

RESEARCH ARTICLE

IL-1 Contributes to the Anti-Cancer Efficacy of Ingenol Mebutate

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OPEN ACCESS

Citation: Le TT, Skak K, Schroder K, Schroder WA, Boyle GM, Pierce CJ, et al. (2016) IL-1 Contributes to the Anti-Cancer Efficacy of Ingenol Mebutate. PLoS ONE 11(4): e0153975. doi:10.1371/journal.pone.0153975

Editor: Irina V Lebedeva, Columbia University, UNITED STATES

Received: October 7, 2015

Accepted: April 6, 2016

Published: April 21, 2016

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Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: The research was primarily conducted at QIMR Berghofer Medical Research Institute and was largely funded by Leo Pharma, and in part by a grant from the Australian National Health & Medical Research Council (APP1043104). Leo provided support in the form of salaries for TTL and consumables, was involved in the initial study design, but was not involved in data collection and analysis. Leo was involved in the decision to publish, but provided minimal input into the writing of the manuscript. K. Schroder is supported by an

Abstract

Ingenol mebutate is approved for the topical treatment of actinic keratoses and may ultimately also find utility in treating skin cancers. Here we show that relapse rates of subcutaneous B16 melanoma tumours treated topically with ingenol mebutate were not significantly different in C57BL/6 and Rag1^{-/-} mice, suggesting B and T cells do not play a major role in the anti-cancer efficacy of ingenol mebutate. Relapse rates were, however, significantly increased in MyD88^{-/-} mice and in C57BL/6 mice treated with the anti-IL-1 agent, anakinra. Ingenol mebutate treatment induces a pronounced infiltration of neutrophils, which have been shown to have anti-cancer activity in mice. Herein we provide evidence that IL-1 promotes neutrophil recruitment to the tumour, decreases apoptosis of infiltrating neutrophils and increases neutrophil tumour killing activity. These studies suggest IL-1, via its action on neutrophils, promotes the anti-cancer efficacy of ingenol mebutate, with ingenol mebutate treatment causing both IL-1β induction and IL-1α released from keratinocytes.

Introduction

Ingenol mebutate (Picato[®]) is an approved topical drug for field treatment of actinic keratoses (AKs) [1,2,3]. AKs are lesions usually found on sun-exposed skin caused by UV exposure and are characterised by atypically proliferating keratinocytes [4]. In about 8% of cases, AKs can progress to invasive squamous cell carcinomas (SCCs) [5], the second most common form of skin cancer. Removal of AKs and mutated keratinocytes by ingenol mebutate treatment thus seeks to reduce the subsequent risk of developing SCCs [1,6,7,8]. Ingenol mebutate may ultimately also find utility in treating other skin cancers [9,10], with efficacy shown in human studies for SCCs and basal cell carcinomas [11,12,13]. Ingenol mebutate is derived from the sap of *Euphorbia peplus* and activates protein kinase C (PKC) [14,15]. *In vivo* topical drug application induces primary necrosis of (i) keratinocytes, including patches of keratinocytes bearing p53 mutations and (ii) subcutaneous tumours cells growing beneath the treatment site [16,17]. Topical treatment in humans [7,18,19] and mice [16] results in a rapid onset transient

Australian Research Council Future Fellowship (FT130100361), and the Queensland Smart Futures Fund. AS is a Principal Research Fellow with the National Health & Medical Research Council, Australia (APP1058391).

Competing Interests: K. Skak is an employee of Leo Pharma. AS was a paid consultant for Leo Pharma in 2011. AS is an inventor on a series of patents covering ingenol mebutate (Picato), but receives no royalties or financial (or other) benefits as a result. These commercial affiliations do not alter the authors' adherence to PLOS ONE policies on sharing data and materials.

erythema and a pronounced recruitment of neutrophils [17,20,21]. Neutrophils recruited and stimulated by ingenol mebutate also appear to mediate anti-cancer activity by significantly reducing tumour-relapse rates in mouse tumour models, including the B16 melanoma model [21].

To investigate the role of adaptive and innate immune responses to the anti-cancer efficacy of ingenol mebutate, we investigated relapse rates after topical treatment of subcutaneous B16 tumours with ingenol mebutate in a series of genetically modified mice. Although T and B cell responses did not appear to play a major role, we provide evidence that IL-1 plays a significant role in the anti-cancer efficacy of ingenol mebutate.

Materials and Methods

Animal ethics statement

All mouse work was conducted in accordance with good animal practice as defined by the National Health and Medical Research Council of Australia. The mouse work was approved by the QIMR Berghofer Medical Research Institute animal ethics committee. Mice were euthanized when tumours reached 100 mm², a tumour size that does not cause excessive distress to the animals.

Mice, tumour inoculation and ingenol mebutate treatment

Female mice, 8–12 weeks old were injected with B16 melanoma cells (4–5x10⁵ cells/mouse in 50 µl medium) by shallow s.c. injection. B16 cells were obtained from the ATCC (CRL-6322) passaged no more than 10 times and shown to be Mycoplasma negative by MycoAlert™ Mycoplasma Detection Kit (Lonza, Basel, Switzerland) and Hoechst staining [22]. When the mean tumour size reached 10–20 mm² (2–4 days post injection) mice were chosen so that each group had a similar mean tumour size and tumour size distribution; mice with tumour sizes that could not be appropriately matched were removed. The tumour sites were then treated topically once with 10 µl placebo or 0.1% ingenol mebutate gel daily for 2 consecutive days (treatment initiation deemed to be day 0). When tumours reached 100 mm² mice were euthanized using carbon dioxide gas. The 0.1% gel comprises 1 mg/g of ingenol-3 angelate in an excipient containing isopropyl alcohol, hydroxyethylcellulose, citric acid monohydrate, sodium citrate, benzyl alcohol and purified water. This dose and schedule was chosen as previous work had shown it provided a 50–100% regression rate in the B16 mouse model [16,21].

C57BL/6 mice were purchased from ARC, Perth, Australia. MyD88^{-/-} (B6.129P2(SJL)-Myd88^{tm1.1Defr/J}), Rag1^{-/-} (B6.129S7-Rag1^{tm1Mom/J}) and µMT (B6.129S2-Igh-6^{tm1Cgn/J}) mice on a C57BL/6 background were obtained from Jackson Laboratory (Bar Harbor, ME, USA), and were bred in-house at QIMR B. FcR-common gamma chain-deficient mice (FcγR^{-/-}) (B6.129P2-FcγR1g^{tm1Rav} N12) on a C57/BL6 background were purchased from Taconic (Hudson, NY, USA).

Anakinra treatment

Kineret (anakinra) SOBI (Swedish Orphan Biovitrum) was given i.p. at 1 mg/mouse (in 100 µl in PBS) daily, day 0 (3–4 hours before ingenol mebutate treatment initiation) through to day 7. The same volume of PBS was given to control mice.

Histology and immunohistochemistry

Hematoxylin and eosin (H&E), ApoTag staining (ApoTag Plus Peroxidase *In situ* Apoptosis Detection Kit, Millipore) and anti-Ly6G staining (using rat anti-mouse Ly6G; catalogue

number NMP-R14; Abcam, Cambridge, MA, USA) were undertaken as described previously [23,24]. Slides were digitally scanned by using a Scan ScopeXT digital slide scanner (Aperio, Vista, CA) and image analyses of sections in duplicate were undertaken using Aperio ImageScope software (v10) and the Positive Pixel Count v9 algorithm using default settings.

Killing assays

B16 cells (5×10^3) were seeded into 96 well flat plates in 100 μ l DMEM supplemented with 5% fetal calf serum and 10 mM HEPES pH 7.2. The next day (day 1) bone marrow cells were collected, washed once, and added within 45 mins of collection at the indicated effector to target ratios with 6 replicates. Anakinra (100 μ g/ml final) or IL-1 β (Shenandoah Biotechnology Inc., Warwick, PA, USA) (40 ng/ml final) was then added, followed by ingenol mebutate (40 ng/ml final). The next day (day 2) suspension cells were removed by inverting the plates onto tissue paper and fresh media was added. After a further 2 days incubation (day 4), medium was removed by inverting the plates onto tissue paper and the cells were fixed and stained with crystal violet. The plates were dried, 100 μ l methanol added per well and absorbance measured at 595 nm. Killing was calculated relative to B16 cells without effectors after subtraction of background.

