

# **HHS Public Access**

Author manuscript *Cell Oncol (Dordr).* Author manuscript; available in PMC 2016 April 05.

Published in final edited form as:

Cell Oncol (Dordr). 2016 April; 39(2): 97-106. doi:10.1007/s13402-016-0268-6.

# PSA and beyond: alternative prostate cancer biomarkers

# Sharanjot Saini, Ph.D.<sup>1,2</sup>

Sharanjot Saini: Sharanjot.Saini@ucsf.edu <sup>1</sup>Department of Urology, Urology Research (112J), Veterans Affairs Medical Center, 4150 Clement Street, San Francisco, CA 94121, USA

<sup>2</sup>University of California San Francisco, San Francisco, CA, USA

# Abstract

**Background**—The use of biomarkers for prostate cancer screening, diagnosis and prognosis has the potential to improve the clinical management of the patients. Owing to inherent limitations of the biomarker prostate-specific antigen (PSA), intensive efforts are currently directed towards a search for alternative prostate cancer biomarkers, particularly those that can predict disease aggressiveness and drive better treatment decisions.

**Methods**—A literature search of Medline articles focused on recent and emerging advances in prostate cancer biomarkers was performed. The most promising biomarkers that have the potential to meet the unmet clinical needs in prostate cancer patient management and/or that are clinically implemented were selected.

**Conclusions**—With the advent of advanced genomic and proteomic technologies, we have in recent years seen an enormous spurt in prostate cancer biomarker research with several promising alternative biomarkers being discovered that show an improved sensitivity and specificity over PSA. The new generation of biomarkers can be tested via serum, urine, or tissue-based assays that have either received regulatory approval by the US Food and Drug Administration or are available as Clinical Laboratory Improvement Amendments-based laboratory developed tests. Additional emerging novel biomarkers for prostate cancer, including circulating tumor cells, microRNAs and exosomes, are still in their infancy. Together, these biomarkers provide actionable guidance for prostate cancer risk assessment, and are expected to lead to an era of personalized medicine.

# Keywords

Prostate cancer; Biomarkers; Prognostic; Predictive; Diagnostic

# 1 Prostate cancer: epidemiology, clinical burden and dilemmas

Prostate cancer is the most commonly diagnosed male malignancy. It is estimated that prostate cancer alone will account for about one quarter of the cancer diagnoses among men in the USA in 2015, with an estimated 220,800 new cases [1]. It is the second leading cause

Correspondence to: Sharanjot Saini, Sharanjot.Saini@ucsf.edu.

Compliance with ethical standards The manuscript complies with ethical standards.

Conflict of interest The author declares that there are no conflicts of interest.

of cancer-related mortality among men with an estimate of 27,540 deaths in 2015 [1]. Although novel therapies with proven survival benefits have been developed in the last few years [2, 3], the increases in survival rates are marginal. Prostate cancer is a remarkably heterogeneous disease. Prostate tumors can be indolent or very aggressive, often metastasizing to bone and other organs, thereby causing significant morbidity and mortality [4]. A major clinical challenge in prostate cancer is the inability of current diagnostic tests, including prostate-specific antigen (PSA) screening and histopathological grading, to distinguish between aggressive and indolent tumors [5]. PSA is present in normal prostatic secretions and its levels are often elevated in prostate cancer patients [6, 7]. Serum PSA levels have been utilized as a prostate cancer biomarker for over 20 years and PSA screening has revolutionized the clinical management of the disease [8]. However, PSA has inherent limitations, including lack of specificity, leading to over-diagnosis and over-treatment of prostate cancer. In view of this, intensive efforts are currently focused on searching for alternative prostate cancer biomarkers, particularly those that can predict the aggressiveness of the disease and drive better treatment decisions.

Also, histopathological grading performed by Gleason scoring has been used as an important prognostic indicator of prostate cancer [9–11]. Gleason-based grading classifies tumors from 1 to 5 according to their relative degree of differentiation (from most to least differentiated, respectively). In prostate cancer, multifocal lesions usually exist [12] and Gleason grades can, therefore, be heterogeneous within or between different tumor foci. In view of this, a combined Gleason score that sums up the two most prevalent patterns within the prostate gland is used [9–11]. Although a Gleason score is a powerful prognostic indicator, it cannot accurately predict the aggressiveness of the disease since it has been found that tumors with similar histological patterns may have different clinical outcomes [11, 13]. Hence, there is a critical need for the development of alternative prostate cancer biomarkers with a better diagnostic and prognostic potential.

### 2 Prostate cancer biomarkers: a bird's eye view

The National Cancer Institute (NCI) defines 'biomarker' as a biological molecule found in blood, other body fluids, or tissues that can be objectively measured and evaluated as a sign of a normal/abnormal biological process and a pathogenic condition/disease. A biomarker may be used for screening purposes, for disease diagnosis and prognosis, for the evaluation of disease disposition and for the prediction/monitoring of treatment responses to various therapeutic interventions [14–16]. Since several years, the use of biomarkers for prostate cancer screening, diagnosis and prognosis has revolutionized its clinical management [8]. The first prostate cancer biomarker, prostatic acid phosphatase (PAP), was described in the 1930s and has since then been used as a clinical marker for prostate cancer progression, since serum PAP levels were found to be elevated in metastatic cases [15, 17]. This biomarker was replaced by prostate-specific antigen (PSA) in the 1980s [15, 18]. PSA is a kallikrein-related serine protease produced by the epithelial cells of the prostate gland [5] that is present in normal prostatic secretions and is often elevated in prostate cancer [6, 7]. Since the 1980s, PSA screening has significantly improved prostate cancer disease management, including its survival rates. However, owing to inherent limitations of PSA as a biomarker (see below), the field of prostate cancer biomarkers is evolving rapidly. Also,

Page 3

recent years an enormous increase in our understanding of prostate cancer biology that, in turn, has contributed to the development of novel biomarkers for prostate cancer diagnosis and prognosis. In this review, I will discuss the current state of our understanding of prostate cancer biomarkers - primarily diagnostic and predictive - thereby highlighting recent advances that have the potential to improve the clinical management of the patients. These biomarkers are blood-/urine-/tissue-based and have been either approved by the US Food and Drug Administration (FDA) or are available as Clinical Laboratory Improvement Amendments (CLIA) based laboratory developed tests (LDTs) [8, 19]. In addition, I will discuss emerging novel biomarkers for prostate cancer (including circulating tumor cells, microRNAs and exosomes) that are still in their infancy. Imaging-based tests such as computerized tomography and transrectal ultrasound have been developed as well, but these are not discussed here and are reviewed elsewhere [20, 21]. Considering the recent spurt in prostate cancer biomarker research, I foresee that promising alternative biomarkers will become available to clinicians to support a better diagnosis, prognosis and clinical management of prostate cancer patients (summarized in Fig. 1).

# 3 Prostate cancer biomarkers in clinical use: US FDA approved

The development of disease biomarkers as clinical diagnostic tools with US Food and Drug Administration (FDA) approval is often a long and regulated process that includes significant validations [19, 22]. Hence, only a few prostate cancer biomarkers have as yet been approved by the US FDA and these are discussed below.

