

Pancreatic cancer from bench to bedside: molecular pathways and treatment options

Christoforos Kosmidis¹, Konstantinos Sapalidis¹, Efstathios Kotidis¹, Nikolaos Mixalopoulos¹, Paul Zarogoulidis², Drosos Tsavlis², Sofia Baka³, Yan-Gao Man⁴, John Kanellos¹

¹3rd Department of Surgery, AUTH, University General Hospital of Thessaloniki AHEPA, Thessaloniki, Greece; ²Pulmonary Department-Oncology Unit, "G. Papanikolaou" General Hospital, Aristotle University of Thessaloniki, Thessaloniki, Greece; ³Department of Oncology, Interbalkan Medical Center of Thessaloniki, Thessaloniki, Greece; ⁴Research Laboratory and International Collaboration, Bon Secours Cancer Institute, VA, USA

Contributions: (I) Conception and design: All authors; (II) Administrative support: All authors; (III) Provision of study materials or patients: All authors; (IV) Collection and assembly of data: All authors; (V) Data analysis and interpretation: All authors; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: Paul Zarogoulidis, MD, PhD. Pulmonary Department-Oncology Unit, "G. Papanikolaou" General Hospital, Aristotle University of Thessaloniki, Thessaloniki, Greece. Email: pzarog@hotmail.com.

Abstract: In the last forty years the pancreatic cancer treatment has made advances, however; still novel drugs are needed. It is known that the five year survival rate remains around 5%. The best treatment option still remains surgery, if patients are diagnosed early. In the last decade the biology of pancreatic cancer has been vastly explored and novel agents such as; tyrosine kinase agents, or vaccines have been added as a treatment perspective. The big challenge is now to translate this knowledge in better outcomes for patients. In this current review we will present information from pancreatic cancer diagnosis to molecular pathways and treatment options; current and future.

Keywords: Pancreatic cancer (PC); molecular pathways; targeted treatment

Submitted Mar 19, 2016. Accepted for publication Mar 24, 2016.

doi: 10.21037/atm.2016.05.11

View this article at: <http://dx.doi.org/10.21037/atm.2016.05.11>

Introduction

It is known that pancreatic cancer (PC) is a lethal disease with a 5-year survival rate of 1.2–6% encompassing the pancreatic ductal adenocarcinoma that accounts for 85–95% of all pancreatic malignancies (1,2). Current data indicate that it is the seventh and fourth leading cause of cancer-related death worldwide. In the near future by 2030 it will become the second leading cause in the United States (3). During the past 30 years the incidence of pancreatic cancer has increased in developed countries, along with an increase in mortality rates. Increased mortality rates are observed in both sexes (4-7). Current statistical data indicate that there is an increased need for efficient and well-tolerated treatment options in pancreatic cancer. Below we will present current treatment options and the molecular pathways currently investigated. Also, we will provide all

novel clinical trials and agents under development that may provide a rationale for future investigations.

Clinical presentation, signs, and symptoms

Unfortunately, pancreatic cancer is usually diagnosed at advanced stage. The most common early disease symptoms are: weight loss, back pain, abdominal pain, nausea and vomiting, dyspepsia, new-onset diabetes, bloating, changes in bowel habit, lethargy, pruritus, shoulder pain, and jaundice (8). In a previous published study, lethargy [OR 1.42 (1.25–1.62)], back pain [odds ratio (OR) 1.33; (95% CI 1.18–1.49)], and new-onset diabetes [OR 2.46 (2.16–2.80)] were observed features of pancreatic cancers (8). The following five symptoms were observed to occur more than 6 months before diagnosis: dysphagia, back pain, changes in bowel habit, shoulder pain, and lethargy (8). In

a previous published study it was observed that lethargy, and depression as the first symptoms in about 38–45% of patients with pancreatic cancer (9). On another occasion a recent review reported nine presenting symptoms in patients with advanced pancreatic cancer (10). In this study abdominal pain and diabetes, were frequently reported in advanced pancreatic cancer (11). Usually in several studies up to 25% of the patients report upper abdominal discomfort up to six months prior to diagnosis (12).

Current management of pancreatic cancer

Surgical resection

Only 20% of patients will be diagnosed with early pancreatic cancer and for these patients, surgical resection is the treatment of choice (13,14). However; after complete resection adjuvant chemotherapy with gemcitabine or 5-fluorouracil and/or chemo-radiation has to follow and still prognosis remains disappointing (15). In the past years randomized controlled trials (16-18) have demonstrated increased overall survival (OS) with adjuvant therapy and this observation is considered the most important advancement in the treatment of pancreatic cancer (13). There are also cases where neoadjuvant chemotherapy is proposed to improve surgical margins in order for the patient to become operable (19-22) (Table 1).

Locally advanced and metastatic disease

The chemotherapy agent gemcitabine or gemcitabine-based combination chemotherapy has been vastly used and is the most acceptable first line treatment for advanced pancreatic cancer. However, still the median survival (MS) remains approximately 9 mo (13,23,24). In a recent study “FOLFIRINOX” (folinic acid, 5-fluorouracil, irinotecan, oxaliplatin) an advantage in survival and quality of life was observed when compared to gemcitabine alone. In specific; this combination significantly improved the OS, progression-free survival in pancreatic cancer patients (25). Almost the same effectiveness was observed with nab-paclitaxel plus gemcitabine (26). It has been observed that almost 10% of patients who have received have survived two years, which is a rare event in advanced disease (27). However, an issue that has to be taken under consideration is the fact that these regimens have increased toxicities and therefore they have to be administered to patients with good performance status (25,26). The care of patients with poor performance

status or metastatic disease remains palliative, since gemcitabine based therapies have limited efficacy, however, local therapies can be considered (28).

