

Concentrations of cadmium and selected essential elements in malignant large intestine tissue

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Abstract

Introduction: Colorectal cancer is one of the most common cancers worldwide. Incidence rates of large intestine cancer indicate a role of environmental and occupational factors. The role of essential elements and their interaction with toxic metals can contribute to the explanation of a complex mechanism by which large intestine cancer develops. Bearing this in mind, determining the levels of essential and toxic elements in tissues (organs), as well as in body fluids, seems to shed light on their role in the mode of action in malignant disease.

Aim: Determination of the levels of cadmium, zinc, copper, selenium, calcium, magnesium, and iron in large intestine malignant tissue.

Material and methods: Two intraoperative intestine sections were investigated: one from the malignant tissue and the other one from the normal tissue, collected from each person with diagnosed large intestine cancer. Cadmium, zinc, copper, calcium, magnesium, and iron levels were determined with atomic absorption spectrometry, and selenium levels by spectrofluorimetric method.

Results: The levels of copper, selenium, and magnesium were higher in the malignant than in normal tissues. In addition, the zinc/copper and calcium/magnesium relationship was altered in malignant tissue, where correlations were lower compared to non-malignant tissue.

Conclusions: The results seem to demonstrate disturbed homeostasis of some essential elements. However, it is hard to confirm their involvement in the aetiology of colorectal cancer.

Introduction

Colorectal cancer is the fourth most common cancer in men and the third most common cancer in women, worldwide. The highest incidence rates are recorded in Europe, North America, and Oceania, and the lowest in South America, Africa, and Asia (e.g. about 4 cases per 100 000 population in India), although select registries in Asia (i.e. Japan, Singapore, and Israel) also record high rates. The great variation in the incidence rates of large intestine cancer observed in different parts of the world indicates a significant role of environmental factors (e.g. diet, physical activity) and so-called “western-style living” [1]. Modification of risk factors, primarily by promoting a well balanced diet, regular physical activity, and maintenance of normal body mass, can re-

duce cancer incidence by as much as 50–75% [2]. It has been also discussed whether occupational factors are responsible for increased risk of colon cancer. A higher risk of colorectal cancer has been reported *alter alia* among workers exposed to asbestos or wood dust, or those working in the textile industry. Moreover, some pesticides have also been associated with excess risk of colon cancer among pesticide applicators [3].

Large intestine cancer is the final stage of a dynamic process of dysplastic transformation, occurring most frequently in adenoma. Most colon cancers probably arise from benign neoplastic polyps [4]. In large intestine cancer, like in other cancers, early diagnosis and adequate therapeutic procedures provide a good chance of complete recovery. According to the literature data, an early diagnosis can ensure a large proportion of re-

covery, reaching as much as 90%. However, the chance of recovery diminishes proportionally to cancer progression. It should be noted that its detection in the most advanced state of malignancy limits 5-year survival to 5% [5].

Growing interest in concentrations of essential elements (e.g. magnesium (Mg), calcium (Ca), copper (Cu), zinc (Zn), iron (Fe), and selenium (Se)) in normal and malignant tissues and differences in their distribution have been observed for a number of years. Numerous reports stress the importance of determining serum copper and zinc levels in assessing activity and prognosis of the disease. Some studies have shown increased serum copper and decreased serum zinc levels in patients with sarcoma, lung cancer, and carcinoma of the digestive organs [6, 7]. However, the mechanisms by which the level of these elements decreases and increases in various cancerous conditions, as well as the issue of whether the altered serum copper and zinc levels are causative factors of the malignant state or its sequel, have not yet been elucidated. Nevertheless, it is obvious that in their attempts to identify possible causes of cancers and thus to facilitate their early diagnosis, researchers also try to analyse the role played by the elements in neoplastic processes.

Reports on the role of essential and toxic elements in the aetiology of large intestine cancer are rather scarce in the literature available to date. The role of copper and its metabolism in the human body has been the subject of much interest among numerous researchers. Its significant involvement in the synthesis of the connective tissue and haemoglobin, and in the normal functioning of the peripheral nervous system has already been evidenced. Tissue and plasma copper levels differ considerably depending on the place of residence, and thus on their content in the diet [8].

