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Frequency of Germline Mutations in 25 Cancer Susceptibility Genes in a Sequential Series of Patients With Breast Cancer

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See accompanying editorial on page 1433 and article on page 1455

ABSTRACT

Purpose

Testing for germline mutations in BRCA1/2 is standard for select patients with breast cancer to guide clinical management. Next-generation sequencing (NGS) allows testing for mutations in additional breast cancer predisposition genes. The frequency of germline mutations detected by using NGS has been reported in patients with breast cancer who were referred for BRCA1/2 testing or with triple-negative breast cancer. We assessed the frequency and predictors of mutations in 25 cancer predisposition genes, including *BRCA1/2*, in a sequential series of patients with breast cancer at an academic institution to examine the utility of genetic testing in this population.

Methods

Patients with stages I to III breast cancer who were seen at a single cancer center between 2010 and 2012, and who agreed to participate in research DNA banking, were included ($N = 488$). Personal and family cancer histories were collected and germline DNA was sequenced with NGS to identify mutations.

Results

Deleterious mutations were identified in 10.7% of women, including 6.1% in *BRCA1/2* (5.1% in non-Ashkenazi Jewish patients) and 4.6% in other breast/ovarian cancer predisposition genes including $CHEK2$ (n = 10), ATM (n = 4), $BRIP1$ (n = 4), and one each in $PALB2$, PTEN, NBN, RAD51C, RAD51D, MSH6, and PMS2. Whereas young age ($P < .01$), Ashkenazi Jewish ancestry ($P < .01$), triplenegative breast cancer ($P = .01$), and family history of breast/ovarian cancer ($P = .01$) predicted for BRCA1/2 mutations, no factors predicted for mutations in other breast cancer predisposition genes.

Conclusion

Among sequential patients with breast cancer, 10.7% were found to have a germline mutation in a gene that predisposes women to breast or ovarian cancer, using a panel of 25 predisposition genes. Factors that predict for BRCA1/2 mutations do not predict for mutations in other breast/ovarian cancer susceptibility genes when these genes are analyzed as a single group. Additional cohorts will be helpful to define individuals at higher risk of carrying mutations in genes other than BRCA1/2.

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INTRODUCTION

Testing for mutations in high-penetrance breast cancer predisposition genes, particularly BRCA1 and BRCA2, has become standard practice for patients with breast cancer. Lifetime estimates of breast cancer risk in BRCA1 or BRCA2 (BRCA1/2) carriers range from 36% to 90% and of ovarian cancer risk range from 24% to 59% and 8% to 35% in BRCA1 and BRCA2 carriers, respectively. $1-5$ Identification of BRCA1/2 mutations permits the implementation of prevention strategies, including

magnetic resonance imaging screening or risk-reducing surgeries, which improves survival.^{[6,7](#page-7-0)} Genetic testing for other high-risk breast cancer susceptibility genes, such as TP53 (Li-Fraumeni syndrome), PTEN (Cowden's syndrome), and CDH1 (hereditary diffuse gastric cancer), is also standard in appropriate patients.

More recently, next-generation sequencing (NGS) has enabled simultaneous testing for mutations in these high-penetrance genes and for other, more moderate-risk genes. Multigene panels are now commercially available and are increasingly being used in cancer risk assessment.

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Terms in blue are defined in the glossary, found at the end of this article and online at www.jco.org.

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Compared with high-penetrance genes, for which inherited mutations confer a five-fold or greater breast cancer risk, mutations in moderate-penetrance genes are associated with a two- to four-fold increase in risk. Cancer risks associated with mutations in these less potent predisposition genes are still being investigated. For example, mutations in PALB2, initially thought to confer a moderate risk of breast cancer, now seem to be associated with a five-fold or greater risk.^{[8](#page-7-0)} NGS also allows simultaneous testing for other hereditary cancer risks, such as Lynch syndrome, in individuals with and without a suggestive family history. In addition, germline mutations in DNA repair genes such as BRIP1, RAD51C, and RAD51D are associated with an increased risk of ovarian cancer (Table 1). $9-15$

To date, studies evaluating the prevalence of mutations in moderate-penetrance breast cancer predisposition genes have been conducted in select breast cancer populations including African Americans,^{[16](#page-7-0)} patients with triple-negative breast cancer (TNBC),^{[17](#page-7-0)} and cases seen in high-risk genetic clinics. $18-21$ $18-21$ $18-21$ The prevalence of mutations among patients with breast cancer, who are unselected for specific risk factors such as age at diagnosis, breast cancer subtype, or family cancer history, is unknown.