Targeted therapy

Targeted treatment based on the genome of the tumor has revolutionized current cancer treatment. Pharmacogenetics seems to be one of the treatment options for pancreatic cancer (29). Unfortunately genetically heterogeneity has been observed in pancreatic cancer (30), however; there are several targeted therapies as treatment options, such as; small molecule inhibitors and monoclonal antibodies which inhibit constitutively-active cell surface signaling molecules. Currently results of phase I–III clinical trials presented by Seicean *et al.* (31) did not present favorable results due to the resistance from *KRAS2* mutations and upregulation of alternate signaling pathways (13,32). Until now only the tyrosine kinase inhibitor of epidermal growth factor receptor, “erlotinib”, has been approved agent, in combination with gemcitabine, and offers increase in survival of two weeks (33) (Figure 1).

Cell metabolism in pancreatic cancer

In order for normal cells to alter their proliferation rate several metabolic pathways are reprogrammed (34). The increased metabolism of cancer cells is one of the most important features of this disease that currently is being targeted as treatment (35). Cancer cells adapt their metabolic needs based on their environment. Recent studies have enlightened several metabolic routes and signaling pathways that control tumor progression (36). The most important pathways are considered the following: glutamine regulatory enzymes, Ras signaling, lipid metabolism and autophagy (37-39). Pancreatic cancer as most cancer types are characterized by a high increase in glucose uptake and metabolism (40) and high glycolytic rates the so called “Warburg effect”. Moreover; it is known that cancer cells compared to normal cells have specific metabolic dependencies, they have increased need of the amino acid glutamine (41). Kras is responsible for reprogramming glutamine metabolism in PDAC (42). Targeting the Warburg effect has shown that the p53 status of PDAC determined the response to inhibitors of the enzyme lactate dehydrogenase-A (43). Recently it has been investigated and observed that cancer cells not only fuel from glucose and glutamine but also these cells utilize amino acids as well

Table 1 Pancreatic cancer staging (criteria defining tumour resectability status according to NCCN guidelines, version 2.2015)

Resectability status	arterial involvement	venous involvement
Resectable		
No contact with coeliac axis (CA), superior mesenteric artery (SMA), or common hepatic artery (CHA)		
No contact with the superior mesenteric vein (SMV), or portal vein (PV) or $>180^\circ$ contact without vein contour irregularity		
Borderline resectable		
Pancreatic head/uncinate process		
Solid tumour in contact with CHA without extension to CA or hepatic artery bifurcation allowing for safe and complete resection and reconstruction		
Solid tumour contact with the SMA of $>180^\circ$		
Presence of variant arterial anatomy (ex: accessory right hepatic artery.) and the presence and degree of tumour contact should be noted if present as it may affect surgical planning		
Pancreatic body/tail		
Solid tumour contact $>180^\circ$ with the CA		
Solid tumour contact $>180^\circ$ with the CA without involvement of the aorta and with intact and uninvolved gastroduodenal artery (some members prefer this criteria to be in the unresectable category)		
Solid tumour in contact $>180^\circ$ with the SMV or PV, or in contact $>180^\circ$ with contour irregularity of the vein or thrombosis of the vein but with suitable vessels proximal and distal to the site of involvement allowing for safe and complete resection and vein reconstruction		
Solid tumour contact with the inferior vena cava (IVC)		
Unresectable $>180^\circ$ distant metastasis		
Head/uncinate process		
Solid tumour contact with SMA $>180^\circ$		
Solid tumour contact with the CA $>180^\circ$		
Solid tumour contact with the first jejunal SMA branch		
Body and tail		
Solid tumour contact $>180^\circ$ with the SMA or CA		
Solid tumour contact with the CA and aortic involvement		
Head/uncinate process		
Unreconstructible SMV/PV due to tumor involvement or occlusion (can be due to tumor or bland thrombus)		
Contact with most of the proximal draining jejunal branch into the SMV		
Body and tail		
Unreconstructible SMV/PV due to tumour involvement or occlusion (can be due to tumour burden)		

Abbreviations: CA, coeliac axis; SMA, superior mesenteric artery; CHA, common hepatic artery; SMV, superior mesenteric vein; PV, portal vein; IVC, inferior vena cava.

as lipids and protein (44). It has been observed that early breakdown of proteins and subsequent increase in plasma levels of branched-chain amino acids are early events in pancreatic cancer progression as of course observed in other cancer types (45). In a recent published work it was

suggested that mitochondrial respiration is another potential functional target to manage pancreatic cancer (46). In the study by Viale *et al.* it was revealed that tumors can relapse 2–4 months after Kras mutation withdrawal. However, it was observed that after Kras mutation extinction, they found a

