

miRNA expression profiling of Epstein–Barr virus-associated NKTL cell lines by Illumina deep sequencing

Julia Alles¹, Jennifer Menegatti¹, Natalie Motsch^{1,*}, Martin Hart¹, Norbert Eichner², Richard Reinhardt³, Gunter Meister² and Friedrich A. Grässer¹

1 Institute of Virology, Saarland University Medical School, Homburg/Saar, Germany

2 Biochemistry Center Regensburg (BZR), Laboratory for RNA Biology, University of Regensburg, Regensburg, Germany

3 Max Planck Genome Centre Cologne, Cologne, Germany

Keywords

CUL5; ebv-miR-BART16; Epstein-Barr virus; hsa-miR-148a; microRNA; S1PR1

Correspondence

F. A. Grässer, Institut für Virologie, Universitätsklinikum des Saarlandes, Kirrbergerstrasse, Haus 47, 66421 Homburg/Saar, Germany
Fax: +49 (0) 6841 1623980
Tel: +49 (0) 6841 1623983
E-mail: Friedrich.Graesser@uks.eu

*Present address

Boehringer Ingelheim, Birkendorfer Strasse 65, D-88397 Biberach, Germany

(Received 14 September 2015, revised 15 December 2015, accepted 17 December 2015)

doi:10.1002/2211-5463.12027

The aim of this work was to establish the microRNA profile of SNK6 and SNT16, two Epstein–Barr virus (EBV)-infected cell lines derived from nasal NK/T-cell lymphoma (NKTL). The oncogenic EBV is strongly associated with the pathogenesis of nasal and extranodal NK/T-cell lymphoma and expresses 44 mature microRNAs and two noncoding EBV-encoded RNAs (EBERs). miRNAs are 19–25nt noncoding RNAs that affect host and viral gene expression post-transcriptionally. Deregulated miRNA patterns are frequently linked to a variety of human cancers including lymphomas. miRNA profiling of the two NK/T cell lines vs. primary cells revealed 10 and 4 up-regulated and 10 and 12 down-regulated miRNAs in SNK6 and SNT16 cells respectively. The results were validated by qRT-PCR for selected miRNAs. Target gene analyses confirmed cullin 5 (CUL5) and sphingosin-1-phosphate receptor 1 (S1PR1) as targets for the down-regulated hsa-miR-148a and viral ebv-miR-BART16 respectively. As recently demonstrated for the regulation of IL1-alpha by miR-142-3p, coexpression of the EBERs selectively exerted corepression of S1PR1 by BART16 but not of CUL5 by miR-148a, indicating selective corepression by the EBERs.

The Epstein–Barr virus (EBV) infects approximately 95% of the adult human population worldwide and generally establishes a symptomless, latent infection [1]. Under certain conditions, however, it induces malignant tumours of B- or T-cell origin, including Burkitt's lymphoma (BL), Hodgkin's lymphoma (HL), as well as epithelial tumours such as nasopharyngeal carcinoma (NPC) and gastric carcinoma (GC) [2]. The tumorigenic potential is reflected in its ability to read-

ily transform B-cells into permanently growing cell lines (lymphoblastoid cell lines, LCLs), the *in vitro* equivalent of post-transplant lymphoproliferative disease (PTLD) that can arise in immunosuppressed patients [3]. EBV is also strongly associated with a type of non-Hodgkin's Lymphoma: the very rare NK/T-cell lymphoma (NKTL). Those tumours derive from NK- and/or T cells [4] and occur predominantly in Asia and Central-/South America. For nasal NKTL

Abbreviations

BL, Burkitt's lymphoma; DLBCL, diffuse large B-cell lymphoma; EBER, Epstein–Barr virus-encoded RNA; EBV, Epstein–Barr virus; GC, gastric carcinoma; GI, gastrointestinal; HL, Hodgkin lymphoma; LCL, lymphoblastoid cell line; miR-/miRNA, microRNA; mut, mutated; NKTL, NK/T-cell lymphoma; NPC, nasopharynx carcinoma; nt, nucleotide; PTLD, post-transplant lymphoproliferative disease; RISC, RNA-induced silencing complex; wt, wild-type.

the upper aerodigestive tract is often affected by a high grade of necrosis [5] as a consequence of perforin [6] or granzyme B expression [7]. Cases of extranodal NKTL of the gastro-intestinal tract, skin, testis, lung, eye or soft tissue have also been reported [8–12]).

In addition to protein-encoding genes, EBV was the first virus where microRNAs (miRNAs) were described [13] and these play important roles in transformation by EBV [14–16]. MiRNAs are conserved, small non-coding RNAs of approximately 22 nt length. They repress gene expression through binding to partially complementary sequences usually located in the 3' untranslated region (UTR) of target mRNAs [17]. To carry out their regulatory functions, miRNAs are incorporated into RNA-induced silencing complexes (RISC) where they directly interact with a member of the Argonaute (Ago) protein family [18]. Upon binding to their distinct target sites, miRNA-Argonaute complexes recruit a member of the glycine-tryptophan-rich motif (GW) protein family, which recruits the deadenylation machinery leading to poly(A) tail shortening and finally mRNA decay. At early stages of repression, however, the GW protein coordinates translational repression of the mRNA without considerably affecting mRNA stability [19–21]. Like cellular miRNAs, viral miRNAs can be secreted in endosome-derived exosomes, and these show an enrichment for 3' end uridinylated isoforms [22]. RISC complexes are associated with endosomal membranes [23,24] and knock down of GW182 reduces exosomal miRNA secretion [25], linking the mechanisms of miRNA activity and release. EBV also encodes two non-polyadenylated RNAs (Epstein-Barr virus Encoded RNA; EBER) [26]. These are transcribed by RNA polymerase III and are of 167 (EBER1) and 172 (EBER2) nucleotides (nt) length. The EBER transcripts are expressed at up to 10^6 copies per cell in all EBV-transformed tumours and cell lines [26,27]. We have recently shown the EBERs exert an additional, miRNA-specific down-regulation on miRNA targets: co-expression of the EBERs further down-regulated the protein expression of interleukin 1 α and RAC1 reporters by miR-142-3p and also the protein expression of interleukin 1 α , but not of ADCY9, another known target of miR-142-3p [28]. Further, the EBERs had no effect on the down-regulation of TOMM22, a known target for the EBV-encoded miRNA ebv-miR-BART16 [29].

Due to its high aggressiveness and accompanying necrosis, the amount of primary tumour tissues available is very limited. We therefore chose to compare normal CD56+/CD3+ cells from healthy donors with the NKTL lines SNK6 and SNT16. Furthermore, we validated new targets for viral and deregulated host

miRNAs. For the sphingosin-1-phosphate receptor 1 (S1PR1), we find that co-expression of the EBERs further represses down-regulation of both a 3'UTR reporter and the protein by ebv-miR-BART16, while the EBERs show no additional effect on the repression of a 3'UTR cullin-5 (CUL5) reporter by miR-148a.

Materials and methods

Cell culture

All cell lines were cultured as described previously [30–33]. The LCL cell lines AM 29 and AM 58 established with a complete and an EBER-deleted EBV genome [34] were a generous gift from Sankar Swaminathan, University of Utah, Salt Lake City, Utah, USA. SNK6 [35] and SNT16 [36] cells were a kind gift from Martin Rowe, University of Birmingham, Birmingham, UK.

Isolation of CD56+/CD3+ primary cells

PBMC were isolated from buffy coats (Blutspendezentrale Saar-Pfalz GmbH, Homburg, Germany) by Ficoll separation. CD56+/CD3+ cells were isolated from PBMC using the human CD56+/CD3+ NKT MACS cell isolation kit according to the manufacturer's instructions (Miltenyi Biotec, Bergisch Gladbach, Germany).

RNA sequencing

RNA-Seq libraries were prepared according to supplier recommendations (TruSeq DNA/RNA sample preparation v2 guide: <https://www.neb.com/protocols/1/01/01/library-preparation-e7300>). Libraries were quantified by fluorometry, immobilized and processed onto a flow cell with a cBot followed by sequencing as a 100 bp single read using TruSeq v3 chemistry on HiSeq2500 (all components by Illumina, San Diego, CA, USA). Data analysis was performed using in-house written scripts. In short, after initial quality check and adapter-trimming the remaining valid reads were mapped against human and EBV miRNAs listed in MIRBASE version 20 (June 2013; www.mirbase.org). The minimum length of annotated reads was set to 18nt and no mismatches were allowed. Annotated miRNA reads were reviewed for multiple insert annotations and then normalized to the number of valid reads in the corresponding library. Finally, the normalized values of SNK6 and SNT16 cell lines were compared to CD56+/CD3+ to calculate fold changes.