Abbreviations: APC, adenomatous polyposis coli; ATM, ataxia telangiectasia
mutated; BARD1, BRCA1-associated RING domain 1; BMPR1A, bone mormutated; *BARD1*, BRCA1-associated RING domain 1; *BMPR1A*, bone mor-
phogenetic protein receptor, type 1A; *BRCA1/2*, early-onset breast cancer genes
BRCA1 and *BRCA2; BRIP1,* BRCA1 interacting protein C-terminal helica CDH1, E-cadherin; CDK4, cyclin-dependent kinase 4; CDKN2A, cyclin-dependent kinase inhibitor 2A; CHEK2, checkpoint kinase 2; EPCAM, epithelial cell adhesion molecule; MLH1, mutL homolog 1; MSH2, mutS homolog 2; MSH6, mutS homolog 6; MUTYH, biallelic mutY homolog; NBN, nibrin; PALB2, partner and localizer of BRCA2; PMS2, PMS2 postmeiotic segregation increased 2; PTEN, phosphatase and tensin homolog; RAD51C, RAD51 paralog C; RAD51D, RAD51 paralog D; RR, relative risk; SMAD4, SMAD family member 4; STK11, serine/ threonine kinase 11; TP53, tumor protein 53.

*Only tumors with biallelic MUTYH mutations were considered for this analysis.

Evidence-based guidelines for BRCA1/2 testing in patients with breast cancer have been established. Criteria include young age at diagnosis, TNBC, Ashkenazi Jewish ancestry, or a significant family history of breast, ovarian, or other related cancers.^{[22,23](#page-7-0)} Existing recommendations for mutation detection in other high-penetrance genes are based on specific syndrome features. 22 22 22 Criteria for testing of moderate-penetrance predisposition genes do not yet exist because predictive factors have not been identified and clinical utility is still being evaluated.

In this study, we assessed the frequency of deleterious germline mutations in 25 cancer susceptibility genes in a population of consecutive patients with breast cancer who presented to an academic cancer center. Our goals were to better understand the contribution of inherited mutations in moderate- and high-risk genes in a breast cancer cohort unselected for family history, breast cancer subtype, ethnicity, or age at diagnosis and to evaluate any clinical or pathologic factors that predict for mutations in moderate-risk genes.

METHODS

Patient Selection

All women with stages I to III breast cancer seen at the Dana-Farber Cancer Institute (Boston, MA) between April 2010 and July 2012, who consented to DNA banking for clinical research, were eligible. Patients with a previous breast cancer were excluded. Cases were identified retrospectively and blood samples were obtained from the Dana-Farber/ Harvard Cancer Center Specialized Program of Research Excellence (SPORE) in breast cancer biobank. Clinical and pathologic data abstracted from medical records as part of the Clinical Outcomes for Research Information Service program included personal and family cancer histories, cancer histology, stage and receptor status, ancestry, and history of genetic testing. All breast cancers were reviewed by breast pathologists in the Department of Pathology at Brigham and Women's Hospital. ASCO/ College of American Pathologists guidelines were used to define estrogen receptor (ER), progesterone receptor, and human epidermal growth factor receptor 2 (HER2) positivity. Women with bilateral breast cancer were eligible, provided their first breast cancer was diagnosed between April 2010 and July 2012; only features of the initial breast cancer were assessed. Genetic test results from this analysis were considered research and were not returned to study participants or used for clinical decision making. Specimens were collected for research purposes only and therefore did not comply with Clinical Laboratory Improvement Amendments chain of custody regulations for clinical testing. This study was approved by the institutional review board of the Dana-Farber/Harvard Cancer Center.