Zinc is an activator of several hundred enzymes participating, among others, in RNA replication and DNA repair. Zn deficiency impairs cellular and humoral immunity, and also limits the cytokine production [9]. Zn has also been recognised as an antioxidant. An effective anti-oxidative defence depends greatly on the regular metabolism of essential elements; therefore, it is not surprising that a great body of studies have been focused on their role in the development of carcinomas and large intestine adenocarcinoma. In the experimental adenocarcinoma, selenium stimulates DNA repair [10]. The protective role of Se in cancer is supported also by clinical and epidemiological studies [11], where an inverse association between Se supplementation and the risk of colorectal cancer was observed [12].

Epidemiological studies have demonstrated an association between the risk of colon cancer and low intake

of Mg. Moreover, a diet rich in Mg was found to reduce the occurrence of this cancer. Animal studies established that Mg could be a protective agent in the early stages of carcinogenesis. Nevertheless, it could also promote the growth of existing malignancies at later stages [13, 14]. It has been shown that Ca, Zn, and Cu can also reduce the risk of colon and rectum cancers [15].

Cadmium is one of the known factors responsible for the disturbed homeostasis of essential metals. According to Elinder *et al.*, cadmium, along with nephrotoxic and carcinogenic (lung cancer) effects, induces changes in Zn, Cu, Fe, Mg, Ca, and Se interaction-based distribution, which can be manifested by the deficiency of these compounds in individual tissues, diminished haematological indices (iron, haemoglobin, haematocrit), disturbed metabolism of carbohydrates (decreased insulin secretion), and induced lipid peroxidation [16].

To be able to shed more light on the role of essential elements in their mode of action in malignant disease, the content of each element in the tumour tissue itself should be determined. Apparently the identification of the role of elements and their interaction with toxic metals (cadmium) can significantly contribute to the explanation of a complex mechanism by which inter alia large intestine cancer develops.

Aim

In view of the limited data available to date, the need to carry out studies aimed at determining the levels of metals and essential and toxic elements in tissues (organs) and body fluids seems to be well founded, so the aim of this study was to determine the levels of cadmium, zinc, copper, selenium, calcium, magnesium, and iron in intraoperative sections of large intestine malignant tissues and in normal tissues taken as the control.

Material and methods

The study was carried out in a group of 25 persons (16 men and 9 women) with diagnosed large intestine cancer (*Adenocarcinoma mucinosum*). Two intraoperative intestine sections, one from the malignant tissue and the other from normal tissue, located at a maximum distance from the focus of malignancy, were collected from each person. All of the tissues were examined morphologically to confirm the diagnosis. The samples were stored in polyurethane containers at -70°C until examination. None of the patients from whom the tissues were collected had previously been subjected to radiotherapy or bio-element supplementation. All of the patients were matched for age and cancer progression, had not been occupationally exposed

Table I. Cadmium (Cd), zinc (Zn), copper (Cu), selenium (Se), magnesium (Mg), calcium (Ca), and iron (Fe) concentrations (mean \pm SD; $\mu\text{g/g}$ wet tissue) in normal and malignant tissues of large intestine

Element	Malignant tissue	Normal tissue
Cd	0.04 \pm 0.03	0.04 \pm 0.02
Zn	17.50 \pm 5.40	15.35 \pm 6.30
Cu	1.10 \pm 0.57*	0.64 \pm 0.39
Se	0.12 \pm 0.07*	0.05 \pm 0.03
Mg	76.78 \pm 31.41*	43.81 \pm 18.91
Ca	58.53 \pm 9.25	47.94 \pm 13.18
Fe	31.92 \pm 15.18	34.06 \pm 11.56

*Statistically significant difference ($p < 0.05$).

to heavy metals, and were non-smokers. This study was approved by the Ethics Committee for Scientific Research at the Medical University in Lodz (Resolution No RNN/156/02/KE).

Cadmium levels were determined with graphite furnace absorption spectrometry (HITACHI Z8270), and zinc, copper, calcium, magnesium, and iron levels with flame atomic absorption spectrometry (GBC Avanta PM) following mineralisation. Selenium levels were determined using the spectrofluorimetric method (HITACHI F4500) [17].

The limits of detection, calculated as concentrations corresponding with the value of absorption equal to a threefold standard deviation of the signal for the lowest standard, were respectively 0.001 $\mu\text{g/ml}$ for cadmium, 0.02 $\mu\text{g/ml}$ for zinc, 0.02 $\mu\text{g/ml}$ for copper, 0.15 $\mu\text{g/ml}$ for selenium, 0.01 $\mu\text{g/ml}$ for calcium, 0.01 $\mu\text{g/ml}$ for magnesium, and 0.01 $\mu\text{g/ml}$ for iron.