IL-1 α and IL-1 β protein measurements

Mice with B16 tumours were treated with ingenol mebutate day 0 and 1, treatment sites were excised at the indicated times, homogenized (using Precellys24 tissue homogenizer and ceramic beads) in PBS supplemented with 0.1% Igepal CA-630 nonionic detergent (Sigma) and protease inhibitor cocktail (Roche). IL-1 α and IL-1 β levels were measured in the supernatants using BD™ Cytometric Bead Array Flex Set reagents (BD Biosciences).

Keratinocyte cultures and anti-IL-1 α immunoblotting

Human adult epidermal keratinocytes (HEKa-APF, Cascade Biologics, OR, USA) were cultured as per supplier's recommendations on coating matrix and in low calcium medium (Epi-Life with S7) without serum (Gibco). Cells were grown to confluence in 6 well plates, calcium was then added to a final concentration of 1 mM and the cells cultured for 48 hours. Cells were then treated with the indicated concentrations of ingenol mebutate for 16 hours in 1 ml of Epi-Life medium containing 1 mM calcium. The supernatants were harvested, spun at 1000 g for 5 mins, and proteins in the supernatant precipitated using methanol as described [25]. Precipitates were then analysed by Western blotting using an anti-IL-1 α antibody (Abcam; catalogue number 9614) and goat anti-rabbit HRP secondary antibody (Merck Millipore). Blots were scanned using ImageQuant LAS 500.

Statistics

Statistical analysis was performed using IBM SPSS Statistics (version 19). Survival and relapse rates are represented as Kaplan-Meier plots, with statistical significance determined by log-rank (Mantel-Cox) tests. For other data sets the t test was used if the difference in the variances was <4 , skewness was >-2 , and kurtosis was <2 ; where the data was nonparametric and difference in variances was <4 , the Mann Whitney U test was used, if >4 the Kolmogorov-Smirnov test was used. A 2 way ANOVA was used for ApoTag staining, with data showing a parametric distribution and differences in variance <4 .

Results

Relapse rates in mice deficient in T and B cell responses

Both anti-cancer T cells and anti-cancer antibodies are induced after topical ingenol mebutate treatment (and regression) of a number of subcutaneously grown tumours in mice [9,10,21]. Experiments using the LK2 squamous cell carcinoma line grown in *Foxn1^{nu}* mice also suggesting a role for antibodies and antibody dependent cellular cytotoxicity (ADCC) in preventing relapse [21]. To investigate further the contribution of adaptive immune responses to the anti-cancer activity of ingenol mebutate, B16 tumours were grown subcutaneously to 10–20 mm² in *Rag1^{-/-}* mice (C57BL/6 background), which are unable to generate adaptive B or T cell responses, and in control C57BL/6 mice. The tumour sites were then treated twice topically (day 0 and day 1) with ingenol mebutate or placebo gel, and tumour growth was monitored over time. Relapse rates were not significantly different between *Rag1^{-/-}* and C57BL/6 mice (Figure A in [S1 File](#)), suggesting neither B nor T cells play a major role in the anti-tumour efficacy of ingenol mebutate in the B16 model.

Further experiments to investigate the role of (i) ADCC [21] using Fc receptor common gamma chain deficient mice (*FcγR^{-/-}*) mice (Figure B in [S1 File](#)) and (ii) antibodies using B cell deficient μ MT mice (Figure C in [S1 File](#)), were inconclusive as growth rates of B16 (in the absence of ingenol mebutate treatment) were different in these mice.

Increased relapse rates in *MyD88^{-/-}* mice

To investigate further the contribution of innate immune responses [21] to the anti-cancer efficacy of ingenol mebutate treatment, B16 cells were grown subcutaneously to 10–20 mm² in *MyD88^{-/-}* and C57BL/6 mice and were then treated twice (day 0 and day 1) with ingenol mebutate or placebo gel, and tumour growth monitored over time.

The relapse rates in *MyD88^{-/-}* mice were significantly higher (84%) than in C57BL/6 mice (50%) post ingenol mebutate treatment (Fig 1A), with an ensuing significant reduction in survival from 50% in C57BL/6 to 16% in *MyD88^{-/-}* mice (Fig 1B).

In contrast to a previous report [26], we observed no significant differences in the growth of B16 in *MyD88^{-/-}* and C57BL/6 mice in placebo treated mice (Figure D in [S1 File](#)). Survival rates were actually marginally (but not significantly) increased in *MyD88^{-/-}* mice when compared with C57BL/6 in the absence of ingenol mebutate (Fig 1B, compare placebo groups). The latter two observations indicate that the increased relapse rates in *MyD88^{-/-}* mice post ingenol mebutate treatment were not due to inherently more robust growth of B16 tumours in *MyD88^{-/-}* mice.

Anakinra treatment increased relapse

MyD88 is involved in signal transduction for both Toll-like receptors and the IL-1 receptor, and IL-1 β mRNA is induced in mice after topical ingenol mebutate treatment [21]. Keratinocytes express high levels of constitutive IL-1 α , which is released after topical application of phorbol ester [27]; phorbol ester, like ingenol mebutate, activates PKC [14]. The use of IL-1 deficient mice is complicated by the very different growth rates of B16 in these animals when compared with wild-type mice [28]. Thus to investigate the role of IL-1, mice were treated with anakinra, a recombinant IL-1 receptor antagonist used in the treatment of rheumatoid arthritis. Anakinra treatment significantly increased relapse rates from 0% to 50% (Fig 2A) and decreased survival from 100% to 0% (Fig 2B), suggesting that IL-1 plays a role in the anti-tumour efficacy of ingenol mebutate. Anakinra treatment did not affect the growth of established B16 tumours *in vivo* (Figure E in [S1 File](#)), consistent with previous findings [29,30,31].

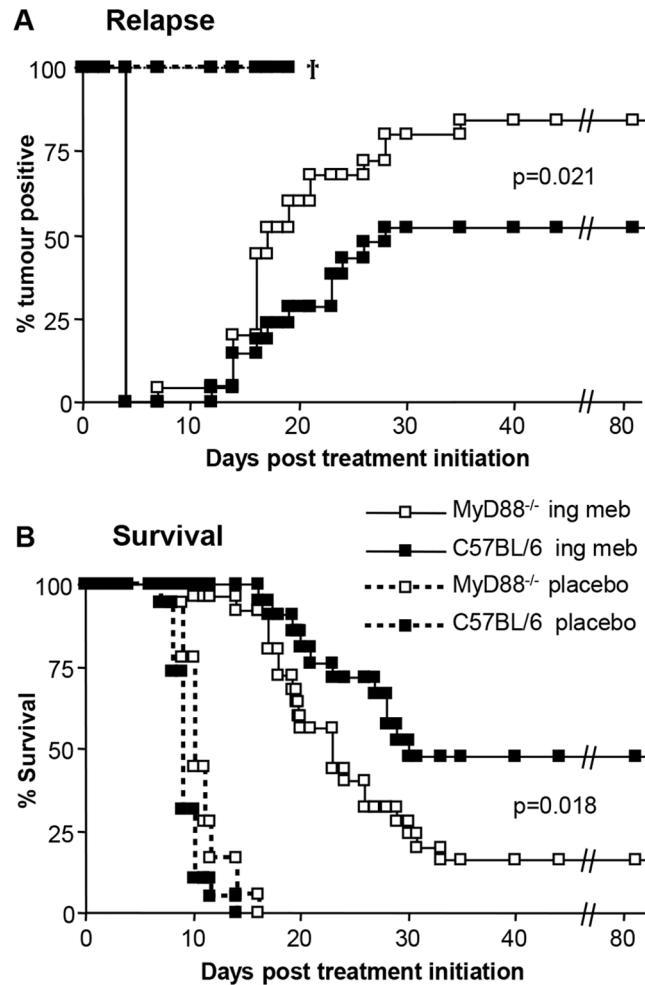


Fig 1. Relapse and survival following ingenol mebutate treatment of B16 tumours grown in MyD88^{-/-} and C57BL/6 mice. (A) Relapse rates following (i) ingenol mebutate treatment of B16 tumours grown in MyD88^{-/-} mice (n = 25), (ii) ingenol mebutate treatment of B16 tumours grown in C57BL/6 mice (n = 21), (iii) placebo treatment of B16 tumours grown in MyD88^{-/-} mice (n = 18) and (iv) placebo treatment of B16 tumours grown in C57BL/6 mice (n = 19). Mice were scored positive when a tumour was clearly visible ($\geq 1-2$ mm in diameter). Data from two independent experiments. Ingenol mebutate treatment groups were significantly different $p = 0.021$, log-rank (Mantel-Cox) test. (B) Survival rates of the same mice described in A; mice were euthanized when tumours reached 100 mm². Ingenol mebutate treatment groups were significantly different $p = 0.018$, log-rank (Mantel-Cox) test.