Reverse transcription and quantitative real-time PCR

DNase I-treated RNA was reverse transcribed using the miScript II RT Kit (Qiagen, Hilden, Germany). Semi-quant-

titative RT-PCR was conducted with the LightCycler 1.5 System (Roche Diagnostics, Mannheim, Germany). miRNAs were amplified using the LightCycler® FastStart DNAMaster^{PLUS} SYBR Green I Kit and reverse Primer 5'-GCG AGC ACA GAA TTA ATA CGA C-3' with miRNA-specific forward primers: qRT-miR-21-5p: 5'-TAG CTT ATC AGA CTG ATG TTG A-3', qRT-miR-148a-3p: 5'-TCA GTG CAC TAC AGA ACT TTG T-3', qRT-miR-150-5p: 5'-TCT CCC AAC CCT TGT ACC AGT G-3', qRT-miR-155-5p: 5'-TTA ATG CTA ATC GTG ATA GGG GTA A-3'. For relative quantification, the '5.8sRNA'-primer: 5'-CTA CGC CTG TCT GAG CGT CGC TT-3' was used [32].

Dual-luciferase assays

Dual-luciferase assays employing 3'UTR reporters in pMIR-RNLTK (a dual firefly and renilla luciferase vector) were carried out in HEK293T cells as described [33]. Typically, 10⁵ HEK293T cells were seeded in 24-well format and transfected using PolyFect (Qiagen) with 0.2 µg-well⁻¹ reporter vector and 0.8 µg effector plasmid. The ratio of firefly (reporter)/renilla (control) luciferase for each sample was determined (%RLU) and assays conducted in duplicate. Statistical significance was tested using Student's *t*-test.

Plasmids

The ebv-miR-BART16 and EBER expression plasmids had been described previously [28,29,37]. The miR-148a precursor was cloned into pSG5 using the following oligonucleotides: 5'-EcoRI-miR-148a: 5'-CGG AAT TCT GTT GGG CAT TTG CTC CTG C-3', 3'-BamHI-miR-148a: 5'-CGC GGA TCC CAG TCA GGA GTC CAC CAG GG-3'. The 3'UTRs of S1PR1 (NM_001400.4) and CUL5 (NM_003478.3) were cloned into the modified dual-luciferase reporter vector pMIR-RNLTK [38] using the following oligonucleotides: 5'-SpeI-S1PR1: 5'-GAC TAG TCG CAG CAA ATC GGA CAA TTC-3', 3'-SacI-S1PR1: 5'-CGA GCT CGC TGG ACT GAA AAG TTT CTG GCG-3', 5'-SpeI-CUL5: 5'-GGA CTA GTG TAA TGC TCA GCT GCA GAC-3', 3'-SacI-CUL5: 5'-CGA GCT CCT GCA ATC ATA ATG ACC TAC C-3'. Mutation of miRNA binding sites within the 3'UTRs was carried out by overlap extension PCR and the following oligonucleotides: 5' SspI-SalI-S1PR1mutBART16: 5'-GCA TAA GGA AGA ATA TTG TCG ACA AAT GAT ATT ATG CC-3', 3' SspI-SalI-S1PR1mutBART16: 5'-GGC ATA ATA TCA TTT GTC GAC AAT ATT CTT CCT TAT GC-3', 5' PmlI-CUL5mut148a: 5'-GAT CTT CAG ATA TTC ACA CGT GCA AAA AAT GCT GTT ATC-3', 3' PmlI-CUL5mut148a: 5'-GAT AAC AGC ATT TTT TGC ACG TGT GAA TAT CTG AAG ATC-3'.

Western blotting

HEK293T cells were transfected with 2 µg plasmid(s) 24 h after seeding 4 × 10⁵ cells in 6-well dishes. Cells were harvested 48 h later. Protein extracts (30 µg) were separated by 8.75% (CUL5) and 12.5% (S1PR1) PAGE and transferred to ProtranTM membranes (Roth, Karlsruhe, Germany). After blocking, membranes were incubated with the following primary antibodies: anti-human-EDG1 (S1PR1) clone H-60 (Santa Cruz), anti-human-CUL5 clone H-300 (Santa Cruz, Heidelberg, Germany), anti-human-β-actin clone 14C10 (Sigma-Aldrich, Munich, Germany) and anti-human-GAPDH (Cell Signalling, Leiden, the Netherlands). After incubation with secondary antibodies coupled to horseradish peroxidase (Sigma-Aldrich), the membranes were developed using ECL (Life Technologies, Braunschweig, Germany).

Northern blotting

Total RNA from transfected HEK293T cells was extracted using peqGOLD Trifast (Peqlab, Erlangen, Germany) according to the manufacturer's description without washing steps. 20 µg RNA was separated on a 12% urea-polyacrylamid gel and subsequently blotted onto a nylon membrane (Amersham, Freiburg, Germany) followed by chemical cross-linking [28,39]. After hybridization with UT³²P-labelled antisense probes (miRVana probe construction kit; Thermo Fisher (Ambion), Braunschweig, Germany) overnight and washing steps, the blots were exposed to phosphor screens and developed using a Typhoon Scanner (Amersham).

Results

miRNA profiling of primary CD56+/CD3+ cells vs. SNK6/SNT16 cell lines

We had previously compared the miRNA profiles of primary nasal NK/T-cell lymphoma (NKTL) with EBV-negative T-cell lymphoma and normal thymus [33]. Since CD56+/CD3+ primary cells are the most likely precursor cells for NKTL [40], we established in the present study the miRNA profile of CD56+/CD3+ cells from healthy donors in comparison with the EBV-positive NKTL lines SNK6 and SNT16. From a total of 5 393 355 valid sequences obtained from the CD56+/CD3+ cells, 985 467 reads (18.27%) could be assigned to human miRNAs, with only 54 reads obtained from viral sequences. Of the 5 674 444 reads from the SNT16 cell line, 1 201 368 reads (21.17%) represented human and 549 121 reads (9.68%) were from EBV miRNAs. For the SNK6 cell line, we obtained 2 327 119 valid sequences with 931 115 reads

from human miRNAs (40.01%) and 160 749 EBV miRNA reads (6.91%). Viral miRNAs thus represented 31.37% and 14.72% of the total miRNA count in the SNK6 and SNT16 cells respectively (Tables S1 and S2).

Of the EBV miRNAs, BART10, -22, -8 and -1 were present at the highest levels, while no BHRF1-derived miRNAs were detectable (Fig. 1). The EBV miRNAs were compared with our previous profiling (Fig. S1). The absence of BHRF1-derived miRNAs is consistent with prior observations that these miRNAs are only present in cells in type III latency (for review, see [16,41]). Similar results were reported for the EBV miRNAs of the two cell lines by Ling and colleagues who showed that the two lines do only marginally express the BHRF1 miRNAs [42]. However, the relative levels of miRNAs detected in that publication differed from the data presented here in that we find (by sequencing) the highest expression for miRNAs BART10, -22, -8 and 19 while they report (by microarray analysis) highest expression for miRNAs BART17, -7, -1 and 16. We assume that the different methods (sequencing vs. microarray) are responsible for these discrepancies. However, both methods come to the conclusion that the BHRF1 miRNAs are only poorly expressed if at all.

The relative up- or down-regulation of cellular miRNAs in the SNK6/SNT16 vs. the primary CD56+/CD3+ cells was determined (Fig. 2A,B respectively). For this comparison, only miRNAs were considered that represented at least 0.1% of all reads in one of the libraries and that showed a relative change of at least 1.5 relative to the CD56+/CD3+ normal cells. In both cell lines the highest relative up-regulation was observed for miR-155-5p and the strongest down-regulation was found for miR-150-5p. Following the above criteria, there were 10 up-regulated and 10 down-regulated miRNAs in the SNK6 line, while only four miRNAs (miR-155, -27b, -21, and -20a) were up-regulated and 12 others were down-regulated in the SNT16 line. The relative differences in the levels of miR-21, -148a, 150 and -155 were determined by qRT-PCR which confirmed the sequencing results for miR-148a, -150 and -155 but not for miR-21 (Fig. 3). miRNAs miR-222-3p and miR-423-5p were up-regulated in the SNK6 cells but down-regulated in SNT16. Conversely, miR-27-3p was down- in SNK6 but up-regulated in SNT16. The relative levels of miRNAs from the present analysis were compared with two earlier studies establishing the expression profile of miRNAs in normal thymus tissue vs. primary NKTL [33], and in primary murine NK