#### 3.1 Prostate-specific antigen

Currently, PSA is the most prevalently used prostate cancer biomarker by clinicians. In 1994, the US FDA approved PSA testing for prostate cancer screening in conjunction with a digital rectal exam (DRE). A PSA level of 4 ng/ml was defined as the upper normal limit. Prostate cancer cases with elevated PSA levels typically undergo biopsy for assessment of prostate cancer. However, more recent studies have shown that  $\sim 20$  % of men with PSA levels<4 ng/ml have prostate cancer and that many men with higher levels do not have prostate cancer [8, 23-25]. Although organ specific, PSA is not cancer specific and circulating PSA levels often rise in the presence of conditions that disrupt prostate basal membrane epithelial cells such as prostatitis, benign prostatic hyperplasia (BPH), prostate biopsies and surgeries [5, 8]. Owing to these shortcomings, PSA screening has led to overdiagnosis and over-treatment of low-risk prostate cancer [8, 26]. In view of this, recent changes in recommendations include later and less frequent PSA screening [27, 28]. This notion highlights the need for the development of novel improved prostate cancer biomarkers. However, despite its lack of specificity, PSA is still the most clinically accepted biomarker used for prostate cancer, especially for monitoring progression and disease recurrence after curative therapy [15, 29]. In fact, PSA screening has contributed to a significant decline (45–70 %) in age-adjusted prostate cancer mortality since the early 1990s [8, 30].

PSA exists in blood either in its free form (fPSA) or in a complex with serum protease inhibitors. fPSA accounts for 5-35 % of total PSA (tPSA) and can occur in three molecular

forms: pro-PSA, benign PSA (bPSA) and intact PSA (iPSA) [5]. The ratio of fPSA over tPSA (percentage free PSA) has been found to be lower in prostate cancer [31] and has been shown to improve the specificity of cancer detection in men with tPSA values of 4–10 ng/ml and a normal DRE [32]. However, due to the instability of fPSA compared to complexed PSA, the percentage of fPSA exhibits a wide analytical variability and is, therefore, not used as primary screening tool [8, 33].

Pro-PSA is the inactive proenzyme form of PSA which contains a seven amino acid leader peptide that is normally cleaved by human kallikrein (hK) enzymes, i.e., hK-2 and hK-4, to PSA. However, roughly one-third of fPSA exists as pro-PSA that is more likely to be associated with prostate cancer [5, 19]. Several truncated forms/isoforms of pro-PSA exist in serum with varying lengths of the pro-leader peptide. One isoform, [-2] proPSA, consisting of PSA with a serinearginine pro-leader peptide, has emerged as a prominent biomarker for prostate cancer [8, 34, 35]. This isoform is the most prevalent biomarker that is correlated with cancer rather than BPH, and is used for the early detection and for determining the aggressiveness of the disease [34–39]. It outperforms both PSA and the percentage fPSA in terms of diagnostic ability, specificity and positive or negative predictive value [8, 40, 41]. It was shown to be the best biomarker for prostate cancer detection, particularly in cases with 2–10 ng/ml PSA [42].

#### 3.2 Prostate health index

In 2012 the US FDA approved another test - the prostate health index (PHI) - that includes three biomarkers and is calculated for each patient as ([-2] proPSA/fPSA)  $\times$  PSA<sup>1/2</sup>. This test was developed by Beckman Coulter in partnership with the NCI Early Detection Research Network [19] and intends to distinguish cancerous and benign prostatic conditions in men aged 50 years with a normal DRE and PSA levels of 4–10 ng/ml [42]. This test determines the need of biopsy in cases with total PSA levels between 4 and 10 ng/ml, thereby reducing unnecessary biopsies (Fig. 1). Several studies suggest that the use of PHI along with [-2] proPSA significantly improves prostate cancer detection in cases with a Gleason score 7 [19, 38, 39] and improves the accuracy of established prostate cancer predictors at biopsy compared to PSA and fPSA [40, 41].

#### 3.3 Prostate cancer antigen 3

Prostate cancer antigen 3 (PCA3, also referred to as DD3) is a prostate cancer-specific antigen that is encoded by a gene on chromosome 9q21-22 [5, 43, 44]. It is a long noncoding RNA of unknown function that was first reported by Bussemakers et al. in 1999 [5, 43]. PCA3 is not detected in normal prostate tissue, exhibits a low expression level in prostatic hyperplastic tissues and is highly expressed in prostate cancer [5, 43]. Owing to its restricted expression profile, PCA3 RNA serves as a useful biomarker for prostate cancer that is highly expressed in >95 % of the primary and metastatic cases. An in vitro amplification test called 'Progensa PCA3 test' (Hologic) was approved by the US FDA in 2012 for use in possible prostate cancer cases based on a negative biopsy result and/or negative PSA level and/or DRE results [19]. Through this assay the PCA3 score is determined in post DRE urine specimens of prostate cancer patients. The PCA3 score is obtained by dividing PCA3 RNA by PSA RNA levels. PCA3 scores less than 25 are

associated with a decreased likelihood of prostate cancer [45–47]. The PCA3 score has a higher specificity and a better predictive value (positive and negative) than serum PSA, although its sensitivity is lower [47]. The sensitivity of PCA3 is variable (ranging from 20 to 96 %) depending on the threshold cutoff used for the PCA3 levels [8, 48]. The PCA3 score, along with PSA and other risk factors, could be incorporated into a nomogram for improved risk stratification. The most widely studied utility of the PCA3 score has been its ability to predict malignancy in men with an elevated PSA and a prior negative biopsy (Fig. 1) [5, 8, 49]. Unlike PSA, the PCA3 score is independent of prostate volume [48]. Overall, PCA3 is a useful adjunct to the currently used methods for prostate cancer diagnosis, including PSA and DRE.

# 4 Prostate cancer biomarkers: non-FDA approved, Clinical Laboratory Improvement Amendments-based laboratory developed tests

In the following section biomarkers are discussed that have been used/developed recently and that are available as Clinical Laboratory Improvement Amendments-based laboratory developed tests [8, 19]. These tests have been commercially developed for prostate cancer but are not approved by the US FDA.

