

A Robust Cross-Linking Strategy for Block Copolymer Worms Prepared via Polymerization-Induced Self-Assembly

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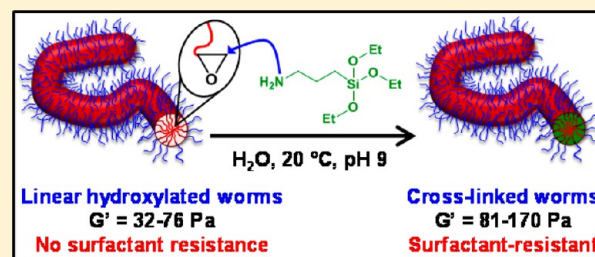
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Supporting Information

ABSTRACT: A poly(glycerol monomethacrylate) (PGMA) chain transfer agent is chain-extended by reversible addition–fragmentation chain transfer (RAFT) statistical copolymerization of 2-hydroxypropyl methacrylate (HPMA) with glycidyl methacrylate (GlyMA) in concentrated aqueous solution via polymerization-induced self-assembly (PISA). A series of five free-standing worm gels is prepared by fixing the overall degree of polymerization of the core-forming block at 144 while varying its GlyMA content from 0 to 20 mol %. ¹H NMR kinetics indicated that GlyMA is consumed much faster than HPMA, producing a GlyMA-rich sequence close

to the PGMA stabilizer block. Temperature-dependent oscillatory rheological studies indicate that increasing the GlyMA content leads to progressively less thermoresponsive worm gels, with no degelation on cooling being observed for worms containing 20 mol % GlyMA. The epoxy groups in the GlyMA residues can be ring-opened using 3-aminopropyltriethoxysilane (APTES) in order to prepare core cross-linked worms via hydrolysis-condensation with the siloxane groups and/or hydroxyl groups on the HPMA residues. Perhaps surprisingly, ¹H NMR analysis indicates that the epoxy–amine reaction and the intermolecular cross-linking occur on similar time scales. Cross-linking leads to stiffer worm gels that do not undergo degelation upon cooling. Dynamic light scattering studies and TEM analyses conducted on linear worms exposed to either methanol (a good solvent for both blocks) or anionic surfactant result in immediate worm dissociation. In contrast, cross-linked worms remain intact under such conditions, provided that the worm cores comprise at least 10 mol % GlyMA.



INTRODUCTION

Over the past fifty years or so, block copolymer self-assembly has become a well-recognized and widely adopted route for the production of organic nanoparticles in a wide range of solvents. Many copolymer morphologies have been reported in the literature.^{1–5} However, there have been relatively few studies of block copolymer worms, cylinders, or rods via traditional post-polymerization processing routes, such as a solvent switch in dilute solution.^{2,6–13} This is presumably because such highly anisotropic morphologies typically occupy relatively little phase space, which means that the range of required block compositions tends to be rather narrow. In contrast, polymerization-induced self-assembly (PISA) has recently enabled the rational synthesis of block copolymer worms in the form of highly concentrated dispersions in a wide range of polar and non-polar solvents.^{14–32}

The worm morphology is particularly interesting for various potential applications. Discher and co-workers have shown that

poly(ethylene oxide)–poly(caprolactone) diblock copolymer worms exhibit substantially extended *in vivo* circulation times compared to the equivalent spherical morphology.⁹ Armes and co-workers have recently demonstrated the advantages offered by highly anisotropic worms when deployed as Pickering emulsifiers:³³ they are much more strongly adsorbed at the oil–water interface compared to the equivalent spheres, yet retain a relatively high specific surface area.^{34,35} Several research groups have studied the rheological properties of block copolymer worms,^{31,32,36–40} with thermoreversible gelation being observed in aqueous solution,^{14,31} polar solvents such as ethanol,⁴¹ and non-polar solvents such as *n*-alkanes.^{19,32,42}

Many strategies have been explored for the covalent stabilization of block copolymer nano-objects. Core cross-

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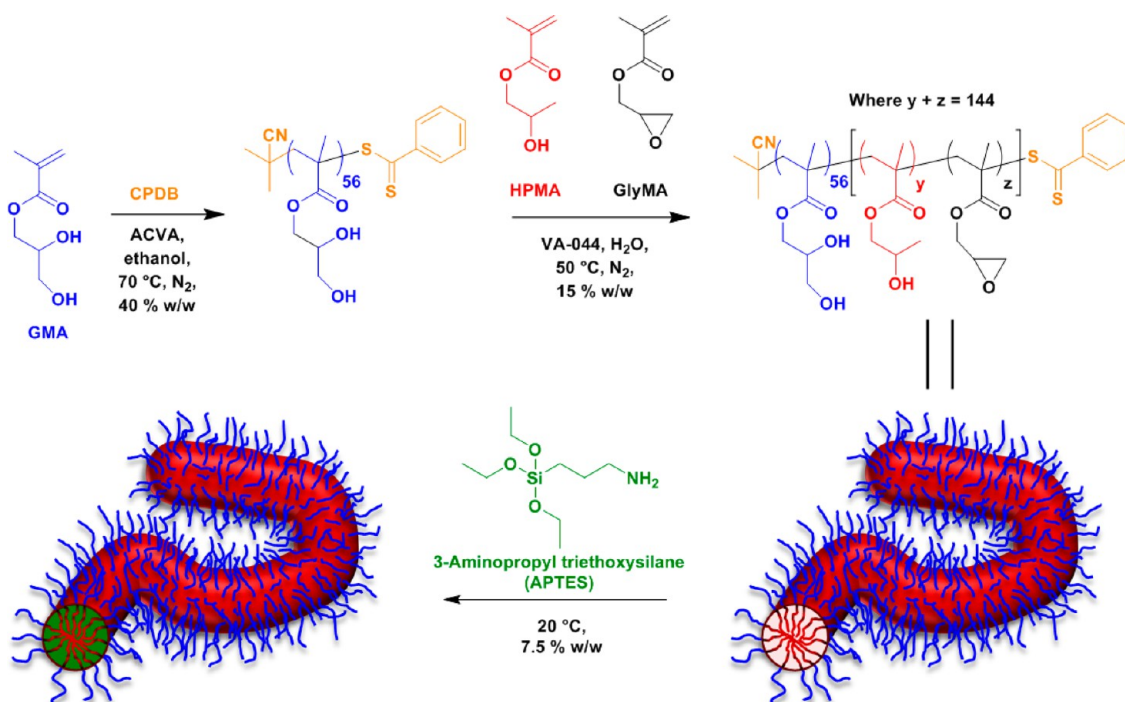


Figure 1. Synthesis of a PGMA₅₆ macro-CTA via RAFT solution polymerization of GMA in ethanol using a CPDB RAFT agent and its subsequent chain extension via statistical copolymerization of varying molar ratios of HPMA and GlyMA to form diblock copolymer worms in aqueous solution via polymerization-induced self-assembly (PISA). Such worms are then cross-linked using APTES in a two-step post-polymerization process involving (i) an epoxy-amine reaction with the GlyMA residues and (ii) hydrolysis-condensation reaction with the hydroxyl groups on the HPMA residues.

linked spherical micelles have been reported by various groups,^{43–46} while Wooley^{47–51} and Armes^{52–55} have worked extensively on shell cross-linked micelles. Both Antonietti et al.⁵⁶ and Bates and co-workers^{7,37} have cross-linked polybutadiene-based block copolymer worms in dilute solution using γ radiation or redox chemistry, respectively. In contrast, Liu's group has developed various photochemical strategies based on cinnamoyl side groups.^{57,58} In the context of PISA formulations, cross-linked block copolymer spheres, worms, and vesicles have been reported by copolymerizing small amounts of divinyl comonomers such as ethylene glycol di(meth)acrylate or poly(ethylene glycol) diacrylate.^{32,34,59–62} However, this strategy is somewhat problematic for the worm morphology, since relatively small perturbations in the block composition can result in the formation of mixed phases, rather than pure worms. An alternative post-polymerization approach was reported by Chambon et al. for cross-linked block copolymer vesicles, whereby pendent epoxy groups were reacted with small molecule or oligomeric diamines.⁶³ Similarly An and co-workers prepared poly(poly(ethylene oxide)methyl ether methacrylate)-poly(2-(acetoacetoxy)ethyl methacrylate) (PPEOMA-PAEMA) diblock copolymer vesicles using PISA via RAFT dispersion polymerization in ethanol.⁶⁴ These vesicles were subsequently cross-linked using *O,O'*-1,3-propanediylbisoxylamine dihydrochloride, which reacted with ketone groups in the PAEMA core-forming block. Very recently, the same team chain-extended a poly(*N,N*-dimethylacrylamide) (PDMA) macro-CTA using a binary mixture of diacetone acrylamide (DAAM) and an asymmetric cross-linker allyl acrylamide (ALAM) to prepare vesicles.⁶⁵ The acrylamide groups in DAAM and ALAM have similar reactivities, whereas the allyl group in ALAM reacts significantly more slowly. This

leads to *in situ* cross-linking of the vesicles toward the end of the copolymerization.

Generally speaking, there are relatively few literature reports describing the synthesis and cross-linking of diblock copolymer worms.^{6–8,57,58,66–69} Herein we describe the facile preparation of core cross-linked diblock copolymer worms. More specifically, a series of hydroxyl-functional methacrylic diblock copolymer worms containing varying amounts of glycidyl methacrylate (GlyMA) in the core-forming block are prepared in aqueous solution via PISA. Such worms are then covalently stabilized via cross-linking of the core-forming block using 3-aminopropyltriethoxysilane (APTES) (see Figure 1). The physical properties of aqueous dispersions of these cross-linked worms are compared to those of the linear worm precursors using various characterization techniques, including transmission electron microscopy (TEM), dynamic light scattering (DLS), and oscillatory rheology.

