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# **Ultrasound Elastography and MR Elastography for Assessing Liver Fibrosis: Part 1, Principles and Techniques**

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# **Abstract**

**OBJECTIVE—**The purpose of this article is to provide an overview of ultrasound and MR elastography, including a glossary of relevant terminology, a classification of elastography techniques, and a discussion of their respective strengths and limitations.

**CONCLUSION—**Elastography is an emerging technique for the noninvasive assessment of mechanical tissue properties. These techniques report metrics related to tissue stiffness, such as shear-wave speed, magnitude of the complex shear modulus, and the Young modulus.

# **Keywords**

elasticity; elastography; liver fibrosis; MR elastography (MRE); MRI; shear stiffness; shear wave; ultrasound; viscoelasticity; viscosity

> Imaging-based elastography is an emerging technology that uses imaging to noninvasively assess mechanical tissue properties. Elastography techniques have evolved significantly over the last 2 decades and have now been implemented on clinical ultrasound and MR systems [1–4]. These techniques provide the capability to examine by imaging what once could be examined only by direct palpation, which is likely to open new opportunities to noninvasively diagnose disease, guide management, and improve outcomes.

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In this first article of a two-part series (see part 2 elsewhere in this issue [5]), we provide a brief overview of elastography, including a glossary of commonly used terms (Table 1), a classification of techniques, a description of the basic principles, definitions of mechanical properties, examples of ultrasound and MR elastography, and a discussion of their respective strengths and limitations.

# **Key Learning Points**

First, imaging-based elastography techniques have been developed to measure stiffness and other mechanical properties noninvasively for both research and clinical indications. In general, these techniques cannot measure stiffness directly; instead, they assess stiffness indirectly by measuring the speed of shear waves propagating in the tissue of interest. The underlying concept is that shear-wave speed is related to tissue stiffness: shear waves travel faster in stiff tissues and slower in soft tissues.

Second, by measuring shear-wave speed, ultrasound elastography and MR elastography techniques estimate tissue stiffness. Depending on the technique, various stiffness-related parameters may be reported. The most commonly reported parameters and corresponding units are shear-wave speed in meters per second, magnitude of complex shear modulus in kilopascals (commonly described in the literature as "shear stiffness"), and the Young elastic modulus in kilopascals (commonly described in the medical literature as "elasticity"). The lack of uniformity in the reported parameters complicates comparisons across techniques. These elastography techniques may also measure parameters other than stiffness, including shear-wave attenuation and tissue viscosity, although these additional parameters are considered investigational and have not yet been reported clinically.

Third, shear waves may be generated by applying a mechanical vibration to the surface of the body or by focusing an acoustic radiation force (acoustic push pulse) inside the tissue of interest. Some ultrasound-based techniques use mechanical vibration for shear-wave generation, whereas others use acoustic radiation force. Commercial MRI-based techniques use only the former. The duration of the shear waves also is variable. Shear waves are generated transiently for ultrasound elastography and continuously for MR elastography.

Fourth, ultrasound elastography techniques track shear waves by using ultrasound-tracking beams. Some ultrasound-based techniques display parametric maps called "elastograms" that display the spatial distribution of the stiffness-related parameter of interest; others provide only numeric results.

Finally, MR elastography tracks shear waves by acquiring images with wave motionsensitized phase-contrast sequences. Tissue motion caused by the shear waves during the scan are encoded into the phase of the MR signal. This phase information is further processed to generate wave images depicting shear wave displacements within the liver. Subsequent processing of the wave images produces elastograms.

# **Classification**

As summarized in Figure 1, elastography techniques may be classified according to the source (static, quasistatic, or dynamic) and duration (transient or continuous) of tissue deformation and the modality used for tracking (ultrasound or MRI). Techniques also may be classified according to the device type (stand alone or adjunct to an imaging scanner), wave generation method (external vibrator or internally focused acoustic radiation force), inversion algorithm (1D, 2D, or 3D), reported parameters (shear-wave speed, magnitude of complex shear modulus, and the Young modulus), or output display (purely numeric, Mmode image, or parametric imaging map).

