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Cytokine-Stimulated Phosphoflow of Whole Blood Using CyTOF Mass Cytometry

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Abstract

The ability to assess the function of a range of cytokine, antigen receptor, and Toll-like receptor (TLR) signaling pathways in a range of immune cells could provide a kind of fingerprint of the state of the human immune system. The mass cytometry or CyTOF, platform allows for the parallel application of about 40 labeled antibodies to a single sample, creating the possibility to read out many cell types and signaling pathways in a single small blood sample. We developed such a mass cytometry panel, consisting of 22 antibodies to cell surface lineage markers and 8 antibodies to phospho-specific epitopes of signaling proteins. These antibodies were chosen to discriminate all major white blood cell lineages, to a level of detail that includes subsets such as naïve, central memory, effector memory, and late effector CD4+ and CD8+T cells, naïve, transitional, and switched memory B cells, plasmablasts, myeloid and plasmacytoid dendritic cells, CD16+ and CD16+CD56+ NK cells, CD16+ and classical monocytes etc. 32 such cell subsets are defined in our standard gating scheme. The eight phospho-specific antibodies were chosen to represent major signaling nodes responsive to cytokine, TLR, and antigen receptor signaling. This antibody panel is used with 8 standard stimulation conditions (unstimulated, IFN α , IL-6, IL-7, IL-10, IL-21, LPS, PMA+ ionomycin), although other stimuli can be added. Comparison of healthy controls to subjects with immune deficiencies of unknown etiology may help elucidate the mechanisms of such deficiencies. Phosphorylation of tyrosine, serine, and threonine residues is critical for the control of protein activity involved in various cellular events. An assortment of kinases and phosphatases regulate intracellular protein phosphorylation in many different cell signaling pathways, such as T and B cell signaling, those regulating apoptosis, growth and cell cycle control, plus those involved with cytokine, chemokine, and stress responses. Phosphoflow assays combine phospho-specific antibodies with the power of flow cytometry to enhance phospho protein study. In our assay, peripheral blood mononuclear cells are stimulated by cytokines, fixed, surface-stained with a cocktail of antibodies labeled with MAXPAR (Brand

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¹Staining volumes are kept at a total of 50 to 60 μ l.

²Washing steps are critical to minimize background. Aspirate such that only ~ 100 μ l volume is left after each wash. Vortex before the next wash.

³Homogeneous staining is aided by thorough resuspension via pipetting up and down.

⁴Sample dilution is critical to collecting an optimal number of single cell events; aim for about 300 total events/sec.

⁵Do not leave sample buffer or wash buffers exposed to air, as dust accumulation can cause clogs.

⁶Each sample should be resuspended in water and filtered through nylon mesh just prior to running; do not let samples sit on water, as cells will lyse and clump, causing clogs and loss of events.

Name) metal-chelating polymers and permeabilized with methanol. They are then stained with intracellular phospho-specific antibodies.

We use a CyTOF™ mass cytometer to acquire the ICP-MS data. The current mass window selected is approximately AW 103-203, which includes the lanthanides used for most antibody labeling, as well as iridium and rhodium for DNA intercalators. Subsequent analysis of the dual count signal data using FlowJo software allows for cell types to be analyzed based on the dual count signal in each mass channel. The percentage of each cell type is determined and reported as a percent of the parent cell type. Median values are reported to quantitate the level of phosphorylation of each protein in response to stimulation. Comparing the level of phosphorylation between samples can offer insight to the status of the immune system. Whole blood stimulation is the closest to the *in vivo* condition and it allows for assessment of granulocyte population as well as lymphocytes and monocytes.

Materials and Reagents

1. Whole blood from patient or donor
2. Cytokine aliquots (IFN α , IL-6, IL-7, IL-10, IL-21, LPS, PMA/Ionomycin *etc.*)
 - a. IFN α (PBL Interferon source, catalog number: 11105-1)
 - b. IFN γ 2 (BD Biosciences, catalog number: 554617)
 - c. IL6 (BD Biosciences, catalog number: 550071)
 - d. IL7 (BD Biosciences, catalog number: 554608)
 - e. IL10 (BD Biosciences, catalog number: 554611)
 - f. IL21 (Life Technologies, Gibco®, catalog number: PHC0214)
 - g. IL2 (BD Biosciences, catalog number: 554603)
 - h. CD3 (BD Biosciences, catalog number: 555329)
 - i. CD28 (BD Biosciences, catalog number: 555725)
 - j. LPS (Sigma-Aldrich, catalog number: L7770)
 - k. IL5 (Pepro Tech, catalog number: 200-05)
 - l. IL17A (Pepro Tech, catalog number: 200-17)
 - m. IL17E (Pepro Tech, catalog number: 200-24)
3. Methanol (Thermo Fisher Scientific, catalog number: A452SK-1)
4. Dulbecco's Phosphate-buffered saline (Ca²⁺, Mg²⁺)
5. Phenotyping and phosphoprotein antibodies filtered with 0.1 μ m spin filters to get even staining of markers
6. Ir-intercalator stock solution from Fluidigm Sciences (Rh103-intercalator can be used)