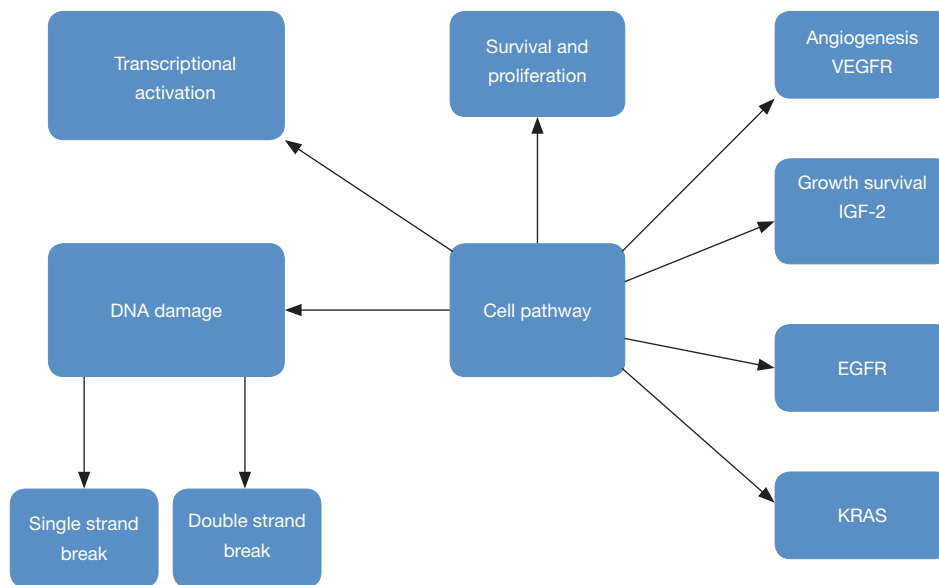


Figure 1 Molecular pathways and treatment. VEGFR, vascular endothelial growth factor receptor; IGF-2, insulin like growth factor-2; EGFR, epidermal growth factor receptor; KRAS, V-Ki-ras2 Kirsten rat sarcoma viral oncogene homolog, is a GTPase that in humans is encoded by the KRAS.

sub-population of surviving cells that which did not depend on Kras and anabolic glucose metabolism as the primary tumor but they had a strong reliance on mitochondrial energy production (46). Based on this observation it is suggested that in order to eliminate pancreatic tumors and prevent disease relapse we should target both populations, with inhibitors of glucose and mitochondrial metabolism. Mitochondrial metabolism regulation will be the next treatment option for pancreatic cancer. In pancreatic cancer high levels of autophagy are observed due to a transcriptional program that regulates lysosome biogenesis and nutrient scavenging (47,48). Blockage of autophagy in pancreatic cancer was achieved with genetic inhibition and drug administration both *in vitro* and *in vivo* (48,49). However, it has been observed that tumor progression in pancreatic cancer is genotype-dependent and autophagy blockage is not the best treatment option (50). Nevertheless, it is still not established whether p53 status affects PDAC response to autophagy inhibition (51). In pancreatic cancer micropinocytosis of protein and lysophospholipids as source of fatty acids has been observed as a food source. Macropinocytosis is a process where extracellular fluids through specialized vesicles named macropinosomes are inserted into the cells. It has been observed that Ras proteins regulate this process by which cancer cells

internalize extracellular proteins, glutamine still remains the most valuable amino acid (52,53). Moreover; Ras-dependent mechanism of scavenging of fatty acid has been shown in pancreatic cancer cells (54). It has been observed that due to the high demand of fatty acids by cancer cells, these cells have found mechanisms of fatty acid supply, such as the uptake of extracellular lipids, in order to face cancer growth requirements. Until now there was an increasing interest in glucose metabolism and aerobic glycolysis, however; it is becoming clearly evident that the alteration of lipid metabolism is critical for cancer cell metabolism (55). Enzymes involved in lipogenesis and lipolysis have been found overexpressed in pancreatic cancer, in particular fatty acid synthase (FASN) that catalyses the final steps of fatty acid synthesis and ATP citrate lyase (39). It has been observed that elevated levels of FASN protein in cancer cells and in serum of pancreatic cancer patients are associated with poor prognosis (39). In cancer cells hyper activation of lipogenesis is observed and characterized by an increase in the degree of lipid saturation compared with non-lipogenic tumors (56). Therefore, the rise in saturated and monounsaturated lipids in cancer cell membranes increases the resistance to oxidative stress, since polyunsaturated lipids are more susceptible to lipid peroxidation. Pancreatic cancer cells use alternative routes to lipogenesis in order to

obtain fatty acids, primarily through uptake of extracellular lipids derived from diet, liver synthesis or released by the adipose tissue. Moreover; current evidences suggest that pancreatic cancer is highly dependent on cholesterol (57). It has been observed that the elevated requirement of cholesterol by pancreatic cancer cells can be supplied by *de novo* synthesis, receptor mediated uptake of cholesterol-rich low-density-lipoproteins (LDL) by the LDL receptor (LDLR), or by hydrolysis of cholesteryl esters (CE) that accumulate in specific lipid droplets (LDs) that are thought to act as storage of triacylglycerol and CE (57). It has been observed that cholesterol uptake, is activated in tumors (57). Interestingly an increase in the amount of cholesterol and overexpression of the low-density lipoprotein receptor (LDLR) in pancreatic tumor cells is observed (57). Based on this observation LDLR has been proposed as a novel metabolic target to limit pancreatic cancer progression. It has been previously observed in pancreatic tumor cells that shRNA silencing of LDLR reduces considerably cholesterol uptake and alters its distribution, decreases tumor cell proliferation, and limits the activation of ERK1/2 survival pathway. Moreover; with cholesterol uptake blockage the effect of chemotherapy on pancreatic cancer regression to chemotherapeutic agents is increased. It has been observed that high LDLR expression in pancreatic cancer is not associated with tumor stage, however; it has been shown to correlate to a higher risk of disease recurrence. Based on these findings it is suggested that pancreatic cancer cells are highly dependent on cholesterol uptake, and that either this process or LDLR is a promising metabolic target to use in combination with chemotherapy. Lipid metabolism in pancreatic cancer is that pancreatic cancer cells do not rely solely in *de novo* synthesis of lipids but also utilize circulating and diet derived lipids. Therefore it can be said that high dietary intake and obesity is a risk factor in pancreatic cancer.

