated that transcription factor TAF1 binding might be influenced according to the Encyclopedia of DNA Elements (ENCODE) ChIP-Seq data from human A549 cells (35), we did not find any significant association of this SNP with *FGFR1OP* mRNA expression levels in either model (Figure 4).

Discussion

The centrosome is the crucial microtubule-organizing center in animal cells and comprises of a pair of centrioles surrounded by the pericentriolar material (36). Centrosome deficiencies, including the occurrence of extra centrioles, and increased ability to nucleate microtubules, are common in a high percentage of many tumors. While centrosome deficiencies in cancer may arise as a result of tumorigenesis, early centrosome aberrations may also lead to increased risk of malignancy (37). Our

study firstly attempted to systematically examine the effects of genetic variants in the centrosomal genes on lung cancer susceptibility. Notably, we found two potentially functional SNPs in *FGFR1OP* to be associated with lung cancer risk in the final meta-analysis of 14 463 lung cancer patients and 44 188 control subjects. Our current study is the first to provide an important insight into the association of variants in 6q27 and carcinogenesis, suggesting that *FGFR1OP* (rs151606 and rs12212247) could represent a novel lung cancer locus in Caucasian populations.

FGFR1OP (fibroblast growth factor receptor 1 oncogene partner) is a centriolar satellite cargo protein, by which centriolar satellites play the role of key mediators of centrosome functions (38). It has been reported that *FGFR1OP* is involved in G1/S transition and thus necessary for cell-cycle progression and survival (39). Two chromosomal translocations, fusing this gene with the *FGFR1* and *RET* genes, have been found in hematopoietic cancers (40,41). Published GWASs have identified several SNPs in the nearby region (6q27) of *FGFR1OP* to be associated with the risk of multiple autoimmune diseases, including Crohn's disease (rs2301436 in *FGFR1OP*) (42), Graves' disease (rs9355610 near *RNASET2*) (43), rheumatoid arthritis (rs3093024 and rs3093023 near *CCR6*) (44,45)

Table 3. Summary of the association results of two SNPs in the eight lung cancer GWASs

Study population	Sample size		Imputation quality	rs151606 A > T		Imputation quality	rs12212247 T > C	
	Cases	Controls		OR (95% CI)	P		OR (95% CI)	P
TRICL combined ^a	12 160	16 838		1.12 (1.07–1.17)	1.18E–06		0.93 (0.90–0.97)	8.93E–05
ICR ^b	1952	5200	0.74	1.19 (1.09–1.30)	1.24E–04	0.99	0.93 (0.86–1.00)	4.44E–02
MDACC ^c	1150	1134	0.45	1.10 (0.92–1.32)	3.10E–01	0.96	0.95 (0.84–1.07)	3.94E–01
IARC ^d	2533	3791	0.44	1.04 (0.93–1.17)	5.23E–01	0.98	0.97 (0.90–1.04)	3.52E–01
NCI ^e	5713	5736	0.72	1.10 (1.03–1.18)	3.28E–03	0.99	0.94 (0.89–0.99)	1.88E–02
Toronto ^f	331	499	0.70	1.19 (0.91–1.57)	2.06E–01	0.99	0.82 (0.66–1.02)	7.60E–02
GLC ^g	481	478	0.52	1.16 (0.89–1.50)	2.78E–01	0.98	0.78 (0.65–0.94)	9.02E–03
Replication combined ^a	2303	27 350		1.02 (0.93–1.11)	7.19E–01		0.93 (0.87–1.00)	3.90E–02
Harvard ^h	984	970	0.75	1.15 (0.95–1.39)	1.50E–01	1.00	0.88 (0.76–1.01)	5.91E–02
DeCODE ⁱ	1319	26 380	0.75	0.98 (0.89–1.08)	7.50E–01	1.00	0.95 (0.88–1.03)	1.96E–01
All combined ^a	14 463	44 188		1.10 (1.05–1.14)	8.03E–06		0.93 (0.90–0.96)	9.59E–06

GWAS, genome-wide association study; SNP, single-nucleotide polymorphism.

^aThe combined OR and P value were estimated using a fixed effects model.