#### **Static or Quasistatic Elastography**

Static and quasistatic elastography assess stiffness by measuring the deformation (i.e., strain) in response to an applied force (i.e., stress) [6]. In static elastography, the stress is produced by manual compression of the tissue, whereas in quasistatic elastography, time-dependent stresses are produced by physiologic vibrations (e.g., the heart [7] or blood vessels [8]).

The assessment is qualitative, as opposed to quantitative, because the applied force cannot be known with certainty: manual compression in static elastography is operator dependent, and stresses caused by physiologic motion in quasistatic elastography are uncontrolled. Current static and quasistatic elastography techniques are ultrasound based. Static and quasistatic elastography will not be further discussed because of their limited applications for liver fibrosis.

#### **Dynamic Elastography**

Dynamic elastography techniques, also known as shear-wave imaging, assess stiffness and stiffness-related parameters by tracking shear waves propagating through media. Shear-wave speed is related to tissue stiffness; for instance, shear waves travel faster in stiff (inflamed, fibrotic, or cirrhotic) liver and slower in soft (normal or fatty) liver [9]. By measuring shearwave speed, the stiffness may be inferred. For most biologic tissues, the shear-wave speed and, hence, the inferred stiffness are frequency dependent: all other things being equal, shear-wave speed and inferred stiffness are greater if the shear waves are applied at higher frequency. Because the shear-wave frequencies used by different techniques differ, the stiffness-related values obtained with various techniques are not directly comparable.

#### **Basic Principles**

#### **Types of Waves**

Body waves, which travel inside organs, include compression and shear waves. With compression waves, tissue moves back and forth in a direction parallel to wave propagation. In contrast, shear waves produce tissue (particle) motion in a direction perpendicular to wave propagation [10]. Compression waves travel orders of magnitude faster than shear waves; in human soft tissues, compression waves propagate at around 1500 m/s, whereas shear waves propagate at around 1–10 m/s. Because compression waves propagate so rapidly through tissues, their speed and related properties cannot be reliably assessed by current imaging

techniques. For this reason, all current imaging elastography techniques are based on tracking shear waves, not compression waves.

#### **Wave Generation**

Shear waves may be generated by either applying external mechanical vibration to the surface of the body or focusing acoustic radiation force impulses inside the tissue of interest. Some ultrasound-based techniques use the former method of shear wave generation, whereas others use the latter. MRI-based techniques use only mechanical vibration.

With ultrasound- and MRI-based external mechanical vibration techniques, the vibrator typically oscillates perpendicular to the body surface at a precisely controlled frequency. Compression waves are applied to the body surface; some of the energy in these compression waves is converted to shear waves through a process called mode conversion [11]. With external vibration, the shear-wave frequency can be controlled precisely and the associated energy absorption by tissue is minimal, but wave delivery into the tissue of interest may be inefficient.

With ultrasound-based acoustic radiation force techniques, acoustic compression pulses are focused inside the liver; some of the acoustic energy is absorbed and released in the form of shear waves [12]. Thus, shear waves are generated locally within the liver, which improves the efficiency of wave delivery into the area of interest. Compared with external vibrationinduced waves, however, internally focused acoustic radiation forces are associated with higher power output and greater energy absorption, and the shear-wave frequency is more difficult to control.

#### **Key Concepts of Dynamic Elastographic Examination**

Regardless of the technique, dynamic elastography generally requires the following steps: baseline imaging, generation of shear waves, tracking of shear waves, and measurement of shear-wave speed or other mechanical parameter of interest.

#### **Mechanical Properties and Parameters**

Current clinically available techniques report the shear-wave speed, the magnitude of the complex shear modulus, or the Young modulus. The modulus parameters are often referred to as "stiffness" in the medical elastography literature, but strictly speaking do not have exactly the same meaning as stiffness, which is a qualitative property assessed through palpation. The term "stiffness" is permanently embedded in the medical elastography literature, however, and to maintain consistency with that literature, we also use the term to describe elastography measurements that approximate tissue response to palpation. Two such elastography parameters are elasticity and viscosity. Elasticity is the characteristic of a material that tends to return to its initial shape after a deformation. Viscosity is the characteristic of a material that resists rapid movement or deformation. Biologic tissues are considered to be viscoelastic, meaning that they have both viscous and elastic properties.