EXPERIMENTAL SECTION

Materials. Glycerol monomethacrylate (GMA; 99.8% purity) was kindly donated by GEO Specialty Chemicals (Hythe, UK) and used without further purification. 2-Hydroxypropyl methacrylate (HPMA) was purchased from Alfa Aesar and used as received. 2,2'-Azobis[2-(2-imidazolyl-2-yl)propane] dihydrochloride (VA-044) was purchased from Wako Pure Chemical Industries (Japan) and used as received. Glycidyl methacrylate (GlyMA), 2-cyano-2-propyl benzodithioate (CPDB), 4,4'-azobis(4-cyanopentanoic acid) (ACVA; V-501; 99%), 3-aminopropyltriethoxysilane (APTES), *d*₄-sodium trimethylsilyl propanoate (TMSP), sodium dodecyl sulfate (SDS), deuterated methanol-*d*₄, ethanol (99%, anhydrous grade), methanol and dichloromethane were purchased from Sigma-Aldrich UK and were used as received. All solvents were of HPLC-grade quality.

Synthesis of Poly(glycerol monomethacrylate) (PGMA₅₆) Macro-CTA via RAFT Solution Polymerization in Ethanol. A

typical protocol for the synthesis of PGMA₅₆ macro-CTA was as follows: GMA (203.0 g, 1.268 mol), CPDB (6.03 g, 0.020 mol; target DP = 63), ACVA (1.14 g, 4.07 mmol; CPDB/ACVA molar ratio = 5.0), and anhydrous ethanol (156.0 g, 3.38 mol) were added to a round-bottomed flask to afford a 55% w/w solution. The resulting pink solution was purged with N₂ for 20 min, before the sealed flask was immersed into an oil bath set at 70 °C. After 140 min (69% conversion as judged by ¹H NMR) the polymerization was quenched by immersion of the flask into an ice bath and exposing the reaction mixture to air. The crude polymer was then precipitated into a 10-fold excess of dichloromethane and washed three times using this nonsolvent to remove residual unreacted GMA monomer before being dried under high vacuum for 3 days at 40 °C. ¹H NMR studies indicated a mean degree of polymerization of 56 via end-group analysis (the integrated aromatic RAFT end-group signals at 7.1–7.4 ppm were compared to those assigned to the two oxymethylene protons at 3.5–4.4 ppm). Taking into account the target DP of 63 and the GMA conversion of 69%, this indicated a CTA efficiency of 76%. GPC studies (DMF eluent, refractive index detector; calibrated against a series of near-monodisperse poly(methyl methacrylate) standards) indicated an M_n of 15 000 g mol⁻¹ and an M_w/M_n of 1.11.

Synthesis of PGMA₅₆–PHPMA₁₄₄ Diblock Copolymer Worms via RAFT Aqueous Dispersion Polymerization. A typical protocol for the chain extension of PGMA₅₆ macro-CTA with 144 units of HPMA via RAFT aqueous dispersion polymerization was as follows: PGMA₅₆ macro-CTA (0.399 g, 0.043 mmol), HPMA monomer (0.90 g, 6.0 mmol), and VA-044 (3.50 mg, 0.011 mmol; PGMA₅₆ macro-CTA/VA-044 molar ratio = 4.0) were added to a 25 mL round-bottomed flask, prior to addition of water to produce a 15% w/w aqueous solution. The reaction solution was purged under nitrogen for 30 min at 20 °C prior to immersion into an oil bath set at 50 °C. The reaction mixture was stirred for 105 min to ensure almost complete conversion of the HPMA monomer (>99% by ¹H NMR analysis), and then the HPMA polymerization was quenched by exposure to air followed by cooling to ambient temperature. The resulting dispersion was diluted with deionized water to give a free-standing 7.5% w/w worm gel that was characterized by DLS, TEM, and rheology without further purification.

Synthesis of PGMA₅₆–P(HPMA_y-stat-GlyMA_z) Diblock Copolymer Worms via RAFT Aqueous Emulsion/Dispersion Polymerization. A typical protocol for chain extension of PGMA₅₆ macro-CTA with 122 units of HPMA and 22 units of GlyMA via RAFT aqueous dispersion/emulsion polymerization was as follows: PGMA₅₆ macro-CTA (0.418 g, 0.046 mmol), HPMA monomer (0.800 g, 5.5 mmol), GlyMA monomer (0.140 g, 1.0 mmol), and VA-044 (3.70 mg, 0.011 mmol; PGMA₅₆ macro-CTA/VA-044 molar ratio = 4.0) were added to a 25 mL round-bottomed flask, prior to addition of sufficient water to afford a 15% w/w aqueous solution. This reaction solution was purged under nitrogen for 30 min at 20 °C prior to immersion into an oil bath set at 50 °C. The reaction mixture was stirred for 105 min to ensure almost complete conversion of the HPMA and GlyMA comonomers (>99% by ¹H NMR analysis). Then the copolymerization was quenched by exposure to air, followed by cooling to ambient temperature. The resulting dispersion was immediately diluted with deionized water to 7.5% w/w solids, yielding a free-standing worm gel that was characterized by DLS, TEM, and rheology without further purification.

Post-Polymerization Cross-Linking of a 7.5% w/w Aqueous Dispersion of PGMA₅₆–P(HPMA_y-stat-GlyMA_z) Worm Gel Using APTES. A typical protocol for the covalent cross-linking of PGMA₅₆–P(HPMA₁₂₂-stat-GlyMA₂₂) diblock copolymer worm gel at 7.5% w/w solids using APTES was as follows. APTES (0.111 g, 0.5 mmol, APTES/GlyMA molar ratio = 1.0) was added to 9.1 g of a 7.5% w/w aqueous dispersion of PGMA₅₆–P(HPMA₁₂₂-stat-GlyMA₂₂) diblock copolymer worms, and the epoxy–amine reaction was allowed to proceed for 24 h at 20 °C with continuous stirring of the shear-thinning worm gels.

Instrumentation. *NMR Spectroscopy.* ¹H NMR spectra were recorded using a 400 MHz Bruker Avance-500 spectrometer with 64 scans being averaged per spectrum.

Gel Permeation Chromatography (GPC). Polymer molecular weights and polydispersities were determined using a DMF GPC setup operating at 60 °C and comprising two Polymer Laboratories PL gel 5 μm Mixed-C columns connected in series to a Varian 390-LC multidetector suite (with only the refractive index detector being used) and a Varian 290-LC pump injection module. The GPC eluent was HPLC-grade DMF containing 10 mM LiBr at a flow rate of 1.0 mL min⁻¹. DMSO was used as a flow-rate marker. Calibration was conducted using a series of ten near-monodisperse poly(methyl methacrylate) standards ($M_n = 625$ – $2\,480\,000$ g mol⁻¹). Chromatograms were analyzed using Varian Cirrus GPC software (version 3.3).

Dynamic Light Scattering (DLS). Studies were conducted using a Malvern Zetasizer NanoZS instrument on 0.10% w/w copolymer dispersions in either water, methanol, or 1.0% w/w SDS aqueous solution at 25 °C before and after cross-linking in a glass cuvette at a fixed backscattering angle of 173°. Intensity-average hydrodynamic diameters were calculated via the Stokes–Einstein equation using a non-negative least-squares (NNLS) algorithm. All data were averaged over three consecutive runs.

Aqueous Electrophoresis. Aqueous electrophoresis studies were performed on 0.10% w/w aqueous copolymer dispersions (containing 10⁻³ mol dm⁻³ NaCl as background electrolyte) using a Malvern Zetasizer NanoZS instrument at 25 °C. The pH of the copolymer dispersion was initially basic and was adjusted using HCl. Zeta potentials were calculated from the Henry equation using the Smoluchowski approximation. All data were averaged over three consecutive runs.

Transmission Electron Microscopy (TEM). As-prepared copolymer dispersions were diluted 150-fold at 20 °C in either methanol or water to generate 0.10% w/w dispersions. Copper/palladium TEM grids (Agar Scientific, UK) were surface-coated in-house to yield a thin film of amorphous carbon. The grids were then plasma glow-discharged for 30 s to create a hydrophilic surface. A micropipet was used to place droplets (12 μL) of aqueous copolymer dispersions onto freshly glow-discharged grids for 1 min, followed by careful blotting with filter paper to remove excess sample. To stain the aggregates, a 0.75% w/w uranyl formate solution (9 μL) was soaked on the sample-loaded grid for 20 s and then carefully blotted to remove excess stain. Each grid was then carefully dried using a vacuum hose. Imaging was performed using a FEI Tecnai Spirit microscope fitted with a Gatan 1kMS600CW CCD camera operating at 80 kV.

Rheology Studies. Storage (G') and loss (G'') moduli were determined between 4 and 25 °C for diblock copolymer worm gels both before and after covalent cross-linking using a TA Instruments AR-G2 rheometer at a fixed strain of 1.0% and an angular frequency of 1.0 rad s⁻¹. The copolymer concentration was fixed at 7.5% w/w for all experiments. A cone-and-plate geometry (40 mm 2° aluminum cone) was used for these measurements.