#### 4.1 TMPRSS2-ERG gene fusion test

Recurrent gene fusions involving ETS transcription factor family member genes (usually ERG, a v-ets erythroblastosis virus E26 oncogene homolog) with the androgen regulated gene TMPRSS2 (transmembrane serine protease isoform 2) are frequently encountered in prostate tumors ( $\sim$ 50 % tumors) [42, 43]. Laxman et al. demonstrated the occurrence of these gene fusions in urine of clinically localized prostate cancer patients post-DRE [44]. These gene fusions serve as potential non-invasive urinary biomarkers that have subsequently been tested in additional studies. TMPRSS2-ERG fusions in urinary sediments have been associated with a high specificity (93 %) and positive predictive value (94 %), although its sensitivity has been reported to be low (37 %) [12, 45]. The drawback of this fusion gene as a biomarker is the associated tumor heterogeneity, as most prostate tumors contain multiple foci [12]. Also, the prognostic implications of this gene fusion are not clearly defined. Some studies suggest that TMPRSS2-ERG gene fusion-positive cases are associated with a higher prostate cancer aggressiveness, metastasis and mortality [13, 46], while others have reported a lack of correlation between this fusion and clinical outcome [47]. Also, the frequency of this gene fusion is low in some populations. Hence, it is difficult to identify an appropriate cutoff across populations [13]. In view of its shortcomings, this prostate cancer-specific biomarker has been combined with PCA3 for developing a urinebased test for prostate cancer. Combining TMPRSS2-ERG and PCA3 has been shown to significantly improve the sensitivity for prostate cancer diagnosis [49, 50], with the sensitivity of PCA3 increasing from 68 to 76 % [45]. Additionally, combined TMPRSS2-ERG and PCA3 scores have been reported to improve the performance of serum PSA for predicting prostate cancer and high-grade prostate cancer at biopsy [50].

#### 4.2 Mi-Prostate score test

Considering the inherent heterogeneity of prostate cancer, a panel of markers rather than a single marker has been suggested to be more suitable for its diagnosis and prognosis [51]. Mi-Prostate Score (MiPS) is a test offered by the University of Michigan MLabs that incorporates blood PSA levels and urinary levels of *TMPRSS2-ERG* and PCA3 for prostate cancer risk assessment [19]. This validated test improves the utility of the PSA blood test and allows risk stratification of prostate cancer, while avoiding unnecessary biopsies [19, 52].

#### 4.3 Oncotype DX test

Genomic Health Inc. offers the Oncotype DX prostate cancer test, which is a multi-gene expression assay developed for formalin-fixed paraffin-embedded (FFPE) diagnostic prostate needle biopsies containing as little as 1 mm of prostate tumor. This assay assesses the activity of a set of 12 cancer-related genes to reveal the underlying biology of the tumor. The 12 cancer-related genes represented in the assay are involved in four different biological pathways, including the androgen pathway (*AZGP1, KLK2, SRD5A2, RAM13C*), proliferation (*TPX2*), cellular organization (*FLNC, GSN, TPM2, GSTM2*) and stromal response (*BGN, COL1A1* and *SFRP4*). Five reference genes have been included to normalize the data and to control for variability. These measurements are algorithmically combined to calculate a Genomic Prostate Score (GPS). This assay has been analytically and clinically validated as a predictor of aggressive prostate cancer [19, 53] and enables prostate cancer risk stratification to guide clinicians in making treatment decisions.

#### 4.4 ProMark test

ProMark, offered by Metamark, is a protein based prognostic test for predicting prostate cancer aggressiveness, particularly in patients with Gleason scores 3 + 3 or 3 + 4 [54]. This test measures the expression of eight proteins (DERL1, CUL2, SMAD4, PDSS2, HSPA9, FUS, phosphorylated S6, YBOX1) in biopsy tissue sections employing an automated immunofluorescence method. Based on the expression levels of these eight proteins, an independent risk score is calculated on a scale of 0–1 that provides a personalized prediction of disease aggressiveness. A risk score of 0.33 is 'favorable' and a score of >0.80 is 'unfavorable'. These scores are associated with false negative and false positive rates of 10 % and 5 %, respectively. This risk score was clinically validated as an independent predictor of prostate cancer aggressiveness relative to current risk stratification tools [54] and can aid in stratifying patients for active surveillance versus therapeutic intervention.

#### 4.5 ConfirmMDx test

ConfirmMDx, offered by MDx Health Inc., is a test that detects an epigenetic field effect or "halo" associated with a cancerization process at the DNA methylation level in cells adjacent to cancer foci [19, 50, 52, 55]. This test has the ability to diagnose prostate cancer cases that are histologically normal/benign. For this test, core specimens collected during a 12-core biopsy are employed with a minimum requirement of eight cores derived from the apex, mid and left/right base, and two additional locations of the prostate [19]. This test helps to distinguish patients who have a true-negative biopsy from those who may have

occult cancer with a 90 % negative predictive value (NPV). Its further clinical utility includes the identification of high-risk men who may need a repeat biopsy and, potentially, treatment.

#### 4.6 Prolaris test

Myriad Genetics Inc. offers Prolaris, a genomic test for predicting prostate cancer aggressiveness in conjunction with clinical parameters such as Gleason score and PSA. This test measures the expression of a set of 31 cell cycle progression genes and 15 housekeeping genes to predict disease progression [56–58]. The expression of the cell cycle-related genes is correlated with the proliferation of prostate cancer and serves as a risk stratification tool enabling a better treatment/monitoring strategy for patients at the time of diagnosis. A low expression of these genes is associated with a low risk of progression and those men may be candidates for active surveillance, while a high expression is associated with a higher risk of disease progression in men who may be treated. This test is significantly more prognostic than the currently used clinicopathological variables. It may also be useful in estimating the disease recurrence risk in post-prostatectomy patients.

#### 4.7 Prostate Core Mitomic test

The Prostate Core Mitomic test offered by Mitomics is based on mitochondrial DNA (mtDNA) alterations in prostate cancer biopsies. It is a highly sensitive test that yields accurate and reliable results. This test can detect large-scale mtDNA deletions (3.4 kb) in normal/benign appearing tissues obtained by biopsy via a 'cancerization' field effect [19, 59, 60]. In this assay, a section of the needle core sample from the negative biopsy is employed to assess the occurrence of 3.4 kb mtDNA deletions using real time PCR. The sensitivity of the assay is 85 % and the specificity is 54 %, with a negative predictive value of 92 %. This test helps to identify men who do not require a repeat biopsy [61].

#### 4.8 4K score test

The 4K score test, offered by Opko Health Inc., measures the blood plasma levels of a panel of four prostate-derived kallikrein proteins, i.e., total PSA, fPSA, intact PSA and human kallikrein 2 (hK2) [19, 62, 63]. These biomarkers are combined with other parameters such as age, DRE status and prior biopsy status to calculate the risk of pathologically insignificant versus aggressive prostate cancer. This test helps to identify patients eligible for biopsy based on the probability of having aggressive prostate cancer, and helps to avoid unnecessary biopsies in low-risk patients.

#### 4.9 Prostarix test

Prostarix is a test developed by Metabolon Inc. and marketed by Bostwick Laboratories. This test is based on a panel of four metabolites excreted into urine, including sarcosine, that are increased during prostate cancer progression [64]. This metabolic test uses a quantitative liquid chromatography-mass spectroscopy (LC-MS) method to accurately measure the concentrations of four amino acids in urine samples collected after DRE. From these measurements, a Prostarix score is generated. In clinical trials, the Prostarix test has shown an increased sensitivity and specificity compared to PSA. This test provides physicians with

a better risk stratification tool leading to more informed patient management decisions, especially for patients with a negative DRE or a modestly elevated serum PSA level.