Investigational techniques may also calculate other parameters, but these additional measures are currently not reported in clinical settings. A full discussion of the numerous

parameters is beyond the scope of this review. A brief summary is provided in the data supplement and Table S1, which both can be viewed in the  $A$ *JR* electronic supplement to this article, available at www.ajronline.org.

# **Elastography Techniques**

Current elastography techniques are compared in Table 2 and illustrated in Figure 2.

# **Ultrasound-Based Dynamic Elastography**

All current ultrasound-based dynamic elastography techniques use transient shear-wave excitations at a frequency of 50–400 Hz depending on the technique [2, 13, 14]. Numerous ultrasound-based techniques have become commercially available. To describe these techniques, we have adopted the terminology proposed by the European Federation of Societies for Ultrasound in Medicine and Biology [15], as summarized in Table 3.

#### **One-Dimensional Transient Elastography**

The first ultrasound-based shear-wave measurement technique based on the concept of transient elastography was commercialized as FibroScan (Echosens). A single-element pistonlike ultrasound transducer mounted on a vibrating actuator generates a transient vibration ("punch") of short duration  $( $30 \text{ ms}$ )$  [16] at a frequency of 50 Hz. The mechanical impulse generates a shear wave that propagates symmetrically with respect to the axis of the single-element transducer [2, 16–20]. The displacements induced by the shear wave are tracked using ultrasonic waves emitted and received at very high frequency (6 kHz) by the single-element ultrasound transducer. A compression wave is also produced by the mechanical impulse, but because of its high speed, it moves beyond the location at which the shear waves are tracked and does not interfere with the shear-wave measurements.

Ultrasound transducers are available targeted for pediatric (S1 and S2), nonobese adult (M), and obese adult (XL) patients. The S probes (S1 and S2) use a pulse-tracking frequency of 5 MHz at window depths of 15–40 mm and 20–50 mm, respectively [21]. The M probe uses a pulse-tracking frequency of 3.5 MHz at a window depth of 20–60 mm, and the XL probe uses a lower pulse-tracking frequency of 2.5 MHz at a window depth of 35–75 mm [19, 22]. The transient elastography device does not provide an anatomic image but instead provides M-mode and A-mode graphs to locate the optimal measurement point. The shear-wave propagation graph (Figs. 2A and 2B) is displayed after each acquisition. The acquired data are used to measure the shear-wave speed in the examined area. The results are converted to the Young modulus and reported in kilopascals. The M probe was the first to be available commercially and there is more extensive experience with this probe than the others.

**Advantages—**The technique is relatively inexpensive, highly portable, and widely available, and it has been independently validated in numerous centers worldwide. The instrument is easy to learn and the measurement is highly standardized and rapid to perform. For these reasons, it can be used by clinicians at the point of service, which is a major advantage [23]. Young modulus thresholds using the M-probe have been proposed for different causes of chronic liver disease, which facilitates interpretation of results [24]. The

shear-wave frequency is controlled at 50 Hz, which ensures comparable measurements. The power output is low because the shear wave is generated mechanically, and there is little energy absorption by tissues.