doi:10.1371/journal.pone.0153975.g001

Anakinra and neutrophil recruitment and apoptosis

Topical ingenol mebutate treatment results in a pronounced recruitment of neutrophils to the treatment site [9,17,20,21], which herein was quantitated using anti-Ly6G antibody staining, immunohistochemistry and image analysis (Aperio Positive Pixel count). As expected, significant recruitment of neutrophils to the treatment site (peaking on day 2 post treatment initiation) was observed (Fig 3A, Treatment site, compare day 0 with day 2 PBS). Although IL-1 has been shown to promote neutrophil recruitment in several settings [32,33,34] anakinra treatment did not significantly affect recruitment to the treatment site (Fig 3A, Treatment site, compare day 2, PBS versus anakinra). However, in sections where the tumour could be clearly seen,

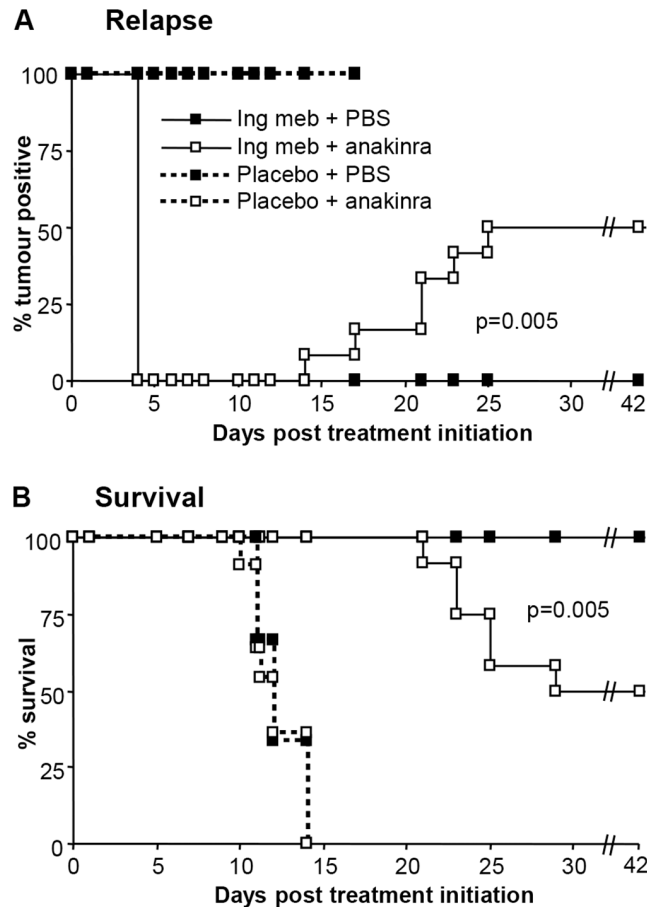


Fig 2. Relapse and survival following ingenol mebutate treatment of B16 tumours grown in C57BL/6 mice treated with anakinra. (A) Relapse rates after C57BL/6 mice bearing B16 tumours were treated with ingenol mebutate or placebo, and received daily injections of PBS or anakinra, days 1–7. (n = 9–12 mice per group). Mice were scored positive when a tumour was clearly visible (≥ 1 –2 mm in diameter). Statistics compared + anakinra with + PBS in ingenol mebutate treated groups using the log-rank (Mantel-Cox) test. (B) Survival of the mice described in A; mice were euthanized when tumours reached 100 mm². Statistics as in A.

doi:10.1371/journal.pone.0153975.g002

the density of neutrophils within ≈ 200 μ m of the tumour mass was marginally (>2 fold), but significantly, lower in anakinra treated animals (Fig 3A, Within ≈ 200 μ m of tumour, compare day 2, PBS versus anakinra). Examples of the latter are shown in Fig 3B, with neutrophils staining brown and the tumour mass readily identifiable by the black melanosomes (Fig 3B, white ovals). Anakinra thus appeared to reduce neutrophil recruitment to the tumour, with tumour-derived IL-1 β previously reported to recruit anti-cancer neutrophils to the tumour [35].

IL-1 has been reported to inhibit neutrophil apoptosis [36,37,38]. Parallel sections to those described above were therefore stained with ApoTag and image analysis (as above) of neutrophil rich areas undertaken to measure the extent of apoptosis. Examples of the Apotag staining are shown in Figure F in S1 File. Anakinra treatment more than doubled the ApoTag staining density in neutrophil rich areas in treatment sites in the dermis 2 days post ingenol mebutate treatment initiation (Fig 3C), suggesting IL-1 promotes the survival of neutrophils recruited by ingenol mebutate treatment.