RESULTS AND DISCUSSION

Block Copolymer Worm Syntheses. A well-defined near-monodisperse poly(glycerol monomethacrylate) (PGMA) macro-CTA was prepared via reversible addition–fragmentation chain transfer (RAFT) solution polymerization of GMA in ethanol at 70 °C using 2-cyano-2-propylbenzodithioate (CPDB) as the chain transfer agent (CTA).^{70,71} ¹H NMR spectroscopy studies suggested a mean degree of polymerization (DP) of 56, as judged by end-group analysis. Moreover, gel permeation chromatography (GPC) studies conducted in DMF against near-monodisperse poly(methyl methacrylate) (PMMA) standards indicated that the PGMA₅₆ macro-CTA possessed a relatively narrow molecular weight distribution ($M_n = 15\,000$ g mol⁻¹ and $M_w/M_n = 1.11$). This homopolymer precursor was then chain-extended via statistical copolymerization of 0, 5, 10, 15, or 20 mol % of glycidyl methacrylate (GlyMA) with 2-hydroxypropyl methacrylate (HPMA) at 15% w/w solids using a PISA formulation (see Figure 1). This

protocol produced a series of free-standing copolymer worm gels after cooling to room temperature. For each PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymer synthesis, a mean DP of 144 was targeted for the core-forming block (i.e., $y + z = 144$). High monomer conversions (>99%) were achieved in all cases, as judged by the disappearance of monomer vinyl signals between 5.9 and 6.1 ppm in the ¹H NMR spectra.

Covalent Stabilization of Block Copolymer Worms.

The as-prepared 15% w/w aqueous dispersions of block copolymer worms described above were diluted to 7.5% w/w solids to allow efficient stirring when conducting post-polymerization derivatization reactions using 3-aminopropyl-triethoxysilane (APTES) (see Figure 1). The primary amine group on this siloxane reagent reacts with the pendent epoxide groups⁷² located within the core-forming blocks, with APTES ingress aided by the partially hydrated nature of the HPMA-rich worm cores.¹⁴ In principle, the triethoxysilane component of the grafted APTES molecules should then undergo hydrolysis-condensation reactions, both with each other and also with the pendent secondary (and primary) hydroxyl groups on the HPMA residues, resulting in core cross-linked worms.⁷³ Post-polymerization cross-linking was undertaken in order to minimize the possibility of *in situ* cross-linking during PISA, which might otherwise prevent the formation of worms or perhaps cause inter-worm aggregation. After cross-linking at 7.5% w/w solids, the diblock copolymer worm gels were expected to be dispersible in a good solvent for both blocks (e.g., methanol) and also possess enhanced resistance toward the presence of ionic surfactants, which are known to cause rapid dissociation of closely related PGMA-PPHMA *linear* diblock copolymer nano-objects.⁶³ Furthermore, the rheological properties of the worm gels were investigated both before and after cross-linking. Recently, Lovett and co-workers reported that similar PGMA-PPHMA diblock copolymer worms prepared using a carboxylic acid-based RAFT CTA undergo worm-to-sphere transitions upon a pH switch as a result of end-group ionization.⁷⁴ Thus, it was important to employ a *non-ionic* CTA in the present study in order to prevent such order-order morphological transitions on addition of the strongly basic APTES reagent.

Synthesis and Characterization of PGMA-P(HPMA-stat-GlyMA) Diblock Copolymer Worms. Previous syntheses of similar PGMA₅₅-P(HPMA₂₄₇-stat-GlyMA₈₂) diblock copolymer vesicles were conducted by Chambon et al., with full conversion being attained after 4 h at 70 °C.⁶³ According to ¹H NMR studies, around 90% of the epoxide groups on the GlyMA residues survived these conditions, with 10% undergoing hydrolysis with water (to afford GMA residues) and/or pendent hydroxyl groups in HPMA resulting in partial *in situ* cross-linking. In the present study, diblock copolymer syntheses were conducted at 50 °C for 105 min in order to minimize such side reactions. ¹H NMR studies confirmed the success of this modified protocol, with approximately 98% of epoxide groups surviving at full comonomer conversion (see Figure S1). The (co)polymerization kinetics for the synthesis of PGMA₅₆-PPHMA₁₄₄, PGMA₅₆-P(HPMA₁₃₀-stat-GlyMA₁₄), and PGMA₅₆-P(HPMA₁₁₅-stat-GlyMA₂₉) at 50 °C were monitored by ¹H NMR (see Figure 2). Aliquots of reaction mixtures were extracted at regular time intervals and diluted prior to NMR analysis using CD₃OD, which is a good solvent for all monomeric and copolymer species. Kinetic studies of the PGMA₅₆-PPHMA₁₄₄ diblock copolymer formulation indicated that full conversion was achieved after 90 min. After a brief

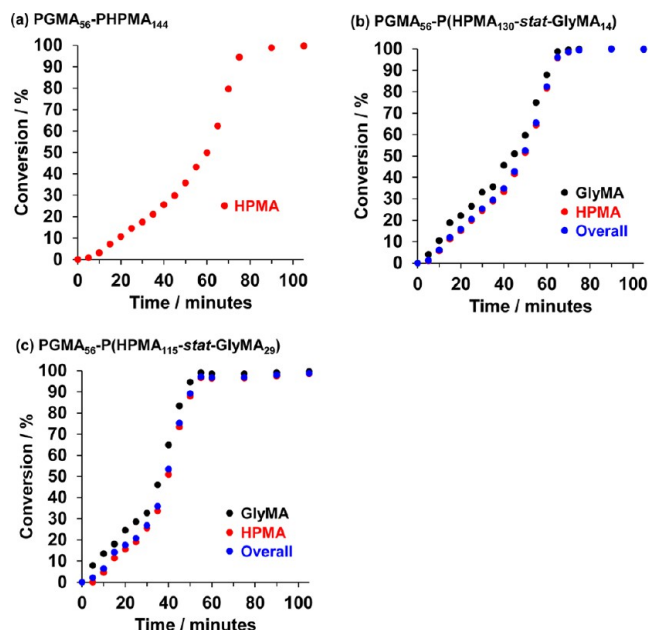


Figure 2. Conversion vs time curves obtained by ¹H NMR for the (co)polymerization of HPMA (red circles), GlyMA (black circles), and the overall comonomer mixture (blue circles) at 50 °C using a PGMA₅₆ macro-CTA when targeting diblock copolymer compositions of (a) PGMA₅₆-PPHMA₁₄₄, (b) PGMA₅₆-P(HPMA₁₃₀-stat-GlyMA₁₄), and (c) PGMA₅₆-P(HPMA₁₁₅-stat-GlyMA₂₉). All syntheses were conducted at 15% w/w solids.

induction period, consumption of the water-miscible HPMA monomer was relatively slow for 35 min. This may be the result of mild retardation, which is not fully understood.⁷⁵ After 65 min (or 62% conversion, which corresponds to a PPHMA DP of 89), the rate of polymerization increases by an order of magnitude (see Figure S2). This is the result of micellar nucleation, which heralds a switch from RAFT solution polymerization to RAFT aqueous dispersion polymerization, as judged by both visual inspection and dynamic light scattering (DLS) (see Figure S2). According to Blanz and co-workers, unreacted HPMA migrates into the micelle cores, increasing the local monomer concentration and hence leading to a faster rate of polymerization.⁷⁶ A similar rate enhancement is also observed when HPMA is partially replaced by GlyMA (see Figure S2). However, in this case the water-immiscible GlyMA comonomer is consumed via aqueous *emulsion* polymerization. Interestingly, ¹H NMR studies indicate significantly faster initial consumption of GlyMA compared to HPMA. For example, in the case of PGMA₅₆-P(HPMA₁₁₅-stat-GlyMA₂₉), 13% GlyMA was consumed after 10 min whereas only 5% HPMA had reacted on the same time scale. Similar observations have been recently reported by Ratcliffe and co-workers for the RAFT statistical copolymerization of *water-immiscible* 4-hydroxybutyl methacrylate (HBMA) with *water-miscible* 2-hydroxyethyl methacrylate (HEMA) in aqueous solution.⁷⁷ In the present study, this leads to a GlyMA-rich sequence for the core-forming block close to its junction with the PGMA stabilizer block. This is important because it has a significant effect on the physical properties of the resulting worm gel, as discussed in more detail below. Visual inspection of the reaction mixture indicates that a homogeneous solution is obtained within 10 min, which suggests that the remaining GlyMA concentration becomes sufficiently low for the statistical copolymerization to proceed

as an aqueous dispersion polymerization before micellar nucleation occurs. This is consistent with the temperature-dependent water solubility of GlyMA reported by Ratcliffe and co-workers.⁷⁸

As expected, partial replacement of HPMA with increasing amounts of GlyMA within the core-forming block induces micellar nucleation at shorter reaction times. For example, nucleation occurs after approximately 55 min when targeting PGMA₅₆-P(HPMA₁₃₀-*stat*-GlyMA₁₄) but after only 40 min when targeting PGMA₅₆-P(HPMA₁₁₅-*stat*-GlyMA₂₉).

The enhanced rate of copolymerization achieved under heterogeneous conditions leads to essentially full monomer conversion within relatively short time scales. More specifically, the synthesis of PGMA₅₆-P(HPMA₁₃₀-*stat*-GlyMA₁₄) was complete after 75 min, while more than 99% conversion was observed for PGMA₅₆-P(HPMA₁₁₅-*stat*-GlyMA₂₉) after only 60 min. In view of these kinetic data, it was decided to conduct these diblock copolymer syntheses for 105 min at 50 °C. These conditions were chosen to ensure very high (>99%) comonomer conversions while minimizing loss of pendent epoxide groups to side reactions, as discussed above.

At the end of each copolymerization, each of the five PGMA₅₆-P(HPMA_{*y*}-*stat*-GlyMA_{*z*}) diblock copolymer dispersions were immediately diluted to 7.5% w/w solids to aid efficient mixing of the APTES cross-linker. Once fully dispersed, these 7.5% w/w dispersions were split into two batches. The first batch was used to determine the physical properties of the linear worms obtained prior to cross-linking, while the second batch was used to examine worm core cross-linking with APTES.