**Limitations—**The technique may be limited in patients with obesity, narrow intercostal space, and ascites. The failure rate is lower with the XL probe than with the M probe (1.1% vs 16%) and reliability is higher (73% vs 50%) [25]. However, the performance of 1D transient elastography with the XL probe in obese subjects is controversial, with some studies reporting unreliability and failure rates as high as 23% [26, 27] and others as low as 6% [28]. This is problematic because obesity-associated fatty liver disease is emerging as the most common cause of chronic liver disease in Western nations. Other elastography techniques also can be challenging in obesity [29]. Further research is required to better understand the performance of all elastographic techniques in the obese population. Although the various probes produce similar results in phantoms, the XL probe provides lower stiffness estimates than the M probe in humans [25, 30]. A plausible explanation is that different ROIs (window depths) are being measured and that, in obese patients, the region examined by the M mode may include subcutaneous tissues, which causes the stiffness to be overestimated. Regardless, stiffness thresholds for staging liver fibrosis reported with the M probe may not be directly applicable to the XL probe [30]. Because anatomic images are not captured, the exact measurement location is not recorded, which may introduce sampling variability for monitoring changes in stiffness over time. Without the imaging capability, the device has narrow applications beyond liver examination.

#### **Point Shear-Wave Elastography**

Point shear-wave elastography relies on a high-frequency spheric compression wave (acoustic radiation force impulse or acoustic push pulse) focused on a spot [12], which is then absorbed as acoustic energy. The absorbed acoustic energy causes the tissue to expand [31], which creates shear waves perpendicular to the ultrasound beam axis [1]. The shearwave displacement created by the push pulse is recorded by a 2D ultrasound probe using a series of tracking pulses. From these data, the shear-wave speed can be derived. There are several implementations of point shear-wave elastography. These can be integrated onto existing ultrasound imaging scanners by adding the needed hardware (appropriate transducers and electronic components) and software (shear wave–tracking algorithms).

On the commercial Acuson S2000 and S3000 systems (Siemens Healthcare), acoustic radiation force impulses are implemented for qualitative (not further discussed) and quantitative applications. With the quantitative technique (Virtual Touch Quantification, Siemens Healthcare), a rectangular ROI ( $10 \times 6$  mm) is placed on a B-mode anatomic image (Figs. 2C and 2D). A transient shear wave is generated within the region. The shear wave is tracked and its speed measured. Results are reported as shear-wave speed in meters per second, at a range of 0.5–5 m/s in abdominal applications [32].

**Advantages—**This technique permits selection by the operator of an ROI in a representative area of the liver. The ROI is saved and, in principle, an ROI in a similar location could be selected in follow-up studies to permit more reliable monitoring. Point

shear-wave elastography techniques are more robust than 1D transient elastography because shear waves are produced locally inside the liver and can be transmitted even in patients with a large body habitus or ascites. Although an entire ultrasound scanner may be more expensive than a 1D transient elastography device, the incremental cost of adding the required software to an existing scanner is low.

**Limitations—**More expertise is needed than with 1D transient elastography. A radiologist or sonographer usually is required, and the technique is less suitable for point of service. Point shear-wave elastography is less validated than 1D transient elastography and currently is available on fewer scanners. Nevertheless, there are sufficient publications for a pooled meta-analysis on the diagnostic performance of point shear-wave elastography for staging of liver fibrosis [33]. The shear wave frequency is difficult to control precisely, which may introduce variability in measurements between different probes and manufacturers. Compared with 1D transient elastography, there is greater energy absorption by tissue.

#### **Shear-Wave Elastography**

The Supersonic Shear Imaging technique (Aixplorer) combines a cone-shaped quasi-planar wave front and an ultrafast imaging technique to track shear-wave displacements across an entire imaging plane [34]. An acoustic radiation force is focused at successively greater depths on an axial line to produce multiple sequential spheric wave fronts. These interfere constructively to create a Mach cone with greater displacement magnitudes than those produced by the individual wave fronts. By analogy with supersonic planes, the Mach cone is produced because the rate of sequential wave front production is greater than the speed of the resulting shear waves. In the commercial implementation of shear-wave elastography, several Mach cones are produced at different lateral positions of the image. An ultra-high frame rate (up to 15,000 images per second) is used to scan the entire imaging plane in one acquisition with high temporal resolution [13]. The combination of Mach cone generation and fast imaging allows real-time generation of elastograms. The results are reported as meters per second or converted to the Young modulus in kilopascals.