^bICR: the Institute of Cancer Research Genome-wide Association Study, UK.

^cMDACC: the MD Anderson Cancer Center Genome-wide Association Study, USA.

^dIARC: the International Agency for Research on Cancer Genome-wide Association Study, France;

^eNCI: the National Cancer Institute Genome-wide Association Study, USA.

^fToronto: the Lunenfeld-Tanenbaum Research Institute Genome-wide Association Study, Toronto, Canada.

^gGLC: German Lung Cancer Study, Germany.

^hHarvard: Harvard Lung Cancer Study, USA.

ⁱdeCODE: Icelandic Lung Cancer Study, Iceland.

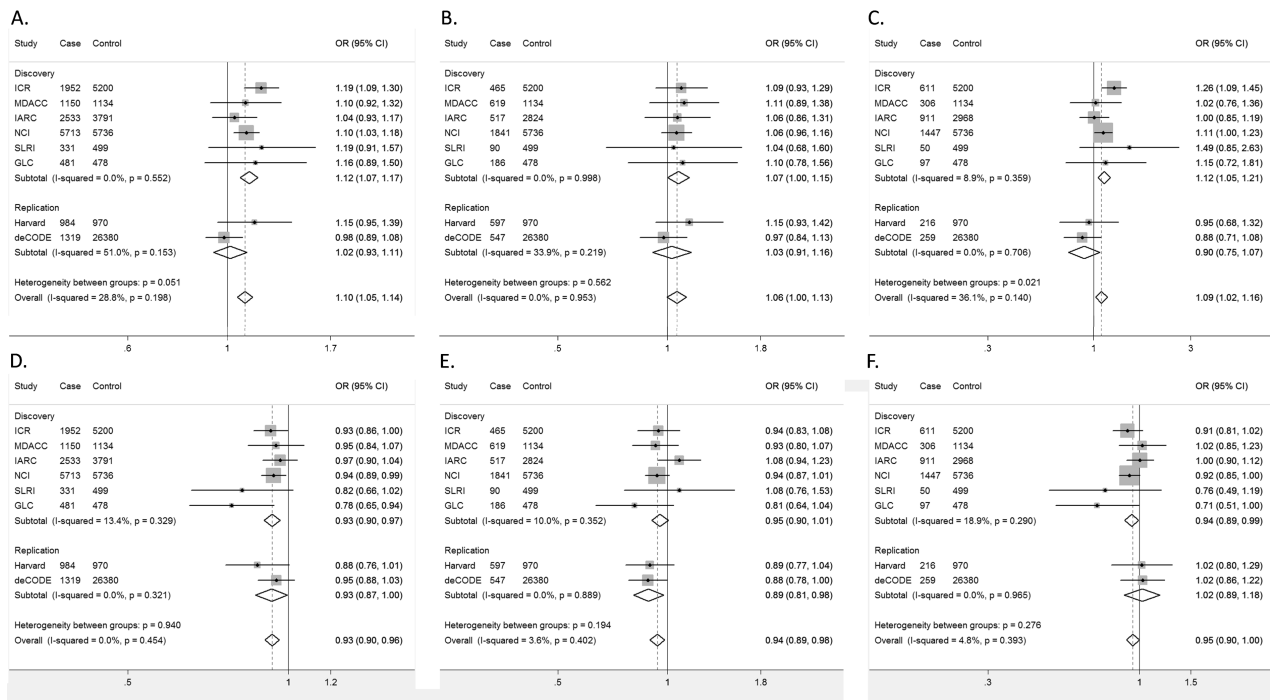


Figure 3. Forest plots of effect size and direction for tagSNPs with all cases from TRICL consortium. *FGFR1OP* rs151606 in all cases, $P_{\text{combined}} = 8.032 \times 10^{-6}$ (A); *FGFR1OP* rs151606 in adenocarcinoma cases, $P_{\text{combined}} = 0.058$ (B); *FGFR1OP* rs151606 in squamous cell carcinoma cases, $P_{\text{combined}} = 0.011$ (C); *FGFR1OP* rs12212247 in all cases, $P_{\text{combined}} = 9.589 \times 10^{-6}$ (D); *FGFR1OP* rs12212247 in adenocarcinoma cases, $P_{\text{combined}} = 0.007$ (E); *FGFR1OP* rs12212247 in squamous cell carcinoma cases, $P_{\text{combined}} = 0.043$ (F). Each box and horizontal line represent the OR point estimate and 95% CI derived from the additive model. The area of each box is proportional to the statistical weight of the study. Diamonds represent the summary ORs obtained from the combined analysis with 95% confidence intervals indicated by their widths. The meta-analysis includes eight GWASs [the Institute of Cancer Research (ICR) GWAS, the MD Anderson Cancer Center (MDACC) GWAS, the International Agency for Research on Cancer (IARC) GWAS, the National Cancer Institute (NCI) GWAS, the Lunenfeld-Tanenbaum Research Institute (Toronto) GWAS, German Lung Cancer Study (GLC), Harvard lung cancer study (Harvard) and Icelandic Lung Cancer Study (deCODE)].

and vitiligo (rs2236313 in *RNASET2* and rs6902119 near *CCR6*) (46,47). Recent studies have also suggested a link between autoimmune diseases and some cancers (48–50). However, little is known about the association between the genetic variants in *FGFR1OP*

and cancer risk. In this study, we observed that *FGFR1OP* rs151606 T allele was associated with decreased levels of mRNA expression and increased risk of lung cancer. These results indicate that the association between SNP rs151606 and lung cancer risk might due