#### 4.10 Decipher test

Decipher is a genomic test offered by Genome Dx Biosciences that assesses the disease progression risk after radical prostatectomy. This test evaluates the expression levels of a set of 22 RNA biomarkers involved in multiple biological pathways related to the development and progression of prostate cancer. The expression of these biomarkers is used to calculate the probability of clinical metastasis within 5 years after radical prostatectomy, and within 3 years after biochemical recurrence. The prognostic value of this test has been validated in a high risk surgical cohort where a high genomic classifier (GC) score was associated with metastasis incidence rates of>25 %. Conversely, a low GC score was found to be associated with a good prognosis in over 70 % of the high risk patients [65, 66].

# 5 Other prostate cancer biomarkers

#### 5.1 a-Methylacyl coenzyme A racemase (AMACR)

α-Methylacyl coenzyme A racemase (AMACR) is over-expressed in prostate cancer and has been used as a biomarker in needle biopsy specimens [67]. AMACR protein expression analysis in 94 prostate needle biopsy specimens revealed a 97 % sensitivity and a 100 % specificity for detecting prostate cancer [67]. In addition, AMACR expression has been found to be down-regulated in prostate cancer recurrences and metastases [68]. It is useful as a tissue biomarker in interpreting prostate needle biopsy specimens that are diagnostically challenging [67]. Although it is used as a biomarker, it is not specific to prostate cancer [15, 69].

#### 5.2 PTEN gene deletions

*PTEN* is a tumor suppressor gene located on chromosome 10 that is commonly lost in prostate cancer. This loss leads to activation of the phosphoinositide-3-kinase (PI3K) signaling pathway, which plays a cardinal role in cell growth, proliferation and metastasis and, in addition, to inhibition of the androgen receptor (AR) signaling pathway [70, 71]. *PTEN* deletions are associated with a poor prognosis and a hormone-refractory disease, and are used as a predictive biomarker for response to therapy [72, 73]. *PTEN* deletions combined with *TMPRSS2-ERG* gene fusions have a better prognostic potential. Tumors with *PTEN* loss and ERG gene fusions are associated with a poor outcome, while those with only *PTEN* loss or an *ERG* fusion exhibit intermediate-poor or intermediate outcomes, respectively [72, 74–76].

# 6 Promising alternative prostate cancer biomarkers: in infancy

Apart from the biomarkers described in the preceding sections, recent studies suggest the potential utility of additional novel biomarkers in prostate cancer as discussed in this section.

#### 6.1 Circulating tumor cells

Circulating tumor cells (CTCs) in whole blood are emerging biomarkers for cancer detection and prognosis [15, 77–79]. CTCs are malignant in origin and carry information about molecular alterations such as *TMPRSS2-ERG* fusions, *PTEN* deletions and *AR* copy number changes [15, 80]. An increased level of CTCs in blood of castration-resistant prostate cancer patients may predict a worse overall survival [15, 78, 81]. Though CTCs yield useful information, current CTC detection techniques are labor-intensive, expensive and lack sensitivity [15, 66]. Based on current detection techniques, ~50 % of the patients present with undetectable CTC levels [66] pointing at a need for a further improvement of these techniques.

#### 6.2 MicroRNA biomarkers

Prominent among alternative prostate cancer biomarkers are microRNAs (miRNAs) that constitute an evolutionarily conserved class of small (18–25 nt long) regulatory RNAs [82]. miRNA expression patterns have been found to serve as phenotypic signatures of various cancers [83], including prostate cancer. In primary prostate cancers, widespread deregulation of miRNAs has been reported compared to normal prostate tissues. Altered miRNA expression profiles have been associated with prostate cancer progression, aggressiveness, metastasis and recurrence, thereby highlighting the prognostic potential of miRNAs [83-86]. miRNAs have emerged as putative diagnostic and prognostic biomarkers that can distinguish tumor from normal tissues, aid in stratification of tumors and monitor treatment responses [83, 87]. Owing to their small size and resistance to endogenous RNase activity, miRNAs are stable biomarkers that can be readily detected in formalin-fixed tissues [88], as also in body fluids such as serum, plasma and urine [89]. Due to these attributes, miRNAs can be potentially used as both tissue biomarkers and as non-invasive cancer biomarkers [83, 90, 91]. It has been found that expression profiles of a set of 15 miRNAs (up-regulated: miR-16, -31, -125b, -145, -149, -181b, -184, -205, -221, -222; down-regulated: miR-96, -182, -182\*, -183, -375) can discriminate benign prostate tissue from prostate cancer with 84 % accuracy [92], and that serum miR-141 levels can distinguish prostate cancer patients from healthy individuals [93]. miRNA profiling in sera from healthy men and untreated prostate cancer patients of low, intermediate and high risk groups based on CAPRA scores revealed miRNA signatures for prostate cancer diagnosis and prognosis [94]. miR-107 and miR-574-3p levels were found to be higher in urine of men with prostate cancer compared to controls [95]. Clearly, miRNAs are emerging as promising alternative biomarkers for prostate cancer, although further studies are required to ultimately develop miRNAs into clinically useful biomarkers [83, 85].

#### 6.3 Exosomal biomarkers

Exosomes are small extracellular vesicles (30–100 nm in size) [96] that are gaining significant interest as alternative disease biomarkers that can be detected non-invasively in blood, urine or semen [87]. Exosomes are a promising source of protein and RNA/miRNA biomarkers for the early diagnosis and prognosis of prostate cancer [97]. Prostate-derived microvesicles (50–150 nm) or prostasomes [98] have been reported to be higher in prostate cancer patients and to correlate with increasing Gleason scores [15, 98]. Two known prostate

cancer biomarkers, PCA3 and *TMPRSS2-ERG* (see above), could be detected in exosomes isolated from urine of prostate cancer patients, showing its potential for prostate cancer diagnosis [99]. Plasma-derived exosomal survivin has been found to be high in prostate cancer patients, suggesting its use as a biomarker for the early detection of prostate cancer [100]. Increasing evidence indicates that also circulating miRNAs in body fluids are present in exosomes [101, 102]. An analysis of miRNAs in serum-derived exosomes revealed an association of miR-141 and miR-375 with metastatic prostate cancer [95].

# 7 Conclusions and perspectives

Since several years, serum PSA has been used as a prostate cancer biomarker [8]. However, due to its inherent limitations, including lack of specificity, PSA screening has proven controversial leading to an intensive search for alternative prostate cancer biomarkers with a better diagnostic and predictive potential. In particular, there is a quest for biomarkers that can distinguish between aggressive and indolent tumors, thereby leading to better treatment decisions. With the advent of advanced genomic and proteomic technologies, we have in recent years experienced an enormous increase in our understanding of prostate cancer biology. As a result, prostate cancer biomarker research has expanded its horizons with several promising alternative biomarkers being discovered. Several US FDA approved and clinical laboratory-based tests have been developed that show an improved sensitivity and specificity over PSA. However, in most recently developed tests, serum PSA is still being used in conjunction with other parameters, highlighting the fact that PSA remains an indispensable tool in the clinical management of prostate cancer. The emerging alternative biomarkers may continue to supplement or, possibly, replace PSA over time. These alternative biomarkers appear to have better detection/diagnostic and prognostic abilities/ predictive values (summarized in Fig. 1). The new generation of prostate cancer biomarkers seems promising and provides actionable guidance to risk assessment and treatment decisions. The clinical implementation of several newly discovered biomarkers has, however, often been hindered by the erroneous design of preclinical trials, inappropriate statistical analyses etc. [15]. Careful validation of emerging biomarkers such as exosomal biomarkers, CTCs and miRNAs may help to fulfill the so far unmet clinical challenges and guide clinicians to better diagnoses and better treatment options for prostate cancer. With the armamentarium of novel alternative biomarkers, prostate cancer is poised to enter an era of personalized medicine.