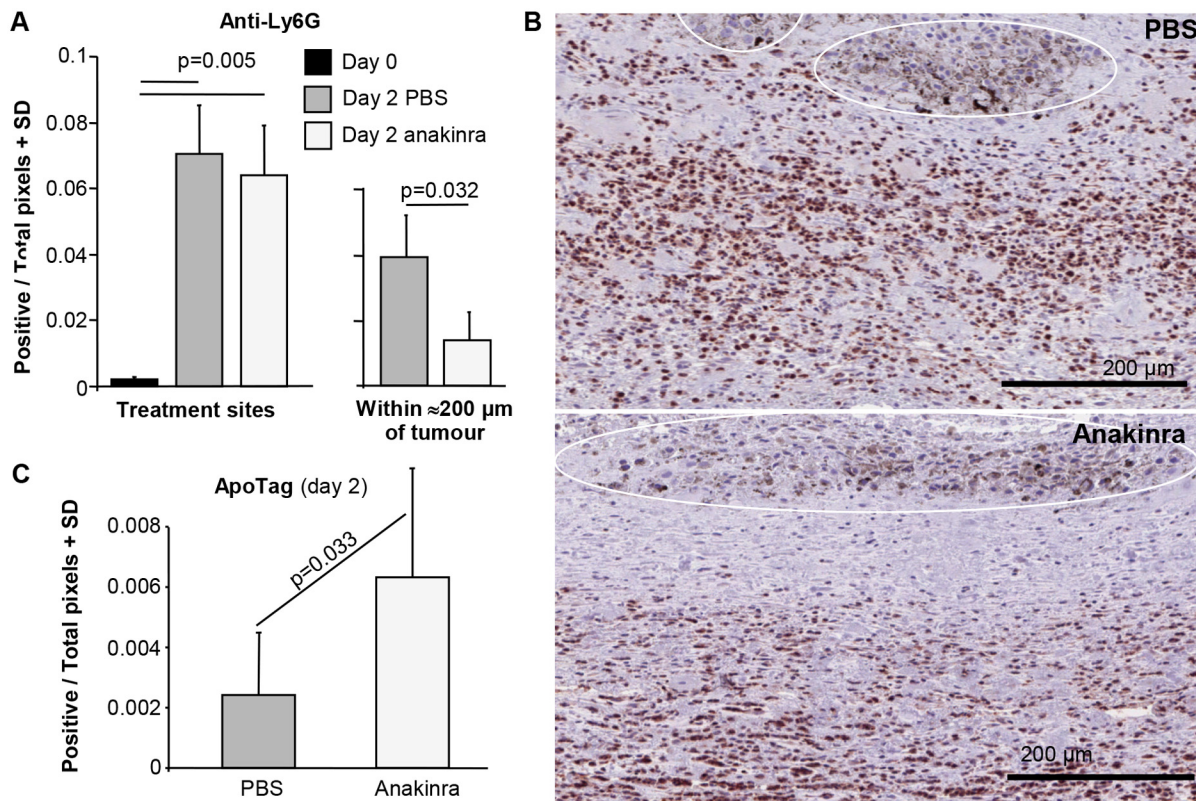


Fig 3. Neutrophil recruitment and apoptosis. (A) Neutrophil recruitment to treatments sites (left bar chart) and to within $\approx 200 \mu\text{m}$ of tumour (right bar chart). Treatment sites; day 2 post initiation of ingenol mebutate treatment, treatment sites were excised and processed for immunohistochemistry and stained with the neutrophil marker, anti-Ly6G. Slides were scanned and analysed by Aperio Pixel count software for brown staining (default settings) excluding areas containing tumour (as melanosomes provide a false positive signal). Two sections per mouse, 6 mice per group. Statistics by Kolmogorov-Smirnov tests (differences in variance between groups was >4). Within $\approx 200 \mu\text{m}$ of tumour; in sections where the tumour mass could be readily identified (by the presence of black melanosomes), brown staining surrounding the tumour (within $\approx 200 \mu\text{m}$) was quantitated as above. One section per mouse, 4–5 mice per group. Statistics by Mann Whitney U test (non-parametric data distribution and differences in variance <4). (B) Images illustrating the reduced density of anti-Ly6G staining neutrophils (brown stain) within $\approx 200 \mu\text{m}$ of the tumour mass in anakinra versus PBS treated mice. The tumours are delineated by white lines and identified by the presence of black melanosomes. Sections are oriented with the skin (not shown) at the top, with the tumours located in the dermis. (C) ApoTag staining of the sections described in A. Six mice per group, 2/3 sections per mouse, statistics by 2 way ANOVA (parametric data distribution and differences in variance <4 , drug and mouse as fixed factors, 2/3 sections per mouse as dependent variables). (Examples of the staining are shown in Figure F in [S1 File](#)).

doi:10.1371/journal.pone.0153975.g003

Ingenol mebutate and IL-1 β promote killing of B16 cells *in vitro*

We have previously shown that human neutrophils show enhanced killing of human melanoma cells *in vitro* in the presence of ingenol mebutate [21]. To investigate further the ability of ingenol mebutate to promote tumour killing by neutrophils, we developed a murine system using B16 cells and bone marrow cells, which comprise $\approx 40\%$ neutrophils, $\approx 40\%$ B cells, $\approx 10\%$ monocytic cells and a small number of other cell types. Ingenol mebutate was consistently able significantly to promote killing of B16 cells by these cells by ≥ 2 fold (Fig 4A, $p = 0.005$ and 0.002). This killing was not inhibited in the presence of anakinra (Fig 4A, dashed lines) suggesting that neither ingenol mebutate-stimulated neutrophils [39] nor B16 cells generate sufficient IL-1 to stimulate tumour cell killing [35]. However, in the presence of exogenously added IL-1 β , killing was marginally but consistently, and often significantly, increased in both the presence and absence of ingenol mebutate (Fig 4B). These data support the view

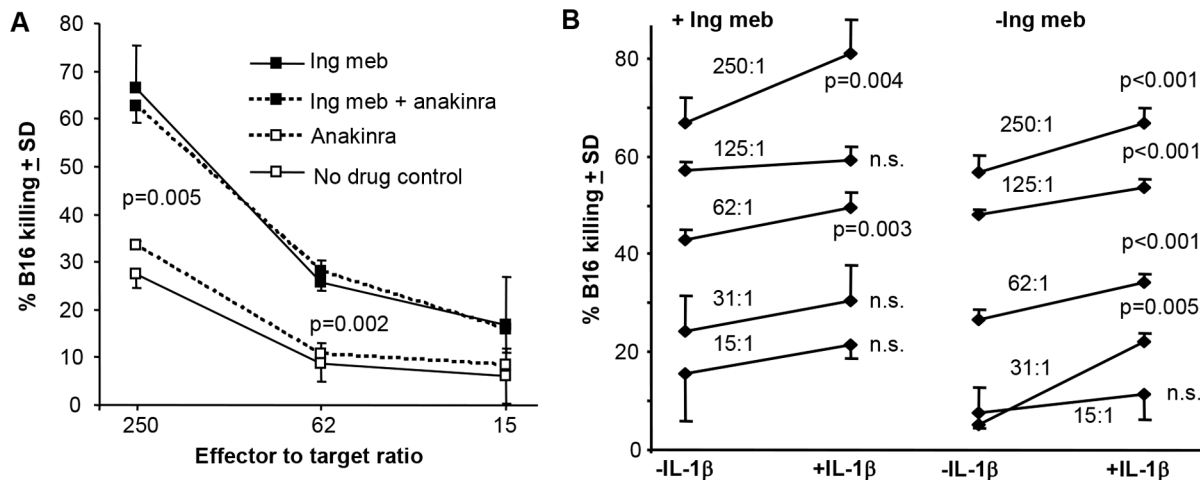


Fig 4. *In vitro* killing of B16 tumour cells in the presence of ingenol mebutate, anakinra and/or IL-1 β . (A) Bone marrow cells (\approx 40% neutrophils) were incubated with B16 cells in 96 well plates (6 replicates) at the indicated effector to target ratios and cell killing calculated relative to B16 cells with no drug or effectors. Statistics by Kolmogorov-Smirnov (difference in variance >4) $p = 0.005$, and Mann Whitney U test (non-parametric distribution and difference in variance <4) $p = 0.002$. Ingenol mebutate (40 ng/ml final) and/or anakinra (100 μ g/ml final) was present during the assay. (B) As for A except a large range of effector to target ratios (i.e. 15:1 to 250:1) were used and IL-1 β (40 ng/ml final) was added. Statistics by Kolmogorov-Smirnov (for data sets where difference in variance was >4) and t tests (for data sets with parametric distribution and differences in variance <4).

doi:10.1371/journal.pone.0153975.g004

that IL-1 improves the anti-cancer activity of neutrophils *in vitro*, consistent with previous reports showing that IL-1 can promote degranulation and respiratory burst activity in human neutrophils [40,41]. (We were unable formally to exclude the possibility that other cells in bone marrow were responsible for the anti-cancer cell activity, as purification of murine neutrophils resulted in inconsistent results).

IL-1 protein levels at the ingenol mebutate treatment sites

The data so far supports the view that IL-1 promotes the anti-cancer efficacy of ingenol mebutate by promoting neutrophil anti-cancer activity, with MyD88 required for IL-1 receptor

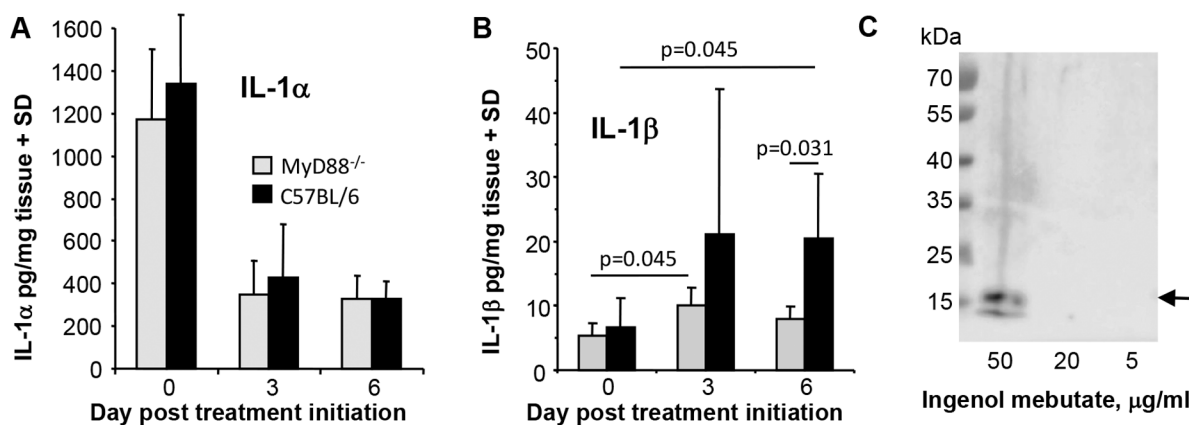


Fig 5. IL-1 α and IL-1 β protein levels after ingenol mebutate treatment. (A, B) C57BL/6 and MyD88 $^{-/-}$ mice with B16 tumours were treated topically with ingenol mebutate day 0 and 1, treatment sites were excised and IL-1 α and IL-1 β levels were measured in extracts using BD BDTM Cytometric Bead Array (n = 6 mice per group and time point). Statistics by Kolmogorov-Smirnov tests (differences in variance >4). (C) Cultured adult human keratinocytes were treated with the indicated concentration of ingenol mebutate for 16 hours and the supernatants analysed by Western using an anti-IL-1 α antibody. Arrow indicates the position of the 18 kDa bioactive form of IL-1 α .