to its effect on the expression of *FGFR10P*, as suggested the eQTL evidence for a role of *FGFR10P* in the assembling of mitotic spindle (10,13,51). However, the exact molecular mechanism underlying the associations among lower expression of *FGFR10P* on spindle multipolarity, centrosome amplification and lung cancer risk warrants additional investigation.

Except for dysregulation of gene expressions, exposure to environmental risk factors has also been shown to induce centrosome abnormality (52). Several studies have reported that exposure to chrysotile asbestos fibers, chromate particles, arsenite, and benzo[a]pyrene diol epoxide, which is a carcinogen present in tobacco smoke as well as in environmental pollution, might cause centrosome abnormalities and multipolar spindles in human lung cells (53–57). Early occurrence of centrosome alterations have been found in the uninvolved lung tissues adjacent to the tumor of NSCLCs, representing an early event in pulmonary carcinogenesis by increasing chromosome instability (58,59). Based on these previous reports and our current findings, analysis of an interaction between SNPs and main environmental risk factors (i.e. smoking status) in future

studies might provide novel evidence for the effects of genetic variants in centrosomal genes on lung cancer risk.

Previous studies have identified several genetic variants in centrosomal genes (*NIN*, *TUBG1* and *APC*) that contributed to breast (60) or pancreatic (61) cancer susceptibility among the Caucasian populations. However, in the present study, we did not find any significance for those SNPs but identified four other SNPs within *NIN* (rs67977855, rs45558232, rs59096640 and rs4534750) showing moderate associations with lung cancer risk ($P \leq 0.05$) based on the six TRICL GWASs results (data not shown). Such inconsistency might due to the heterogeneity among cancers or populations.

There are some potential limitations of the present study. Firstly, the gene-set selection mainly depended on the quality and integrity of curated biological pathways. As the biological understanding of the function of diverse genes in pathways has been steadily improved, the emerging new data on functions of these genes will inevitably expand the currently available lists of ‘canonical’ pathways. Some important centrosomal genes might be omitted in this study. Considering this, we comprehensively