**Advantages—**This technique, which also relies on acoustic radiation force to generate shear waves, has similar strengths as point shear-wave elastography. Fast imaging permits generation of quantitative elastograms. Multiple ROIs then can be positioned on the elastograms, thereby reducing some of the sampling variability that can occur with 1D transient elastography and point shear-wave elastography [35].

**Limitations—**Shear-wave elastography has the same limitations as point shear-wave elastography with regard to the use of acoustic radiation force impulses. Compared with transient elastography and point shear-wave elastography, shear-wave elastography has restricted product availability, and, currently, there are fewer studies on its diagnostic performance for assessment of liver fibrosis [13, 36, 37].

#### **Newer Ultrasound-Based Systems**

GE Healthcare and Philips Healthcare just released commercial shear-wave elastography packages for some of their imaging systems. These use acoustic radiation force to generate transient shear waves. Other manufacturers are following suit.

# **MR Elastography**

MR elastography has been implemented on 1.5- and 3-T clinical scanners. The measured stiffness does not depend on field strength [38] but, as discussed earlier, does depend on mechanical excitation frequency [39]. Hence, mechanical properties acquired at different field strengths should be comparable as long as they are obtained with the same mechanical excitation frequency.

Because the spatial encoding of the shear-wave data is done over multiple wave cycles, MR elastography usually cannot measure transient shear waves. Instead, MR elastography techniques use continuous waves. MR elastography requires five components: a driver system to generate oscillatory mechanical waves continuously at a fixed frequency, a phasecontrast multiphase pulse sequence with motion-encoding gradients that are synchronized to the mechanical waves, processing of phase-sensitive MR images to depict wave amplitudes (shear-wave displacement images or, simply, wave images), further postprocessing (using an inversion algorithm) to generate elastograms (Fig. 3), and analysis of the elastograms.

#### **Driver System**

Several driver systems have been designed to produce shear waves for MR elastography [40], including pneumatic, electromagnetic, piezoelectric, and focused ultrasound systems. The current commercial system is an acoustic driver with two components, an active and a passive driver: the active driver in the equipment room generates compression waves, which are transmitted through a flexible tube to the passive driver positioned against the right anterior chest wall of the subject in the scan room and secured using an elastic band. The passive driver usually is made of plastic and contains a flexible membrane that transmits the compression waves into the patient. The active driver does not need to be MRI compatible. These compression waves are then converted inside the patient's body to shear waves by mode conversion.

In MR elastography, compression waves are generated continuously throughout the entire pulse sequence. The frequency used is typically around 60 Hz, because this low-frequency range results in waves with better propagation than higher-frequency waves while still yielding measurable shear wavelengths ranging from millimeters to centimeters [41].

#### **Pulse Sequence**

Various motion-sensitized MRI sequences, including gradient-recalled echo [4], spin-echo [42], echo-planar imaging [39, 43], and steady-state free precession [44], may be used to track the shear waves traversing the tissue of interest. A trigger pulse keeps the motionencoding gradient synchronized with the compression waves generated by the driver. In most implementations, the frequency of the motion-encoding gradients matches the

frequency of the waves. In the commercial 2D MR elastography sequence, the motionencoding gradient is only applied in the z direction (orthogonal to the patient axial plane), which suffices for 2D inversion of the wave component propagating along the x-y plane. In investigational 3D MR elastography sequences, motion-encoding gradients are successively applied in the  $x$ ,  $y$ , and  $z$  directions. A generic MR elastography pulse sequence diagram designed to detect the propagation of shear waves is illustrated in Figure 4.

#### **Image Acquisition**

Images are acquired with a modified phase-contrast technique that generates both magnitude and phase images. Because the motion-encoding gradients are synchronized with the frequency of the shear waves (phase locked), tiny particle displacements caused by the shear waves can be detected on the phase image. To acquire snapshots of the shear-wave propagation, phase images are acquired at different time offsets between the motion sensitization gradients and the shear waves (called phase offsets). Typically, four phase offsets are acquired (although 3D inversion techniques in development, as described later, use three phase offsets) to sample different spatial displacements of the wave cycle. Images at each phase offset are acquired through the liver at several slice locations to sample different portions of the liver. The total acquisition time is about 1 minute, typically divided into four separate approximately 15-second breath-holds (one for each slice location), acquired in end expiration if possible.