DMF GPC analysis of these five diblock copolymers prior to addition of the APTES cross-linker suggested minimal intrinsic cross-linking occurred during their synthesis, since no high molecular weight shoulder was observed at shorter retention times (see Figure 3). This was not unexpected, since the reaction of epoxy groups with the (mainly) secondary hydroxyl

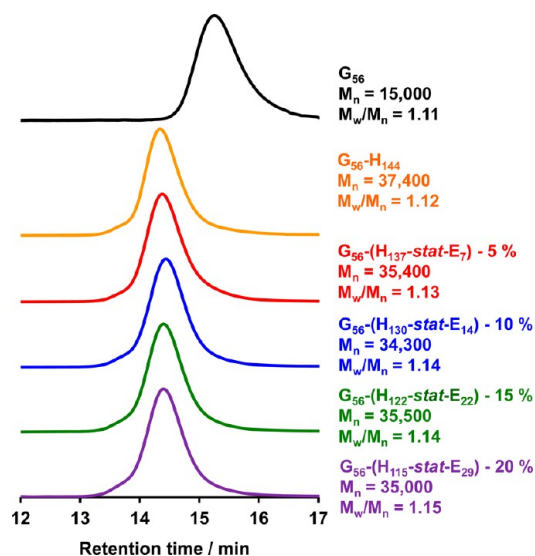


Figure 3. DMF GPC curves obtained for PGMA₅₆ macro-CTA (black curve) and the corresponding traces for four PGMA₅₆-P(HPMA_{*y*}-*stat*-GlyMA_{*z*}) (where $y + z = 144$; these copolymers are denoted as G₅₆-(H_{*y*}-*stat*-E_{*z*}) for brevity) diblock copolymers prepared at 50 °C. Molecular weights are expressed relative to a series of near-monodisperse poly(methyl methacrylate) calibration standards.

groups on the HPMA residues should be negligible at 50 °C. Furthermore, these GPC studies indicated relatively high blocking efficiencies, narrow molecular weight distributions, and similar number-average molecular weights for all five diblock copolymers. In striking contrast, DMF GPC analysis of PGMA₅₅-P(HPMA₂₄₇-*stat*-GlyMA₈₂) vesicles prepared at 70 °C for 4 h performed by Chambon et al. indicated relatively high polydispersities and a prominent high molecular weight shoulder.⁶³ This suggests that epoxide-based cross-linking occurs when such statistical copolymerizations are conducted over longer reaction times at elevated temperatures, although in principle differences in the levels of dimethacrylate impurity in the HPMA comonomer could be an alternative explanation.^{59,76}

DLS and transmission electron microscopy (TEM) studies were conducted on dilute (0.1% w/w) dispersions of the five diblock copolymer worms prior to cross-linking in order to assess their colloidal stability in both water and methanol. DLS studies of dilute aqueous dispersions indicate that these worms possessed sphere-equivalent hydrodynamic diameters of 100–210 nm and relatively high polydispersities (>0.20), which compares well with literature data reported for such nano-objects.^{14,74} Moreover, relatively intense light scattering (derived count rates exceeding 30 000 kcps) was recorded in all cases, which is consistent with the presence of nano-objects (see Table 1). TEM images obtained for dried aqueous copolymer dispersions confirmed the presence of highly anisotropic worms in all cases. Image analysis of 50 worms per copolymer sample indicated well-defined worm widths of approximately 20–25 nm but highly variable worm lengths of 100–1200 nm (see Figure 4). In contrast, TEM studies of the same diblock copolymer dispersions diluted using methanol prior to drying confirmed the absence of any well-defined nano-objects (see Figure S3) while only very weak light scattering (<300 kcps) was observed by DLS. Both observations are consistent with *molecular dissolution* of copolymer chains in methanol, which is a good solvent for both blocks.^{73,79}

Previous rheological studies on a similar PGMA₅₄-PHPMA₁₄₀ diblock copolymer worm gel have shown that degelation occurs on cooling to 5 °C.^{14,36} TEM and small-angle X-ray scattering (SAXS) studies confirmed that this is the result of a worm-to-sphere order–order morphological transition. This is driven by surface plasticization of the PHPMA core-forming block, which causes a shift in the packing parameter, P , from worm phase space ($0.33 < P < 0.5$) to spherical phase space ($P < 0.33$).³ As expected, the linear PGMA₅₆-PHPMA₁₄₄ diblock copolymer worm gel prepared in this study is similarly thermoresponsive. Its critical gelation temperature (CGT) was determined to be 13.5 °C on cooling to 5 °C, as judged by the point of cross-over of the storage modulus (G') and loss modulus (G'') curves in temperature-dependent rheological studies (see Figure 5a). PGMA₅₆-P(HPMA_{*y*}-*stat*-GlyMA_{*z*}) diblock copolymer worm gels possess similar thermoresponsive degelation when up to 15 mol % GlyMA ($z = 22$) is incorporated into the core-forming block, as judged by rheology (see Figure 5 and Figure S4). However, increasing the GlyMA content suppresses the thermoresponsive behavior of the diblock copolymer worm gels, with lower CGTs being observed. As previously discussed, GlyMA is consumed faster than HPMA during the RAFT statistical copolymerization of these two comonomers. This results in a GlyMA-enriched block junction. However, as GlyMA residues are more hydrophobic than HPMA residues, progressively lower temper-

Table 1. Summary of Dynamic Light Scattering (DLS) Data Obtained Both before and after APTES Cross-Linking for PGMA₅₆-P(HPMA_y-stat-GlyMA_z) Diblock Copolymers (Where $y + z = 144$; These Copolymers Are Denoted as G₅₆-(H_y-stat-E_z) for Brevity) in Pure Water, Methanol, and in a 1.0% w/w Aqueous SDS Solution

| copolymer composition | before cross-linking | | | | after cross-linking | | | | | |
|--|----------------------|-------------------------|---------------|-------------------------|---------------------|-------------------------|---------------|-------------------------|---------------|-------------------------|
| | water | | methanol | | water | | methanol | | SDS | |
| | diam/nm (PDI) | derived count rate/kcps | diam/nm (PDI) | derived count rate/kcps | diam/nm (PDI) | derived count rate/kcps | diam/nm (PDI) | derived count rate/kcps | diam/nm (PDI) | derived count rate/kcps |
| G ₅₆ -H ₁₄₄ | 102 (0.184) | 39 600 | 9 (0.216) | 140 | nd | nd | nd | nd | nd | nd |
| G ₅₆ -(H ₁₃₇ -stat-E ₇) | 150 (0.210) | 56 300 | 38 (0.252) | 250 | 152 (0.272) | 22 400 | 66 (0.169) | 3 600 | 48 (0.281) | 5 740 |
| G ₅₆ -(H ₁₃₀ -stat-E ₁₄) | 122 (0.206) | 51 400 | 14 (0.246) | 280 | 172 (0.345) | 36 300 | 266 (0.411) | 24 400 | 200 (0.255) | 31 700 |
| G ₅₆ -(H ₁₂₂ -stat-E ₂₂) | 128 (0.238) | 39 200 | 13 (0.354) | 210 | 235 (0.404) | 32 800 | 251 (0.295) | 30 300 | 231 (0.269) | 32 300 |
| G ₅₆ -(H ₁₁₅ -stat-E ₂₉) | 203 (0.286) | 41 500 | 61 (0.212) | 210 | 200 (0.412) | 28 900 | 220 (0.242) | 40 500 | 173 (0.238) | 34 000 |

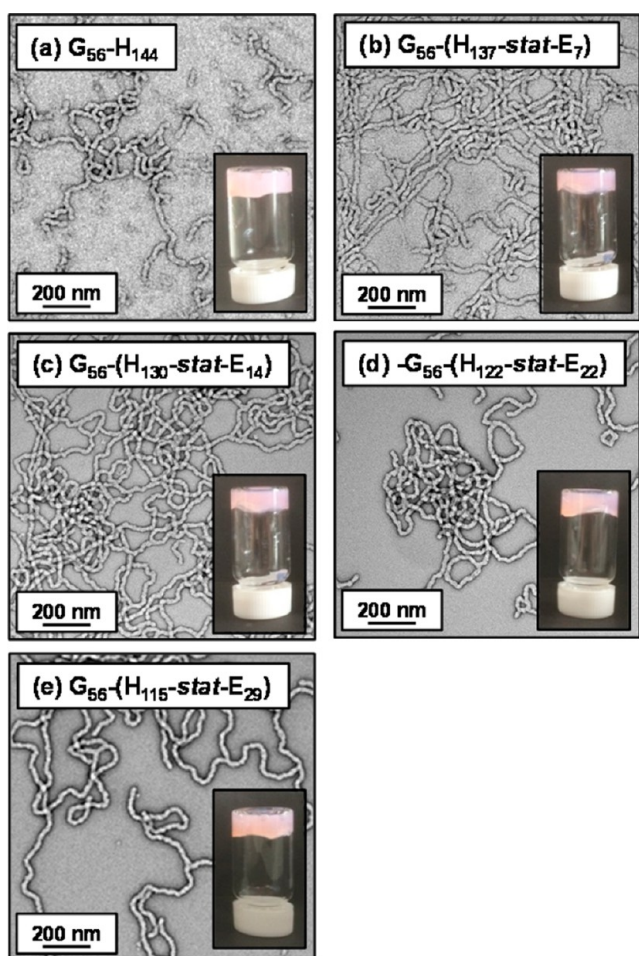


Figure 4. Representative TEM images obtained for dried 0.1% w/w aqueous dispersions of PGMA₅₆-P(HPMA_y-stat-GlyMA_z) linear diblock copolymers prior to cross-linking (where $y + z = 144$; these copolymers are denoted as G₅₆-(H_y-stat-E_z) for brevity). Digital photographic images of the corresponding free-standing gels recorded at 7.5% w/w solids are shown as insets.