The intra-laboratory quality control was based on reference material SRM 1577b – lyophilised bovine liver

(National Institute of Standards & Technology, Gaithersburg, Germany), with certified measurements of the following elements [$\mu\text{g/g}$]: Cd (0.5 \pm 0.03), Zn (127 \pm 16), Cu (160 \pm 8), Se (0.73 \pm 0.06), Ca (116 \pm 4), Mg (601 \pm 28), and Fe (184 \pm 15). Mean discrepancies between the obtained results, compared with certified values expressed as RSD, were: Cd \pm 2.1%, Zn \pm 0.3%, Cu \pm 8.3%, Se \pm 3.5%, Ca \pm 6.7%, Mg \pm 7.2%, and Fe \pm 5.8%. The error of repeatability did not exceed 10% in any of the study samples.

Results

The results of the study are presented in Table I. The concentration of selenium in malignant tissue was more than two times higher than in normal tissue, and this was the highest statistically significant difference in our study. Levels of copper and magnesium were also significantly higher in malignant tissue – respectively, 170% and 180% of value determined in normal tissue. Cancerous transformation does not seem to affect zinc, calcium, cadmium, and iron levels, which were similar in malignant and non-malignant tissue.

The determined values of elements were also analysed in terms of their interrelations and correlations. On the basis of adequate calculations, a high correlation (almost 1) was observed between Zn and Cu concentrations in the normal tissue, but it was disturbed in the malignant tissue, where it was much lower (Figure 1). Similarly large differences between correlations were noted between Ca and Mg concentrations. Although the correlation between these elements in normal tissue was moderate; in the malignant tissue there seemed to be no relationship at all (Figure 2).

Discussion

The majority of published literature data focus on the determination of essential elements in blood se-

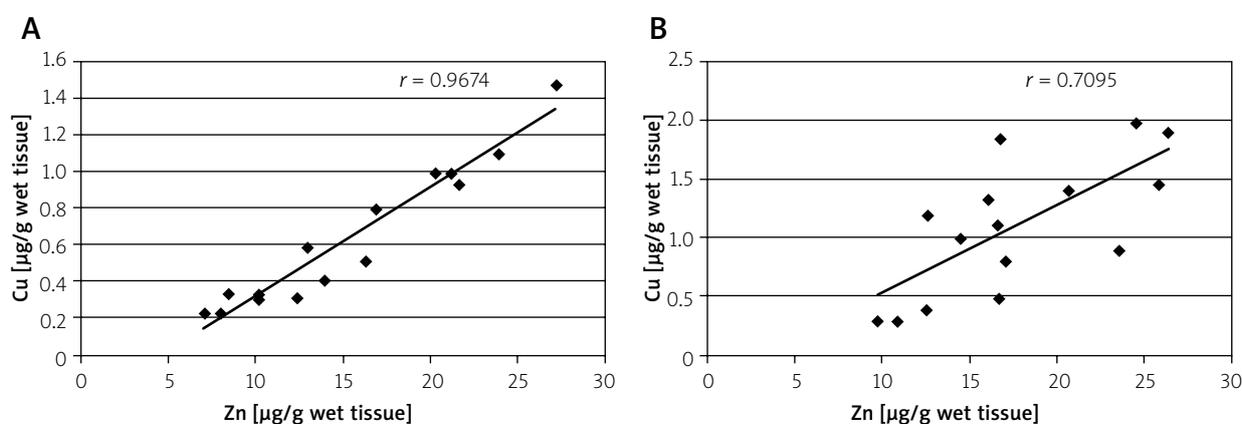


Figure 1. Correlation between zinc (Zn) and copper (Cu) concentrations in normal (A) and malignant (B) tissues of large intestine