NGS Assay

Sample preparation for NGS was performed from frozen DNA using the RainDance microdroplet polymerase chain reaction (PCR) system (RainDance Technologies, Billerica, MA). Briefly, PCR products representing exons and proximal splicing elements of patient DNA were amplified in merged droplets consisting of fragmented patient DNA and select target enrichment primers. These PCR products were subsequently tagged with barcodes and sequencing adaptors for NGS on the Illumina HiSeq platform (Illumina, San Diego, CA). To circumvent highly homologous pseudogenes, modified sample preparation with long-range and nested PCR, followed by NGS on the Illumina MiSeq platform, was used for portions of the CHEK2 and PMS2 genes. All clinically actionable variants identified by NGS, as well as regions that did not meet our preset NGS quality metrics, were independently confirmed with orthogonal sitespecific Sanger sequencing.

To detect exonic deletions and duplications, NGS dosage, microarray comparative genomic hybridization, multiplex ligation-dependent probe amplification, or a combination of these analyses was performed, 24 with all positive results confirmed by an orthogonal method. Gene variants deemed deleterious or suspected as deleterious were considered mutations. Analyzed genes were categorized into two groups [\(Table 1\)](#page-1-0).

Variant Classification

Variants were classified using American College of Medical Genetics and Genomics recommendations, with supporting linkage, biochemical, clinical, functional, and statistical data used for specific missense and intronic alterations. $25-27$

Statistics

Participant characteristics and sequencing results were summarized with descriptive statistics, including medians, means, and standard deviations for continuous data. For categorical data, proportions with 95% CIs were calculated by the Clopper-Pearson method. Demographic, clinical, and pathologic characteristics were compared using the χ^2 test (categorical variables) and the t test/analysis of variance (continuous variables). P values $< .05$ were considered significant.

RESULTS

Study Population

During the study period, 69.8% of patients with breast cancer seen at Dana-Farber Cancer Institute agreed to use of their clinical data and specimens for research. Sixty-one percent of blood samples were collected within 90 days of the initial breast cancer diagnosis and 94% within 1 year of diagnosis. The median time from diagnosis to blood sample collection was 77 days. Blood samples from 582 cases were analyzed and 87 failed due to insufficient DNA quantity or poor DNA quality. Six cases were excluded due to a prior breast cancer diagnosis and one was excluded for lack of clinical data, resulting in 488 cases which comprised the study cohort. Clinical and tumor pathologic features for study participants are provided in Table 2. The mean age at diagnosis was 50.3 years (range, 28 to 88 years); 7.8% of the study population were Ashkenazi Jewish and 81.4% were non-Ashkenazi white. Nearly 18% of women had TNBC, and 82.6% had stage I or II disease. Further, 49.0% of patients reported having a first- or second-degree relative with breast or ovarian cancer.

Frequency of Deleterious Mutations

Among 488 patients with breast cancer, 55 deleterious mutations were identified in 52 (10.7%) women ([Table 3](#page-3-0); Appendix [Table A1](#page-11-0), online only). Thirty (6.1%) women had a germline BRCA1/2 mutation; 18 in BRCA1 and 12 in BRCA2. In addition, 20 (4.1%) women had a total of 21 deleterious mutations in non-BRCA1/2 breast cancer predisposition genes including CHEK2 (n = 10), ATM (n = 4), BRIP1 (n = 4), and one each in PALB2, PTEN, and NBN. One ATM mutation was identified in a woman with a BRCA2 mutation, and one patient had both an ATM and a CHEK2 mutation. Four (0.8%) women carried deleterious mutations in genes unrelated to breast cancer; two in Lynch-related genes (one each in MSH6 and PMS2), and one each in RAD51C and RAD51D. The patient with a PMS2 mutation also had a BRCA1 mutation. Thus, 49 (10.0%) women had an inherited mutation in

Abbreviations: HER2, human epidermal growth factor receptor 2; HR, hormone receptor; SD, standard deviation, TNBC, triple-negative breast cancer.

*Tumor grade was missing for one patient.

†Excludes in situ cancers and nonmelanoma skin cancers.