Pancreatic cancer and its precursors

Pancreatic cancer is usually diagnosed at late disease and therefore it has a poor prognosis with a mortality rate almost equaling to the incidence rate (58,59). Therefore tools for early disease are in need. Moreover; deciphering the factors important for pancreatic cancer progression will help to identify novel treatment options as mentioned in the previous paragraphs. It has been observed that pancreatic cancer can develop from three established precursor lesions (60): (I) intraductal papillary mucinous neoplasm (IPMN); (II) pancreatic intraepithelial neoplasia (PanIN)

and the cystic lesions; and (III) mucinous cystic neoplasm (MCN). The majority of pancreatic cancer is thought to arise from PanINs and less frequently from IPMN, whereas MCNs are rare (61). There is indirect evidence for PanINs as precursors for pancreatic cancer, which is largely based on the fact that pancreatic cancer is often associated with advanced PanIN, and both share common tumor promoting genetic alterations. On the other hand, cystic lesions can directly be identified as the origin for PDA on histological examination and imaging techniques such as MRI scan or endoscopic ultrasound (EUS). Due to more frequent and better diagnostic imaging as well as physicians' awareness, IPMN lesions are increasingly identified in the pancreas, the ideal management of these patients is still an ongoing debate (62). It has been observed that pancreatic cancer that is associated with IPMNs has a much more favorable prognosis than pancreatic cancer that arises from PanINs (63-65). A possible explanation could be different genetic mutations during evolution of pancreatic cancer from its precursors. It has been observed that *KRAS* mutation occurs nearly universally during PanIN initiation (66), *KRAS* is less frequently mutated in IPMNs (67,68). IPMNs but not PanINs frequently harbor mutations in *GNAS* and *RNF43* (69-71). Moreover; common genetic alterations in both precursors are found in *TP53* and *CDKN2A* [reviewed by Xiao (72) and Gnoni *et al.* (61)]. Furthermore; the different biology of PanIN and IPMN-associated with pancreatic cancer could be the different cellular origin of the precursors. Recent evidence from genetically engineered mouse models (GEMM) revealed that the cellular origin of PanINs and IPMNs might be different (73-75).

Targeted therapy in PDAC

Epidermal growth factor receptor (EGFR) pathway inhibitors

Epidermal growth factor receptor (EGFR) is a transmembrane receptor member of the ErbB family with a tyrosine kinase domain that is activated by many ligands including epidermal growth factor (EGF), amphiregulin, epiregulin, tumor growth factor- α (TGF- α), heparin-binding EGF, betacellulin and neuregulin. EGFR is involved in cell cycle regulation, adhesion, differentiation and cell survival, through activation of the Ras/MAP kinase, Janus kinase/Stat, phospholipase C/protein kinase C and phosphatidylinositol 3'-kinase (PI3K)/Akt pathways. In previous studies it was observed that EGFR is overexpressed

in up to 90% of pancreatic cancer samples. Based on this findings tyrosine kinase inhibitor targeting EGFR have been considered a promising therapeutic agent (76). Erlotinib is a tyrosine kinase inhibitor (TKI) molecule that competes with ATP for binding to the kinase domain, thereby blocking downstream signal transduction of EGFR. A large phase III trial, enrolling 569 chemotherapy naïve patients with locally advanced or metastatic pancreatic adenocarcinoma randomized to receive gemcitabine plus placebo or gemcitabine plus erlotinib 100–150 mg daily was performed. The median overall survival (mOS) and progression free survival (PFS) were modestly, but statistically significantly, improved in the combination arm, 6.24 vs. 5.91 mo (P=0.038) and 3.75 vs. 3.55 mo (P=0.004), respectively (33). Both EGFR status nor KRAS status analysed in the subgroup of patients treated with erlotinib was associated with survival benefit in patients receiving the combination schedule (77).

For now erlotinib has been approved by the FDA in combination with gemcitabine as a first-line treatment for advanced pancreatic adenocarcinoma. Cetuximab is a monoclonal antibody binding the extracellular domain of epidermal growth factor receptor. Unfortunately the encouraging results in a phase I trial, were not observed again in subsequent studies (gemcitabine-based chemotherapy) failed to demonstrate any survival benefit (78,79). Gefitinib; another tyrosine kinase inhibitor of ATP binding to the intracellular kinase domain of EGFR, in combination with gemcitabine in inoperable or metastatic pancreatic cancer patients was administered in a phase II trial. The combination demonstrated promising activity with a mOS and PFS in the combination arm of 7.3 and 4.1 mo, respectively, however; other evidence supporting a role of gefitinib in pancreatic adenocarcinoma treatment is lacking (80). HER-2, which is another ErbB family of transmembrane tyrosine kinase receptors which has been observed overexpressed in 11% of pancreatic adenocarcinoma cases. It has been observed that HER2-positive status is correlated with shorter survival (81). Trastuzumab plus gemcitabine was tested in 34 metastatic pancreatic cancer patients with HER-2 overexpression as determined by immunohistochemistry, and partial responses were observed in 6% of cases (82). Harder *et al.* (83) in a multicentre phase II study, investigated the efficacy and toxicity of the HER2 antibody, trastuzumab, plus capecitabine in patients with pancreatic cancer and HER2 overexpression, however; this study did not present favourable results to either PFS or OS compared with standard chemotherapy. Lapatinib another HER-2 inhibitor was approved by