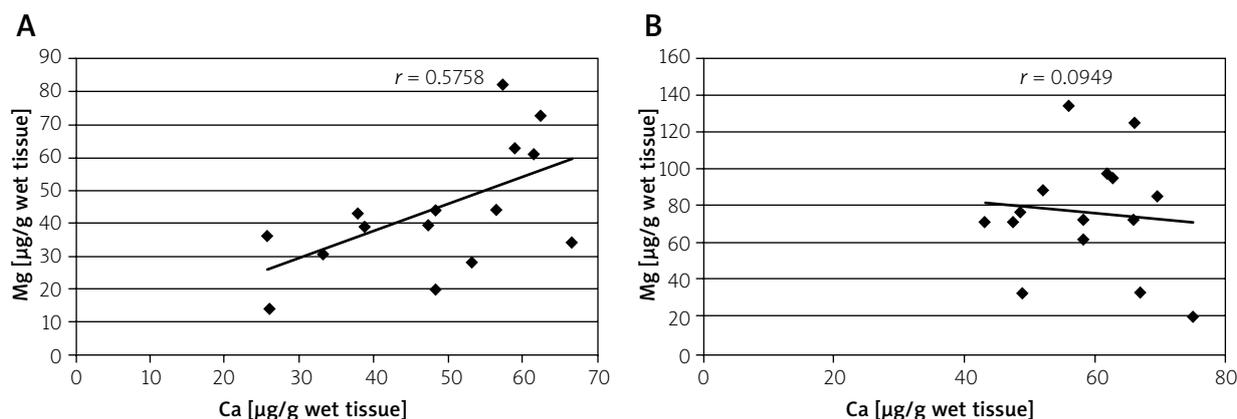


Figure 2. Correlation between calcium (Ca) and magnesium (Mg) concentrations in normal (A) and malignant (B) tissues of large intestine

rum to identify a biomarker useful in early detection of malignant pathologies. Studies carried out to observe changes in concentrations of these elements in organs and tissues already affected by neoplastic lesions are less numerous. It is well known that the observed alterations in element concentrations in blood serum result, among others, from changes in homeostasis of essential elements in malignant tissues.

The literature data published in the last decade indicate that disturbed levels of elements, mostly zinc and copper, can play a significant role in the mechanism responsible for uncontrolled cellular growth. It is thought that Cu plays an essential role in the activation of endothelium cells by stimulating angiogenic growth factors, which results in stimulation of the proliferation process [18]. Copper involvement in the carcinogenic process may also be linked to its ability to bind to some proteins, thus stimulating their angiogenic activity. This can explain the fact that in a number of cancers even a several-fold increase in Cu level in serum and malignant tissues was observed (in our study almost twofold), whereas others do not confirm this observation, especially according to tissue levels [19, 20]. Due to long-term monitoring of serum Cu concentration, it is feasible that the absence of changes in its concentration for a long period of time indicates a very good prognosis, whereas the increase in its concentration may show the recurrence of the carcinogenic process.

Zinc plays an important role in the carcinogenic process as it takes part in each stage of cellular cycle, regulation, and expression of genes as well as in DNA synthesis. Numerous authors have shown in *in vitro* studies that Zn exerts anti-proliferating and pro-apoptotic effects *inter alia* in prostate gland cells [21]. It is also suggested that the protective effect of Zn results from its competing with other essential elements,

mainly Cu and Fe. Based on the literature, it is also well known that the blood serum Zn concentration undergoes changes in the state of increased metabolic activity of the organism. A diminished Zn concentration is observed in cancers of different sites. The progression of neoplastic transformation is linked with decreasing Zn concentration in serum [22].

Numerous publications provide evidence that Se compounds are characterised by anticancer properties. Se may act in cancer prevention via numerous mechanisms and on all stages of cancer progression [11]. It has been found that Se induces apoptosis in malignant tissues [23]. An increased Se concentration in malignant tissues may point to the body's immune reaction and its attempt to induce apoptosis in aplastic cells through increasing the number of free radicals and thus enhancing oxidative stress. The mechanism by which free radicals are produced, and the results from this oxidative stress induced by selenium, is favourable to the body on the one hand, in view of anticancer effect, and unfavourable on the other as an Se toxic effect cannot be ruled out [24].

Unbalanced Mg homeostasis is often noticed in malignant cells, which accumulate this element, behaving like a magnesium trap. Moreover, high affinity of neoplastic cells for Mg was also observed in cell cultures with low Mg concentration [13].

The results of our study partly confirm observations of other researchers (Tables II and III), who found increased Cu levels in the large intestine malignant tissues. The same concerns Zn, levels of which are often only slightly departed from those found in normal tissue [6, 19, 25–28]. The observed inconsistency in element levels both in normal and malignant tissue are difficult to elucidate. It may be influenced by many factors, e.g. diet, different determination methods, and differences in the end outcomes (µg/g dry or wet tissue).