#### Acknowledgments

**Funding** The author is supported by the National Cancer Institute at the National Institutes of Health (Grant Number RO1CA177984).

# References

- 1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. CA Cancer J Clin. 2015; 65:5–29. [PubMed: 25559415]
- 2. Fong MK, Hare R, Jarkowski A. A new era for castrate resistant prostate cancer: a treatment review and update. J Oncol Pharm Pract. 2012; 18:343–354. [PubMed: 22343966]
- 3. Rodrigues DN, Butler LM, Estelles DL, de Bono JS. Molecular pathology and prostate cancer therapeutics: from biology to bedside. J Pathol. 2013; 232:178–184. [PubMed: 24108540]

- Loberg RD, Logothetis CJ, Keller ET, Pienta KJ. Pathogenesis and treatment of prostate cancer bone metastases: targeting the lethal phenotype. J Clin Oncol. 2005; 23:8232–8241. [PubMed: 16278478]
- 5. Romero Otero J, Garcia Gomez B, Campos Juanatey F, Touijer KA. Prostate cancer biomarkers: an update. Urol Oncol. 2014; 32:252–260. [PubMed: 24495450]
- 6. Lilja H. Testing new PSA subforms to enhance the accuracy of predicting cancer risk and disease outcome in prostate cancer. Clin Chem. 2008; 54:1248–1249. [PubMed: 18593967]
- 7. Lilja H, Ulmert D, Vickers AJ. Prostate-specific antigen and prostate cancer: prediction, detection and monitoring. Nat Rev Cancer. 2008; 8:268–278. [PubMed: 18337732]
- Cary KC, Cooperberg MR. Biomarkers in prostate cancer surveillance and screening: past, present, and future. Ther Adv Urol. 2013; 5:318–329. [PubMed: 24294290]
- 9. Epstein JI. An update of the Gleason grading system. J Urol. 2010; 183:433–440. [PubMed: 20006878]
- Mellinger GT, Gleason D, Bailar J 3rd. The histology and prognosis of prostatic cancer. J Urol. 1967; 97:331–337. [PubMed: 6018430]
- Shen MM, Abate-Shen C. Molecular genetics of prostate cancer: new prospects for old challenges. Genes Dev. 2010; 24:1967–2000. [PubMed: 20844012]
- Squire JA, Park PC, Yoshimoto M, Alami J, Williams JL, Evans A, et al. Prostate cancer as a model system for genetic diversity in tumors. Adv Cancer Res. 2011; 112:183–216. [PubMed: 21925305]
- Schoenborn JR, Nelson P, Fang M. Genomic profiling defines subtypes of prostate cancer with the potential for therapeutic stratification. Clin Cancer Res. 2013; 19:4058–4066. [PubMed: 23704282]
- Ilyin SE, Belkowski SM, Plata-Salaman CR. Biomarker discovery and validation: technologies and integrative approaches. Trends Biotechnol. 2004; 22:411–416. [PubMed: 15283986]
- 15. Prensner JR, Rubin MA, Wei JT, Chinnaiyan AM. Beyond PSA: the next generation of prostate cancer biomarkers. Sci Transl Med. 2012; 4:127rv123.
- 16. Sawyers CL. The cancer biomarker problem. Nature. 2008; 452:548–552. [PubMed: 18385728]
- Lowe FC, Trauzzi SJ. Prostatic acid phosphatase in 1993. Its limited clinical utility. Urol Clin North Am. 1993; 20:589–595. [PubMed: 8273267]
- Ercole CJ, Lange PH, Mathisen M, Chiou RK, Reddy PK, Vessella RL. Prostatic specific antigen and prostatic acid phosphatase in the monitoring and staging of patients with prostatic cancer. J Urol. 1987; 138:1181–1184. [PubMed: 2444720]
- Sartori DA, Chan DW. Biomarkers in prostate cancer: what's new? Curr Opin Oncol. 2014; 26:259–264. [PubMed: 24626128]
- Kelloff GJ, Choyke P, Coffey DS. Challenges in clinical prostate cancer: role of imaging. AJR Am J Roentgenol. 2009; 192:1455–1470. [PubMed: 19457806]
- Mazaheri Y, Shukla-Dave A, Muellner A, Hricak H. MRI of the prostate: clinical relevance and emerging applications. J Magn Reson Imaging. 2011; 33:258–274. [PubMed: 21274967]
- Bensalah K, Montorsi F, Shariat SF. Challenges of cancer biomarker profiling. Eur Urol. 2007; 52:1601–1609. [PubMed: 17919807]
- 23. Hernandez J, Thompson IM. Prostate-specific antigen: a review of the validation of the most commonly used cancer biomarker. Cancer. 2004; 101:894–904. [PubMed: 15329895]
- 24. Thompson IM, Pauler DK, Goodman PJ, Tangen CM, Lucia MS, Parnes HL, et al. Prevalence of prostate cancer among men with a prostate-specific antigen level < or =4.0 ng per milliliter. N Engl J Med. 2004; 350:2239–2246. [PubMed: 15163773]
- 25. Catalona WJ, Smith DS, Ornstein DK. Prostate cancer detection in men with serum PSA concentrations of 2.6 to 4.0 ng/mL and benign prostate examination. Enhancement of specificity with free PSA measurements. JAMA. 1997; 277:1452–1455. [PubMed: 9145717]
- Walter LC, Bertenthal D, Lindquist K, Konety BR. PSA screening among elderly men with limited life expectancies. JAMA. 2006; 296:2336–2342. [PubMed: 17105796]
- Brooks DD, Wolf A, Smith RA, Dash C, Guessous I. Prostate cancer screening 2010: updated recommendations from the American Cancer Society. J Natl Med Assoc. 2010; 102:423–429. [PubMed: 20533778]