7. 10× phosphate-buffered saline (Rockland, catalog number: MB-008)
8. Smart tube 1× thaw-lyse buffer (Smart Tube Inc.)
9. Complete RPMI (see Recipes)
10. CyFACS buffer (see Recipes)

Equipment

1. Nunc Coded Cryobank Vials (Cluster tubes, catalog number: 374078)
2. 37 °C water bath
3. Biosafety cabinet
4. Centrifuge
5. CO₂ incubator at 37 °C
6. Calibrated pipettes
7. 8 or 12 pin aspirator (V&P Scientific, model: Inc VP187A)
8. Smart tube proteomic stabilizer (Smart Tube Inc.)

Procedure

- A. Prepare stimulations in cluster tubes
 1. Prepare cytokines at 5× concentrations in Complete RPMI, with enough volume to pipette 50 µl into a well for each sample and control. See chart below for dilution for a full plate.
 2. The Cluster tubes with the aliquoted stimulants can be frozen away at −80 °C until further use.
- B. Example of a full plate

1	2	3	4	5	6	7	8	9	10	11	12
unstim	unstim	unstim	unstim	unstim	unstim	unstim	unstim	unstim	unstim	unstim	unstim
IFN α	IFN α	IFN α	IFN α	IFN α	IFN α	IFN α	IFN α	IFN α	IFN α	IFN α	IFN α
IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6
IL-7	IL-7	IL-7	IL-7	IL-7	IL-7	IL-7	IL-7	IL-7	IL-7	IL-7	IL-7
IL-10	IL-10	IL-10	IL-10	IL-10	IL-10	IL-10	IL-10	IL-10	IL-10	IL-10	IL-10
IL-21	IL-21	IL-21	IL-21	IL-21	IL-21	IL-21	IL-21	IL-21	IL-21	IL-21	IL-21
LPS	LPS	LPS	LPS	LPS	LPS	LPS	LPS	LPS	LPS	LPS	LPS
PMA /I ω	PMA /I ω	PMA /I ω	PMA /I ω	PMA /I ω	PMA /I ω	PMA /I ω	PMA /I ω	PMA /I ω	PMA /I ω	PMA /I ω	PMA /I ω

C. Example of cytokine stimulations

IFN α : Final concentration of stimulation used=10,000 units/ ml

IFN γ 2: Final concentration of stimulation used= 50 ng/ ml

IL6: Final concentration of stimulation used= 50 ng/ ml

IL7: Final concentration of stimulation used= 50 ng/ ml

IL10: Final concentration of stimulation used= 50 ng/ ml

IL21: Final concentration of stimulation used= 50 ng/ ml

IL2: Final concentration of stimulation used= 50 ng/ ml

CD3 = 2.5 μ l in 990 μ l (Final concentration 500 ng/ml)

CD28 = 10 μ l in above media (Final concentration 2,000 ng/ml)

LPS: Final concentration of stimulation used= 1 μ g/ ml

PMA: 10 ng/ml final concentration /ml

Ionomycin: 1,000 ng/ml final concentration /ml

IL5: Final concentration of stimulation used= 10 ng/ ml

IL17A: Final concentration of stimulation used= 50 ng/ ml

IL17E: Final concentration of stimulation used= 50 ng/ ml

D. Stimulation

1. Rest the blood collected from donors in incubator at 37 °C in CO₂ incubator for 1 h. Just before use, take out the required number of cluster tubes with the stimulant from the -80 °C freezer and let warm at 37 °C water bath for 5–10 min.
2. Aliquot 200 μ l of whole blood into column 1 of the cluster tube plate using a multichannel pipette. Change tips between each patient.
3. Repeat with all the columns of tubes depending on the number of donors.
4. Work as rapidly as possible.
5. Tap plate to mix, and incubate 15 min at 37 °C in CO₂ incubator.
6. Remove stimulated whole blood from incubator at 15 min and using a multichannel pipette, add 250 μ l of 1 \times Proteomic Stabilizer to each column of patient samples in the Cluster tube block. Pipette up and down to mix for each patient. Change tips between patients. Add the stabilizer in the same order that you added the whole blood for stimulation.
7. Incubate for 10 min at room temperature. At this point it can be frozen away at -80 °C until it is ready to be Thaw-lysed and surface stained.