#### **Postprocessing**

The source phase images are postprocessed to produce wave displacement images. A mathematic filter called the curl filter separates the compression from the shear-wave components. In the clinical implementation adopted by MRI vendors, four additional phase offsets are interpolated to double the number of phase offset steps at each level to produce a fluid cine loop with eight wave images. Color maps are typically applied to these wave images, in which red and blue hues indicate opposite wave polarity and color saturation indicates wave amplitude.

These wave images are further processed to create elastograms, which depict the magnitude of the complex shear modulus. Waves propagate in three dimensions. The conversion from shear-wave displacement images to parametric maps requires solving an inverse problem (i.e., what distribution of stiffness, or related parameter, explains the observed wave patterns). With 2D inversion algorithms, only two-directional components within a slice of the 3D wave propagation are analyzed. With 3D inversion algorithms, a more complete analysis of the full wave motion is possible. Mathematic inversion algorithms currently require simplifying assumptions about tissue properties [45], including that it has a uniform density of 1 g/cm<sup>3</sup> and that it is purely viscoelastic with locally (microscopically) homogeneous, isotropic, and linear mechanical properties. The color elastograms represent the shear modulus with scales of 0–8 kPa and 0–20 kPa. In an advanced implementation, an elastogram confidence mask is automatically created that displays the portions of the elastogram in which the wave data are considered reliable.

#### **Image Analysis**

To obtain a single reportable value from the multislice parametric maps, the MR elastography–generated magnitude images, wave images, and elastogram confidence masks are viewed together. Geographic ROIs are drawn manually on portions of the liver parenchyma with reliable data, while avoiding edges of liver, large blood vessels, and regions with multipath wave interference. Because ROIs are drawn manually, some subjectivity (operator dependence) is unavoidable. Automated analysis techniques are in development; these are expected to reduce operator dependence.

#### **Parameters Reported**

In most publications using a now commercially available implementation of MR elastography, quantitative results have been reported as "shear stiffness" in kilopascals. This is taken to mean the magnitude of the complex modulus, also in kilopascals [46–49]. See example elastograms in Figure 5. Publications using investigational implementations have reported additional parameters, such as storage  $(G)$  and loss modulus  $(G')$  [39] or elasticity and viscosity, by assuming a Kelvin-Voigt model [50].

**Advantages—**Although numerous investigative teams have developed different MR elastography techniques, using different hardware and software, only one technique is being adopted for clinical implementation by the major MRI scanner manufacturers, which potentially will enable reproducibility of results. The hardware and software for this technique are produced by Resoundant and are made available by the scanner manufacturers as commercial elastography packages. MR elastography provides a high diagnostic accuracy for advanced fibrosis. This technique is robust, because it is feasible in larger patients or even those with ascites. Compared with ultrasound-based techniques, MR elastography assesses a larger proportion of the liver, which may potentially reduce sampling variability for longitudinal monitoring. The incremental cost of hardware and software is less than the cost of a new 1D transient elastography device but higher than the cost of purchasing shearwave elastography software for an existing ultrasound system. Power output and energy absorption by tissues are minimal, because the shear waves are generated externally.

**Limitations—**Because of the reliance on gradient-recalled echo sequences, quality in the current commercial implementation may be degraded in patients with iron deposition. Investigational sequences may permit MR elastography measurements even in patients with significant iron deposition. MR elastography requires postprocessing and offline analysis to obtain valid average stiffness measurements. The calculation involves some subjectivity inherent to the selection of the ROIs. MR elastography has limited availability outside of academic centers. Additional time is required for positioning the passive transducer, and the tension with which the transducer is secured is not yet standardized. The transducer causes discomfort in some patients, which may be alleviated by flexible traducers in development. Liver MR elastography is acquired with different breath-holds; if breath-holds are not consistent, this may potentially introduce errors that are not yet well characterized. Finally, shear waves tend to attenuate in normal (soft) livers; consequently, wave propagation may be poor in patients with normal livers, and large ROIs are not always obtainable.