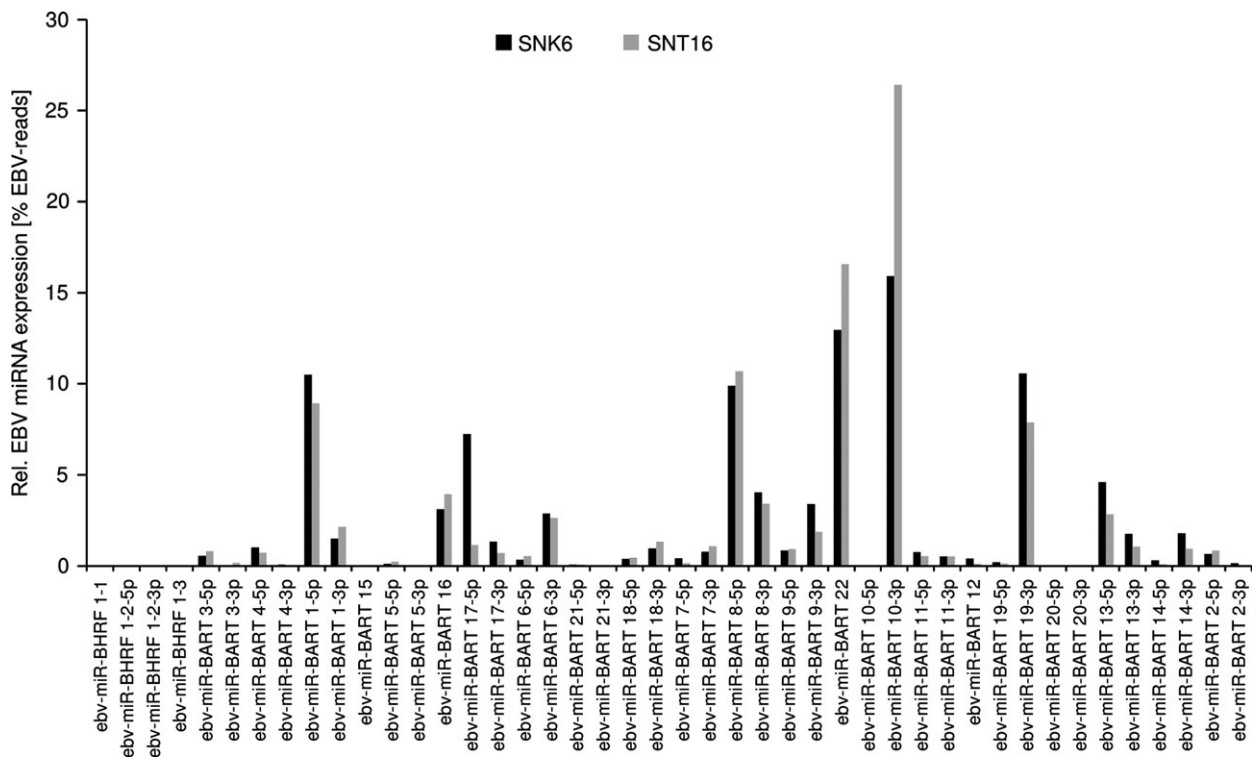


Fig. 1. Comparison of EBV-miRNA expression levels in NKTL-cell lines. ebv-miRNA expression in the SNK6 vs. SNT16 cell lines. The relative levels of the ebv-miRNAs obtained by sequencing of the two NK/T-lymphoma cell lines were compared with each other. Black: SNK6, grey: SNT16.

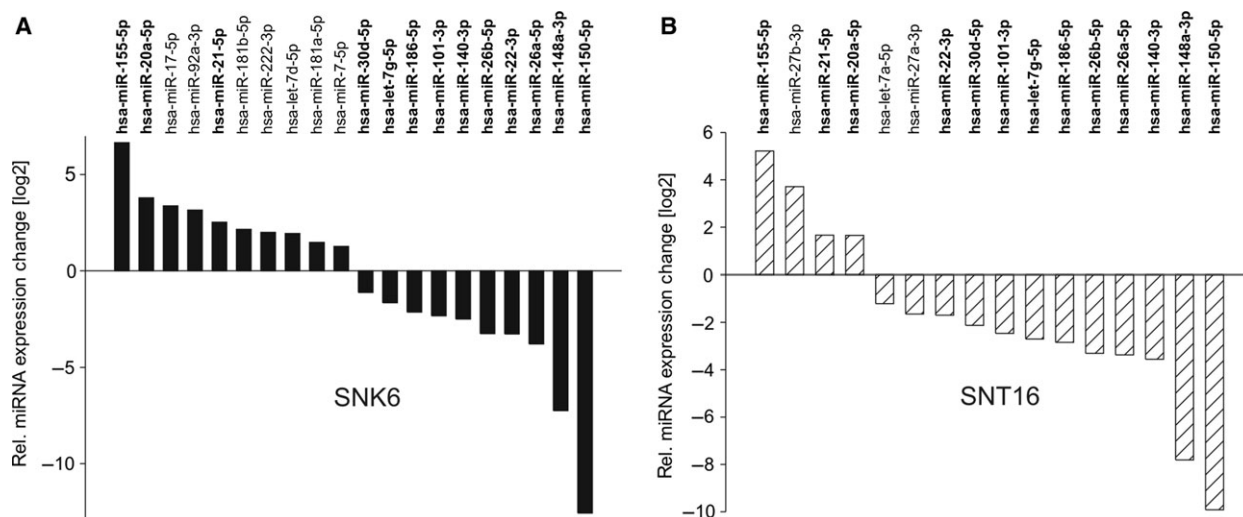


Fig. 2. MicroRNA profiling of primary CD56+/CD3+ NK/T cells and the NK/T cell lines SNK6 (A) and SNT16 (B). MiRNAs up- or down-regulated at least 1.5 fold in SNK6 or SNT16 cells compared to primary CD56+/CD3+ NK/T cells, with a representation of at least 0.1% in one of the cDNA libraries are depicted.

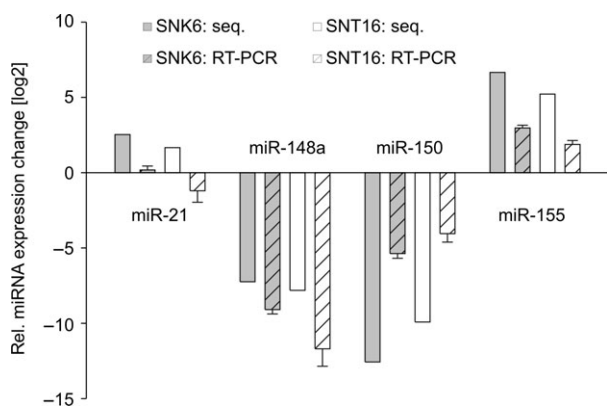


Fig. 3. Validation of the Illumina results. The relative levels (SNK6 or SNT16/primary cells) of the four miRNAs hsa-miR-21 and hsa-miR-155 (determined as up-regulated by sequencing) and hsa-miR-148a and hsa-miR-150 (determined as down-regulated by sequencing), analysed by qRT-PCR are shown. The graph represents the results of at least three independent experiments. Error bars show SD.

cells [43] (Fig. 4). Greater divergence is seen in the values obtained from thymus, as compared to the human primary NK/T and mouse primary NK cells, which likely reflects the fact that the thymus is composed of different cell types. The SNK6/SNT16 cell lines were also compared with the previous sequencing of primary NKTL [33]. The 10 miRNAs with the highest expression levels in each entity are shown in Table 1. In the two cell lines, eight of the 10 miRNAs were identical, while only three highly expressed miRNAs from the primary tumour samples were also among the top 10 found in the SNK6/SNT16 cell lines.

Identification of new targets for viral and deregulated host miRNAs

A bioinformatical analysis using the ‘TargetScan Human Custom’ algorithm (http://www.targetscan.org/vert_50/seedmatch.html) predicted the 3’UTR of the sphingosin-1-phosphate receptor 1 (S1PR1) to be a target of ebv-miR-BART16 (Fig. 5A). In this experiment, curiously, expression of BART16 showed reduced luciferase activity with the empty reporter vector. Nevertheless, inclusion of the S1PR1 3’UTR in the reporter down-modulated activity to a significantly greater degree ($P < 0.0001$) and this enhanced down modulation was abrogated by mutation of the potential binding site, demonstrating its specificity (Fig. 5B). We have recently shown that coexpression of the EBV-encoded EBER RNAs co-represses certain targets such as down modulation of IL-1 α by miR-142-3p but had no effect on the regulation of TOMM22 by ebv-miR-BART16 [28]. Expression of EBER1/2 also down modulated the empty vector and coexpression with BART16 increased this. Significantly, EBER1/2 cooperated with BART16 in the specific down modulation of the S1PR1 3’ UTR ($P = 0.002$). In conjunction with the previous report showing that BART16 had no effect on TOMM22 [28], this again illustrates that the effect of the EBERs are specific for a given miRNA vis-à-vis its mRNA target.

Using the miRecords database (http://c1.accurascience.com/miRecords/prediction_query.php), a combination of 11 miRNA target prediction programs,

an interaction for the strongly down-regulated hsa-miR-148a with the cullin 5 (CUL5) 3'UTR was predicted in 6 out of 11 algorithms (Fig. 6A). In reporter assays, miR-148a overexpression resulted in a significant decrease in the relative luciferase activity using the wt-CUL5 reporter vector ($P = 0.0005$), but this effect was abrogated when the CUL5 reporter with the mutation of the potential binding site was cotransfected. Coexpression of the EBERs had no additional effect (data not shown). Successful expression of ebv-miR-BART16 and hsa-miR-148a in 293T cells used in the luciferase assays was verified by northern blotting (Fig. S2).