doi:10.1371/journal.pone.0153975.g005

signal transduction [42]. To gain further insights into IL-1 production, tissue IL-1 α and IL-1 β protein levels at the treatment sites were quantitated before and after ingenol mebutate treatment. IL-1 α protein levels at the treatment sites were high prior to treatment (Fig 5A), with IL-1 α expression restricted to skin rather than the B16 tumour (Figure G in S1 File). This observation is consistent with the known high constitutive pro-IL-1 α protein expression in keratinocytes [27]. IL-1 α protein levels dropped significantly (≈ 3 fold) post treatment (Fig 5A), likely associated with extensive keratinocyte necrosis induced within 6–24 h of ingenol mebutate treatment [17]. This pattern of IL-1 α protein expression was similar in MyD88^{-/-} mice (Fig 5A, MyD88^{-/-}; Figure G in S1 File).

IL-1 β protein levels in C57BL/6 mice were low prior to ingenol mebutate treatment and increased (≈ 3 fold) after ingenol mebutate treatment, with this increase reaching significance on day 6 (Fig 5B, C57BL/6) consistent with previous mRNA data [21]. On day 6 IL-1 β protein levels were also significantly higher in ingenol mebutate treated C57BL/6 mice when compared with ingenol mebutate treated MyD88^{-/-} mice (Fig 5B, day 6). Thus MyD88^{-/-} mice have a defect in IL-1 β protein induction after ingenol mebutate treatment, consistent with the MyD88-dependent IL-1 β induction seen in several settings [43,44].

The bead based IL-1 assays, used herein on tissue lysates, are unable to differentiate between inactive (pro-IL-1) and cleaved-active IL-1 species, with recent data suggesting pro-IL-1 α also needs to be cleaved to the 18 kDa form to be active at physiological concentrations [45]. Thus IL-1 α released from keratinocytes [27] and/or IL-1 β induction may provide IL-1 after ingenol mebutate treatment.

Ingenol mebutate causes IL-1 α release from keratinocytes *in vitro*

The considerably higher levels of IL-1 α , when compared with IL-1 β (Fig 5A vs 5B), might argue that IL-1 α is a major player in this setting. The drop in total IL-1 α levels after ingenol mebutate treatment (Fig 5A) (contemporaneous with keratinocyte necrosis [17]) suggests keratinocyte IL-1 α may be released, given that other topically applied PKC activators cause IL-1 α release from keratinocytes [27]. Keratinocytes constitutively express high levels of intracellular pro-IL-1 α [27], which is bound to IL-1 receptor 2 (IL1R2) [46], with IL1R2 actually up-regulated after ingenol mebutate treatment [15]. Production of physiologically active IL-1 α is believed to require (i) release of pro-IL-1 α from IL-1R2, a process mediated by caspase-1-mediated cleavage of IL-1R2, and (ii) subsequent cleavage of the released pro-IL-1 α by calpain to generate physiologically bioactive 18 kDa IL-1 α [45].

To determine whether ingenol mebutate treatment of keratinocytes causes the release of IL-1 α , cultured primary human keratinocytes were treated with ingenol mebutate *in vitro* for 16 hours and the supernatant analysed for the presence of the 18 kDa species of IL-1 α by Western. Treatment with ingenol mebutate at 50 μ g/ml, but not 20 or 5 μ g/ml, resulted in the release of 18 kDa IL-1 α (Fig 5C, arrow). The 50 μ g/ml concentration causes increases in cytosolic calcium, but is just below the concentration that causes overt cytotoxic effects in keratinocytes [47]; (we also observed no overt cell death in these cultures).

Discussion

Herein we provide evidence that the anti-cancer efficacy of topical ingenol mebutate treatment requires both MyD88 and IL-1. Increased tumour relapse rates were seen in MyD88^{-/-} mice, and in C57BL/6 mice following anakinra (anti-IL-1) treatment, with MyD88 required for IL-1 receptor signalling [42]. Topical ingenol mebutate treatment is known to cause a pronounced recruitment of neutrophils to the treatment site [9,17,20,21], with IL-1 previously shown to promote the anti-cancer activities of neutrophils in the B16 model [35,48]. (IL-1 β has no direct

effect on B16 growth [35]). Neutrophils are also known to mediate anti-cancer activities in other systems [49,50]. We show herein that IL-1 affects three aspects of neutrophil behaviour following ingenol mebutate administration; (i) increased recruitment to the tumour (Fig 3B), consistent with the widely reported role of IL-1 in promoting neutrophil recruitment [32,33,34,35,51,52,53], (ii) reduced neutrophil apoptosis (Fig 3C), with IL-1 previously reported to inhibit neutrophil apoptosis [36,37,38], and (iii) increased tumour killing, consistent with the reported role of IL-1 in promoting neutrophil activation [40,41]. IL-1 and IL-1 receptor signalling thus appear to play a role in reducing relapse rates post ingenol mebutate treatment by promoting the anti-cancer activities of neutrophils.

Treatment of keratinocytes *in vitro*, with a dose of ingenol mebutate that is capable of raising intracellular calcium [47], resulted in the release of the 18 kDa bioactive form of IL-1 α . Keratinocyte-derived IL-1 α may thus (at least initially) be a major source of IL-1 after ingenol mebutate treatment. Lower ingenol mebutate doses did not induce IL-1 α release, suggesting PKC activation (which occurs at concentrations as low as 10 ng/ml [21]) is not sufficient for IL-1 α release. Increased intracellular calcium [16] might be expected to activate calpain [54], with active calpain required for cleavage of pro-IL-1 α to the 18 kDa IL-1 α species [45]. However, pro-IL-1 α must first be released from IL-1R2, a process believed to be mediated by caspase-1-mediated cleavage of IL-1R2 [45]. Release of 18 kDa IL-1 α release may suggest activation of keratinocyte caspase-1, and caspase 1 induction and activation can be detected in skin after ingenol mebutate treatment (Figure H in S1 File). Furthermore, permeabilization of the plasma membrane may be sufficient for inflammasome (and thus caspase 1) activation [55], with ingenol mebutate able to induce plasma membrane permeabilization [16,47]. However, activation of keratinocyte caspase-1 (and perhaps pyroptosis [56]) after topical ingenol mebutate treatment remains to be formally demonstrated, and is complicated by the abundance of neutrophils, which also express caspase 1.

The keratinocyte-derived IL-1 α may be involved in (MyD88-dependent) induction of IL-1 β (Fig 5B), with IL-1 β induction by IL-1 reported previously [57,58] and tumour-derived IL-1 β also shown to recruit neutrophils to the tumour [35]. As B16 cells do not make IL-1 [35], non-malignant cells in/around the tumour (e.g. stromal cells, fibroblasts and/or macrophages) may represent the source of IL-1 β . After ingenol mebutate treatment *in vivo* (i) B16 tumour tissue show a \approx 1.5 fold induction of IL-1 β mRNA [21] and (ii) tumour treatment sites show a \approx 2.5 fold increase in IL-1 β protein levels (Fig 5B). Stimulated neutrophils can also make IL-1 β [39,59]. Toll-like receptor engagement could also be involved in IL-1 β induction [43,44], with induction of primary necrosis and haemorrhage in the tumour (evident after ingenol mebutate treatment in mice [60], see also Figure I in S1 File) potentially providing self-derived Toll-like receptor agonists [61].