Table II. Zinc (Zn), copper (Cu), and selenium (Se) concentrations (mean \pm SD; $\mu\text{g/g}$ wet tissue) in malignant and normal tissues of the large intestine according to different authors

Authors, methods	Zn		Cu		Se	
	Malignant tissue	Normal tissue	Malignant tissue	Normal tissue	Malignant tissue	Normal tissue
Margalioth <i>et al.</i> , 1983 [6], AAS	18.6 \pm 6.34	18.3 \pm 3.8	1.90 \pm 0.6	1.53 \pm 0.35	–	–
Gregoriadis <i>et al.</i> , 1983 [25], PIXRF	14.3 \pm 2.7	15.2 \pm 3.1	1.70 \pm 0.64	1.34 \pm 0.33	–	–
Drake and Sky-Peck, 1989 [19], ultramicroEDXRF	98.2 \pm 18.7	64.1 \pm 20.5	8.9 \pm 3.4	14.1 \pm 4.4	1.26 \pm 0.28	1.53 \pm 0.33
Witkowski <i>et al.</i> , 1993 [26], AAS	–	–	5.18 \pm 2.63	5.76 \pm 1.54	–	–
Arriola <i>et al.</i> , 1999 [29], INNA	8.08 \pm 0.60	7.53 \pm 0.50	0.27 \pm 0.04	0.40 \pm 0.06	1.54 \pm 0.25	1.31 \pm 0.20
Kucharzewski <i>et al.</i> , 2003 [27], TRXRF	14.80 \pm 0.82	–	3.87 \pm 0.27	–	0.86 \pm 0.19	–
Daragó <i>et al.</i> , 2005 [28], AAS	17.44 \pm 5.60	15.54 \pm 6.73	1.09 \pm 0.63	0.64 \pm 0.42	0.12 \pm 0.07	0.05 \pm 0.03
Milde <i>et al.</i> , 2005 [30], AAS	69.20 \pm 21.03	–	6.08 \pm 3.78	–	1.17 \pm 0.73	–
Majewska <i>et al.</i> , 2007 [4], TXRF	14.8 \pm 9.63	–	3.55 \pm 2.36	–	0.816 \pm 0.557	–
Lavilla <i>et al.</i> , 2009 [31], ICP-OES, ICP-MS	89	79	8.5	6.2	1.6	0.9
Szewczyk <i>et al.</i> , 2013 [7], AAS	1.61 \pm 0.93*	3.22 \pm 1.91*	10.3 \pm 3.0*	6.3 \pm 1.8*	–	–

* $\mu\text{g/g}$ protein. AAS – atomic absorption spectrometry, PIXRF – photon-induced X-ray fluorescence, EDXRF – energy dispersive X-ray fluorescence, TRXRF – total reflection X-ray fluorescence, INNA – instrumental neutron activation analysis, ICP-OES – inductively coupled plasma optical emission spectrometry, ICP-MS – inductively coupled plasma mass spectrometry.

Table III. Calcium (Ca), iron (Fe), cadmium (Cd), and magnesium (Mg) concentrations (mean \pm SD; $\mu\text{g/g}$ wet tissue) in malignant and normal tissues of the large intestine according to different authors

Authors, methods	Fe		Mg		Ca		Cd	
	Malignant tissue	Normal tissue	Malignant tissue	Normal tissue	Malignant tissue	Normal tissue	Malignant tissue	Normal tissue
Drake and Sky-Peck, 1989 [19], ultramicroEDXRF	129 \pm 41.2	189 \pm 50.5	–	–	393 \pm 195	591 \pm 253	–	–
Arriola <i>et al.</i> , 1999 [29], INNA	120.24 \pm 4.09	131.45 \pm 4.21	20.96 \pm 0.82	19.76 \pm 0.73	9.07 \pm 0.83	11.18 \pm 0.93	–	–
Kucharzewski <i>et al.</i> , 2003 [32], TRXRF	46.1 \pm 4.27	–	–	–	–	–	–	–
Milde <i>et al.</i> , 2005 [30], AAS	–	–	753.59 \pm 310.54	–	–	–	–	–
Daragó <i>et al.</i> , 2005 [28], AAS	–	–	–	–	–	–	0.04 \pm 0.02	0.04 \pm 0.02
Majewska <i>et al.</i> , 2007 [4], TXRF	45.00 \pm 33.40	–	–	–	–	–	–	–
Szewczyk <i>et al.</i> , 2009 [20], colorimetrically	–	–	6.77 \pm 2.4	6.40 \pm 1.7	–	–	–	–
Lavilla <i>et al.</i> , 2009 [31], ICP-OES, ICP-MS	194	125	664	323	1462	459	0.14	0.23

Abbreviations as in Table II.

Conclusions

The results of our study seem to demonstrate disturbed homeostasis of some essential elements, mostly Mg and Cu. However, it is hard to confirm their involvement in the aetiology of colorectal cancer.

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Conflict of interest

The authors declare no conflict of interest.

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