atures are required for the surface plasticization necessary to induce a worm-to-sphere transition (and hence degelation). Furthermore, more pronounced hysteresis is observed on returning to 25 °C. This is because the worm-to-sphere transition is relatively fast compared to the sphere-to-worm transition, since the latter process is highly cooperative. These rheological studies also indicate a reduction in storage modulus (G') from 86 to 11 Pa at 25 °C on increasing the GlyMA

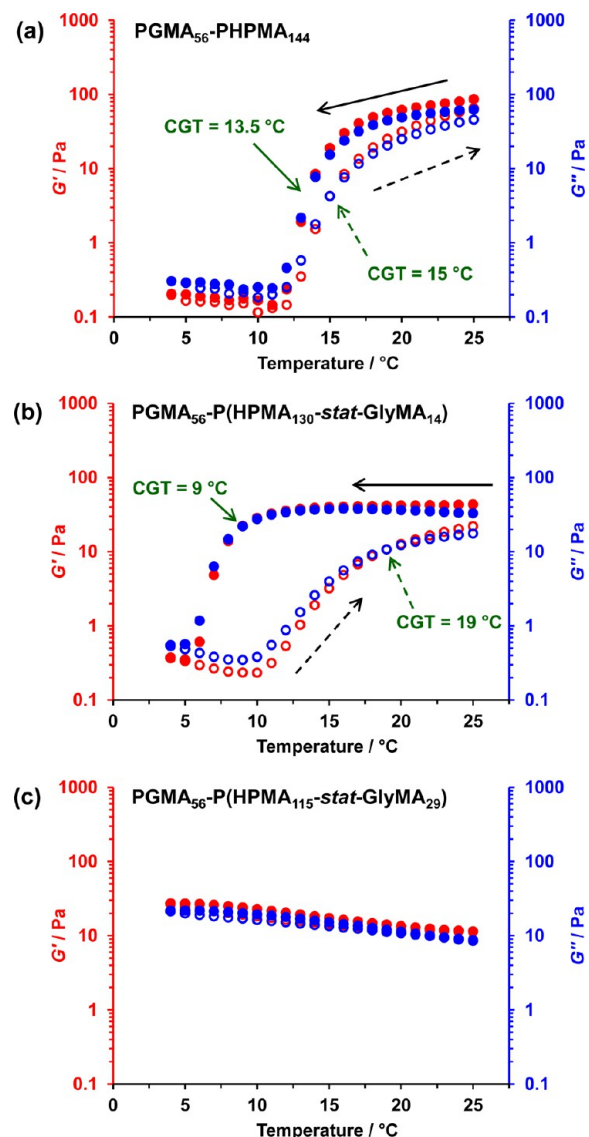


Figure 5. Variation of the storage modulus (G' ; denoted by red data set) and the loss modulus (G'' ; denoted by blue data set) as a function of temperature (closed circles denote a 25 to 5 °C temperature sweep, and open circles denote a 5 to 25 °C temperature sweep) for a 7.5% w/w aqueous dispersion of (a) PGMA₅₆-PPHMA₁₄₄, (b) PGMA₅₆-P(HPMA₁₃₀-stat-GlyMA₁₄), and (c) PGMA₅₆-P(HPMA₁₁₅-stat-GlyMA₂₉) worms before cross-linking. Conditions: angular frequency = 1.0 rad s⁻¹, applied strain = 1.0%, and rate of cooling/heating = 0.50 °C min⁻¹.

content in the core-forming block from 0 to 20 mol %. Rheological studies of PGMA₅₄-PHPMA_y diblock copolymer worms reported by Verber and co-workers over a range of y values indicated that block compositions closer to the worm/sphere phase boundary formed weaker gels.³⁶ Thus, it is possible that incorporating more GlyMA into the core-forming statistical block shifts the worm morphology toward this phase boundary.

It is worth considering why these worm gels are so soft at 25 °C. If the mean worm width and length for the linear PGMA₅₆-PHPMA₁₄₄ worms are 24 and 245 nm, respectively, and assuming a worm density of approximately 1.10 g cm⁻³, then the mean worm mass is estimated to be 1.22×10^{-16} g. Assuming a worm concentration of 7.5% w/w, then the worm number density is 6.2×10^{20} m⁻³. If we assume a mean copolymer molecular weight of 30 000 g mol⁻¹, then the mean aggregation number (or average number of copolymer chains per worm) is estimated to be 2450. It is well-known that $G' = \nu_e kT$.⁸⁰ If G' is 86 Pa (see Figure 5a), then the number density of elastically effective chains (ν_e) is estimated to be 2.1×10^{22} m⁻³. This means that each worm contributes 34 elastically effective chains. Thus, there is one just elastically effective chain per 72 copolymer chains. This explains why these worm gels are so soft: the copolymer chains are used rather inefficiently within the dissipative network. This is in part because the chains have some degree of mobility within the worms, especially when subjected to strain. In this context, it is noteworthy that variable temperature ¹H NMR studies reported by Blanz and co-workers provide direct experimental evidence for partial solvation of the PHPMA core-forming blocks.¹⁴ When the worm cores are covalently cross-linked using APTES, an increase in gel modulus is observed (see later). This means that the number of copolymer chains per elastically effective chain is reduced because core cross-linking links individual copolymer chains together, leading to the more effective distribution of stress within the worm gel.

Post-Polymerization Cross-Linking of PGMA-P-(HPMA-*stat*-GlyMA) Diblock Copolymer Worms. On reaching full conversion, the aqueous worm gels were immediately diluted from 15 to 7.5% w/w to lower the gel viscosity. Once a homogeneous dispersion was achieved, APTES was added (APTES/GlyMA molar ratio = 1.0), and the shear-thinning gel was stirred overnight at 20 °C. As discussed earlier, the primary amine of the APTES reacts with the pendent epoxide groups in the GlyMA residues while the siloxane groups undergo multiple hydrolysis–condensation reactions that lead to highly cross-linked worm cores (see Figure 6). In reality, cross-linking is likely to be even more complex because the secondary amines formed via ring-opening of the epoxide group can in principle react with a second epoxide. One interesting question here is the following: to what extent does the time scale for the epoxy–amine reaction differ from that of the hydrolysis–condensation reactions? To address this point, the rate of reaction of APTES with the epoxide groups and the rate of hydrolysis–condensation for the PGMA₅₆-P(HPMA₁₁₅-*stat*-GlyMA₂₉) diblock copolymer worms were monitored by ¹H NMR using *d*₄-sodium trimethylsilylpropanoate (TMSP) as an internal standard (see Figure 7). Aliquots of the aqueous reaction mixture were extracted at regular intervals and diluted using CD₃OD prior to NMR analysis. This choice of diluent enables chemical changes in the core-forming block to be monitored up to relatively high degrees of cross-linking. The rate of ring-opening by the

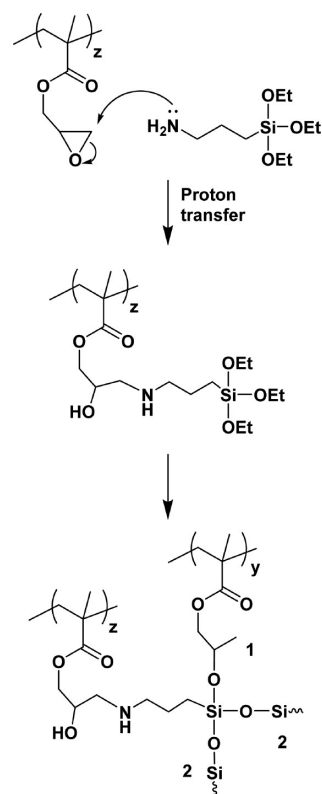


Figure 6. Reaction scheme illustrating worm core cross-linking chemistry by (i) epoxy ring-opening via nucleophilic attack with APTES and (ii) intermolecular cross-linking via hydrolysis–condensation. The latter step involves either reaction of the APTES with hydroxyl groups on HPMA residues on another copolymer chain (denoted as 1) and/or condensation with other APTES groups (denoted as 2). In reality, ¹H NMR studies indicate that these two steps occur more or less simultaneously, rather than consecutively as shown (see main text for details). Moreover, the chemistry is likely to be more complex than that shown as the secondary amine species may react further.

nucleophilic APTES was determined by monitoring the disappearance of the characteristic epoxy proton signals at 3.0 ppm in the ¹H NMR spectra relative to the internal standard (see blue data set shown in Figures 7b and 7a for the corresponding ¹H NMR spectra). The integrated epoxy signal is reduced to 6% of its original value after 8 h (and to just 3% after 24 h). As the hydrolysis–condensation reaction proceeds, the chemical cross-links lead to worm core *swelling* in CD₃OD, rather than worm *dissolution*. At higher degrees of cross-linking, the worm cores become solid-like and hence no longer solvated by the CD₃OD; thus, signals associated with the P(HPMA-*stat*-GlyMA) core-forming block gradually become undetectable by ¹H NMR. This can be used to infer the relative degree of cross-linking by determining either the normalized reduction in the methyl group signal assigned to the methacrylic backbone at 0.9 ppm (green data set in Figure 7) or that of the pendent methyl group assigned to the HPMA residues (see red data set in Figure 7 and Figure S5 for the corresponding ¹H NMR traces). However, the latter method is preferred because in the former method data analysis is made more complicated by overlapping backbone methyl group signals arising from the PGMA stabilizer block, which remains soluble (and hence detectable) even after core cross-linking is complete. It was originally anticipated that the epoxy–amine reaction would occur prior to