‡Ten patients were missing family history information. These patients were excluded from analysis. If age at diagnosis was unavailable, it was conservatively considered to be older than 50 years.

a breast cancer predisposition gene and 52 (10.7%) in a gene associated with breast or ovarian cancer risk. Eight of the 10 CHEK2 mutations identified were 1100delC. No mutations in

NOTE. No mutations were identified in the following genes: BARD1; CDH1; STK11; TP53; APC; BMPR1A; CDK4; CDKN2A p14; CDKN2A p16; EPCAM; MLH1; MSH2; MUTYH (biallellic); and SMAD4.

*One patient had deleterious mutations in both BRCA2 and ATM. Another patient had deleterious mutations in both BRCA1 and PMS2. Another patient had deleterious mutations in both ATM and CHEK2.

†Eight of 10 CHEK2 mutations were 1100delC (Appendix [Table A1\)](#page-11-0).

BARD1, CDH1, STK11, TP53, APC, BMPR1A, CDK4, CDKN2A p14, CDKN2A p16, EPCAM, MLH1, MSH2, MUTYH (biallelic), or SMAD4 were detected. Four patients with I1307K APC variants and nine women with monoallelic MUTYH mutations were identified but not included in this analysis due to lower associated cancer risk. Specific mutations identified and associated patient characteristics are provided in Appendix [Table A1.](#page-11-0)

Of the 30 patients with a BRCA1/2 mutation, four (13.3%) had not been clinically identified after diagnosis but did meet National Comprehensive Cancer Network (NCCN) criteria for BRCA1/2 testing.

Variants of Uncertain Significance

At least one variant of uncertain significance (VUS) was identified in 162 (33.2%) women, with as many as three variants found per patient. Fifteen patients with a VUS also had a deleterious mutation. All VUSs identified are listed in Appendix [Table A2](#page-13-0) (online only).

Predictors of Deleterious Mutations

Age. For BRCA1/2, the prevalence of deleterious mutations decreased with age at breast cancer diagnosis, with a frequency of 12.2%, 3.0%, and 1.8% for women diagnosed at age \leq 45 years, 46 to 60 years, and older than 60 years, respectively ([Table 4](#page-4-0)). In contrast, for these same age groups, the frequency of mutations in other genes related to breast cancer ranged from 3.7% to 4.4%, irrespective of age at diagnosis ([Table 4](#page-4-0)).

Breast cancer subtype. [Table 5](#page-4-0) illustrates the prevalence of deleterious mutations according to breast cancer subtype. The highest prevalence of BRCA1/2 or of any mutations was in women with TNBC. Among 87 women with TNBC, 15 (17.2%) had a

deleterious germline mutation, with 12 of these (13.8%) in BRCA1/2 (11 BRCA1, 1 BRCA2). Two (2.3%) women had a mutation in another breast cancer predisposition gene (one each in BRIP1 and NBN) and one (1.1%) in RAD51D. Among 301 women with ERpositive HER2-negative breast cancer, 26 (8.6%) had at least one mutation, with 15 (5.0%) in BRCA1/2 (5 BRCA1, 10 BRCA2) and 11 (3.7%) in another breast/ovarian cancer predisposition gene including CHEK2 ($n = 4$), ATM ($n = 3$), and one each in PALB2, BRIP1, PTEN, RAD51C, and MSH6. One woman had mutations in both ATM and CHEK2. Among 37 women with ER-negative HER2 positive disease, two (5.4%) had a BRCA1 mutation and two (5.4%) had a CHEK2 mutation. Eleven percent of 63 women with ERpositive HER2-positive breast cancer had a mutation, one (1.6%) in BRCA2 (also in ATM) and six (9.5%) in other breast cancer predisposition genes including *CHEK2* ($n = 4$) and *BRIP1* ($n = 2$).

All predictors. Factors that significantly predicted for a BRCA1/2 mutation included younger age at breast cancer diagnosis $(P < .01)$; Ashkenazi Jewish heritage $(P < .01)$; TNBC $(P = .01)$; tumor histologic grade 3 ($P < .01$); and family history of breast cancer diagnosed at age younger than 50 years, male breast cancer, or ovarian cancer ($P < .01$; [Table 6\)](#page-5-0). No factors predicted for a mutation in other breast cancer predisposition genes when these genes were analyzed as a single group.