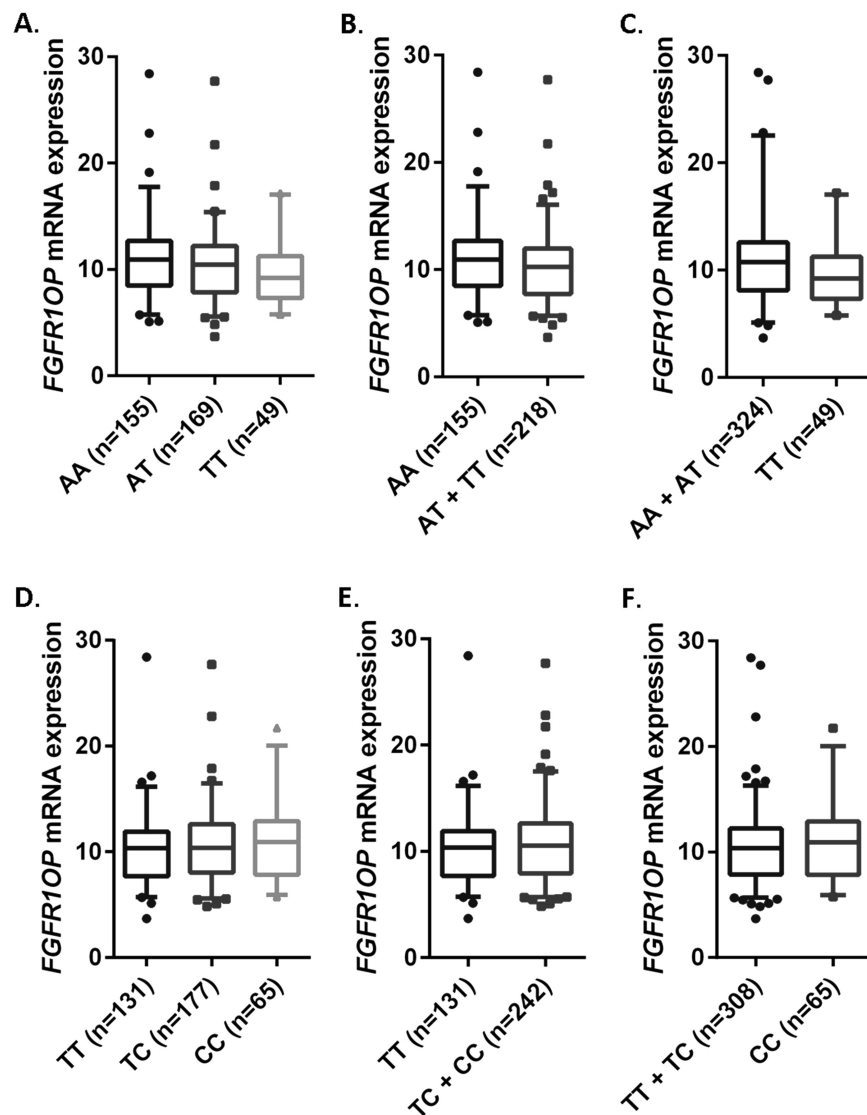


Figure 4. The correlations between identified SNPs and *FGFR10P* mRNA expression. rs151606 (A, additive model, $P = 0.022$; B, dominant model, $P = 0.088$; C, recessive model, $P = 0.031$) and rs12212247 (D, additive model, $P = 0.344$; E, dominant model, $P = 0.465$; F, recessive model, $P = 0.405$).

considered multiple centrosomal gene sets and managed to make an all-inclusive listing as possible as we could. Secondly, defining whether a particular SNP is of biological function was limited by the applied *in silico* tools. To overcome this limitation, we combined the prediction results of multiple bioinformatics tools to explain our data. Thirdly, our eQTL analyses were limited to lymphoblastoid cell lines with publically available data, and gene expression analysis in tissues will give biologically plausible and more convincing results of the effect of those two identified SNPs.

In summary, the present study found significant associations between SNPs in *FGFR10P* and lung cancer risk. The results suggested that the SNPs within the centrosomal genes may serve as potential markers to predict lung cancer risk in European populations. Our discoveries shed some new light on the role of centrosome abnormalities in human carcinogenesis. Further validation and functional evaluation of these genetic variants are needed to verify our findings.