the FDA and clinical trials have been initiated to test the effect of this inhibitor combined with chemotherapy in pancreatic carcinoma. Lapatinib was tested in combination with capecitabine as a second-line treatment in advanced pancreatic cancer with promising initial results. However; more studies are needed to evaluate the real effectiveness and role of this molecule in the treatment of pancreatic adenocarcinoma (84). Nimotuzumab, another anti-EGFR monoclonal antibody, showed promising results (85). In the study by Strumberg *et al.* (86) a phase II trial showed PFS after 1 year of 10.3% and median overall survival of 18.1 wk with a tolerable toxicity profile. Currently afatinib; another TKI of EGFR, HER2 and HER4, is under evaluation in an ongoing phase II trial (87,88).

The KRAS pathway and downstream signalling cascade inhibitors

Current information regarding KRAS mutations indicated that KRAS activating mutations are present in 70% to 90% of pancreatic cancer. K-Ras is a GTPase protein belonging to the Ras protein family. This protein has oncogenic activity, and promotes activation of proliferation and inhibits apoptosis through the RAF/MEK/ERK and PIK3/AKT pathways. It is known that K-Ras pathway is very difficult to target, and currently there are no inhibitors that can actually block this pathway in clinical practice (89). In a preclinical study it was observed that that farnesylation is an important post-translational modification required for Ras activation, allowing the protein to be attached to the plasma membrane for signal transduction (90). Although farnesyl-transferase inhibitors, particularly tipifarnib, presented anti-proliferative activity in pancreatic tumour cell lines, it failed to improve overall survival either as a single agent or in combination with gemcitabine in a phase III trial (91,92). Therefore it is suggested for now to block targets downstream of KRAS, such as the protein kinase MEK. Therefore selumetinib an oral small molecule that inhibits MEK1/2 was produced. This agent was administered in a phase II trial, where patients were randomized to receive single-agent capecitabine or selumetinib as a second-line treatment for advanced pancreatic cancer. The selumetinib arm showed a median overall survival of 5.4 vs. 5.0 mo in the capecitabine arm, but this result was not statistically significant (93). Trametinib, another MEK1/2 inhibitor, has been tested in pancreatic cancer in combination with gemcitabine against a regimen of gemcitabine plus placebo in a phase II randomized multicentre study. However; again no significant advantages

were demonstrated in terms of overall survival or PFS (94). Rigosertib, a first-in-class Ras mimetic and small molecule inhibitor of multiple signalling pathways, including polo-like kinase 1 and phosphoinositide 3-kinase (PI3K), was administered in combination with gemcitabine in patients with treatment-naïve metastatic pancreatic adenocarcinoma in a phase II/III randomized study. In the combination regimen no improved of survival or response was observed, as recently presented at the 2015 ASCO Annual Meeting (95). Research in this field is continuing.

IGFR pathway inhibitors

Insulin like growth factor 1 receptor (IGFR-1) is another possible target for pancreatic cancer, which is highly expressed in pancreatic cells. It has been observed that upon ligand binding, it activates several pathways involved in cell proliferation and cell survival such as the PIK3/AKT pathway (96). Moreover, other agents such as; monoclonal antibodies against IGFR (cixutumumab, ganitumab) were evaluated in pancreatic cancer treatment, however; they failed to show a statically significant survival benefit (97). In a phase III trial assessing ganitumab in combination with gemcitabine, the study closed early based on a pre-planned futility analysis. The median overall survival was 7.1 mo in the maximum dose ganitumab arm *vs.* 7.2 mo in the placebo arm (HR 0.97, P=0.397) (98).

Angiogenesis pathway inhibitors

Neo-angiogenesis is known to be an essential metastatization and tumor progression mechanism. Vascular endothelial growth factor (VEGF) is known to stimulate proliferation of endothelial cells and it has been observed to be overexpressed in human pancreatic cancer. However; current data indicate that neo-angiogenesis inhibitors, particularly VEGF inhibitors, have not presented favorable overall survival in combination with gemcitabine in advanced pancreatic cancer. In two phase III trials that tested the efficacy of bevacizumab in association with gemcitabine alone, or gemcitabine plus erlotinib, did not confirm previous findings (99,100). A new recombinant fusion protein, aflibercept which has extracellular portions of VEGFR-1 and VEGFR-2, which binds VEGF-A, VEGF-B and placental growth factors 1 and 2 thereby inhibiting VEGF-ligand-dependent signaling (101) processes, was observed to suppress tumor growth in pancreatic cell lines and xenografts. However; in a phase III study adding

aflibercept to gemcitabine did not improve OS in metastatic pancreatic cancer patients (102). Similarly sorafenib, an oral multikinase inhibitor of Raf-kinase, VEGF-R2/-R3 and PDGFR- β , and axitinib, an anti-angiogenesis agent did not present any statistically significant efficacy in advanced pancreatic adenocarcinoma (103,104). Currently there are phase II studies combining chemotherapy with promising new anti-angiogenic molecular agents, such as TL-118, which is a nonsteroidal anti-inflammatory oral medication, or necuparanib, which is re-engineered from heparin with possible anti-tumor activity. The results of these studies will be published soon (105,106).