Previous data showing increased relapse of LK2 tumours in SCID mice (that make little or no immunoglobulin) versus *Foxn1*tm mice (that retain the ability to make IgM), suggested a role for anti-cancer antibodies and neutrophil ADCC in reducing relapse rates [21]. However, the data presented herein using B16 tumours grown in *Rag1*^{-/-} mice, suggests neither antibodies (nor T cells) play a major role in reducing relapse rates. LK2 tumours were generated in C3H/HeNcr mice (H2^k) [62], but were grown in SCID and *Foxn1*tm mice on a Balb/c (H2^d) background [21]. The discrepancy between the LK2 and B16 models may thus be explained (at least in part) by allogeneic IgM responses directed against LK2 in *Foxn1*tm mice, which would not be present in the syngeneic (and poorly immunogenic) B16 C57BL/6 model. Although IgG responses specific for the tumour are generated after ingenol mebutate treatment of B16 tumours [21] (and T7 squamous cell carcinomas [9]), the *Rag1*^{-/-} experiments would suggest such responses have minimal impact on relapse rates of the ingenol mebutate treated tumours. Injection of anti-B16 anti-serum (raised against B16 lysates) also did not reduce relapse rates

after ingenol mebutate treatment (Figure J in [S1 File](#)), providing further support for this contention. In summary, no major role for antibodies in the anti-cancer efficacy of ingenol mebutate was apparent in the B16 C57BL/6 model.

Although many of the complex processes and interactions involved in the anti-cancer activity of ingenol mebutate remain to be established, this paper provides evidence that IL-1 (and its effect on neutrophils) is involved in the anti-cancer efficacy of ingenol mebutate. Our study also highlights the question of whether other inflammatory cytokines, for instance, TNF or IL-6 (both also induced by ingenol mebutate [21]), are also involved in the anti-cancer efficacy of ingenol mebutate.

Supporting Information

S1 File. Supporting Information. B16 relapse rates in Rag1^{-/-} mice after ingenol mebutate treatment (**Figure A**). B16 relapse, survival and growth in FcγR^{-/-} mice after ingenol mebutate (**Figure B**). B16 relapse rates in μMT^{-/-} mice after ingenol mebutate treatment (**Figure C**). Growth of B16 tumours in placebo treated MyD88^{-/-} and C57BL/6 mice (**Figure D**). Anakinra did not affect B16 growth (**Figure E**). High resolution images of ApoTag staining (**Figure F**). IL-1α levels in skin and B16 tumours (**Figure G**.) Capase 1 in skin after ingenol mebutate treatment (**Figure H**). Haemorrhage post ingenol mebutate treatment (**Figure I**). The effects of anti-B16 anti-serum on relapse rates of B16 tumours after ingenol mebutate treatment (**Figure J**).
(PDF)

Acknowledgments

We would like to thank Clay Winterford and Glynn Res (QIMR B) for assistance with histology, and the QIMR B animal house staff for their excellent support. The authors would also like to thank Dr Chris Schmidt for his assistance with the manuscript.

Author Contributions

Conceived and designed the experiments: K. Schroder K. Skak AS WS GB. Performed the experiments: TTL K. Schroder WS CP. Analyzed the data: K. Schroder AS. Contributed reagents/materials/analysis tools: K. Skak GB CP. Wrote the paper: AS.

References

1. Zarchi K, Jemec GB. Ingenol mebutate: from common weed to cancer cure. *Curr Probl Dermatol*. 2015; 46: 136–142. doi: [10.1159/000366549](#) PMID: [25561218](#)
2. Ogbourne SM, Parsons PG. The value of nature's natural product library for the discovery of New Chemical Entities: the discovery of ingenol mebutate. *Fitoterapia*. 2014; 98: 36–44. doi: [10.1016/j.fitote.2014.07.002](#) PMID: [25016953](#)
3. Tzogani K, Nagercoil N, Hemmings RJ, Samir B, Gardette J, Demolis P, et al. The European Medicines Agency approval of ingenol mebutate (Picato) for the cutaneous treatment of non-hyperkeratotic, non-hypertrophic actinic keratosis in adults: Summary of the scientific assessment of the Committee for Medicinal Products for Human Use (CHMP). *Eur J Dermatol*. 2014; 24: 457–463. doi: [10.1684/ejd.2014.2368](#) PMID: [25115145](#)
4. Ackerman AB, Mones JM. Solar (actinic) keratosis is squamous cell carcinoma. *Br J Dermatol*. 2006; 155: 9–22. PMID: [16792746](#)
5. Glogau RG. The risk of progression to invasive disease. *J Am Acad Dermatol*. 2000; 42: 23–24. PMID: [10607353](#)
6. Lebwohl M, Shumack S, Stein Gold L, Melgaard A, Larsson T, Tying SK. Long-term follow-up study of ingenol mebutate gel for the treatment of actinic keratoses. *JAMA Dermatol*. 2013; 149: 666–670. doi: [10.1001/jamadermatol.2013.2766](#) PMID: [23553119](#)