DISCUSSION

To our knowledge, this is the first study of the frequency of germline mutations in BRCA1/2 and other breast cancer predisposition genes retrospectively done in a prospectively collected, sequential series of patients with breast cancer who consented to DNA banking for clinical research. Among 488 patients, we found that 6.1% had a BRCA1/2 mutation (18.4% among Ashkenazi Jewish and 5.1% among non-Ashkenazi) and an additional 3.9% had a mutation in another breast cancer predisposition gene. In addition, four (0.8%) patients had a mutation in a gene linked to ovarian cancer, RAD51C, RAD51D, or a Lynch syndrome gene, one of whom also had a BRCA1 mutation. In total, 10.7% of patients had a deleterious mutation in at least one cancer predisposition gene. The only other report of germline mutations among unselected patients with breast cancer is from The Cancer Genome Atlas, which found that among 507 cases, 5.5% had a germline BRCA1/2 mutation and 4.3% had a mutation in another cancer predisposition gene. The mutation distribution was almost identical to that found in our study. 28 Likewise, the prevalence of mutations in non-BRCA1/2 predisposition genes was 3.7% in more than $1,800$ TNBC cases^{[17](#page-7-0)} and 4.5% in 289 African American women with breast cancer.^{[16](#page-7-0)}

Previous studies of multigene panel testing in patients with breast cancer, who were identified through high-risk clinics, have also consistently reported a prevalence of germline mutations in non-BRCA1/2 breast cancer predisposition genes of approximately 4%, with up to an additional 1% in other cancer susceptibility genes if heterozygous MUTYH mutations are excluded.^{[18](#page-7-0)-[21](#page-7-0)} The prevalence of mutations in moderate-penetrance breast cancer genes seems to be similar in this sequential series of breast cancer cases and in cases from high-risk clinics. Predictive factors for BRCA1/2 and other high-penetrance genes that lead to referral to

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NOTE. No mutations were identified in the following genes: BARD1; CDH1; STK11; TP53; APC; BMPR1A; CDK4; CDKN2A p14; CDKN2A p16; EPCAM; MLH1; MSH2; MUTYH (biallelic); and SMAD4.

Abbreviation: DM, deleterious mutation.

*One patient diagnosed in the \leq 45 years old group had deleterious mutations in both BRCA2 and ATM. Another patient diagnosed in the \leq 45 years old group had deleterious mutations in both BRCA1 and PMS2. Another patient diagnosed in the \leq 45 years old group had deleterious mutations in both ATM and CHEK2.

high-risk clinics do not seem to predict for mutations in moderatepenetrance genes. For example, as expected, the frequency of BRCA1/2 mutations decreased with increasing age at breast cancer diagnosis. However, the frequency of deleterious mutations in non-BRCA1/2 predisposition genes is independent of age at diagnosis

(Table 4). As a result, among women diagnosed with breast cancer between ages 46 and 60 years in our cohort, more than half the mutations are in genes other than BRCA1/2. Among women diagnosed after 60 years of age, 6.4% had a deleterious mutation, with almost three-fourths in genes other than BRCA1/2.

NOTE. No mutations were identified in the following genes: BARD1; CDH1; STK11; TP53; APC; BMPR1A; CDK4; CDKN2A p14; CDKN2A p16; EPCAM; MLH1; MSH2; MUTYH (biallelic); and SMAD4.

Abbreviations: HER2, human epidermal growth factor receptor 2; HR, hormone receptor; TNBC, triple-negative breast cancer.

*One HR-positive/HER2-positive patient had deleterious mutations in both BRCA2 and ATM. One TNBC patient had deleterious mutations in both BRCA1 and PMS2. One HR-positive/HER2-negative patient had deleterious mutations in both ATM and CHEK2.

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Abbreviations: BC, breast cancer; HER2, human epidermal growth factor receptor 2; HR, hormone receptor; SD, standard deviation; TNBC, triple-negative breast cancer.