Embryonic pathway inhibitors

The Hedgehog signaling is known to have a critical role in cell proliferation and survival during the embryonic development. In normal pancreatic cells this pathway is silenced, however; pathological activation is observed in many solid tumors. In pancreatic cancer this signaling pathway is found overexpressed. Hedgehog binds to the extracellular receptor Patched, which, in the absence of Hedgehog, suppresses activation of the G-protein-coupled receptor Smoothened and upregulates glioma associated oncogene homolog1 transcriptional activity (107). In the study by Bailey *et al.* (108) it was observed that Sonic hedgehog (SHH) was detected in precursor lesions and in pancreatic cancer tumor samples which contributed to the formation of the desmoplastic reaction. This characteristic of pancreatic cancer limits the effective delivery of anticancer agents to pancreatic cancer cells. Genetically engineered mouse models demonstrated a depletion of tumor matrix from Sonic hedgehog SHH pathway inhibition this treatment option could be the strategy in pancreatic cancer therapy (109). Currently, vismodegib (GDC-0449), an oral small-molecule inhibitor targeting Smoothened (110), is administered in an open phase II trials in combination with gemcitabine in advanced cancer, in combination with gemcitabine and nab-paclitaxel in metastatic settings. Current results provide favorable outcome (111), and as a single agent in neoadjuvant settings followed by surgery (112-114). The Smoothened inhibitor saridegib (IPI-926) was administered in combination with gemcitabine against gemcitabine plus placebo in a randomized, double-blind, placebo-controlled phase II trial. In this trial patients with metastatic disease were enrolled. The study was terminated due to early decreased of patient survival in the saridegib arm (115). Hedgehog inhibitors are

currently an active research field, and the results of several clinical trials are under way (116). Notch signaling is another embryonic pathway crucial for pancreatic organogenesis, this pathway is silenced after pancreas development, however; it is active only in a stem cell subgroup. This pathway has been observed to be upregulated pancreatic cancer and promotes tumorigenesis. Binding of Notch ligand to its receptor promotes a cascade of proteolytic cleavages, mediated by γ -secretase. The activated form ICN (intra cellular notch) forms part of a transcription complex that, after translocating to the nucleus, regulates transcription of several genes involved in proliferation and differentiation of cells. It has been observed that interacts with other pathways such as Hedgehog, KRAS and NF- κ B signaling (117,118). The selective inhibitor RO4929097 of the γ -secretase enzyme has anti-tumor activity in preclinical studies (119). In a recent phase II single-arm trial RO4929097 was assessed, enrolling 18 previously treated advanced pancreatic cancer patients. The treatment was well tolerated; the median survival was 4.1 mo, and the median progression-free survival was 1.5 mo (120). Encouraging clinical results were observed testing demcizumab, an anti-Delta-like ligand 4 antibody, plus gemcitabine and nab-paclitaxel in advanced PDAC in a phase b trial. However; more studies are in need to confirm these preliminary data (121).

Poly ADP-ribose polymerase (PARP) inhibitors

Mutations that affect breast cancer pathway components, and especially the tumor suppressor gene breast cancer-2 (*BRCA2*) gene, which is associated with hereditary predisposition to breast, ovarian and pancreatic cancer are known to promote deficiency in DNA damage repair mechanisms and also induce genome instability (122). Poly ADP-ribose polymerase (PARP) is a nuclear enzyme recruited to repair cell DNA damage, patients with defects in the homologous DNA recombination pathway may benefit from the use of PARP inhibitors. This treatment can be applied as monotherapy or in combination with radiation or other chemotherapeutic agents. Clinical trials testing poly ADP ribose polymerase (PARP) inhibitors are currently under development phase (123-125).

mTOR and PI3K/Akt pathway inhibitors

It is known that after activation, Ras can phosphorylate PI3K, which in turn activates Akt, a serine/threonine

kinase. Signal transduction by activated PI3K/Akt plays a role in tumor cell proliferation, metabolism and survival. This occurs through several downstream targets, such as; the mammalian target of rapamycin (mTOR) (126). Currently there are trials testing PI3K/AKT axis inhibitors in advanced pancreatic cancer patients after encouraging preclinical model results (127). There are several PI3K/AKT axis inhibitors being tested such as: RX-0201, an Akt antisense oligonucleotide tested in a phase II study plus gemcitabine; BKM120, a PI3K inhibitor tested in combination with the mFOLFOX-6 schedule; and BEZ235, a combined inhibitor of PI3K and mTOR assessed in a phase study in combination with the MEK inhibitor MEK162 (128-130). In the study by Wolpin *et al.* (131) everolimus, an oral mTOR inhibitor, was administered as monotherapy in 33 gemcitabine-refractory pancreatic cancer patients. The PFS and OS observed were 1.8 and 4.5 mo, respectively. In a recent published phase II study (single arm) where everolimus was tested in combination with capecitabine the median OS was 8.9 mo and PFS was 3.6 mo (132). In the recent future we are anticipating the results of a phase I/II study testing everolimus in combination with gemcitabine in advanced settings and the results of a phase II trial testing temsirolimus (mTOR inhibitor) (133,134).