URLs

Transdisciplinary Research In Cancer of the Lung (TRICL), <http://u19tricl.org/>; Genetic Associations and MEchanisms in ONcology (GAME-ON) consortium, <http://epi.grants.cancer.gov/gameon/>; International Lung Cancer Consortium (ILCCO), <http://ilcco.iarc.fr/>; 1000 Genomes Project, <http://www.1000genomes.org/>; IMPUTE2, http://mathgen.stats.ox.ac.uk/impute/impute_v2.html; MaCH, <http://www.sph.umich.edu/csg/abecasis/MACH/>; Minimac, <http://genome.sph.umich.edu/wiki/Minimac/>; MsigDB, <http://www.broadinstitute.org/gsea/msigdb/index.jsp>; LocusZoom, <http://csg.sph.umich.edu/locuszoom/>; RegulomeDB, <http://regulome.stanford.edu/>; SNPinfo, <http://snpinfo.niehs.nih.gov/snpinfo/snpfunc.htm>; F-SNP, <http://compbio.cs.queensu.ca/F-SNP/>.

Supplementary material

Supplementary Tables 1–4 and Figure 1 can be found at <http://carcin.oxfordjournals.org/>

Funding

As Duke Cancer Institute members, Q.W., M.W.O., and K.O. acknowledge support from the Duke Cancer Institute as part of the P30 Cancer Center Support Grant (NIH CA014236). Q.W. was also supported by a start-up fund from Duke Cancer Institute, Duke University Medical Center.

TRICL

Transdisciplinary Research in Cancer of the Lung (TRICL) Study, U19-CA148127 on behalf of the Genetic Associations and Mechanisms in Oncology (GAME-ON) Network. The Toronto study was supported by Canadian Cancer Society Research Institute(020214), Ontario Institute of Cancer and Cancer Care Ontario Chair Award to RH The ICR study was supported by Cancer Research UK (C1298/A8780 and C1298/A8362—Bobby Moore Fund for Cancer Research UK) and NCRN,HEAL and Sanofi-Aventis. Additional funding was obtained from NIH grants (5R01CA055769, 5R01CA127219, 5R01CA133996 and 5R01CA121197). The Liverpool Lung Project (LLP) was supported by The Roy Castle Lung Cancer Foundation, UK. The ICR and LLP studies made use of genotyping data from the Wellcome Trust Case Control Consortium 2 (WTCCC2); a full list of the investigators who contributed to the generation of the data is available from www.wtccc.org.uk. Sample collection for the Heidelberg lung cancer study was in part supported by a grant (70–2919) from the Deutsche Krebshilfe. The work was additionally supported by a Helmholtz-DAAD fellowship (A/07/97379 to MNT)