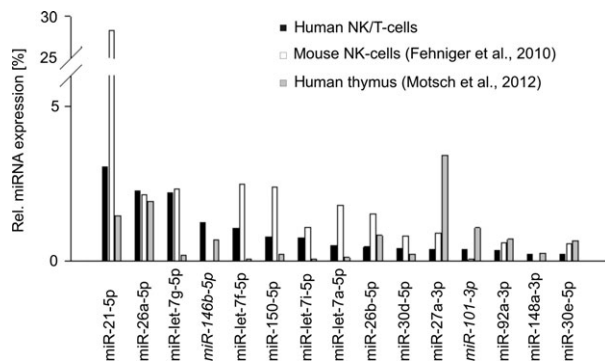


Fig. 4. Top 10 miRNAs in human NK/T cells vs. mouse NK cells vs. thymus. Comparison of the top 10 miRNAs expressed in primary human CD56+/CD3+ NK/T cells (black) to our previous sequencing analysis of thymus tissue (grey) [33] and the sequencing analysis of activated mouse NK cells (white) [43]. (comment on miRNAs in italic: miR-146b: sequencing data from thymus counted for miR-146a+b. miR-101: among mmu-miRNAs there are miR-101a/b/c, in the human miRNA database only miR-101-5p and -3p are listed. Therefore, the sequencing data for mmu-miR-101 do not show up in this graph).

As BART16 exerted a significant effect on the luciferase reporter in the absence of the S1PR1 3'UTR the activity of BART16, with and without the EBERs, on S1PR1 protein levels was assayed directly. Overexpression of BART16 ($P = 0.01$) or the EBERs ($P = 0.02$) in HEK293T cells resulted in a down modulation of the S1PR1 protein and the effect was additive with expression of both BART16 and the EBERs ($P = 0.03$) (Fig. 7A). We then compared the S1PR1 expression levels in the EBV-infected SNK6/SNT16 NK/T cells with primary CD56+/CD3+ NK/T cells isolated from healthy blood donors. The levels of endogenous S1PR1 were significantly lower in the EBV-transformed SNK6 and SNT16 cells compared to primary NK/T cells ($P = 0.03$ and 0.0006 respectively) suggesting that S1PR1 levels may be down modulated by EBV-transformation (Fig. 7B). Successful expression of ebv-miR-BART16 and EBER1/2 in 293T cells used in the western blots was verified by northern blotting. Furthermore, no up-regulation of BART16 by the EBERs was observed, eliminating the potential of a higher ebv-miR-BART16 level causing stronger S1PR1 protein down-regulation in this experiment; in fact, the BART16 levels were slightly lower in the EBER/BART16 co-transfection arguing against the effect of induced BART16 due to the presence of the EBERs (Fig. S3). In summary, our data establish S1PR1 as a novel target for BART16. To further corroborate these results, we compared the S1PR1 protein levels in two LCL lines that were established using the wt-EBV ('LCL AM 29') and the EBER- knock-out virus ('LCL AM 58') [34]. Here, the AM 58 cell line with the EBER deletion had higher S1PR1 levels than the EBER-positive AM 29 cells ($P = 0.008$) (Fig. 7D).

By virtue of the positive luciferase reporter assays for miR-148a and CUL5, we further investigated the CUL5 protein levels as a consequence of miR-148a

Table 1. Top 10 miRNAs expressed in NK/T-cell lines vs. primary NK/T-cell lymphoma.

SNK6		SNT16		NKTL [33]	
miRNA	Rel. expr. (%)	miRNA	Rel. expr. (%)	miRNA	Rel. expr. (%)
hsa-miR-21-5p	17.65	hsa-miR-21-5p	9.67	hsa-miR-16	9.09
hsa-miR-155-5p	7.75	hsa-miR-155-5p	2.85	hsa-miR-23b+23a	7.23
hsa-miR-92a-3p	3.08	hsa-miR-146b-5p	2.24	hsa-miR-21	5.87
hsa-let-7f-5p	1.69	hsa-let-7f-5p	1.00	hsa-miR-27a	5.01
hsa-miR-146b-5p	1.68	hsa-miR-27b-3p	0.59	hsa-miR-26a	4.52
hsa-let-7i-5p	1.05	hsa-let-7i-5p	0.55	hsa-miR-199a-3+199b-3p	4.08
hsa-let-7 g-5p	0.71	hsa-miR-92a-3p	0.43	hsa-miR-27b	3.31
hsa-miR-20a-5p	0.68	hsa-let-7 g-5p	0.34	hsa-miR-15a	2.96
hsa-let-7a-5p	0.52	hsa-let-7a-5p	0.22	hsa-miR-26b	2.9
hsa-miR-30e-5p	0.39	hsa-miR-26a-5p	0.22	hsa-miR-145	2.31

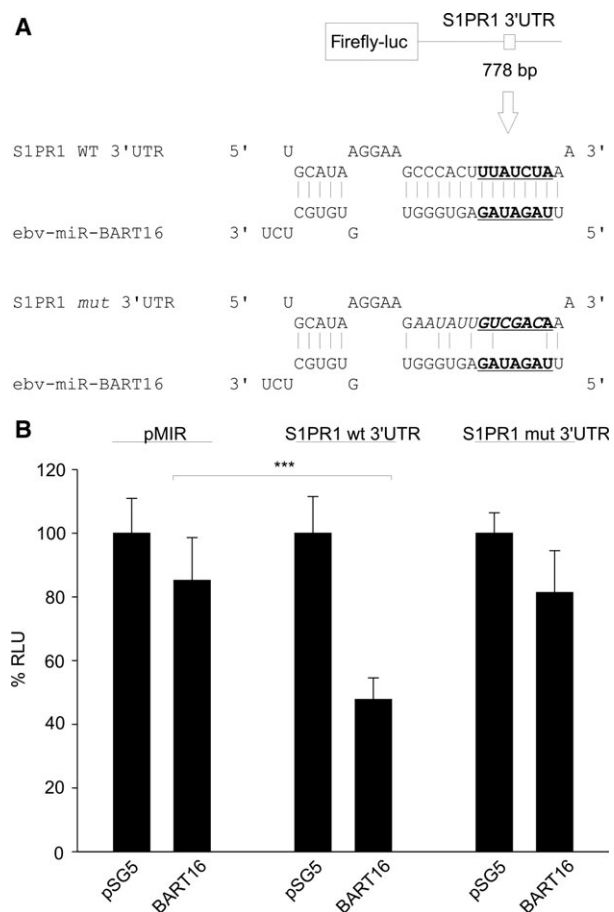


Fig. 5. S1PR1 3'UTR is targeted by ebv-miR-BART16. (A) Schematic representation of the pMIR-S1PR1 3'UTR reporter construct (upper panel) and prediction of the ebv-BART16 binding site. The mutated binding site for BART16 is shown in the lower panel. (B) Dual-luciferase reporter assays for ebv-miR-BART16 and S1PR1 3'UTR. Co-transfection of the S1PR1 WT-reporter construct with pSG5-BART16 results in a significant decrease in luciferase activity compared to the empty pMIR reporter vector. Using the S1PR1 reporter with mutated BART16 binding site, no effect was observed. The graph represents the results of at least three independent experiments carried out in duplicate. Error bars show SD. Stars denote: *** $P < 0.001$.

expression (Fig. 8A/C). Transfection of the miR-148a expression plasmid resulted in a significant reduction in the CUL5 protein level in HEK293T cells ($P = 0.0003$). We then further analysed the endogenous CUL5 protein levels in the EBV-positive SNK6/SNT16 cells compared to primary CD56+/CD3+ NK/T cells. In the NKTL cell lines, in which miR-148a expression was strongly down-regulated, the CUL5 protein level showed an induction of about threefold ($P = 0.0002$ and 0.016 respectively) (Fig. 8B/D). Together, these data establish S1PR1 and CUL5 to be