Tumor stroma inhibitors

In pancreatic cancers a dynamic compartment (stroma) has been observed to be critically involved in tumor formation, progression and metastatic process. Stroma microenvironment and consisting elements are considered a treatment target in addition to previously described trials evaluating Hedgehog signaling inhibitors (135). Currently a phase II trial is recruiting patients in order to administer PEGPH20, a pegylated formulation of recombinant hyaluronidase. The study design is to enrol untreated patients with metastatic disease in order to receive a combination of PEGPH20, nab-naclitaxel and gemcitabine or a combination of nabpaclitaxel and gemcitabine (136,137). Moreover; it has been observed that inhibition of platelet derived growth factor receptor (PDGFR), could be a possible treatment target. This receptor is expressed in stromal cells and has a critical role in recruiting pericytes and in interstitial fluid pressure in the tumor stroma. The importance of this pathway was suggested by preclinical studies using an orthotopic pancreatic tumor mouse model (138). TKI258, a PDGFR inhibitor, is currently under evaluation in a phase I dose

assessment for advanced pancreatic cancer patients (139). Moreover; matrix metalloproteinases (MMPs) are a family of proteolytic enzymes responsible for the degradation of connective tissue proteins. Matrix metalloproteinase proteins aberrant expression is observed in pancreatic adenocarcinoma. Marimastat has been previously tested. However; the results of a phase III trial which used marimastat with gemcitabine in patients with advanced pancreatic cancer did not present favorable results (140).

Oncolytic virotherapy

It has been suggested that viruses could be a strategic tool for targeting tumors. This is due to the observation that viruses activate similar pathways as tumors, and viral infections activate both the innate and adaptive immune responses (75). It has been observed that tumors create a niche of innate and adaptive immune suppression, which not only protect the tumor from the host immune system. Additionally, it limits its ability to respond to viral infection (141). In early case reports tumor regression was observed following a naturally occurring viral infection (142). The propose strategy suggests that tumor-targeted oncolytic viruses (TOVs) could selectively infect, replicate in, and lyse tumor cells, sparing healthy, normal tissues. There are two approaches for tumor-targeted oncolytic viruses the inherently tumor-selective, *i.e.*, are naturally nonpathogenic to humans and sensitive to antiviral signaling (143) or secondly to depend on oncogenic signaling pathways such as constitutively-activated Ras (144,145). TOVs can also be genetically engineered to be tumor-selective. In order to achieve this deletions of genes are required for replication in normal tissues.(144) Deletion could involve deletion of thymidine kinase *UL23* gene in herpes simplex virus (HSV)-1 (146) or thymidine kinase and vaccinia growth factor (VGF) for vaccinia virus (VV) (147). In recently published literature, gene silencing by RNA interference technology to achieve tumor selectivity has also been utilized (145). Furthermore; another strategy used is to involve the insertion of a tumor-specific promoter (148,149), by doing this the expression of a gene necessary for viral replication in order to restrict its replication in tumor cells is overexpressed (145). TOVs can also be engineered to express cell surface receptors unique to tumor cells (150,151), which allow specific tropism (145). TOVs can designed to express immunomodulatory transgenes (144). In contrast to the pharmacokinetics of the usual drug administration, the therapeutic dose of TOVs increases with time as the virus

replicates and spreads to neighboring cells (144). Again as every virus particle carries a therapeutic gene, each viral progeny will also carry the transgene, in this mode it enhances the therapeutic effect (152). TOVs have the ability to directly lyse infected malignant cells and cause acute tumor debulking. However; another major advantage, is the ability of the virus to spread and potentiate an inflammatory response that allows the destruction of a tumor. This feature distinguishes TOVs from vaccines or immune adjuvants (153). Moreover; another unique feature of TOVs is to target silmutaneously multiple cellular pathways and therefore the risk of tumor resistance development is low (144). Another observation suggested that TOVs act synergistically with conventional chemotherapy and radiation (154-156). The first TOV to undergo a clinical trial was ONYX-015 (*dl1520*), an adenovirus deficient in the *E1B* gene (157). The gene product E1B-55 kDa protein was originally thought to sequester p53, inactivating it and as a result, allowing replication in a cell (158). In the study by O'Shea *et al.* (159) later determined that it was for the differential viral RNA export between normal and cancer cells which accounts for ONYX-015's tumor-selectivity. Until now the first approved oncolytic virus to date is H101 (Oncorine; Shanghair Sunway Biotech, Shanghai, China), which was approved for combination with chemotherapy in China in 2005 for the treatment of head and neck cancers (157).