*Three patients with mutations not associated with breast cancer were not included in this analysis.

†One patient with BRCA2 and ATM mutations is included with BRCA1/2 Mutation (not with Other BC Mutation).

‡Excludes in situ cancers and nonmelanoma skin cancers.

§Tumor grade was missing for one patient without a mutation.
∥Ten patients were missing family history information. These patients were excluded from analysis. If age at diagnosis was unavailable, it was conservatively
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As expected, factors known to enrich for BRCA1/2 mutations (such as Ashkenazi heritage, TNBC subtype, and strong family history of breast or ovarian cancer) predicted for BRCA1/2 mutations in our study. However, these factors were not significantly associated with mutations in other breast cancer predisposition genes when these genes were analyzed as a single group. We were unable to identify any factors that predicted for a mutation in non-BRCA1/2 breast cancer genes. Larger cohorts of women with mutations in each gene are required to identify genespecific predictors. For example, the prevalence of ER-positive breast cancer is higher among germline CHEK2 1100delC mutation carriers than noncarriers.²

We found that among non-BRCA1/2 genes, mutations in CHEK2 were most common. This is consistent with results from most studies that have evaluated multigene panels in breast cancer cohorts not enriched for TNBC.^{[19,20](#page-7-0),[30](#page-7-0)} The clinical significance of mutations in moderate-risk breast cancer genes such as CHEK2, ATM, and NBN is still being evaluated. An increased risk of contralateral breast cancer has been associated with germline mutations in PALB2 and CHEK2, suggesting that subsequent screening with breast magnetic resonance imaging may be indi-cated in these patients.^{[29,31](#page-7-0)-[33](#page-7-0)} We identified one mutation in PTEN, a high-penetrance cancer predisposition gene associated with Cowden syndrome and an increased risk of breast, thyroid, endometrial, and other cancers. This result would allow initiation of appropriate surveillance and risk-reduction strategies.

We also identified six mutations in three genes (BRIP1, RAD51C, and RAD51D) that are associated with a six- to 12-fold increased risk of ovarian cancer.^{[10](#page-7-0)-[13,15](#page-7-0)} Mutations were found even in women with breast cancer diagnosed after 60 years of age. Only two of the six mutation carriers had a personal or family history of ovarian cancer. It seems that identification of mutations in more moderate-penetrance ovarian cancer susceptibility genes will require testing of families with less notable cancer histories.^{[34](#page-7-0)} Finally, we found two mutations in genes for Lynch syndrome. Identification of such mutations can lead to increased surveillance for and identification of colorectal, endometrial, ovarian, and other cancers in these patients with breast cancer and their relatives.[18](#page-7-0)

The use of multigene panels for assessment of cancer susceptibility has been increasing rapidly in clinical practice. In addition to including high-penetrance genes with established clinical utility, these panels contain genes for which clinical validity or significance is less certain at this time.^{[35](#page-7-0)} Proper interpretation of results is critical so that appropriate recommendations for risk management are offered. This presents challenges for clinicians, who often lack genetic training, and their patients, who face decisions about screening and prevention strategies.

Whereas the significance of mutations in several of the non-BRCA1/2 predisposition genes is still being studied, the benefit of identifying BRCA1/2 mutations is well established. In addition to cancer prevention strategies, BRCA-associated cancers have a greater response to therapies such as poly-(ADP) ribose poly-merase inhibitors and platinum agents than sporadic cancers.^{[36](#page-7-0)-[39](#page-7-0)} Studying a sequential series of patients with breast cancer allowed us to evaluate how often BRCA mutations might be missed in clinical practice, at least in an academic setting. Only four (13.3%) of the BRCA1/2 mutation carriers were first identified through this study, and all 30 carriers fulfilled NCCN 2015 genetic testing criteria. Thus, clinicians seem to be recognizing patients with breast cancer who are appropriate for BRCA1/2 testing, and the NCCN criteria seem to perform well. Increased clinician education about testing criteria might decrease the frequency of unidentified carriers even further.