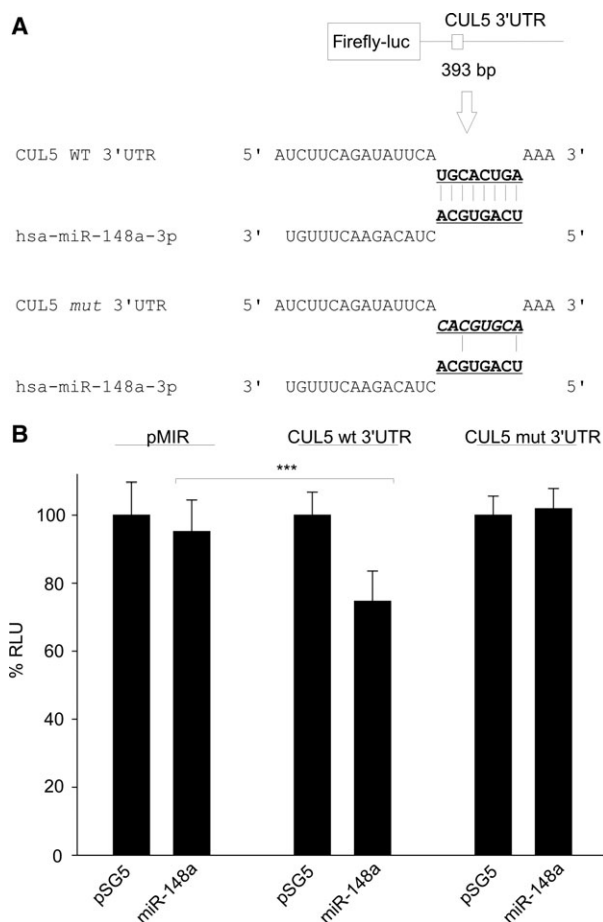


Fig. 6. CUL5 3'UTR is targeted by hsa-miR-148a. (A) Schematic representation of the pMIR-CUL5 3'UTR reporter construct (upper panel) and prediction of the miR-148a binding site. The mutated binding site for miR-148a is shown in the lower panel. (B) Dual-luciferase reporter assays for miR-148a and CUL5 3'UTR. Co-transfection of the CUL5 WT-reporter construct with pSG5-miR-148a results in a significant decrease in luciferase activity compared to the empty pMIR reporter vector. Using the CUL5 reporter with mutated miR-148a binding site, no effect was observed. The graph represents the results of at least three independent experiments carried out in duplicate. Error bars show SD. Stars denote: *** $P = 0.0005$.

the new targets for ebv-miR-BART16 and hsa-miR-148a respectively.

Discussion

Here, we present miRNA profiling of the cell lines SNK6 and SNT16 derived from peripheral NKTL. The up- or down-regulated cellular miRNAs were largely similar in the two cell lines compared to the CD56+/CD3+ primary NK/T cells. However, we observed differences when comparing these data with

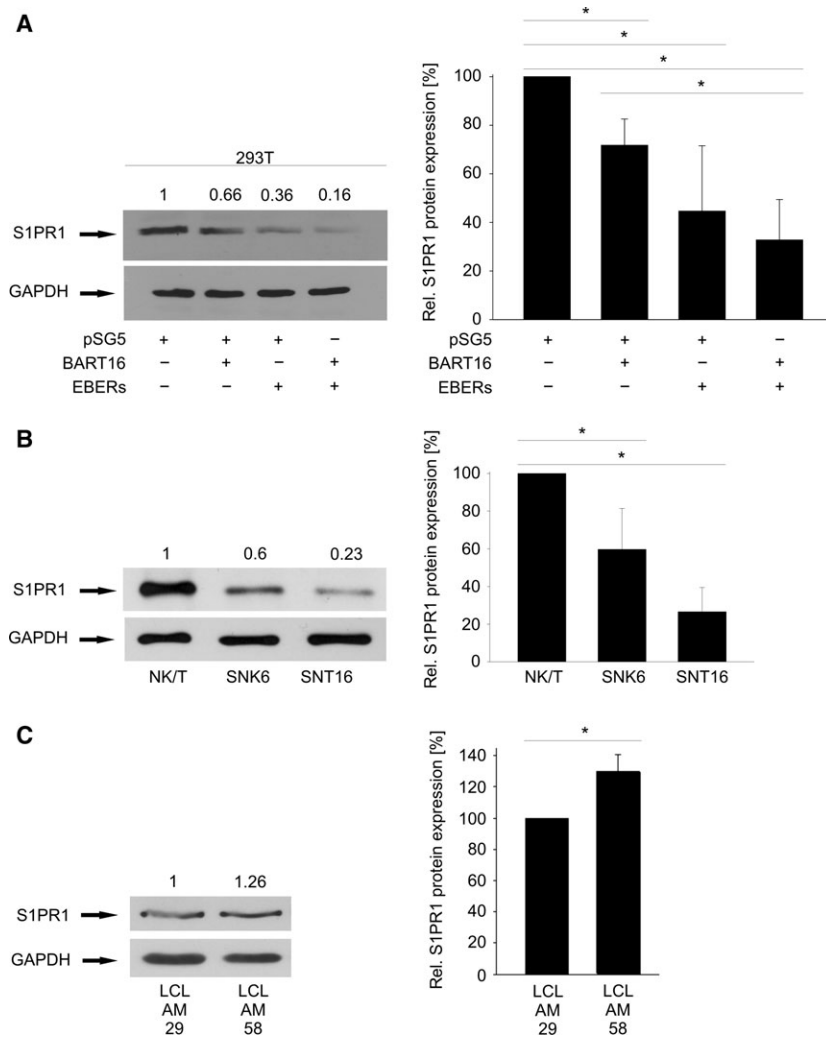


Fig. 7. Down-regulation of S1PR1 protein by ebv-miR-BART16 and EBER. Western Blot analyses of S1PR1 levels in cell lines. (A) The empty pSG5 vector, pSG5-BART16 and pSG5-EBERs were transfected in HEK293T cells. The expression of BART16 and EBERs alone down-regulated S1PR1 levels significantly ($*P = 0.01$, $*P = 0.02$ respectively). Co-expression of BART16 and the EBERs further down-regulated the S1PR1 protein level to a stronger extent than the BART16 plasmid alone ($*P = 0.03$) (B) S1PR1 is down-regulated in NK/T-cell lines (SNK6 and SNT16) compared to nontransformed CD56+/CD3+ primary cells ($*P = 0.03$, $*P = 0.0006$ respectively). (C) S1PR1 levels in lymphoblastoid cell lines established with an EBER-deleted EBV (LCL AM 58) or with the parental EBV (LCL AM 29) ($*P = 0.008$). The quantification of at least three independent experiments is shown on the right side of a representative blot. Error bars show SD.

results obtained in a prior miRNA profiling carried out with primary NKTL [33]. The discrepancies in the absolute values in the cell lines vs. tumour tissue may primarily be explained by the fact that the tumour contains stromal tissue. MiR-205, which was found to be down-regulated in the prior analysis, was strongly up-regulated in the SNK6/SNT16 cell lines compared to the CD56+/CD3+ primary cells. Comparison of the CD56+/CD3+ primary cell data with the profiling of primary murine NK cells [43] showed good concordance. The sequencing of the SNK6/SNT16 cell lines (which are in latency type II, LMP1-positive but no expression of EBNA2 and EBNA3s) revealed the virtual absence of BHRF1-derived miRNAs confirming previous observations (reviewed in [16,41]). A recent report described the down-regulation of miR-15a in primary nasal NKTL [44]. We confirmed the down-regulation of miR-15a, -223, -150 and -342 in our analysis. It was also reported that ebv-miR-BART20-

5p down-regulated T-bet (TBX21) in nasal NKTL [45]. Here, as previously [33], we found only relatively low levels of ebv-miR-BART20-5p. However, in the SNK6 and SNT16 cell lines analysed, the EBV-encoded miRNAs overall represented 31.4% and 14.7% of all miRNAs reads consistent with the notion that EBV-encoded miRNAs play a pivotal role in tumorigenic transformation by EBV [46]. The difference in expression of, i.e. hsa-miR-223-3p, -423-5p or -27-3p, might be due to the fact that SNK6, which is CD3-negative [35] has features of NK cells while SNT16 is presumably of T-cell origin [36].

S1PR1 (also called EDG1), a new target regulated by ebv-miR-BART16, was originally found in differentiating human endothelial cells [47] and represents one of five sphingosin-1-phosphate (SIP)-activated G-coupled receptors (reviewed in [48,49]). SIP inhibits cell migration during endothelial to mesenchymal transformation in cardiac development [50]. In addi-