and by the NIH (U19CA148127). The KORA Surveys were financed by the GSF, which is funded by the German Federal Ministry of Education, Science, Research and Technology and the State of Bavaria. The Lung Cancer in the Young study (LUCY) was funded in part by the National Genome Research Network (NGFN), the DFG (BI576/2-1; BI 576/2-2), the Helmholtzgemeinschaft (HGF) and the Federal office for Radiation Protection (BfS: STSch4454). Genotyping was performed in the Genome Analysis Center (GAC) of the Helmholtz Zentrum Muenchen. Support for the Central Europe, HUNT2/Tromsø and CARET genome-wide studies was provided by Institut National du Cancer, France. Support for the HUNT2/Tromsø genome-wide study was also provided by the European Community (Integrated Project DNA repair, LSHG-CT- 2005–512113), the Norwegian Cancer Association and the Functional Genomics Programme of Research Council of Norway. Support for the Central Europe study, Czech Republic, was also provided by the European Regional Development Fund and the State Budget of the Czech Republic (RECAMO, CZ.1.05/2.1.00/03.0101). Support for the CARET genome-wide study was also provided by grants from the US National Cancer Institute, NIH (R01 CA111703 and UO1 CA63673), and by funds from the Fred Hutchinson Cancer Research Center. Additional funding for study coordination, genotyping of replication studies and statistical analysis was provided by the US National Cancer Institute (R01 CA092039). The lung cancer GWAS from Estonia was partly supported by a FP7 grant (REGPOT245536), by the Estonian Government (SF0180142s08), by EU RDF in the frame of Centre of Excellence in Genomics and Estonian Research Infrastructure's Roadmap and by University of Tartu (SP1GVARENG). The work reported in this article was partly undertaken during the tenure of a Postdoctoral Fellowship from the IARC (for MNT). The Environment and Genetics in Lung Cancer Etiology (EAGLE), the Alpha-Tocopherol, Beta-Carotene Cancer Prevention Study (ATBC), and the Prostate, Lung, Colon, Ovary Screening Trial (PLCO) studies and the genotyping of ATBC, the Cancer Prevention Study II Nutrition Cohort (CPS-II) and part of PLCO were supported by the Intramural Research Program of NIH, NCI, Division of Cancer Epidemiology and Genetics. ATBC was also supported by US Public Health Service contracts (N01-CN-45165, N01-RC-45035 and N01-RC-37004) from the NCI. PLCO was also supported by individual contracts from the NCI to the University of Colorado Denver (N01-CN-25514), Georgetown University (N01-CN-25522), Pacific Health Research Institute (N01-CN-25515), Henry Ford Health System (N01-CN-25512), University of Minnesota (N01-CN-25513), Washington University (N01-CN-25516), University of Pittsburgh (N01-CN-25511), University of Utah (N01-CN-25524), Marshfield Clinic Research Foundation (N01-CN-25518), University of Alabama at Birmingham (N01-CN-75022, Westat, Inc. N01-CN-25476), University of California, Los Angeles (N01-CN-25404). The Cancer Prevention Study II Nutrition Cohort was supported by the American Cancer Society. The NIH Genes, Environment and Health Initiative (GEI) partly funded DNA extraction and statistical analyses (HG-06-033-NCI-01 and R01HL091172-01), genotyping at the Johns Hopkins University Center for Inherited Disease Research (U01HG004438 and NIH HHSN268200782096C) and study coordination at the GENEVA Coordination Center (U01 HG004446) for EAGLE and part of PLCO studies. Funding for the MD Anderson Cancer Study was provided by NIH grants (P50 CA70907, R01CA121197, R01CA127219, U19 CA148127, R01 CA55769 and K07CA160753) and CPRIT grant (RP100443). Genotyping services were provided by the Center for Inherited Disease Research (CIDR). CIDR is funded through a federal contract from the NIH to The Johns Hopkins University (HHSN268200782096C). The Harvard Lung Cancer Study was supported by the NIH (National Cancer Institute) grants CA092824, CA090578 and CA074386.

deCODE

The deCODE project was funded in part by the National Institutes of Health (DA017932). Approval for the deCODE study was granted by the Icelandic National Bioethics Committee (ref. 12-122-V7) and the Icelandic Data Protection Authority (refs. 2001/25 and 2006/518).

Conflict of Interest Statement: None declared.