### **Comparison of Techniques**

The relative strengths and limitations of ultrasound- and MRI-based techniques for staging of liver fibrosis are summarized in Table 4.

# **Conclusion**

Over the past 2 decades, elastography techniques have evolved significantly from investigational prototypes to clinical tools used for patient care. Depending on the indication (screening, diagnosis, or monitoring of liver fibrosis), different modalities may be preferred according to their respective strengths and limitations.

Ultrasound elastography techniques are relatively inexpensive, portable, and increasingly available while providing good diagnostic accuracy but may be unreliable in obese patients and those with narrow intercostal spaces. MR elastography offers excellent diagnostic accuracy that probably slightly exceeds that of ultrasound-based techniques, but quality may be degraded in patients with marked iron deposition, and availability remains comparatively limited. In a research setting, MR elastography may become a surrogate reference standard when liver biopsy is either not feasible or acceptable.

# **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

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Classification of elastographic techniques with focus on current commercial techniques.







# **Fig. 2.**

#### Current elastography techniques

**A–H**, Illustrations (**A**, **C**, **E**, and **G**; all by Tang A) show probe location, source of shearwave generation (blue arrows), direction of shear-wave propagation, and FOV (black outline). Also shown are companion images (**B**, **D**, **F**, and **H**) produced by each elastography technique: 1D transient elastography (**A** and **B**; FibroScan image [**B**] courtesy of Echosens), point shear wave elastography (**C** and **D**), shear-wave elastography (**E** and **F**), and MR elastography (**G** and **H**).



#### **Fig. 3.**

MR elastography experiment works on clinical MRI scanners and requires five components: driver system to generate mechanical waves, phase-contrast multiphase pulse sequence with motion-encoding gradients that are synchronized to mechanical waves, acquisition of phasesensitive MR images that contain raw data on wave motion and can also provide anatomic images, postprocessing to generate wave images and stiffness maps (also known as elastograms), and ROI analysis to produce single stiffness value. (Photographs by Tang A)



#### **Fig. 4.**

Generic MR elastography sequence. Trigger pulse synchronizes mechanical vibration and motion-encoding gradient. In this diagram, motion-encoding gradient is applied using sliceselect gradient. Motion-encoding gradients are applied successively in  $x$ ,  $y$ , and  $z$  directions (when generating data for 3D inversion) and switched in polarity. In this diagram, frequency of motion-encoding gradient matches frequency of cyclic mechanical vibrations. Varying trigger delay shifts phase of mechanical wave relative to pulse sequence (motion-encoding gradient), allowing generation of data that capture propagation of wave displacements and that can be played as cine loop. Typically, four phase offsets are applied. Interpolation is applied to double number of phase offsets, from four to eight wave images to generate fluid cine loop.



# **Fig. 5.**

MR elastography in different subjects correlated with biopsy-documented fibrosis stages. **A–J**, Axial wave cine loops (**A–E**) and corresponding axial elastogram images (**F–J**) are shown for subjects with fibrosis stages 0–4. Color elastogram represents magnitude of complex shear modulus with scale from 0 to 8 kPa.

# **TABLE 1**

# Glossary of Commonly Used Terms in Elastography







A ot all stiffness data are valid on the 2D elastogram. Valid data are considered reliable on the basis of confidence map and with good wave propagation. Not all stiffness data are valid on the 2D elastogram. Valid data are considered reliable on the basis of confidence map and with good wave propagation.

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# **TABLE 3**

Terminology Proposed by European Federation of Societies for Ultrasound in Medicine and Biology (EFSUMB) to Describe Ultrasound-Based Dynamic Elastography Techniques



# **TABLE 4**

Strengths and Limitations of Ultrasound Elastography and MR Elastography Techniques for Staging of Liver Fibrosis