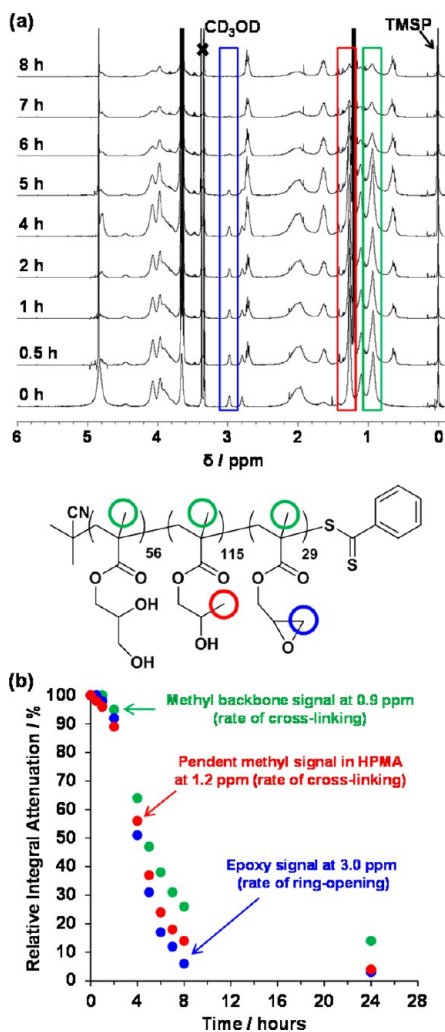


Figure 7. (a) ¹H NMR spectra obtained at various time points following the reaction of APTES with PGMA₅₆-P(HPMA₁₅-stat-GlyMA₂₉) after dilution into CD₃OD. The amine reacts with GlyMA as judged by the reduction in the epoxy signal peak at 3.0 ppm compared to the internal standard TMS. (b) Kinetics of the ring-opening epoxy-amine reaction (blue data set) as judged by the attenuation in the relative integral of the epoxide signal at 3.0 ppm compared to an internal standard by ¹H NMR spectroscopy. Kinetics of worm core cross-linking as judged by the relative attenuation in the integrated pendent methyl group signal at 0.9 ppm assigned to the HPMA residues (red data set) and the relative attenuation in the integrated methyl signal at 1.2 ppm assigned to the methacrylate backbone (green data set) compared to the same internal standard at 0 ppm.

the hydrolysis-condensation reaction, leading to a sequential cross-linking process. However, the data shown in Figure 7 indicate that the relative integral of the HPMA methyl signal is attenuated at a comparable rate as that of the epoxy signals. This indicates that ring-opening of the epoxy groups and the hydrolysis-condensation reactions actually occur over similar time scales, suggesting that cross-linking does not proceed via a two-stage mechanism. However, the precise degree of cross-linking cannot be calculated because further cross-linking may occur that is no longer detectable by ¹H NMR. It is perhaps noteworthy that the reaction times shown in Figure 7 correspond to the times at which each aliquot was taken from the reaction mixture—it does not include the time taken to run each ¹H NMR spectrum. Diluting each aliquot with an

equal volume of CD₃OD may not adequately quench the reaction, so it was important to analyze each aliquot as soon as possible in order to minimize this “dead time”. (In practice, the time required for instrument setup and spectrum acquisition was around 15 min for each sample.) Notwithstanding such minor time domain errors, this spectroscopic study confirmed that an approximate time scale of 24 h is required for extensive cross-linking of each of the four GlyMA-containing diblock copolymer worms at 20 °C under the stated conditions.

In principle, core cross-linking should prevent worm dissolution on dilution in methanol (which is a good solvent for both blocks). DLS studies conducted on 0.1% w/w aqueous worm dispersions (see Table 1) indicates that cross-linking causes a significant increase in the apparent hydrodynamic diameters [from 122 to 172 nm for the PGMA₅₆-P(HPMA₁₃₀-stat-GlyMA₁₄) worms and from 128 to 235 nm for PGMA₅₆-P(HPMA₁₂₂-stat-GlyMA₂₂) worms]. However, it is emphasized that only *sphere-equivalent dimeters* are reported by DLS, so it is difficult to interpret such observations in terms of changes in either worm contour lengths or worm widths. Moreover, this apparent increase in particle dimensions could in principle simply be a result of some degree of inter-particle cross-linking. Nevertheless, TEM images obtained for the four core cross-linked diblock copolymer worms containing 5, 10, 15, or 20 mol % GlyMA (see Figure 8) after dilution to 0.1% w/w

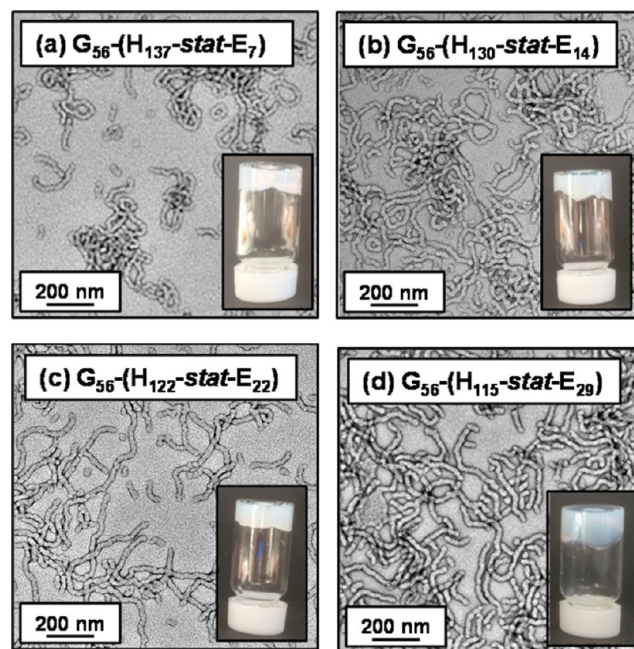


Figure 8. Representative TEM images obtained for dried 0.1% w/w aqueous dispersions of PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymers after APTES cross-linking of 7.5% w/w worm dispersions at 20 °C. Inset digital photographic images were recorded for the same aqueous copolymer dispersions at 7.5% w/w solids; free-standing gels are observed in each case.

aqueous dispersions do not indicate any discernible change in the original worm morphology. Furthermore, all four core cross-linked worm dispersions still form free-standing gels at 7.5% w/w solids, as judged by a tube inversion test (see Figure 8). However, DLS studies conducted on the same four worm dispersions after dilution to 0.1% w/w in methanol suggest that only worms comprising at least 10 mol % GlyMA are fully

resistant to the presence of methanol (see Table 1). In contrast, the PGMA₅₆-P(HPMA₁₃₇-stat-GlyMA₇) diblock copolymer (5 mol % GlyMA) shows a dramatic reduction in apparent hydrodynamic diameter from 152 nm in water to 66 nm in methanol, with a relatively low derived count rate (3600 kcps) being observed in the latter solvent. This suggests that the worms undergo a morphological transition to spheres and/or short worms. However, TEM images obtained (see Figure 9a)

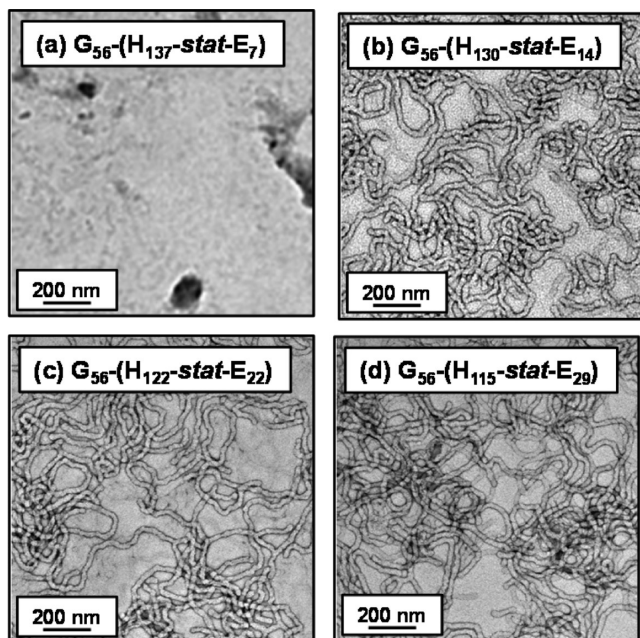


Figure 9. Representative TEM images obtained for core cross-linked PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymers (abbreviated G₅₆-(H_y-stat-E_z) for the sake of brevity) after drying 0.1% w/w methanolic dispersions at 20 °C. (a) No well-defined nano-objects were observed at 5 mol % GlyMA, whereas the original worm morphology persists when core cross-linked worms contain higher proportions of GlyMA; see images (b), (c), and (d).

for this latter diblock copolymer dried as a 0.1% w/w dispersion in methanol suggest that no well-defined particles are present (i.e., worm dissolution most likely occurs under these conditions). Indeed, ¹H NMR studies of this copolymer in CD₃OD confirm a strong signal at around 1.25 ppm corresponding to the pendent methyl groups on the HPMA residues (data not shown). In contrast, DLS studies of the other three diblock copolymer worms (containing 10, 15, or 20 mol % GlyMA) in methanol indicate a much higher derived count rate of at least 22 000 kcps (see Table 1). Moreover, these diblock copolymer worms exhibit an increase in hydrodynamic diameter when dispersed in methanol as opposed to water. This is the result of *swelling* of the cross-linked worm cores because methanol is a good solvent for both blocks, but the relatively low degree of cross-linking is sufficiently high to prevent worm *dissolution*. TEM images obtained for PGMA₅₆-P(HPMA₁₃₀-stat-GlyMA₁₄), PGMA₅₆-P(HPMA₁₂₂-stat-GlyMA₂₂), and PGMA₅₆-P(HPMA₁₁₅-stat-GlyMA₂₉) diblock copolymers dried from 0.1% w/w methanolic dispersions confirmed the persistence of the pure worm morphology in each case (Figure 9b–d).