7. Lebowhl M, Swanson N, Anderson LL, Melgaard A, Xu Z, Berman B. Ingenol mebutate gel for actinic keratosis. *N Engl J Med*. 2012; 366: 1010–1019. doi: [10.1056/NEJMoa1111170](https://doi.org/10.1056/NEJMoa1111170) PMID: [22417254](https://pubmed.ncbi.nlm.nih.gov/22417254/)
8. Alchin DR. Ingenol mebutate: a succinct review of a succinct therapy. *Dermatol Ther (Heidelb)*. 2014; 4: 157–164.
9. Cozzi SJ, Le TT, Ogbourne SM, James C, Suhrbier A. Effective treatment of squamous cell carcinomas with ingenol mebutate gel in immunologically intact SKH1 mice. *Arch Dermatol Res*. 2013; 305: 79–83. doi: [10.1007/s00403-012-1270-0](https://doi.org/10.1007/s00403-012-1270-0) PMID: [22871992](https://pubmed.ncbi.nlm.nih.gov/22871992/)
10. Le TT, Gardner J, Hoang-Le D, Schmidt CW, MacDonald KP, Lambley E, et al. Immunostimulatory cancer chemotherapy using local ingenol-3-angelate and synergy with immunotherapies. *Vaccine*. 2009; 27: 3053–3062. doi: [10.1016/j.vaccine.2009.03.025](https://doi.org/10.1016/j.vaccine.2009.03.025) PMID: [19428919](https://pubmed.ncbi.nlm.nih.gov/19428919/)
11. Ramsay JR, Suhrbier A, Aylward JH, Ogbourne S, Cozzi SJ, Poulsen MG, et al. The sap from *Euphorbia peplus* is effective against human nonmelanoma skin cancers. *Br J Dermatol*. 2011; 164: 633–636. doi: [10.1111/j.1365-2133.2010.10184.x](https://doi.org/10.1111/j.1365-2133.2010.10184.x) PMID: [21375515](https://pubmed.ncbi.nlm.nih.gov/21375515/)
12. Siller G, Rosen R, Freeman M, Welburn P, Katsamas J, Ogbourne SM. PEP005 (ingenol mebutate) gel for the topical treatment of superficial basal cell carcinoma: results of a randomized phase IIa trial. *Australas J Dermatol*. 2010; 51: 99–105. doi: [10.1111/j.1440-0960.2010.00626.x](https://doi.org/10.1111/j.1440-0960.2010.00626.x) PMID: [20546215](https://pubmed.ncbi.nlm.nih.gov/20546215/)
13. Cantisani C, Paolino G, Cantoresi F, Faina V, Richetta AG, Calvieri S. Superficial basal cell carcinoma successfully treated with ingenol mebutate gel 0.05%. *Dermatol Ther*. 2014; 27: 352–354. doi: [10.1111/dth.12148](https://doi.org/10.1111/dth.12148) PMID: [25052730](https://pubmed.ncbi.nlm.nih.gov/25052730/)
14. Kedei N, Lundberg DJ, Toth A, Welburn P, Garfield SH, Blumberg PM. Characterization of the interaction of ingenol 3-angelate with protein kinase C. *Cancer Res*. 2004; 64: 3243–3255. PMID: [15126366](https://pubmed.ncbi.nlm.nih.gov/15126366/)
15. Freiburger SN, Cheng PF, Iotzova-Weiss G, Neu J, Liu Q, Dziunycz P, et al. Ingenol Mebutate Signals via PKC/MEK/ERK in Keratinocytes and Induces Interleukin Decoy Receptors IL1R2 and IL13RA2. *Mol Cancer Ther*. 2015; 14: 2132–2142. doi: [10.1158/1535-7163.MCT-15-0023-T](https://doi.org/10.1158/1535-7163.MCT-15-0023-T) PMID: [26116359](https://pubmed.ncbi.nlm.nih.gov/26116359/)
16. Ogbourne SM, Suhrbier A, Jones B, Cozzi SJ, Boyle GM, Morris M, et al. Antitumor activity of 3-ingenol angelate: plasma membrane and mitochondrial disruption and necrotic cell death. *Cancer Res*. 2004; 64: 2833–2839. PMID: [15087400](https://pubmed.ncbi.nlm.nih.gov/15087400/)
17. Cozzi SJ, Ogbourne SM, James C, Rebel HG, de Gruijl FR, Ferguson B, et al. Ingenol mebutate field-directed treatment of UVB-damaged skin reduces lesion formation and removes mutant p53 patches. *J Invest Dermatol*. 2012; 132: 1263–1271. doi: [10.1038/jid.2011.418](https://doi.org/10.1038/jid.2011.418) PMID: [22189786](https://pubmed.ncbi.nlm.nih.gov/22189786/)
18. Anderson L, Schmieder GJ, Werschler WP, Tschen EH, Ling MR, Stough DB, et al. Randomized, double-blind, double-dummy, vehicle-controlled study of ingenol mebutate gel 0.025% and 0.05% for actinic keratosis. *J Am Acad Dermatol*. 2009; 60: 934–943. doi: [10.1016/j.jaad.2009.01.008](https://doi.org/10.1016/j.jaad.2009.01.008) PMID: [19467365](https://pubmed.ncbi.nlm.nih.gov/19467365/)
19. Siller G, Gebauer K, Welburn P, Katsamas J, Ogbourne SM. PEP005 (ingenol mebutate) gel, a novel agent for the treatment of actinic keratosis: results of a randomized, double-blind, vehicle-controlled, multicentre, phase IIa study. *Australas J Dermatol*. 2009; 50: 16–22. doi: [10.1111/j.1440-0960.2008.00497.x](https://doi.org/10.1111/j.1440-0960.2008.00497.x) PMID: [19178487](https://pubmed.ncbi.nlm.nih.gov/19178487/)
20. Hampson P, Kavanagh D, Smith E, Wang K, Lord JM, Ed Rainger G. The anti-tumor agent, ingenol-3-angelate (PEP005), promotes the recruitment of cytotoxic neutrophils by activation of vascular endothelial cells in a PKC-delta dependent manner. *Cancer Immunol Immunother*. 2008; 57: 1241–1251. doi: [10.1007/s00262-008-0458-9](https://doi.org/10.1007/s00262-008-0458-9) PMID: [18265980](https://pubmed.ncbi.nlm.nih.gov/18265980/)
21. Challacombe JM, Suhrbier A, Parsons PG, Jones B, Hampson P, Kavanagh D, et al. Neutrophils are a key component of the antitumor efficacy of topical chemotherapy with ingenol-3-angelate. *J Immunol*. 2006; 177: 8123–8132. PMID: [17114487](https://pubmed.ncbi.nlm.nih.gov/17114487/)
22. La Linn M, Bellett AJ, Parsons PG, Suhrbier A. Complete removal of mycoplasma from viral preparations using solvent extraction. *J Virol Methods*. 1995; 52: 51–54. PMID: [7539444](https://pubmed.ncbi.nlm.nih.gov/7539444/)
23. Poo YS, Nakaya H, Gardner J, Larcher T, Schroder WA, Le TT, et al. CCR2 deficiency promotes exacerbated chronic erosive neutrophil-dominated chikungunya virus arthritis. *J Virol*. 2014; 88: 6862–6872. doi: [10.1128/JVI.03364-13](https://doi.org/10.1128/JVI.03364-13) PMID: [24696480](https://pubmed.ncbi.nlm.nih.gov/24696480/)
24. Nakaya HI, Gardner J, Poo YS, Major L, Pulendran B, Suhrbier A. Gene profiling of Chikungunya virus arthritis in a mouse model reveals significant overlap with rheumatoid arthritis. *Arthritis Rheum*. 2012; 64: 3553–3563. doi: [10.1002/art.34631](https://doi.org/10.1002/art.34631) PMID: [22833339](https://pubmed.ncbi.nlm.nih.gov/22833339/)
25. Gross O. Measuring the inflammasome. *Methods Mol Biol*. 2012; 844: 199–222. doi: [10.1007/978-1-61779-527-5_15](https://doi.org/10.1007/978-1-61779-527-5_15) PMID: [22262445](https://pubmed.ncbi.nlm.nih.gov/22262445/)
26. Hayashi T, Chan M, Norton JT, Wu CC, Yao S, Cottam HB, et al. Additive melanoma suppression with intralesional phospholipid-conjugated TLR7 agonists and systemic IL-2. *Melanoma Res*. 2011; 21: 66–75. doi: [10.1097/CMR.0b013e328340ce6c](https://doi.org/10.1097/CMR.0b013e328340ce6c) PMID: [21030882](https://pubmed.ncbi.nlm.nih.gov/21030882/)