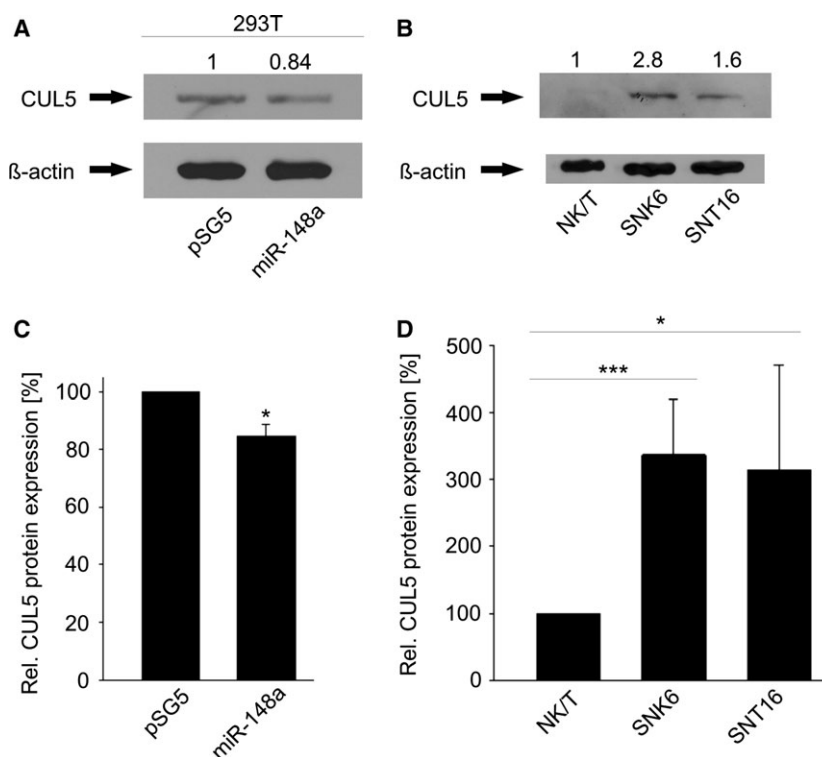


Fig. 8. Down-regulation of CUL5 protein by hsa-miR-148a. Western Blot analyses of CUL5 levels. (A) pSG5-miR-148a and the empty control (pSG5) were transfected in HEK293T cells. miR-148a overexpression down-regulated the CUL5 level significantly ($*P = 0.0003$). (B) CUL5 is up-regulated in NK/T-cell lines (SNK6 and SNT16) compared to nontransformed CD56+/CD3+-primary cells ($***P = 0.0002$, $*P = 0.016$ respectively). (C, D) Western Blot quantification of at least three independent experiments. Error bars show SD.

tion to other cell types, S1PR1 is expressed in cells of lymphoid origin and has now been designated as CD363 [51,52]. Depending on the cell type and co-expression of the other S1P-receptors, S1PR1 may either inhibit or activate cell growth and/or cell motility (reviewed in [53]). Down-regulation of S1PR1 expression was found in breast carcinoma [54] and low levels of S1PR1 were correlated with higher proliferation of glioma cells and are a predictor for poor survival of glioblastoma patients [55]. Down-regulation of S1PR1 was observed in lung carcinoma as compared to normal lung tissue, and the metastatic potential of a lung cancer cell line was correlated with lower S1PR1 levels [56]. In contrast, chronic activation of S1PR1 by S1P, the product of sphingosine kinase 1 (SphK1) in colitis-associated cancer increases the risk for colon cancer [57]. Strong up-regulation of S1PR1 was observed in 40/40 mantle cell lymphoma, while the same report found S1PR1 expression only in 2/37 DLBCLs [58]. Inhibition of S1PR1 retarded DLBCL cell growth via reduction of STAT3 *in vitro* [59]. In Hodgkin's lymphoma, 7/56 cases showed staining for S1PR1. Finally, it was demonstrated that S1PR1 and the chemokine receptor CX3CR1 are down-regulated in nonsmall cell lung tumour-derived NK cells, while CXCR5 and CXCR6 were induced [60]. Our results support the notion that down-regulation of S1PR1 may contribute to tumour formation in NKTL. How-

ever, this issue has to be resolved by determination of S1PR1 levels in primary NKTL tissue which was beyond the scope of this work.

Another novel target in this study validated for the strongly repressed host miR-148a was CUL5 (also known as VACM1). As a member of 7 Cul-RING-E3 ubiquitin ligases (CRLs), CRLs, in combination with different substrate receptor proteins, facilitate the conjugation of ubiquitin residues to target proteins which then were designated to degradation by the proteasome. As their role in cancer and other diseases is dependent on their substrate receptors and target proteins, cullins are nontypical tumour suppressors or oncoproteins. Many publications suggested CUL5 to function as a tumour suppressor, for example, through inhibition of Src-dependent tumorigenesis [61]. As a target for miR-19a/b, CUL5 induces proliferation and invasion in cervical cancer cells [62]. Inhibition by miR-7 triggers G1/S transition in hepatocellular carcinoma cells [63]. Mediating Jak3 degradation together with CUL1 as a consequence of NOTCH activation, CUL5 affects B- and T-cell development [64]. In addition, several viral substrate receptors for CUL5 have been reported: the HIV Vif protein causes CUL5-dependent degradation of the antiviral APOBEC3G [65,66]. The Adenovirus 5 E4orf6 and E1B55K collaborate to target the tumour suppressor p53 for degradation [67]. A similar mechanism is known for the

KSHV LANA, which promotes proteasomal degradation of p53 and VHL in KSHV-infected tumour cells [68]. In EBV-infected cells, CUL5 catalyses p53 poly-ubiquitination due to its binding by the viral substrate receptor BZLF1 [69]. BZLF1 triggers EBV to switch from latent to lytic cycle replication. EBV typically is in a latent type of infection but EBV needs to reactivate and replicate to some extent. Our data suggest that an up-regulation of CUL5 not necessarily maintains tumour growth rather than EBV transmission and spread within NKTL tissues.

A recent study from the Cullen group revealed that the percentage of a miRNA expressed in cells does not necessarily indicate its impact on post-transcriptional target gene repression as the amount of RISC-associated miRNAs better reflects their potential of down-regulating targets [70]. According to that result, the loading of the RISC complex with miRNAs will be an important additional experiment to determine functional relevance. Furthermore, a recent publication showed that the majority of miRNAs in primary, resting cells are localized in non-functional, low-molecular weight complexes while the biologically active, mRNA-associated miRNAs are preferentially found in high-molecular weight complexes [71]. Our results do not include the analysis of RISC association, however, the consequence of our newly confirmed targets for one viral and one strongly down-regulated host miRNA can clearly be seen by the determination of the endogenous level of those proteins in the nontransformed primary CD56+/CD3+ NK/T cells compared to our two EBV-positive lymphoma cell lines.

Conclusions

The high-throughput sequencing of the two NKTL cell lines revealed that the miRNA profiling yields a largely different set of deregulated miRNAs as compared to the profiles obtained when primary NKTL tissues are examined. The observation that the viral miRNAs represent a large fraction of the total miRNA pool implies a relevant contribution to the induction and/or maintenance of the transformed phenotype. In line with previous observations of EBV-positive tumour tissues, the cell lines do not express the EBV-encoded BHRF1 miRNAs. We show that the highly deregulated miRNA hsa-miR-148a down-regulates cullin 5 (CUL5) and that the viral miRNA ebv-miR-BART16 down-regulates the sphingosin-1-phosphate receptor 1 (S1PR1). The potential importance of the down-regulation of S1PR1 is highlighted by the fact that the EBV-encoded EBER RNAs exert a

co-repression on the S1PR1 protein levels. We again show that the co-repression by the EBERs is specific for a given miRNA as we observe no co-repression by the EBERs on CUL5 and hsa-miR-148a or on TOMM22, a previously established target for ebv-miR-BART16.

Acknowledgements

We thank Ruth Nord for expert technical assistance, Claire Shannon-Lowe, Martin Rowe and Sankar Swaminathan for the kind gift of cell lines. G.M. and F.G. were supported by a grant from Deutsche Krebs-hilfe (Project 107166) and the German Ministry for Education and Research (BMBF, RNomics no. 01GS0805). F.G. was also supported by an intramural grant (HOMFOR).

Author contributions

G.M. and F.A.G. planned experiments; J.A., N.M., M.H., N.E., R.R., performed experiments and/or analysed data; J.A. and F.A.G. wrote the paper.

References

- 1 Kieff E and Rickinson AE (2007) Epstein-Barr Virus and its replication. In *Fields Virology*. Vol. 2, 5th edn (Knipe D, Griffin DE, Lamb RA, Strauss SE, Howley PM, Martin MA and Roizman B, eds), pp. 2603–2654. Lippincott Williams & Wilkins, Philadelphia, PA.
- 2 Delecluse HJ, Feederle R, O'Sullivan B and Taniere P (2007) Epstein Barr virus-associated tumours: an update for the attention of the working pathologist. *J Clin Pathol* **60**, 1358–1364.
- 3 Rickinson AB and Kieff E (2007) Epstein-Barr virus. In *Fields Virology*. Vol. 2 (Knipe D, Griffin DE, Lamb RA, Strauss SE, Howley PM, Martin MA and Roizman B, eds), pp. 2655–2700. Lippincott Williams & Wilkins, Philadelphia, PA.
- 4 Suzumiya J (2008) NK-cell lymphoma: pathogenesis and clinical features. *Rinsho Ketsueki* **49**, 545–552.
- 5 Jaffe ES, Chan JK, Su IJ, Frizzera G, Mori S, Feller AC and Ho FC (1996) Report of the Workshop on Nasal and Related Extranodal Angiocentric T/Natural Killer Cell Lymphomas. Definitions, differential diagnosis, and epidemiology. *Am J Surg Pathol* **20**, 103–111.
- 6 Ohshima K, Suzumiya J, Shimazaki K, Kato A, Tanaka T, Kanda M and Kikuchi M (1997) Nasal T/NK cell lymphomas commonly express perforin and Fas ligand: important mediators of tissue damage. *Histopathology* **31**, 444–450.