E. Surface staining

1. Take out the required number of frozen stimulated cluster tubes with whole blood as can be comfortably run on the CyTOF in 1 day. (You can run 3–4 cluster or strip of samples in one day by CyTOF. One cluster or strip has 8 wells). Thaw in cold water for about 15 min.
2. Using a pipette, transfer the fixed whole blood sample (450 μ l) into a labeled deep well plate with 1.2 ml of 1 \times Thaw-Lyse and let it sit for 10 min at room temperature.
3. Centrifuge cells at 548 rcf ($\times g$) for 10 min at room temperature.
4. Aspirate supernatant from the cells.
5. Vortex the pellet and add 1.6 ml, 1 \times Thaw-Lyse, and let it sit for 5–10 min at room temperature. Centrifuge cells at 548 rcf ($\times g$) for 10 min at room temperature.
6. Aspirate supernatant from the cells.
7. Vortex and wash the pellet with \sim 1.8 ml CyFACS.
8. Centrifuge cells at 974 rcf ($\times g$) for 10 min at 4 $^{\circ}$ C.
9. Aspirate supernatant from the cells so that about 50 μ l remains at the bottom of each well.
10. Make cocktail in phosphate buffered saline of metal-chelating polymer-labeled surface antibodies according to previously determined titration. Make sufficient volume for each well to have 20 μ l of cocktail. Pipet into 0.1 μ m spin filter and centrifuge in a tabletop microcentrifuge (RCF=14,000) for 2 min at room temperature. This ensures even staining.
11. Add 20 μ l of antibody cocktail to the cells in the deep well plate, vortex to mix and let it incubate at room temperature for 30 min.
12. Wash cells with 1.8 ml phosphate buffered saline and centrifuge cells at 974 rcf ($\times g$) for 10 min at 4 $^{\circ}$ C. Aspirate.
13. Permeabilize the cells by adding 600 μ l cold methanol to each well of the deep well block using a multichannel pipette. Pipette up and down to mix for each patient. Change tips between patients. Cells are stored overnight at this point at -80 $^{\circ}$ C. This is done for convenience of work flow.
14. Remove samples from freezer. Add 1.0 ml of CyFACS. Centrifuge cells at 974 rcf ($\times g$) for 10 min at 4 $^{\circ}$ C. Aspirate so that about 100 μ l remains in the wells.
15. Wash in 1.8 ml/ well CyFACS buffer.
16. Centrifuge cells at 974 rcf ($\times g$) for 10 min at 4 $^{\circ}$ C. Discard supernatant by aspiration.

F. Intracellular staining

1. Make cocktail in PBS of metal-chelating polymer-labeled intracellular antibodies according to previously determined titration. Make sufficient volume for each sample to have 20 μ l of cocktail. Pipet into 0.1 μ m spin filter and centrifuge in tabletop microcentrifuge (RCF=14,000) for 2 min at room temperature.
2. Add 20 μ l of antibody cocktail to the cells in the deep well plate, vortex to mix and let it incubate at room temperature for 30 min.
3. Add 1.6 ml of PBS. Centrifuge cells at 974 rcf ($\times g$) for 10 min at 4 $^{\circ}$ C. Discard supernatant by aspiration.
4. Make 1:200 dilution in PBS of Ir-intercalator. Add 20 μ l of diluted Ir-intercalator solution to each sample, pipet to mix. Incubate on ice for 20 min.
5. Wash 3 times in MilliQ water. Centrifuge cells at 974 rcf ($\times g$) for 10 min at 4 $^{\circ}$ C. Discard supernatant by aspiration.
6. Bring up the samples in 1 ml of MilliQ water and filter samples through a cell strainer. Acquire samples on the Cytof, after standard instrument setup procedures.

Representative data

Recipes

1. Complete RPMI (Hyclone RPMI-1640 Medium)
RPMI with 10% FBS
Penicillin/streptomycin
Glutamine
2. CyFACS buffer
1 \times CyPBS PBS with 0.1% BSA, and 0.05% Na azide
Made in MilliQ water
Note: Do not use FBS!