27. Murphy JE, Morales RE, Scott J, Kupper TS. IL-1 alpha, innate immunity, and skin carcinogenesis: the effect of constitutive expression of IL-1 alpha in epidermis on chemical carcinogenesis. *J Immunol*. 2003; 170: 5697–5703. PMID: [12759452](#)
28. Voronov E, Shouval DS, Krelin Y, Cagnano E, Benharroch D, Iwakura Y, et al. IL-1 is required for tumor invasiveness and angiogenesis. *Proc Natl Acad Sci U S A*. 2003; 100: 2645–2650. PMID: [12598651](#)
29. Bjorkdahl O, Wingren AG, Hedhund G, Ohlsson L, Dohlsten M. Gene transfer of a hybrid interleukin-1 beta gene to B16 mouse melanoma recruits leucocyte subsets and reduces tumour growth in vivo. *Cancer Immunol Immunother*. 1997; 44: 273–281. PMID: [9247562](#)
30. Bjorkdahl O, Dohlsten M, Sjogren HO. Vaccination with B16 melanoma cells expressing a secreted form of interleukin-1beta induces tumor growth inhibition and an enhanced immunity against the wild-type B16 tumor. *Cancer Gene Ther*. 2000; 7: 1365–1374. PMID: [11059695](#)
31. Triozzi PL, Aldrich W. Effects of interleukin-1 receptor antagonist and chemotherapy on host-tumor interactions in established melanoma. *Anticancer Res*. 2010; 30: 345–354. PMID: [20332438](#)
32. Sonogo F, Castanheira FV, Czaikoski PG, Kanashiro A, Souto FO, Franca RO, et al. MyD88-, but not Nod1- and/or Nod2-deficient mice, show increased susceptibility to polymicrobial sepsis due to impaired local inflammatory response. *PLoS One*. 2014; 9: e103734. doi: [10.1371/journal.pone.0103734](#) PMID: [25084278](#)
33. Feng Y, Zou L, Si R, Nagasaka Y, Chao W. Bone marrow MyD88 signaling modulates neutrophil function and ischemic myocardial injury. *Am J Physiol Cell Physiol*. 2010; 299: C760–769. doi: [10.1152/ajpcell.00155.2010](#) PMID: [20631245](#)
34. Jarchum I, Liu M, Shi C, Equinda M, Pamer EG. Critical role for MyD88-mediated neutrophil recruitment during *Clostridium difficile* colitis. *Infect Immun*. 2012; 80: 2989–2996. doi: [10.1128/IAI.00448-12](#) PMID: [22689818](#)
35. Chen LC, Wang LJ, Tsang NM, Ojcius DM, Chen CC, Ouyang CN, et al. Tumour inflammasome-derived IL-1beta recruits neutrophils and improves local recurrence-free survival in EBV-induced nasopharyngeal carcinoma. *EMBO Mol Med*. 2012; 4: 1276–1293. doi: [10.1002/emmm.201201569](#) PMID: [23065753](#)
36. Watson RW, Rotstein OD, Parodo J, Bitar R, Marshall JC. The IL-1 beta-converting enzyme (caspase-1) inhibits apoptosis of inflammatory neutrophils through activation of IL-1 beta. *J Immunol*. 1998; 161: 957–962. PMID: [9670975](#)
37. Prince LR, Allen L, Jones EC, Hellewell PG, Dower SK, Whyte MK, et al. The role of interleukin-1beta in direct and toll-like receptor 4-mediated neutrophil activation and survival. *Am J Pathol*. 2004; 165: 1819–1826. PMID: [15509550](#)
38. Colotta F, Re F, Polentarutti N, Sozzani S, Mantovani A. Modulation of granulocyte survival and programmed cell death by cytokines and bacterial products. *Blood*. 1992; 80: 2012–2020. PMID: [1382715](#)
39. Bakele M, Joos M, Burdi S, Allgaier N, Poschel S, Fehrenbacher B, et al. Localization and functionality of the inflammasome in neutrophils. *J Biol Chem*. 2014; 289: 5320–5329. doi: [10.1074/jbc.M113.505636](#) PMID: [24398679](#)
40. Smith RJ, Bowman BJ, Speziale SC. Interleukin-1 stimulates granule exocytosis from human neutrophils. *Int J Immunopharmacol*. 1986; 8: 33–40. PMID: [2420732](#)
41. Yagisawa M, Yuo A, Kitagawa S, Yazaki Y, Togawa A, Takaku F. Stimulation and priming of human neutrophils by IL-1 alpha and IL-1 beta: complete inhibition by IL-1 receptor antagonist and no interaction with other cytokines. *Exp Hematol*. 1995; 23: 603–608. PMID: [7601250](#)
42. Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. *Immunity*. 2013; 39: 1003–1018. doi: [10.1016/j.immuni.2013.11.010](#) PMID: [24332029](#)
43. Wu TT, Tai YT, Cherng YG, Chen TG, Lin CJ, Chen TL, et al. GATA-2 transduces LPS-induced il-1beta gene expression in macrophages via a toll-like receptor 4/MD88/MAPK-dependent mechanism. *PLoS One*. 2013; 8: e72404. doi: [10.1371/journal.pone.0072404](#) PMID: [23940812](#)
44. Shaw MH, Kamada N, Kim YG, Nunez G. Microbiota-induced IL-1beta, but not IL-6, is critical for the development of steady-state TH17 cells in the intestine. *J Exp Med*. 2012; 209: 251–258. doi: [10.1084/jem.20111703](#) PMID: [22291094](#)
45. Zheng Y, Humphry M, Maguire JJ, Bennett MR, Clarke MC. Intracellular interleukin-1 receptor 2 binding prevents cleavage and activity of interleukin-1alpha, controlling necrosis-induced sterile inflammation. *Immunity*. 2013; 38: 285–295. doi: [10.1016/j.immuni.2013.01.008](#) PMID: [23395675](#)
46. Rauschmayr T, Groves RW, Kupper TS. Keratinocyte expression of the type 2 interleukin 1 receptor mediates local and specific inhibition of interleukin 1-mediated inflammation. *Proc Natl Acad Sci U S A*. 1997; 94: 5814–5819. PMID: [9159157](#)

47. Stahlhut M, Bertelsen M, Hoyer-Hansen M, Svendsen N, Eriksson AH, Lord JM, et al. Ingenol mebutate: induced cell death patterns in normal and cancer epithelial cells. *J Drugs Dermatol*. 2012; 11: 1181–1192. PMID: [23134983](#)
48. Neville ME, Pezzella KM, Schmidt K, Galbraith W, Ackerman N. In vivo inhibition of tumor growth of B16 melanoma by recombinant interleukin 1 beta. II. Mechanism of inhibition: the role of polymorphonuclear leukocytes. *Cytokine*. 1990; 2: 456–463. PMID: [1966551](#)
49. van Egmond M, Bakema JE. Neutrophils as effector cells for antibody-based immunotherapy of cancer. *Semin Cancer Biol*. 2013; 23: 190–199. doi: [10.1016/j.semcancer.2012.12.002](#) PMID: [23287459](#)
50. Albanesi M, Mancardi DA, Jonsson F, Iannascoli B, Fiette L, Di Santo JP, et al. Neutrophils mediate antibody-induced antitumor effects in mice. *Blood*. 2013; 122: 3160–3164. doi: [10.1182/blood-2013-04-497446](#) PMID: [23980063](#)
51. Martinon F, Petrilli V, Mayor A, Tardivel A, Tschopp J. Gout-associated uric acid crystals activate the NALP3 inflammasome. *Nature*. 2006; 440: 237–241. PMID: [16407889](#)
52. Dostert C, Guarda G, Romero JF, Menu P, Gross O, Tardivel A, et al. Malarial hemozoin is a Nalp3 inflammasome activating danger signal. *PLoS One*. 2009; 4: e6510. doi: [10.1371/journal.pone.0006510](#) PMID: [19652710](#)
53. Yazdi AS, Guarda G, Riteau N, Drexler SK, Tardivel A, Couillin I, et al. Nanoparticles activate the NLR pyrin domain containing 3 (Nlrp3) inflammasome and cause pulmonary inflammation through release of IL-1alpha and IL-1beta. *Proc Natl Acad Sci U S A*. 2010; 107: 19449–19454. doi: [10.1073/pnas.1008155107](#) PMID: [20974980](#)
54. Gross O, Yazdi AS, Thomas CJ, Masin M, Heinz LX, Guarda G, et al. Inflammasome activators induce interleukin-1alpha secretion via distinct pathways with differential requirement for the protease function of caspase-1. *Immunity*. 2012; 36: 388–400. doi: [10.1016/j.immuni.2012.01.018](#) PMID: [22444631](#)
55. Cullen SP, Kearney CJ, Clancy DM, Martin SJ. Diverse Activators of the NLRP3 Inflammasome Promote IL-1beta Secretion by Triggering Necrosis. *Cell Rep*. 2015; 11: 1535–1548. doi: [10.1016/j.celrep.2015.05.003](#) PMID: [26027935](#)
56. Winkler S, Rosen-Wolff A. Caspase-1: an integral regulator of innate immunity. *Semin Immunopathol*. 2015; 37: 419–427. doi: [10.1007/s00281-015-0494-4](#) PMID: [26059719](#)
57. Toda Y, Tsukada J, Misago M, Kominato Y, Auron PE, Tanaka Y. Autocrine induction of the human pro-IL-1beta gene promoter by IL-1beta in monocytes. *J Immunol*. 2002; 168: 1984–1991. PMID: [11823535](#)
58. Lachmann HJ, Lowe P, Felix SD, Rordorf C, Leslie K, Madhoo S, et al. In vivo regulation of interleukin 1beta in patients with cryopyrin-associated periodic syndromes. *J Exp Med*. 2009; 206: 1029–1036. doi: [10.1084/jem.20082481](#) PMID: [19364880](#)
59. Otten MA, Bakema JE, Tuk CW, Glennie MJ, Tutt AL, Beelen RH, et al. Enhanced FcalphaRI-mediated neutrophil migration towards tumour colonies in the presence of endothelial cells. *Eur J Immunol*. 2012; 42: 1815–1821. doi: [10.1002/eji.201141982](#) PMID: [22535639](#)
60. Li L, Shukla S, Lee A, Garfield SH, Maloney DJ, Ambudkar SV, et al. The skin cancer chemotherapeutic agent ingenol-3-angelate (PEP005) is a substrate for the epidermal multidrug transporter (ABCB1) and targets tumor vasculature. *Cancer Res*. 2010; 70: 4509–4519. doi: [10.1158/0008-5472.CAN-09-4303](#) PMID: [20460505](#)
61. Schaefer L. Complexity of danger: the diverse nature of damage-associated molecular patterns. *J Biol Chem*. 2014; 289: 35237–35245. doi: [10.1074/jbc.R114.619304](#) PMID: [25391648](#)
62. Cavanagh LL, Halliday GM. Dendritic epidermal T cells in ultraviolet-irradiated skin enhance skin tumor growth by inhibiting CD4+ T-cell-mediated immunity. *Cancer Res*. 1996; 56: 2607–2615. PMID: [8653705](#)