- 7 Yamashita Y, Yatabe Y, Tsuzuki T, Nakayama A, Hasegawa Y, Kojima H, Nagasawa T and Mori N (1998) Perforin and granzyme expression in cytotoxic T-cell lymphomas. *Mod Pathol* **11**, 313–323.
- 8 Jaffe ES, Krenacs L, Kumar S, Kingma DW and Raffeld M (1999) Extranodal peripheral T-cell and NK-cell neoplasms. *Am J Clin Pathol* **111** (1 Suppl 1), S46–S55.
- 9 Stokkermans-Dubois J, Jouary T, Vergier B, Delaunay MM and Taieb A (2006) A case of primary cutaneous nasal type NK/T-cell lymphoma and review of the literature. *Dermatology* **213**, 345–349.
- 10 Liang DN, Yang ZR, Wang WY, Zhao S, Yang QP, Tang Y, Bi CF and Liu WP (2012) Extranodal nasal type natural killer/T-cell lymphoma of testis: report of seven cases with review of literature. *Leuk Lymphoma* **53**, 1117–1123.
- 11 Ely A, Evans J, Sundstrom JM, Malysz J, Specht CS and Wilkinson M (2012) Orbital involvement in extranodal natural killer T cell lymphoma: an atypical case presentation and review of the literature. *Orbit* **31**, 267–269.
- 12 Kunami N, Takamatsu Y, Fujita M, Katsuya H, Sasaki H, Wakamatsu S, Ishitsuka K, Nabeshima K and Tamura K (2010) Extranodal NK/T-cell lymphoma arising from soft tissue of the left forearm. *Rinsho Ketsueki* **51**, 422–426.
- 13 Pfeffer S, Zavolan M, Grasser FA, Chien M, Russo JJ, Ju J, John B, Enright AJ, Marks D, Sander C *et al.* (2004) Identification of virus-encoded microRNAs. *Science* **304**, 734–736.
- 14 Feederle R, Linnstaedt SD, Bannert H, Lips H, Bencun M, Cullen BR and Delecluse HJ (2011) A viral microRNA cluster strongly potentiates the transforming properties of a human herpesvirus. *PLoS Pathog* **7**, e1001294.
- 15 Seto E, Moosmann A, Gromminger S, Walz N, Grundhoff A and Hammerschmidt W (2010) Micro RNAs of Epstein-Barr virus promote cell cycle progression and prevent apoptosis of primary human B cells. *PLoS Pathog* **6**, e1001063.
- 16 Barth S, Meister G and Grasser FA (2011) EBV-encoded miRNAs. *Biochim Biophys Acta* **1809**, 631–640.
- 17 Skalsky RL and Cullen BR (2010) Viruses, microRNAs, and host interactions. *Annu Rev Microbiol* **64**, 123–141.
- 18 Meister G (2013) Argonaute proteins: functional insights and emerging roles. *Nat Rev Genet* **14**, 447–459.
- 19 Braun JE, Huntzinger E and Izaurralde E (2013) The role of GW182 proteins in miRNA-mediated gene silencing. *Adv Exp Med Biol* **768**, 147–163.
- 20 Pfaff J and Meister G (2013) Argonaute and GW182 proteins: an effective alliance in gene silencing. *Biochem Soc Trans* **41**, 855–860.
- 21 Peters L and Meister G (2007) Argonaute proteins: mediators of RNA silencing. *Mol Cell* **26**, 611–623.
- 22 Koppers-Lalic D, Hackenberg M, Bijnsdorp IV, van Eijndhoven MA, Sadek P, Sie D, Zini N, Middeldorp JM, Ylstra B, de Menezes RX *et al.* (2014) Nontemplated nucleotide additions distinguish the small RNA composition in cells from exosomes. *Cell Rep* **8**, 1649–1658.
- 23 Lee YS, Pressman S, Andress AP, Kim K, White JL, Cassidy JJ, Li X, Lubell K, Lim do H, Cho IS *et al.* (2009) Silencing by small RNAs is linked to endosomal trafficking. *Nat Cell Biol* **11**, 1150–1156.
- 24 Gibbins DJ, Ciaudo C, Erhardt M and Voinnet O (2009) Multivesicular bodies associate with components of miRNA effector complexes and modulate miRNA activity. *Nat Cell Biol* **11**, 1143–1149.
- 25 Yao B, La LB, Chen YC, Chang LJ and Chan EK (2012) Defining a new role of GW182 in maintaining miRNA stability. *EMBO Rep* **13**, 1102–1108.
- 26 Lerner MR, Andrews NC, Miller G and Steitz JA (1981) Two small RNAs encoded by Epstein-Barr virus and complexed with protein are precipitated by antibodies from patients with systemic lupus erythematosus. *Proc Natl Acad Sci USA* **78**, 805–809.
- 27 Howe JG and Shu MD (1988) Isolation and characterization of the genes for two small RNAs of herpesvirus papio and their comparison with Epstein-Barr virus-encoded EBER RNAs. *J Virol* **62**, 2790–2798.
- 28 Alles J, Hasler D, Kazmi SMA, Tesson M, Hamilton A, Schlegel L, Marx S, Eichner N, Reinhardt R, Meister G *et al.* (2015) Epstein-Barr virus EBER transcripts affect miRNA-mediated regulation of specific targets and are processed to small RNA species. *Noncoding RNA* **1**, 170–191.
- 29 Dolken L, Malterer G, Erhard F, Kothe S, Friedel CC, Suffert G, Marciniowski L, Motsch N, Barth S, Beitzinger M *et al.* (2010) Systematic analysis of viral and cellular microRNA targets in cells latently infected with human gamma-herpesviruses by RISC immunoprecipitation assay. *Cell Host Microbe* **7**, 324–334.
- 30 Shannon-Lowe C, Adland E, Bell AI, Delecluse HJ, Rickinson AB and Rowe M (2009) Features distinguishing Epstein-Barr virus infections of epithelial cells and B cells: viral genome expression, genome maintenance, and genome amplification. *J Virol* **83**, 7749–7760.
- 31 Frohn A, Eberl HC, Stohr J, Glasmacher E, Rudel S, Heissmeyer V, Mann M and Meister G (2012) Dicer-dependent and -independent Argonaute2 protein interaction networks in mammalian cells. *Mol Cell Proteomics* **11**, 1442–1456.
- 32 Livak KJ and Schmittgen TD (2001) Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* **25**, 402–408.