References

- Jemal, A. et al. (2011) Global cancer statistics. *CA. Cancer J. Clin.*, 61, 69–90.
- Sun, S. et al. (2007) Lung cancer in never smokers—a different disease. *Nat. Rev. Cancer*, 7, 778–790.
- Hung, R.J. et al. (2008) A susceptibility locus for lung cancer maps to nicotinic acetylcholine receptor subunit genes on 15q25. *Nature*, 452, 633–637.
- Amos, C.I. et al. (2008) Genome-wide association scan of tag SNPs identifies a susceptibility locus for lung cancer at 15q25.1. *Nat. Genet.*, 40, 616–622.
- McKay, J.D. et al. (2008) Lung cancer susceptibility locus at 5p15.33. *Nat. Genet.*, 40, 1404–6.
- Pande, M. et al. (2011) Novel genetic variants in the chromosome 5p15.33 region associate with lung cancer risk. *Carcinogenesis*, 32, 1493–1499.
- Wang, Y. et al. (2011) Variation in TP63 is associated with lung adenocarcinoma in the UK population. *Cancer Epidemiol. Biomarkers Prev.*, 20, 1453–1462.
- Wang, Y. et al. (2014) Rare variants of large effect in BRCA2 and CHEK2 affect risk of lung cancer. *Nat. Genet.*, 46, 736–741.
- Gergely, F. et al. (2008) Multiple centrosomes: together they stand, divided they fall. *Genes Dev.*, 22, 2291–2296.
- Zyss, D. et al. (2009) Centrosome function in cancer: guilty or innocent? *Trends Cell Biol.*, 19, 334–346.
- Fukasawa, K. (2007) Oncogenes and tumour suppressors take on centrosomes. *Nat. Rev. Cancer*, 7, 911–924.
- Ganem, N.J. et al. (2009) A mechanism linking extra centrosomes to chromosomal instability. *Nature*, 460, 278–282.
- Nigg, E.A. et al. (2009) Centrioles, centrosomes, and cilia in health and disease. *Cell*, 139, 663–678.
- Anderhub, S.J. et al. (2012) Centrosome amplification in tumorigenesis. *Cancer Lett.*, 322, 8–17.
- Pihan, G.A. (2013) Centrosome dysfunction contributes to chromosome instability, chromoanagenesis, and genome reprogramming in cancer. *Front. Oncol.*, 3, 277.
- Godinho, S.A. et al. (2014) Oncogene-like induction of cellular invasion from centrosome amplification. *Nature*, 510, 167–171.
- Shinmura, K. et al. (2012) Centrosome abnormality and human lung cancer. *Lung Dis.*, 171–188.
- Timofeeva, M.N. et al. (2012) Influence of common genetic variation on lung cancer risk: meta-analysis of 14 900 cases and 29 485 controls. *Hum. Mol. Genet.*, 21, 4980–4995.
- Su, L. et al. (2006) Genotypes and haplotypes of matrix metalloproteinase 1, 3 and 12 genes and the risk of lung cancer. *Carcinogenesis*, 27, 1024–1029.
- Thorgeirsson, T.E. et al. (2008) A variant associated with nicotine dependence, lung cancer and peripheral arterial disease. *Nature*, 452, 638–642.
- Consortium, G.P. (2012) An integrated map of genetic variation from 1,092 human genomes. *Nature*, 491, 56–65.
- Howie, B.N. et al. (2009) A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS Genet.*, 5, e1000529.
- Li, Y. et al. (2010) MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. *Genet. Epidemiol.*, 34, 816–834.
- Howie, B. et al. (2012) Fast and accurate genotype imputation in genome-wide association studies through pre-phasing. *Nat. Genet.*, 44, 955–959.
- Wang, Y. et al. (2015) Deciphering associations for lung cancer risk through imputation and analysis of 12 316 cases and 16 831 controls. *Eur. J. Hum. Genet.*, 23, 1723–1728.
- Xu, Z. et al. (2009) SNPinfo: integrating GWAS and candidate gene information into functional SNP selection for genetic association studies. *Nucleic Acids Res.*, 37, W600–W605.
- Boyle, A.P. et al. (2012) Annotation of functional variation in personal genomes using RegulomeDB. *Genome Res.*, 22, 1790–1797.
- Lee, P.H. et al. (2008) F-SNP: computationally predicted functional SNPs for disease association studies. *Nucleic Acids Res.*, 36, D820–D824.
- Purcell, S. et al. (2007) PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am. J. Hum. Genet.*, 81, 559–575.
- Higgins, J.P. et al. (2003) Measuring inconsistency in meta-analyses. *BMJ*, 327, 557–560.
- Benjamini, Y. et al. (1995) Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J. Roy. Statist. Soc. B (Methodological)*, 57, 289–300.
- Pruim, R.J. et al. (2010) LocusZoom: regional visualization of genome-wide association scan results. *Bioinformatics*, 26, 2336–2337.
- Barrett, J.C. et al. (2005) Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics*, 21, 263–265.