Acknowledgments

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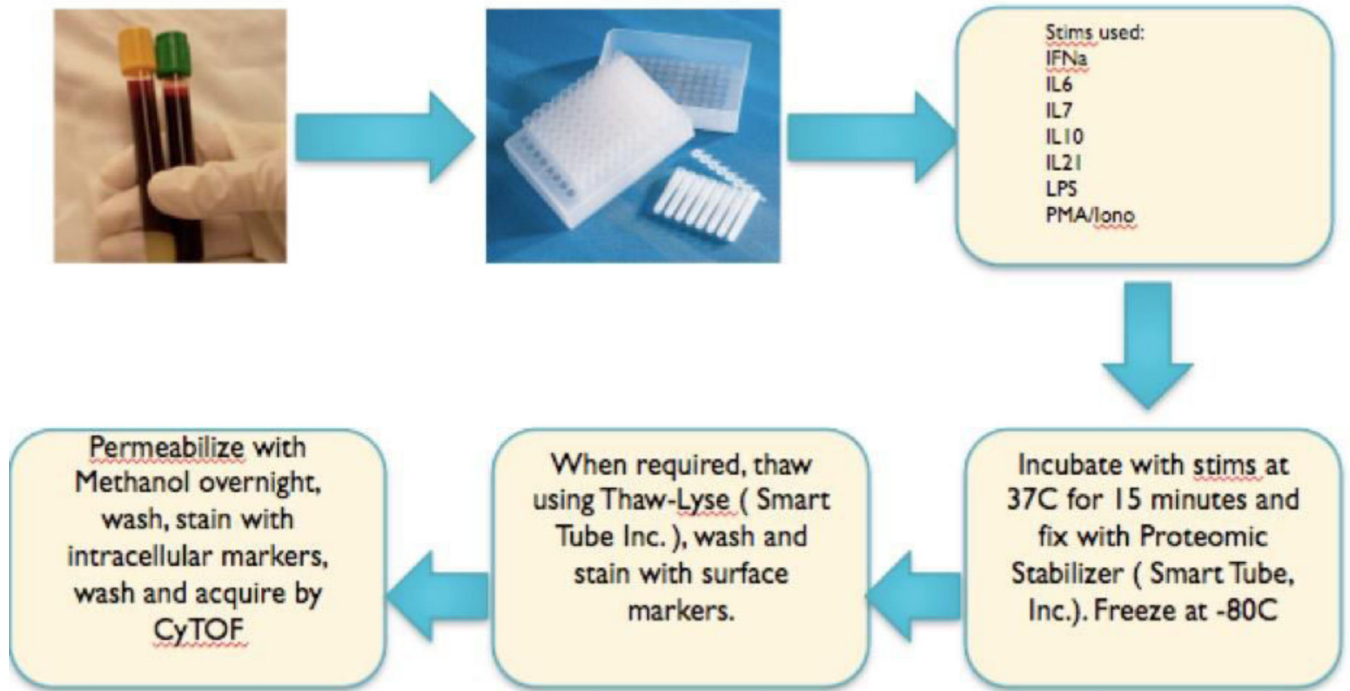