- 33 Motsch N, Alles J, Imig J, Zhu J, Barth S, Reineke T, Tinguely M, Cogliatti S, Dueck A, Meister G *et al.* (2012) MicroRNA profiling of Epstein-Barr virus-associated NK/T-Cell lymphomas by deep sequencing. *PLoS One* **7**, e42193.
- 34 Swaminathan S, Tomkinson B and Kieff E (1991) Recombinant Epstein-Barr virus with small RNA (EBER) genes deleted transforms lymphocytes and replicates in vitro. *Proc Natl Acad Sci USA* **88**, 1546–1550.
- 35 Nagata H, Konno A, Kimura N, Zhang Y, Kimura M, Demachi A, Sekine T, Yamamoto K and Shimizu N (2001) Characterization of novel natural killer (NK)-cell and gammadelta T-cell lines established from primary lesions of nasal T/NK-cell lymphomas associated with the Epstein-Barr virus. *Blood* **97**, 708–713.
- 36 Zhang Y, Nagata H, Ikeuchi T, Mukai H, Oyoshi MK, Demachi A, Morio T, Wakiguchi H, Kimura N, Shimizu N *et al.* (2003) Common cytological and cytogenetic features of Epstein-Barr virus (EBV)-positive natural killer (NK) cells and cell lines derived from patients with nasal T/NK-cell lymphomas, chronic active EBV infection and hydroa vacciniforme-like eruptions. *Br J Haematol* **121**, 805–814.
- 37 Repellin CE, Tsimbouri PM, Philbey AW and Wilson JB (2010) Lymphoid hyperplasia and lymphoma in transgenic mice expressing the small non-coding RNA, EBER1 of Epstein-Barr virus. *PLoS One* **5**, e9092.
- 38 Beitzinger M, Peters L, Zhu JY, Kremmer E and Meister G (2007) Identification of human microRNA targets from isolated argonaute protein complexes. *RNA Biol* **4**, 76–84.
- 39 Pall GS, Codony-Servat C, Byrne J, Ritchie L and Hamilton A (2007) Carbodiimide-mediated cross-linking of RNA to nylon membranes improves the detection of siRNA, miRNA and piRNA by northern blot. *Nucleic Acids Res* **35**, e60.
- 40 Kwong YL (2011) The diagnosis and management of extranodal NK/T-cell lymphoma, nasal-type and aggressive NK-cell leukemia. *J Clin Exp Hematol* **51**, 21–28.
- 41 Klinke O, Feederle R and Delecluse HJ (2014) Genetics of Epstein-Barr virus microRNAs. *Semin Cancer Biol* **26**, 52–59.
- 42 Ramakrishnan R, Donahue H, Garcia D, Tan J, Shimizu N, Rice AP and Ling PD (2011) Epstein-Barr virus BART9 miRNA modulates LMP1 levels and affects growth rate of nasal NK T cell lymphomas. *PLoS One* **6**, e27271.
- 43 Fehniger TA, Wylie T, Germino E, Leong JW, Magrini VJ, Koul S, Keppel CR, Schneider SE, Koboldt DC, Sullivan RP *et al.* (2010) Next-generation sequencing identifies the natural killer cell microRNA transcriptome. *Genome Res* **20**, 1590–1604.
- 44 Komabayashi Y, Kishibe K, Nagato T, Ueda S, Takahara M and Harabuchi Y (2014) Downregulation of miR-15a due to LMP1 promotes cell proliferation and predicts poor prognosis in nasal NK/T-cell lymphoma. *Am J Hematol* **89**, 25–33.
- 45 Lin TC, Liu TY, Hsu SM and Lin CW (2013) Epstein-Barr virus-encoded miR-BART20-5p inhibits T-bet translation with secondary suppression of p53 in invasive nasal NK/T-cell lymphoma. *Am J Pathol* **182**, 1865–1875.
- 46 Kang D, Skalsky RL and Cullen BR (2015) EBV BART microRNAs target multiple pro-apoptotic cellular genes to promote epithelial cell survival. *PLoS Pathog* **11**, e1004979.
- 47 Hla T and Maciag T (1990) An abundant transcript induced in differentiating human endothelial cells encodes a polypeptide with structural similarities to G-protein-coupled receptors. *J Biol Chem* **265**, 9308–9313.
- 48 Rosen H and Goetzl EJ (2005) Sphingosine 1-phosphate and its receptors: an autocrine and paracrine network. *Nat Rev Immunol* **5**, 560–570.
- 49 Takuwa Y, Takuwa N and Sugimoto N (2002) The Edg family G protein-coupled receptors for lysophospholipids: their signaling properties and biological activities. *J Biochem* **131**, 767–771.
- 50 Wendler CC and Rivkees SA (2006) Sphingosine-1-phosphate inhibits cell migration and endothelial to mesenchymal cell transformation during cardiac development. *Dev Biol* **291**, 264–277.
- 51 Matesanz-Isabel J, Sintes J, Llinas L, de Salort J, Lazaro A and Engel P (2011) New B-cell CD molecules. *Immunol Lett* **134**, 104–112.
- 52 Spiegel S and Milstien S (2011) The outs and the ins of sphingosine-1-phosphate in immunity. *Nat Rev Immunol* **11**, 403–415.
- 53 Takuwa N, Du W, Kaneko E, Okamoto Y, Yoshioka K and Takuwa Y (2011) Tumor-suppressive sphingosine-1-phosphate receptor-2 counteracting tumor-promoting sphingosine-1-phosphate receptor-1 and sphingosine kinase 1 – Jekyll Hidden behind Hyde. *Am J Cancer Res* **1**, 460–481.
- 54 Wittmann BM, Wang N and Montano MM (2003) Identification of a novel inhibitor of breast cell growth that is down-regulated by estrogens and decreased in breast tumors. *Cancer Res* **63**, 5151–5158.
- 55 Yoshida Y, Nakada M, Harada T, Tanaka S, Furuta T, Hayashi Y, Kita D, Uchiyama N, Hayashi Y and Hamada J (2010) The expression level of sphingosine-1-phosphate receptor type 1 is related to MIB-1 labeling index and predicts survival of glioblastoma patients. *J Neurooncol* **98**, 41–47.
- 56 Nakano T, Shimizu K, Kawashima O, Kamiyoshihara M, Kakegawa S, Sugano M, Ibe T, Nagashima T, Kaira K, Sunaga N *et al.* (2012) Establishment of a human lung cancer cell line with high metastatic potential to multiple organs: gene expression associated

- with metastatic potential in human lung cancer. *Oncol Rep* **28**, 1727–1735.
- 57 Liang J, Nagahashi M, Kim EY, Harikumar KB, Yamada A, Huang WC, Hait NC, Allegood JC, Price MM, Avni D *et al.* (2013) Sphingosine-1-phosphate links persistent STAT3 activation, chronic intestinal inflammation, and development of colitis-associated cancer. *Cancer Cell* **23**, 107–120.
- 58 Nishimura H, Akiyama T, Monobe Y, Matsubara K, Igarashi Y, Abe M, Sugihara T and Sadahira Y (2010) Expression of sphingosine-1-phosphate receptor 1 in mantle cell lymphoma. *Mod Pathol* **23**, 439–449.
- 59 Liu Y, Deng J, Wang L, Lee H, Armstrong B, Scuto A, Kowolik C, Weiss LM, Forman S and Yu H (2012) S1PR1 is an effective target to block STAT3 signaling in activated B cell-like diffuse large B-cell lymphoma. *Blood* **120**, 1458–1465.
- 60 Gillard-Bocquet M, Caer C, Cagnard N, Crozet L, Perez M, Fridman WH, Sautes-Fridman C and Cremer I (2013) Lung tumor microenvironment induces specific gene expression signature in intratumoral NK cells. *Front Immunol* **4**, 19.
- 61 Laszlo GS and Cooper JA (2009) Restriction of Src activity by Cullin-5. *Curr Biol* **19**, 157–162.
- 62 Xu XM, Wang XB, Chen MM, Liu T, Li YX, Jia WH, Liu M, Li X and Tang H (2012) MicroRNA-19a and -19b regulate cervical carcinoma cell proliferation and invasion by targeting CUL5. *Cancer Lett* **322**, 148–158.
- 63 Ma C, Qi Y, Shao L, Liu M, Li X and Tang H (2013) Downregulation of miR-7 upregulates Cullin 5 (CUL5) to facilitate G1/S transition in human hepatocellular carcinoma cells. *IUBMB Life* **65**, 1026–1034.
- 64 Wu W and Sun XH (2011) A mechanism underlying NOTCH-induced and ubiquitin-mediated JAK3 degradation. *J Biol Chem* **286**, 41153–41162.
- 65 Mehle A, Goncalves J, Santa-Marta M, McPike M and Gabuzda D (2004) Phosphorylation of a novel SOCS-box regulates assembly of the HIV-1 Vif-Cul5 complex that promotes APOBEC3G degradation. *Genes Dev* **18**, 2861–2866.
- 66 Mehle A, Strack B, Ancuta P, Zhang C, McPike M and Gabuzda D (2004) Vif overcomes the innate antiviral activity of APOBEC3G by promoting its degradation in the ubiquitin-proteasome pathway. *J Biol Chem* **279**, 7792–7798.
- 67 Querido E, Blanchette P, Yan Q, Kamura T, Morrison M, Boivin D, Kaelin WG, Conaway RC, Conaway JW and Branton PE (2001) Degradation of p53 by adenovirus E4orf6 and E1B55K proteins occurs via a novel mechanism involving a Cullin-containing complex. *Genes Dev* **15**, 3104–3117.
- 68 Cai X, Lu S, Zhang Z, Gonzalez CM, Damania B and Cullen BR (2005) Kaposi's sarcoma-associated herpesvirus expresses an array of viral microRNAs in latently infected cells. *Proc Natl Acad Sci USA* **102**, 5570–5575.
- 69 Sato Y, Kamura T, Shirata N, Murata T, Kudoh A, Iwahori S, Nakayama S, Isomura H, Nishiyama Y and Tsurumi T (2009) Degradation of phosphorylated p53 by viral protein-ECS E3 ligase complex. *PLoS Pathog* **5**, e1000530.
- 70 Flores O, Kennedy EM, Skalsky RL and Cullen BR (2014) Differential RISC association of endogenous human microRNAs predicts their inhibitory potential. *Nucleic Acids Res* **42**, 4629–4639.
- 71 La Rocca G, Olejniczak SH, Gonzalez AJ, Briskin D, Vidigal JA, Spraggon L, DeMatteo RG, Radler MR, Lindsten T, Ventura A *et al.* (2015) In vivo, Argonaute-bound microRNAs exist predominantly in a reservoir of low molecular weight complexes not associated with mRNA. *Proc Natl Acad Sci USA* **112**, 767–772.

Supporting information

Additional supporting information may be found in the online version of this article at the publisher's web site:
Fig. S1. Comparison of EBV miRNA expression levels.

Fig. S2. Ectopic expression of ebv-miR-BART16 and hsa-miR-148a in 293T cells.

Fig. S3. Transfection control of EBER1, EBER2 and ebv-miR-BART16 in 293T cells.

Table S1. Sequencing data (EBV miRNAs) reads obtained and relative ebv- miRNA expression in SNK6/SNT16 cDNA libraries.

Table S2. Sequencing data (host miRNAs) Reads obtained and relative hsa-miRNA expression in NK/T and SNK6/SNT16 cDNA libraries.