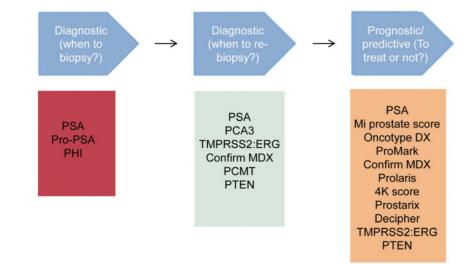
- Wolf AM, Wender RC, Etzioni RB, Thompson IM, D'Amico AV, Volk RJ, et al. American Cancer Society guideline for the early detection of prostate cancer: update 2010. CA Cancer J Clin. 2010; 60:70–98. [PubMed: 20200110]
- D'Amico AV, Chen MH, Roehl KA, Catalona WJ. Preoperative PSA velocity and the risk of death from prostate cancer after radical prostatectomy. N Engl J Med. 2004; 351:125–135. [PubMed: 15247353]
- Etzioni R, Tsodikov A, Mariotto A, Szabo A, Falcon S, Wegelin J, et al. Quantifying the role of PSA screening in the US prostate cancer mortality decline. Cancer Causes Control. 2008; 19:175– 181. [PubMed: 18027095]
- Christensson A, Bjork T, Nilsson O, Dahlen U, Matikainen MT, Cockett AT, et al. Serum prostate specific antigen complexed to alpha 1-antichymotrypsin as an indicator of prostate cancer. J Urol. 1993; 150:100–105. [PubMed: 7685416]
- 32. Catalona WJ, Partin AW, Slawin KM, Brawer MK, Flanigan RC, Patel A, et al. Use of the percentage of free prostate-specific antigen to enhance differentiation of prostate cancer from benign prostatic disease: a prospective multicenter clinical trial. JAMA. 1998; 279:1542–1547. [PubMed: 9605898]
- Khan MA, Sokoll LJ, Chan DW, Mangold LA, Mohr P, Mikolajczyk SD, et al. Clinical utility of proPSA and "benign" PSA when percent free PSA is less than 15 %. Urology. 2004; 64:1160– 1164. [PubMed: 15596190]
- 34. Hori S, Blanchet JS, McLoughlin J. From prostate-specific antigen (PSA) to precursor PSA (proPSA) isoforms: a review of the emerging role of proPSAs in the detection and management of early prostate cancer. BJU Int. 2013; 112:717–728. [PubMed: 22759214]
- Mikolajczyk SD, Marker KM, Millar LS, Kumar A, Saedi MS, Payne JK, et al. A truncated precursor form of prostate-specific antigen is a more specific serum marker of prostate cancer. Cancer Res. 2001; 61:6958–6963. [PubMed: 11559576]
- 36. Chan TY, Mikolajczyk SD, Lecksell K, Shue MJ, Rittenhouse HG, Partin AW, et al. Immunohistochemical staining of prostate cancer with monoclonal antibodies to the precursor of prostate-specific antigen. Urology. 2003; 62:177–181. [PubMed: 12837462]
- Heidegger I, Klocker H, Steiner E, Skradski V, Ladurner M, Pichler R, et al. [-2]proPSA is an early marker for prostate cancer aggressiveness. Prostate Cancer Prostatic Dis. 2014; 17:70–74. [PubMed: 24165692]
- 38. Lazzeri M, Abrate A, Lughezzani G, Gadda GM, Freschi M, Mistretta F, et al. Relationship of chronic histologic prostatic inflammation in biopsy specimens with serum isoform [-2]proPSA (p2PSA), %p2PSA, and prostate health index in men with a total prostate-specific antigen of 4-10 ng/ml and normal digital rectal examination. Urology. 2014; 83:606–612. [PubMed: 24315305]
- 39. Lazzeri M, Haese A, Abrate A, de la Taille A, Redorta JP, McNicholas T, et al. Clinical performance of serum prostate-specific antigen isoform [-2]proPSA (p2PSA) and its derivatives, %p2PSA and the prostate health index (PHI), in men with a family history of prostate cancer: results from a multicentre European study, the PROMEtheuS project. BJU Int. 2013; 112:313–321. [PubMed: 23826841]
- 40. Guazzoni G, Nava L, Lazzeri M, Scattoni V, Lughezzani G, Maccagnano C, et al. Prostate-specific antigen (PSA) isoform p2PSA significantly improves the prediction of prostate cancer at initial extended prostate biopsies in patients with total PSA between 2.0 and 10 ng/ml: results of a prospective study in a clinical setting. Eur Urol. 2011; 60:214–222. [PubMed: 21482022]
- Houlgatte A, Vincendeau S, Desfemmes F, Ramirez J, Benoist N, Bensalah K, et al. Use of [-2] pro PSA and phi index for early detection of prostate cancer: a prospective of 452 patients. Prog Urol. 2012; 22:279–283. [PubMed: 22515924]
- 42. Sokoll LJ, Sanda MG, Feng Z, Kagan J, Mizrahi IA, Broyles DL, et al. A prospective, multicenter, National Cancer Institute Early Detection Research Network study of [-2]proPSA: improving prostate cancer detection and correlating with cancer aggressiveness. Cancer Epidemiol Biomarkers Prev. 2010; 19:1193–1200. [PubMed: 20447916]
- Bussemakers MJ, van Bokhoven A, Verhaegh GW, Smit FP, Karthaus HF, Schalken JA, et al. DD3: a new prostate-specific gene, highly overexpressed in prostate cancer. Cancer Res. 1999; 59:5975– 5979. [PubMed: 10606244]