- Broderick, P. et al. (2009) Deciphering the impact of common genetic variation on lung cancer risk: a genome-wide association study. *Cancer Res.*, 69, 6633–6641.
- Consortium, E.P. (2012) An integrated encyclopedia of DNA elements in the human genome. *Nature*, 489, 57–74.
- Bettencourt-Dias, M. et al. (2007) Centrosome biogenesis and function: centrosomes brings new understanding. *Nat. Rev. Mol. Cell Biol.*, 8, 451–463.
- Marx, J. (2001) Cell biology. Do centrosome abnormalities lead to cancer? *Science*, 292, 426–429.
- Tollenaere, M.A. et al. (2015) Centriolar satellites: key mediators of centrosome functions. *Cell. Mol. Life Sci.*, 72, 11–23.
- Acquaviva, C. et al. (2009) The centrosomal FOP protein is required for cell cycle progression and survival. *Cell Cycle*, 8, 1217–1227.
- Popovici, C. et al. (1999) The t(6;8)(q27;p11) translocation in a stem cell myeloproliferative disorder fuses a novel gene, FOP, to fibroblast growth factor receptor 1. *Blood*, 93, 1381–1389.
- Mulligan, L.M. (2014) RET revisited: expanding the oncogenic portfolio. *Nat. Rev. Cancer*, 14, 173–186.
- Barrett, J.C. et al. (2008) Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. *Nat. Genet.*, 40, 955–962.
- Chu, X. et al. (2011) A genome-wide association study identifies two new risk loci for Graves' disease. *Nat. Genet.*, 43, 897–901.
- Kochi, Y. et al. (2010) A regulatory variant in CCR6 is associated with rheumatoid arthritis susceptibility. *Nat. Genet.*, 42, 515–519.
- Stahl, E.A. et al. (2010) Genome-wide association study meta-analysis identifies seven new rheumatoid arthritis risk loci. *Nat. Genet.*, 42, 508–514.
- Jin, Y. et al. (2010) Common variants in FOXP1 are associated with generalized vitiligo. *Nat. Genet.*, 42, 576–578.
- Quan, C. et al. (2010) Genome-wide association study for vitiligo identifies susceptibility loci at 6q27 and the MHC. *Nat. Genet.*, 42, 614–618.
- Teulings, H.E. et al. (2013) Decreased risk of melanoma and non-melanoma skin cancer in patients with vitiligo: a survey among 1307 patients and their partners. *Br. J. Dermatol.*, 168, 162–171.
- Joseph, C.G. et al. (2014) Association of the autoimmune disease scleroderma with an immunologic response to cancer. *Science*, 343, 152–157.
- Franks, A.L. et al. (2012) Multiple associations between a broad spectrum of autoimmune diseases, chronic inflammatory diseases and cancer. *Anticancer Res.*, 32, 1119–1136.
- Bettencourt-Dias, M. et al. (2011) Centrosomes and cilia in human disease. *Trends Genet.*, 27, 307–315.
- Chan, J.Y. (2011) A clinical overview of centrosome amplification in human cancers. *Int. J. Biol. Sci.*, 7, 1122–1144.
- Cortez, B.A. et al. (2008) Chrysolite effects on human lung cell carcinoma in culture: 3-D reconstruction and DNA quantification by image analysis. *BMC Cancer*, 8, 181.
- Xie, H. et al. (2007) Neoplastic transformation of human bronchial cells by lead chromate particles. *Am. J. Respir. Cell Mol. Biol.*, 37, 544–552.
- Holmes, A.L. et al. (2010) Chronic exposure to zinc chromate induces centrosome amplification and spindle assembly checkpoint bypass in human lung fibroblasts. *Chem. Res. Toxicol.*, 23, 386–395.

-
56. Liao, W.T. et al. (2007) Arsenic promotes centrosome abnormalities and cell colony formation in p53 compromised human lung cells. *Toxicol. Appl. Pharmacol.*, 225, 162–170.
 57. Shinmura, K. et al. (2008) Induction of centrosome amplification and chromosome instability in p53-deficient lung cancer cells exposed to benzo[a]pyrene diol epoxide (B[a]PDE). *J. Pathol.*, 216, 365–374.
 58. Jung, C.K. et al. (2007) Centrosome abnormalities in non-small cell lung cancer: correlations with DNA aneuploidy and expression of cell cycle regulatory proteins. *Pathol. Res. Pract.*, 203, 839–847.
 59. Matsuura, S. et al. (2013) SGOL1 variant B induces abnormal mitosis and resistance to taxane in non-small cell lung cancers. *Sci. Rep.*, 3, 3012.
 60. Olson, J.E. et al. (2011) Centrosome-related genes, genetic variation, and risk of breast cancer. *Breast Cancer Res. Treat.*, 125, 221–228.
 61. Couch, F.J. et al. (2010) Association of mitotic regulation pathway polymorphisms with pancreatic cancer risk and outcome. *Cancer Epidemiol. Biomarkers Prev.*, 19, 251–257.