When APTES is reacted with the epoxy groups on the GlyMA residues, a secondary amine is generated (see Figure 6). Thus, the resulting core cross-linked worms might be expected

to possess weakly cationic character below neutral pH (where the secondary amine groups become protonated). In a control experiment, aqueous electrophoresis studies conducted on a 0.1% w/w aqueous dispersion of linear PGMA₅₆-PHPMA₁₄₄ diblock copolymer worms indicated no cationic character from pH 10 to 3 (see Figure 10). In contrast, APTES-cross-linked

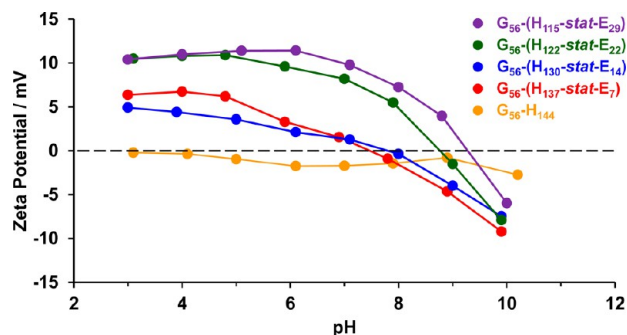


Figure 10. Zeta potential versus pH curves obtained at 25 °C for 0.1% w/w aqueous dispersions of linear PGMA₅₆-PHPMA₁₄₄ diblock copolymer worms and four examples of APTES cross-linked PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymer worms in the presence of 10⁻³ M KCl.

PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymer worms displayed cationic character below pH 7–9 (see Figure 10). However, these particles exhibit only relatively weak cationic character (+5 to +10 mV) at pH 5. In contrast, Penfold et al. have recently reported that linear PGMA₅₀-PHPMA₁₄₀ worms prepared using a morpholine-based RAFT agent exhibit zeta potentials of around +15 mV, even though there is only one terminal morpholine group per stabilizer block in this case.⁸¹ This discrepancy most likely arises because the cationic charge is located within the cores of the cross-linked worms in the present study, rather than in the stabilizer block. Above pH 9, all worms exhibit weakly anionic character (−5 to −10 mV). This most likely indicates the presence of carboxylic acid end-groups on some of the PGMA stabilizer chains resulting from use of ACVA initiator in their RAFT synthesis.⁷⁴

It is noteworthy that the characteristic pink color of the worm gels that arises from the dithiobenzoate-based RAFT CTA is removed during the APTES cross-linking reaction (see Figure 8). This is the result of nucleophilic attack on the dithioester by the strongly basic primary amine groups (after APTES addition, the solution pH increases to pH 9–10).^{82,83} However, as the dithioester chain-ends are located *within* the worm cores, this side reaction is unlikely to adversely affect the physical properties of these copolymer worm dispersions.

Cross-linking also causes the PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymer worms to form stronger gels, as judged by comparing the storage moduli (*G'*) of 7.5% w/w worm gels before and after cross-linking by oscillatory rheology (see Table 2). For example, cross-linking the PGMA₅₆-P(HPMA₁₃₀-stat-GlyMA₁₄) worms leads to an increase in *G'* from 43 to 81 Pa at 25 °C (see Figure 11). Previous work by Bates and co-workers suggest that this is due to worm stiffening, which leads to a longer worm persistence length.⁷ Moreover, temperature-dependent rheological studies indicate that the degelation that is observed on cooling *linear* diblock copolymer worm gels no longer occurs after worm core cross-linking (see Figure 11 and Figure S6). Clearly, covalent stabilization of the PGMA₅₆-P(HPMA_y-stat-GlyMA_z) worms

Table 2. Summary of Data Obtained for PGMA₅₆–P(HPMA_y-stat-GlyMA_z) Diblock Copolymer Worm Gels in the Presence and Absence of Methanol or 1.0% Aqueous SDS Solution before and after APTES Cross-Linking at 20 °C

| copolymer composition | before cross-linking | | | | | after cross-linking | | | | |
|---|---|------------------------|----------------------------------|---|--|--|----------------------------------|---|--|--|
| | M_n / g mol ⁻¹ ^a | M_w/M_n ^a | G' at 25 °C/Pa ^b | thermoreponsive degelation? ^b | stable in the presence of methanol? ^c | stable in the presence of SDS? ^d | G' at 25 °C/Pa ^b | thermoreponsive degelation? ^b | stable in the presence of methanol? ^c | stable in the presence of SDS? ^d |
| G ₅₆ -H ₁₄₄ | 37 400 | 1.12 | 86 | | no | no | | | | |
| G ₅₆ (H ₁₃₇ -stat-E ₇) | 35 400 | 1.13 | 76 | yes | no | no | 170 | no | no | partial* |
| G ₅₆ (H ₁₃₀ -stat-E ₁₄) | 34 300 | 1.14 | 43 | yes | no | no | 81 | no | yes | yes |
| G ₅₆ (H ₁₂₂ -stat-E ₂₂) | 35 500 | 1.14 | 32 | yes | no | no | 119 | no | yes | yes |
| G ₅₆ (H ₁₁₅ -stat-E ₂₉) | 35 000 | 1.15 | 11 | no | no | no | 13 | no | yes | yes |

^aCalculated using DMF GPC against a series of near-monodisperse PMMA calibration standards using a refractive index detector. ^bDetermined for 7.5% copolymer worm gels using oscillatory rheology at an angular frequency of 1.0 rad s⁻¹ and an applied strain of 1.0%. ^cAs judged by DLS and TEM studies conducted on PGMA₅₆–P(HPMA_y-stat-GlyMA_z) diblock copolymer worms diluted to 0.1% w/w in methanol. ^dAs judged by DLS and TEM studies conducted on PGMA₅₆–P(HPMA_y-stat-GlyMA_z) diblock copolymer worms diluted to 0.1% w/w in the presence of 1.0% w/w SDS aqueous solution (i.e., SDS/copolymer mass ratio = 10).

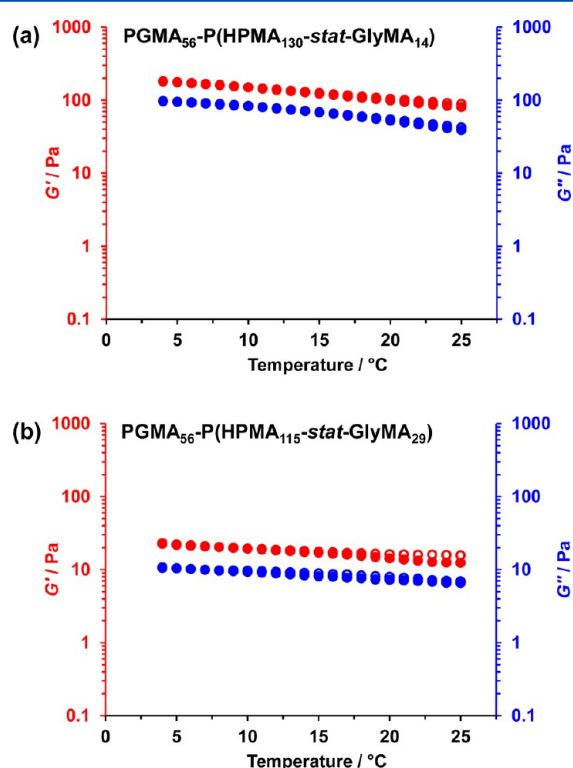


Figure 11. Variation in storage modulus (G' ; red circles) and loss modulus (G'' ; blue circles) as a function of temperature (closed circles denote the cooling temperature sweep and open circles denote the heating temperature sweep) for 7.5% w/w aqueous dispersions of (a) PGMA₅₆–P(HPMA₁₃₀-stat-GlyMA₁₄) and (b) PGMA₅₆–P(HPMA₁₁₅-stat-GlyMA₂₉) after worm core cross-linking using APTES (final solution pH 9–10). Conditions: angular frequency = 1.0 rad s⁻¹; applied strain = 1.0%; heating/cooling rate = 0.5 °C min⁻¹.

prevents their dissociation into spheres at around 5 °C. Moreover, even the relatively lightly cross-linked PGMA₅₆–P(HPMA₁₃₇-stat-GlyMA₇) worm gel is no longer thermoresponsive (see Figure S6a), although DLS and TEM studies indicate that the same APTES-treated worms undergo dissolution when diluted in methanol (see Table 1).