- 44. de Kok JB, Verhaegh GW, Roelofs RW, Hessels D, Kiemeney LA, Aalders TW, et al. DD3(PCA3), a very sensitive and specific marker to detect prostate tumors. Cancer Res. 2002; 62:2695–2698. [PubMed: 11980670]
- 45. Auprich M, Bjartell A, Chun FK, de la Taille A, Freedland SJ, Haese A, et al. Contemporary role of prostate cancer antigen 3 in the management of prostate cancer. Eur Urol. 2011; 60:1045–1054. [PubMed: 21871709]
- 46. Crawford ED, Rove KO, Trabulsi EJ, Qian J, Drewnowska KP, Kaminetsky JC, et al. Diagnostic performance of PCA3 to detect prostate cancer in men with increased prostate specific antigen: a prospective study of 1,962 cases. J Urol. 2012; 188:1726–1731. [PubMed: 22998901]
- 47. Vlaeminck-Guillem V, Ruffion A, Andre J, Devonec M, Paparel P. Urinary prostate cancer 3 test: toward the age of reason? Urology. 2010; 75:447–453. [PubMed: 19586654]
- Deras IL, Aubin SM, Blase A, Day JR, Koo S, Partin AW, et al. PCA3: a molecular urine assay for predicting prostate biopsy outcome. J Urol. 2008; 179:1587–1592. [PubMed: 18295257]
- Tomlins SA, Day JR, Lonigro RJ, Hovelson DH, Siddiqui J, Kunju LP, et al. Urine TMPRSS2:ERG Plus PCA3 for Individualized Prostate Cancer Risk Assessment. Eur Urol. 201510.1016/j.eururo.2015.04.039
- Leyten GH, Hessels D, Jannink SA, Smit FP, de Jong H, Cornel EB, et al. Prospective multicentre evaluation of PCA3 and TMPRSS2-ERG gene fusions as diagnostic and prognostic urinary biomarkers for prostate cancer. Eur Urol. 2014; 65:534–542. [PubMed: 23201468]
- 51. Schiffer E. Biomarkers for prostate cancer. World J Urol. 2007; 25:557–562. [PubMed: 17690889]
- Salami SS, Schmidt F, Laxman B, Regan MM, Rickman DS, Scherr D, et al. Combining urinary detection of TMPRSS2:ERG and PCA3 with serum PSA to predict diagnosis of prostate cancer. Urol Oncol. 2013; 31:566–571. [PubMed: 21600800]
- 53. Knezevic D, Goddard AD, Natraj N, Cherbavaz DB, Clark-Langone KM, Snable J, et al. Analytical validation of the Oncotype DX prostate cancer assay - a clinical RT-PCR assay optimized for prostate needle biopsies. BMC Genomics. 2013; 14:690. [PubMed: 24103217]
- Blume-Jensen P, Berman DM, Rimm DL, Shipitsin M, Putzi M, Nifong TP, et al. Development and clinical validation of an in situ biopsy-based multimarker assay for risk stratification in prostate cancer. Clin Cancer Res. 2015; 21:2591–2600. [PubMed: 25733599]
- 55. Cornu JN, Cancel-Tassin G, Egrot C, Gaffory C, Haab F, Cussenot O. Urine TMPRSS2:ERG fusion transcript integrated with PCA3 score, genotyping, and biological features are correlated to the results of prostatic biopsies in men at risk of prostate cancer. Prostate. 2013; 73:242–249. [PubMed: 22821767]
- 56. Cooperberg MR, Simko JP, Cowan JE, Reid JE, Djalilvand A, Bhatnagar S, et al. Validation of a cell-cycle progression gene panel to improve risk stratification in a contemporary prostatectomy cohort. J Clin Oncol. 2013; 31:1428–1434. [PubMed: 23460710]
- Cuzick J, Berney DM, Fisher G, Mesher D, Moller H, Reid JE, et al. Prognostic value of a cell cycle progression signature for prostate cancer death in a conservatively managed needle biopsy cohort. Br J Cancer. 2012; 106:1095–1099. [PubMed: 22361632]
- Freedland SJ, Gerber L, Reid J, Welbourn W, Tikishvili E, Park J, et al. Prognostic utility of cell cycle progression score in men with prostate cancer after primary external beam radiation therapy. Int J Radiat Oncol Biol Phys. 2013; 86:848–853. [PubMed: 23755923]
- Parr RL, Mills J, Harbottle A, Creed JM, Crewdson G, Reguly B, et al. Mitochondria, prostate cancer, and biopsy sampling error. Discov Med. 2013; 15:213–220. [PubMed: 23636138]
- 60. Verschoor ML, Ungard R, Harbottle A, Jakupciak JP, Parr RL, Singh G. Mitochondria and cancer: past, present, and future. Biomed Res Int. 2013; 2013:612369. [PubMed: 23509753]
- Robinson K, Creed J, Reguly B, Powell C, Wittock R, Klein D, et al. Accurate prediction of repeat prostate biopsy outcomes by a mitochondrial DNA deletion assay. Prostate Cancer Prostatic Dis. 2010; 13:126–131. [PubMed: 20084081]
- 62. Carlsson S, Maschino A, Schroder F, Bangma C, Steyerberg EW, van der Kwast T, et al. Predictive value of four kallikrein markers for pathologically insignificant compared with aggressive prostate cancer in radical prostatectomy specimens: results from the European Randomized Study of Screening for Prostate Cancer section Rotterdam. Eur Urol. 2013; 64:693–699. [PubMed: 23683475]

- 63. Voigt JD, Zappala SM, Vaughan ED, Wein AJ. The Kallikrein panel for prostate cancer screening: its economic impact. Prostate. 2014; 74:250–259. [PubMed: 24166488]
- 64. Sreekumar A, Poisson LM, Rajendiran TM, Khan AP, Cao Q, Yu J, et al. Metabolomic profiles delineate potential role for sarcosine in prostate cancer progression. Nature. 2009; 457:910–914. [PubMed: 19212411]
- 65. Badani K, Thompson DJ, Buerki C, Davicioni E, Garrison J, Ghadessi M, et al. Impact of a genomic classifier ofmetastatic risk on postoperative treatment recommendations for prostate cancer patients: a report from the DECIDE study group. Oncotarget. 2013; 4:600–609. [PubMed: 23592338]
- 66. Crawford ED, Ventii K, Shore ND. New biomarkers in prostate cancer. Oncology (Williston Park). 2014; 28:135–142. [PubMed: 24701701]
- Rubin MA, Zhou M, Dhanasekaran SM, Varambally S, Barrette TR, Sanda MG, et al. alpha-Methylacyl coenzyme A racemase as a tissue biomarker for prostate cancer. JAMA. 2002; 287:1662–1670. [PubMed: 11926890]
- Rubin MA, Bismar TA, Andren O, Mucci L, Kim R, Shen R, et al. Decreased alpha-methylacyl CoA racemase expression in localized prostate cancer is associated with an increased rate of biochemical recurrence and cancer-specific death. Cancer Epidemiol Biomarkers Prev. 2005; 14:1424–1432. [PubMed: 15941951]
- 69. Jiang Z, Fanger GR, Woda BA, Banner BF, Algate P, Dresser K, et al. Expression of alphamethylacyl-CoA racemase (P504s) in various malignant neoplasms and normal tissues: astudy of 761 cases. Hum Pathol. 2003; 34:792–796. [PubMed: 14506641]
- Cairns P, Okami K, Halachmi S, Halachmi N, Esteller M, Herman JG, et al. Frequent inactivation of PTEN/MMAC1 in primary prostate cancer. Cancer Res. 1997; 57:4997–5000. [PubMed: 9371490]
- Carver BS, Chapinski C, Wongvipat J, Hieronymus H, Chen Y, Chandarlapaty S, et al. Reciprocal feedback regulation of PI3K and androgen receptor signaling in PTEN-deficient prostate cancer. Cancer Cell. 2011; 19:575–586. [PubMed: 21575859]
- Bostrom PJ, Bjartell AS, Catto JW, Eggener SE, Lilja H, Loeb S, et al. Genomic predictors of outcome in prostate cancer. Eur Urol. 201510.1016/j.eururo.2015.04.008
- 73. Sircar K, Yoshimoto M, Monzon FA, Koumakpayi IH, Katz RL, Khanna A, et al. PTEN genomic deletion is associated with p-Akt and AR signalling in poorer outcome, hormone refractory prostate cancer. J Pathol. 2009; 218:505–513. [PubMed: 19402094]
- 74. Krohn A, Diedler T, Burkhardt L, Mayer PS, De Silva C, Meyer-Kornblum M, et al. Genomic deletion of PTEN is associated with tumor progression and early PSA recurrence in ERG fusionpositive and fusion-negative prostate cancer. Am J Pathol. 2012; 181:401–412. [PubMed: 22705054]
- 75. Leinonen KA, Saramaki OR, Furusato B, Kimura T, Takahashi H, Egawa S, et al. Loss of PTEN is associated with aggressive behavior in ERG-positive prostate cancer. Cancer Epidemiol. Biomarkers Prev. 2013; 22:2333–2344.
- 76. Yoshimoto M, Joshua AM, Cunha IW, Coudry RA, Fonseca FP, Ludkovski O, et al. Absence of TMPRSS2:ERG fusions and PTEN losses in prostate cancer is associated with a favorable outcome. Mod Pathol. 2008; 21:1451–1460. [PubMed: 18500259]
- 77. Broersen LH, van Pelt GW, Tollenaar RA, Mesker WE. Clinical application of circulating tumor cells in breast cancer. Cell Oncol. 2014; 37:9–15.
- Danila DC, Heller G, Gignac GA, Gonzalez-Espinoza R, Anand A, Tanaka E, et al. Circulating tumor cell number and prognosis in progressive castration-resistant prostate cancer. Clin Cancer Res. 2007; 13:7053–7058. [PubMed: 18056182]
- 79. Kim MJ, Choi NY, Lee EK, Kang MS. Identification of novel markers that outperform EpCAM in quantifying circulating tumor cells. Cell Oncol. 2014; 37:235–243.
- Attard G, Swennenhuis JF, Olmos D, Reid AH, Vickers E, A'Hern R, et al. Characterization of ERG, AR and PTEN gene status in circulating tumor cells from patients with castration-resistant prostate cancer. Cancer Res. 2009; 69:2912–2918. [PubMed: 19339269]