Consistent with previous studies, we found the highest prevalence of BRCA1/2 mutations among cases with TNBC. For patients with ER-negative or ER-positive breast cancer, we found the frequency of BRCA mutations to be lower if the tumor also overexpressed HER2, consistent with previous reports $(Table 5).⁴⁰⁻⁴³$ $(Table 5).⁴⁰⁻⁴³$ $(Table 5).⁴⁰⁻⁴³$ $(Table 5).⁴⁰⁻⁴³$ $(Table 5).⁴⁰⁻⁴³$

Approximately one-third of patients had at least one VUS, as has been reported in other series evaluating NGS panels.^{[18](#page-7-0)} Most of these variants will eventually be reclassified, primarily as benign, but some will likely be deleterious.^{[27](#page-7-0)[,44](#page-8-0)} VUSs should not be used to make clinical decisions.

Our study has limitations. Cases were ascertained from an academic center and may not reflect the breast cancer population or prevalence of mutations in the community. In our cohort, the median age at diagnosis was 50 years compared with 61 years in the United States.^{[45](#page-8-0)} Likewise, compared with the general population, a higher proportion of individuals (7.8%) were of Ashkenazi descent. However, although these factors may increase the prevalence of BRCA1/2 mutations, our results show that they do not seem to increase the frequency of mutations in moderatepenetrance breast cancer susceptibility genes. Indeed, one report found a lower frequency of non-BRCA1/2 mutations in the Ashkenazi population.^{[19](#page-7-0)} Finally, only 11% of women in the study were nonwhite, limiting the generalizability of our findings in more diverse populations. Large population-based studies are needed to establish the true frequency of mutations in these genes.

In conclusion, this is the largest prospective study to date to assess the prevalence of mutations in cancer susceptibility genes among a sequential series of breast cancer cases seen at an academic institution and not otherwise selected for age, family history, ethnicity, or breast cancer subtype. We identified a mutation in 10.7% of patients, 6.1% in BRCA1/2 (5.1% in patients of non-Ashkenazi descent) and 4.6% in other breast/ovarian cancer predisposition genes. The prevalence of mutations in genes other than BRCA1/2 seems to be relatively consistent regardless of breast cancer population tested, when these genes are assessed as a single group. Although clinicians seem to be identifying the majority of patients with breast cancer who have BRCA1/2 mutations, the lack of predictive factors for mutations in other breast/ovarian cancer predisposition genes presents a challenge for identifying these carriers. Until better predictors emerge, it will be necessary to continue casting a wider net for identification of mutations in non-BRCA1/2 genes. This is despite the unclear clinical significance at present for several genes that are included in many commercially available, broad gene panels.

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Disclosures provided by the authors are available with this article at www.jco.org.

AUTHOR CONTRIBUTIONS

Conception and design: Nadine Tung, Nancy U. Lin, Brian A. Allen, Anne-Renee Hartman, Eric P. Winer, Judy E. Garber Financial support: Richard J. Wenstrup

Provision of study materials or patients: Nancy U. Lin, Eric P. Winer, Judy E. Garber

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GLOSSARY TERMS

germline mutation: an inherited variation in the lineage of germ cells. Germline mutations can be passed on to offspring.

NextGen Sequencing: a non-Sanger rapid DNA sequencing method that can be done with greater speed, developed after the first methodologic articles describing relatively rapid DNA sequencing produced by Sanger et al (1977).

penetrance: the likelihood that a given gene mutation will produce disease. This likelihood is calculated by examining the proportion of people with the particular genetic mutation that show symptoms of disease.

triple-negative breast cancer (TNBC): breast tumors that are negative for estrogen and progesterone receptor expression and that also underexpress HER-neu.

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Frequency of Germline Mutations in 25 Cancer Susceptibility Genes in a Sequential Series of Patients With Breast Cancer

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APC NM_000038.5 ATM NM_000051.3 BARD1 NM_000465.3 BMPR1A
NM_004329.2 BRCA1 NM_007294.3 BRCA2 NM_000059.3 BRIP1
NM_032043.2 CDH1 NM_004360.3 CDK4 NM_000075.3 CHEK2 NM_007194.3
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