G' and G'' represent the energy per unit strain that is stored or dissipated, respectively. It is well-known that $\tan \delta = G''/G'$.⁸⁴ If the latter parameter increases, then this indicates greater energy dissipation. For the linear worm gels reported herein,

there are two likely energy dissipation mechanisms: (i) *intra-worm* interactions between the hydrophobic core-forming PHPMA blocks and (ii) *inter-worm* entanglements and/or multiple contacts. In each case energy is efficiently dissipated because such physical interactions are not fixed. Core cross-linking reduces $\tan \delta$, presumably because there is less dissipation via *intra-worm* interactions (see Figure 12). There

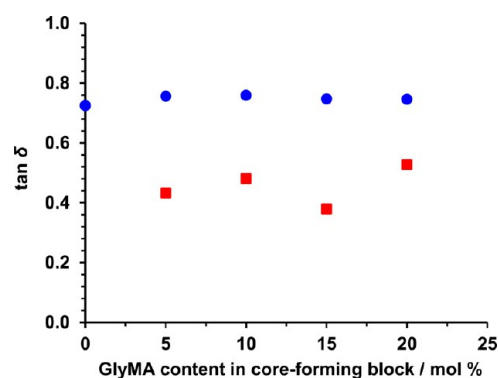


Figure 12. Values for $\tan \delta$ at 25 °C for a series of PGMA₅₆–P(HPMA_y-stat-GlyMA_z) diblock copolymer worms before (blue data set) and after (red data set) after cross-linking. Covalent stabilization of the worms leads to a reduction in $\tan \delta$.

may also be a contribution from the greater persistence length: the cross-linked worms acquire “rod-like” character compared to the highly flexible linear worms, which should reduce dissipation owing to entanglements or multiple contacts. However, $\tan \delta$ values are still relatively high compared to other types of physical gels (e.g., peptide gels^{85–87}), suggesting that cross-linking does not completely suppress dissipation.

Chambon et al. demonstrated that PGMA₅₅–PHPMA₃₃₀ diblock copolymer vesicles fully dissociated to form individual copolymer chains when challenged with an anionic surfactant.⁶³ In contrast, PGMA₅₅–P(HPMA₂₄₇-stat-GlyMA₈₂) diblock copolymer vesicles that had been cross-linked using a small molecule (or polymeric) diamine proved to be surfactant-resistant. In principle, similar findings might be expected for the core cross-linked diblock copolymer worms described herein. Thus, core cross-linking is potentially useful because the resulting worms may be suitable as viscosity modifiers for various commercial surfactant-based home and personal care formulations. In this study, sodium dodecyl sulfate (SDS) was

selected to assess the surfactant resistance of the worms, as this amphiphile was previously demonstrated to be particularly disruptive toward diblock copolymer vesicles.⁶³ The surfactant resistance of all diblock copolymer worms was judged by TEM analysis of 0.1% w/w copolymer dispersions conducted in the absence and presence of 1.0% w/w SDS (i.e., a SDS/copolymer mass ratio of 10). As expected, when the linear PGMA₅₆-PHPMA₁₄₄ worm gels were subjected to an SDS challenge, there was an immediate reduction in turbidity, and DLS studies indicated a relatively low count rate of 260 kcps (see Table 1), suggesting rapid dissociation to form dissolved copolymer chains. This was corroborated by TEM, since no nano-objects could be observed (see Figure 13a). Similarly, linear PGMA₅₆-

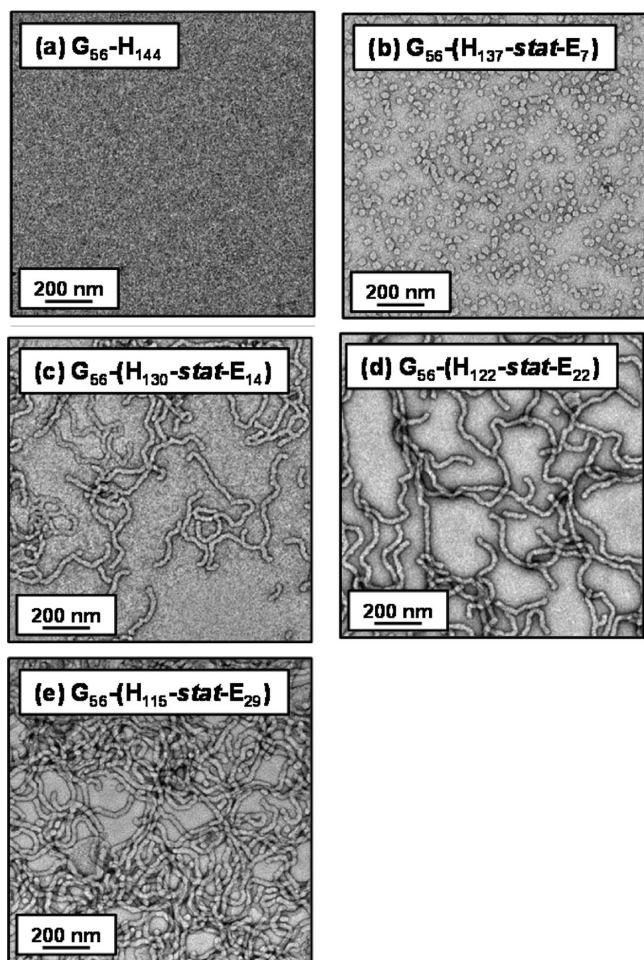


Figure 13. Representative TEM images obtained for dried dispersions of 0.1% w/w APTES cross-linked PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymer worms exposed to the presence of 1.0% w/w SDS at 20 °C.

P(HPMA_y-stat-GlyMA_z) diblock copolymer worms challenged with SDS also undergo immediate dissociation. In all cases no particles could be observed by TEM (see Figure S6). Interestingly, APTES cross-linked PGMA₅₆-P(HPMA₁₃₇-stat-GlyMA₇) worms only exhibit *partial* resistance to this surfactant challenge. Rather than undergoing complete dissolution, a worm-to-sphere transition is instead observed by TEM (see Figure 13b), while DLS indicated a significant reduction in hydrodynamic diameter from 150 to 48 nm in the presence of SDS (see Table 1). However, on increasing the GlyMA content to 10, 15, or 20 mol % (and therefore the degree of core cross-

linking), the worms became completely resistant to the presence of SDS. DLS studies of PGMA₅₆-P(HPMA₁₃₀-stat-GlyMA₁₄), PGMA₅₆-P(HPMA₁₂₂-stat-GlyMA₂₂), and PGMA₅₆-P(HPMA₁₁₅-stat-GlyMA₂₉) diblock copolymer worms in the presence and absence of SDS revealed only minor changes in their apparent sphere-equivalent diameters (see Table 1). Furthermore, TEM images recorded after drying these diluted “worm plus surfactant” dispersions confirm that the original worm morphology is retained over time scales of months in each case (see Figure 13c–e).

The colloidal stabilities of the five PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymer worms (prepared targeting the same overall mean degree of polymerization; $y + z = 144$) before and after cross-linking are summarized in Table 2. Prior to cross-linking, none of the linear PGMA₅₆-P(HPMA_y-stat-GlyMA_z) worms remained intact when challenged with either methanol or SDS. However, APTES treatment can significantly improve worm stability toward either reagent. In particular, for worm cores comprising at least 10 mol % GlyMA, TEM and DLS studies confirm that the worm morphology is preserved in the presence of either methanol or 1.0% w/w aqueous SDS solution. Furthermore, temperature-dependent oscillatory rheology studies demonstrate that worm core cross-linking results in stiffer gels that no longer exhibit thermoresponsive behavior.

CONCLUSIONS

In summary, a series of PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymer worm gels have been conveniently prepared by polymerization-induced self-assembly in concentrated aqueous solution by targeting a constant core-forming block DP of 144 in each case. Increasing the GlyMA content in such linear copolymers affords weaker gels, as judged by rheology studies. ¹H NMR studies of the kinetics of statistical copolymerization of water-immiscible GlyMA with water-miscible HPMA indicate that the former comonomer is more reactive than the latter. Thus, the comonomer composition of the core-forming statistical block becomes GlyMA-rich at its junction with the PGMA stabilizer block. This explains why temperature-dependent rheological studies indicate that worms with higher GlyMA contents gradually become less thermoresponsive, since progressively lower temperatures are required to induce surface plasticization of the worms and hence degelation via a worm-to-sphere transition. Ultimately, thermally-induced degelation is no longer observed at a GlyMA content of 20 mol %. Such diblock copolymer worms can be core cross-linked by adding APTES. Perhaps surprisingly, ¹H NMR studies indicate that the ring-opening and cross-linking reactions occur over similar time scales rather than via a two-stage reaction. The cross-linked worms are expected to be significantly stiffer than the linear worm precursors. Indeed, DLS provides some evidence for longer persistence lengths, and the cross-linked worms also form stronger gels, which in all cases no longer undergo thermally-induced degelation on cooling. TEM studies of dried diluted aqueous worm dispersions confirmed that core cross-linking produced no discernible change in the copolymer morphology. Furthermore, TEM studies conducted prior to cross-linking indicate that all of the linear PGMA₅₆-P(HPMA_y-stat-GlyMA_z) diblock copolymer worms are unstable with respect to the addition of either methanol (a good solvent for both blocks) or an anionic surfactant (SDS). In contrast, the corresponding cross-linked worms remain colloidally stable provided that the

core-forming block contained at least 10 mol % GlyMA. Finally, it is noteworthy that the cross-linking chemistry described herein (i) utilizes cheap commercially available reagents, (ii) can be conveniently conducted at 20 °C in aqueous solution, and (iii) produces secondary amine groups within the worm cores, which results in weakly cationic worms below pH 7, as judged by aqueous electrophoresis.

■ ASSOCIATED CONTENT

■ Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.macromol.6b00422.

Assigned ^1H NMR spectra for PGMA₅₆-P(HPMA₁₁₅-stat-GlyMA₂₉) diblock copolymer; conversion vs time curves, corresponding semilogarithmic plots and DLS data; TEM images for linear PGMA₅₆-P(HPMA₇-stat-GlyMA₂) diblock copolymers in the presence of methanol or SDS; rheological data for PGMA₅₆-P(HPMA₁₃₇-stat-GlyMA₇) and PGMA₅₆-P(HPMA₁₂₂-stat-GlyMA₂₂) diblock copolymer worms before and after cross-linking; ^1H NMR spectra obtained during cross-linking of PGMA₅₆-P(HPMA₁₁₅-stat-GlyMA₂₉) (PDF)

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Notes

The authors declare no competing financial interest.

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