- de Bono JS, Scher HI, Montgomery RB, Parker C, Miller MC, Tissing H, et al. Circulating tumor cells predict survival benefit from treatment in metastatic castration-resistant prostate cancer. Clin Cancer Res. 2008; 14:6302–6309. [PubMed: 18829513]
- Bartel DP. MicroRNAs: target recognition and regulatory functions. Cell. 2009; 136:215–233. [PubMed: 19167326]
- Gordanpour A, Nam RK, Sugar L, Seth A. MicroRNAs in prostate cancer: from biomarkers to molecularly-based therapeutics. Prostate Cancer Prostatic Dis. 2012; 15:314–319. [PubMed: 22333688]
- Hurst DR, Edmonds MD, Welch DR. Metastamir: the field of metastasis-regulatory microRNA is spreading. Cancer Res. 2009; 69:7495–7498. [PubMed: 19773429]
- Saini S, Majid S, Dahiya R. Diet, microRNAs and prostate cancer. Pharm Res. 2010; 27:1014– 1026. [PubMed: 20221895]
- Saini S, Majid S, Yamamura S, Tabatabai L, Suh SO, Shahryari V, et al. Regulatory role of mir-203 in prostate cancer progression and metastasis. Clin Cancer Res. 2011; 17:5287–5298. [PubMed: 21159887]
- Giusti I, Dolo V. Extracellular vesicles in prostate cancer: new future clinical strategies? Biomed Res Int. 2014; 2014;561571. [PubMed: 24707491]
- 88. Xi Y, Nakajima G, Gavin E, Morris CG, Kudo K, Hayashi K, et al. Systematic analysis of microRNA expression of RNA extracted from fresh frozen and formalin-fixed paraffin-embedded samples. RNA. 2007; 13:1668–1674. [PubMed: 17698639]
- 89. Weber JA, Baxter DH, Zhang S, Huang DY, Huang KH, Lee MJ, et al. The microRNA spectrum in 12 body fluids. Clin Chem. 2010; 56:1733–1741. [PubMed: 20847327]
- Dijkstra S, Mulders PF, Schalken JA. Clinical use of novel urine and blood based prostate cancer biomarkers: a review. Clin Biochem. 2014; 47:889–896. [PubMed: 24177197]
- Szczyrba J, Loprich E, Wach S, Jung V, Unteregger G, Barth S, et al. The microRNA profile of prostate carcinoma obtained by deep sequencing. Mol Cancer Res. 2010; 8:529–538. [PubMed: 20353999]
- Schaefer A, Jung M, Mollenkopf HJ, Wagner I, Stephan C, Jentzmik F, et al. Diagnostic and prognostic implications of microRNA profiling in prostate carcinoma. Int J ancer. 2010; 126:1166– 1176.
- 93. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, et al. Circulating microRNAs as stable blood-based markers for cancer detection. Proc Natl Acad Sci U S A. 2008; 105:10513–10518. [PubMed: 18663219]
- 94. Moltzahn F, Olshen AB, Baehner L, Peek A, Fong L, Stoppler H, et al. Microfluidic-based multiplex qRT-PCR identifies diagnostic and prognostic microRNA signatures in the sera of prostate cancer patients. Cancer Res. 2011; 71:550–560. [PubMed: 21098088]
- Bryant RJ, Pawlowski T, Catto JW, Marsden G, Vessella RL, Rhees B, et al. Changes in circulating microRNA levels associated with prostate cancer. Br J Cancer. 2012; 106:768–774. [PubMed: 22240788]
- Mathivanan S, Ji H, Simpson RJ. Exosomes: extracellular or-ganelles important in intercellular communication. J Proteomics. 2010; 73:1907–1920. [PubMed: 20601276]
- 97. Duijvesz D, Luider T, Bangma CH, Jenster G. Exosomes as biomarker treasure chests for prostate cancer. Eur Urol. 2011; 59:823–831. [PubMed: 21196075]
- 98. Tavoosidana G, Ronquist G, Darmanis S, Yan J, Carlsson L, Wu D, et al. Multiple recognition assay reveals prostasomes as promising plasma biomarkers for prostate cancer. Proc Natl Acad Sci U S A. 2011; 108:8809–8814. [PubMed: 21555566]
- Nilsson J, Skog J, Nordstrand A, Baranov V, Mincheva-Nilsson L, Breakefield XO, et al. Prostate cancer-derived urine exosomes: a novel approach to biomarkers for prostate cancer. Br J Cancer. 2009; 100:1603–1607. [PubMed: 19401683]
- 100. Khan S, Jutzy JM, Valenzuela MM, Turay D, Aspe JR, Ashok A, et al. Plasma-derived exosomal survivin, a plausible biomarker for early detection of prostate cancer. PLoS ONE. 2012; 7:e46737. [PubMed: 23091600]

- 101. Cannistraci A, Di Pace AL, De Maria R, Bonci D. MicroRNA as new tools for prostate cancer risk assessment and therapeutic intervention: results from clinical data set and patients' samples. Biomed Res Int. 2014; 2014:146170. [PubMed: 25309903]
- 102. Cortez MA, Bueso-Ramos C, Ferdin J, Lopez-Berestein G, Sood AK, Calin GA. MicroRNAs in body fluids-the mix of hormones and biomarkers. Nat Rev Clin Oncol. 2011; 8:467–477. [PubMed: 21647195]



#### Fig. 1.

Overview of clinically validated and emerging prostate cancer biomarkers that are either approved by the US FDA or are available as Clinical Laboratory Improvement Amendments (CLIA)-based laboratory developed tests. These are blood-/urine-/tissue-based biomarkers that have a diagnostic and/or prognostic/predictive potential. Diagnostic biomarkers include those that help clinicians to decide on when to biopsy or to re-biopsy. In addition, some recently developed biomarkers may have prognostic/predictive potential and may help to guide treatment decisions