Figure 1.
A flow chart of the steps described in our procedure

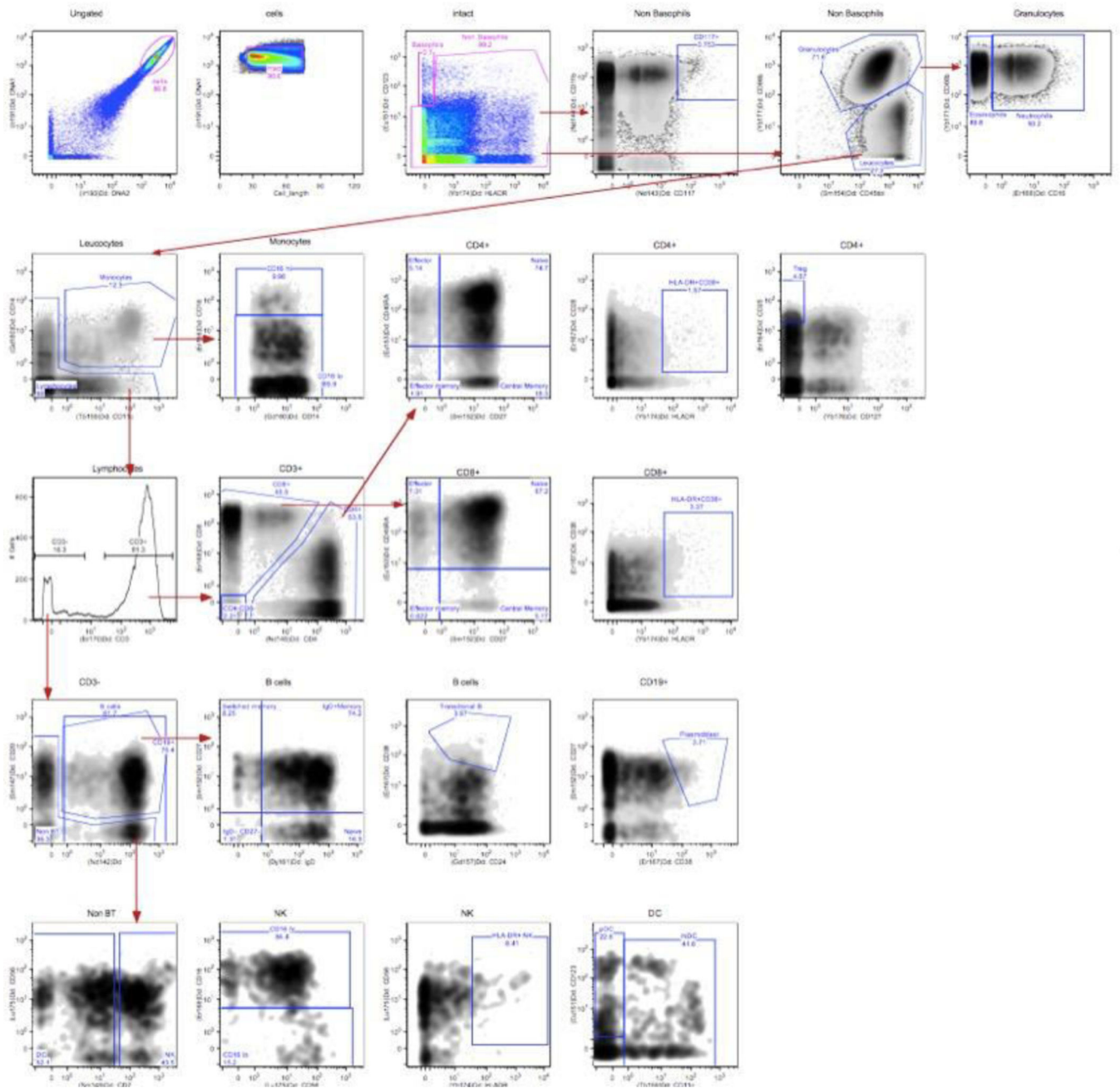


Figure 2.
Whole blood gating scheme

Table 1

Antibodies in the phospho-CyTOF panel

Phospho-CyTOF				
Metal label	Specificity	Clone	Source	Staining step
139La				
141Pr				
142Nd	CD19	HIB19	DVS	Surface
143Nd				
144Nd				
145Nd	CD4	RPA-T4	DVS	Surface
146Nd				
147Sm	CD20	H1	In house	Intracellular
148Nd				
149Sm	CD7	CD7-6B7	In house	Surface
150Nd				
151Eu	CD123	6H6	DVS	Surface
152Sm	CD27	O323	In house	Surface
153Eu	CD45RA	HI100	DVS	Surface
154Sm				
155Gd				
156Gd	pp38	D3F9	DVS	Intracellular
157Gd	CD24	ML-5	In house	Surface
158Gd	pSTAT3	4/P-Stat3	DVS	Intracellular
159Tb	CD11c	Bu15	DVS	Surface
160Gd	CD14	M5E2	DVS	Surface
161Dy	IgD	BD -1A6-2	In house	Surface
162Dy	pErk1/2	Clone 20A	In house	Intracellular
163Dy	IkBtot	CS L35A5	In house	Intracellular
164Dy	CD25	M-A251	In house	Surface
165Ho	pS6	Clone N7-548	In house	Intracellular
166Er	CD16	B73.1	In house	Surface
167Er	CD38	HIT2	DVS	Surface
168Er	CD8	SK1	DVS	Surface
169Tm	pSTAT1	BD -4A	In house	Intracellular
170Er	CD3	UCHT1	DVS	Surface
171Yb				
172Yb	pSTAT5	BD- 47	In house	Intracellular
173Yb	pPLCg2	BD -K860689.37	In house	Intracellular
174Yb	HLA-DR	1243	DVS	Surface
175Lu	CD56	HCD56	In house	Surface

Phospho-CyTOF				
Metal label	Specificity	Clone	Source	Staining step
176Yb	CD127	A019D5	DVS	Surface
Qdot				

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