Discussion

The exploration of the genome of pancreatic cancer is currently the best therapeutic strategy as it has important clinical relevance. However; current efforts to understand the tumor genome profile and identify efficient targeted therapies have not presented favorable results. Several targeted agents which have been mentioned in the previous sections, almost all have failed to demonstrate efficacy in late phase clinical trials. Until now only the tyrosine kinase inhibitor erlotinib has been approved by the FDA for advanced pancreatic cancer treatment, however; the improvement of overall survival was barely 2 weeks compared with gemcitabine alone (33). Pancreatic cancer has extreme genomic heterogeneity and this is the most important reason which blocks the identification of new actionable molecular targets or to testing existing biological therapies which have already been approved for human use for other cancers. Since targeted agent administration as a single agent has not presented favorable results,

multitargeted agents or molecular agent combinations are in the development phase in order to inhibit more than one pathway simultaneously and to prevent or evade resistance. The majority of trials until now have combined target agents with gemcitabine, but actually, the first-line schedules are represented by FOLFIRINOX or gemcitabine plus Nabpaclitaxel. It is suggested that greater efficacy may be obtained from the combination of target agents with those chemotherapeutic drugs. Furthermore, in most studies in which molecular or chemotherapeutic agents in pancreatic cancer were tested, patients from unselected population were enrolled to treat. In the past three years, 116 trials specific for pancreatic cancer systemic therapy were registered of which only about 8% applied criteria to select a patient subset based upon molecular biomarkers (160). In order to stratify patients, the Australian Pancreatic Cancer Genome Initiative has started a pilot study to evaluate the feasibility of assessing a more stratified approach in the management of pancreatic cancer through predefined actionable molecular phenotypes. Patients are enrolled in this trial, called IMPaCT (Individualised Molecular Pancreatic Cancer Therapy). Firstly, a preliminary phenotype screening is performed in order to compare the use of gemcitabine in an unselected population to a stratified approach. The major target of the study is to create a tailored approach to pancreatic cancer treatment, as this seems to be the major challenge for the future of pancreatic cancer treatment (161,162). Moreover; based on the advancements of biotechnology, biological agents can find application in cancer treatment by tumor-targeted delivery of cytotoxic drugs. In the study by Ahn *et al.* (162) a developed antibody fragment was installed in polymeric micelles *via* maleimide-thiol conjugation for selective delivery of platinum drugs to pancreatic tumors. It was observed that this antibody-drug conjugate significantly suppressed the growth of pancreatic tumor xenografts. This technology advancement, which has activity *in vitro* and in a mouse model, could be a promising future strategy in pancreatic cancer therapy not only as a systematic therapy, but also as a local treatment (28,162). In conclusion, the lack of efficacy of targeted therapy in PDAC represents a challenge for the future, and more efforts are needed in order to make pancreatic cancer a curable disease. Future steps include the development of novel conjugates for efficient drug delivery, further exploration of the pancreatic genome and extensive evaluate of combination therapeutic strategies with non-specific cyto-toxic agents and targeted agents in order to maximize clinical benefit (7). The ability

of TOVs to stimulate inflammation, deliver genes and immunomodulatory agents as well as reduce tumor burden by direct cell lysis, could be a novel therapeutic approach, however; more trials are in need. Probably this treatment approach could be used simultaneously with non-specific cyto-toxic agents or as an adjuvant treatment. Again local therapies could be considered to reduce tumor burden.

Acknowledgements

None.

Footnote

Conflicts of Interest: The authors have no conflict of interest to declare.

References

1. Torre LA, Bray F, Siegel RL, et al. Global cancer statistics, 2012. *CA Cancer J Clin* 2015;65:87-108.
2. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. *CA Cancer J Clin* 2015;65:5-29.
3. Rahib L, Smith BD, Aizenberg R, et al. Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States. *Cancer Res* 2014;74:2913-21.
4. Bosetti C, Bertuccio P, Malvezzi M, et al. Cancer mortality in Europe, 2005-2009, and an overview of trends since 1980. *Ann Oncol* 2013;24:2657-71.
5. Malvezzi M, Bertuccio P, Levi F, et al. European cancer mortality predictions for the year 2014. *Ann Oncol* 2014;25:1650-6.
6. Malvezzi M, Bertuccio P, Rosso T, et al. European cancer mortality predictions for the year 2015: does lung cancer have the highest death rate in EU women? *Ann Oncol* 2015;26:779-86.
7. Sharma P, Wagner K, Wolchok JD, et al. Novel cancer immunotherapy agents with survival benefit: recent successes and next steps. *Nat Rev Cancer* 2011;11:805-12.
8. Keane MG, Horsfall L, Rait G, et al. A case-control study comparing the incidence of early symptoms in pancreatic and biliary tract cancer. *BMJ Open* 2014;4:e005720.
9. Cosci F, Fava GA, Sonino N. Mood and anxiety disorders as early manifestations of medical illness: a systematic review. *Psychother Psychosom* 2015;84:22-9.
10. Sharma C, Eltawil KM, Renfrew PD, et al. Advances in diagnosis, treatment and palliation of pancreatic carcinoma:

- 1990-2010. *World J Gastroenterol* 2011;17:867-97.
11. Li J, Li Y, Cao G, et al. Early manifestations of pancreatic cancer: the effect of cancer-nerve interaction. *Med Hypotheses* 2013;81:180-2.
 12. Grahm AL, Andren-Sandberg A. Prospective evaluation of pain in exocrine pancreatic cancer. *Digestion* 1997;58:542-9.
 13. Hidalgo M. Pancreatic cancer. *N Engl J Med* 2010;362:1605-17.
 14. Shaib Y, Davila J, Naumann C, El-Serag H. The impact of curative intent surgery on the survival of pancreatic cancer patients: a U.S. Population-based study. *Am J Gastroenterol* 2007;102:1377-82.
 15. Rossi ML, Rehman AA, Gondi CS. Therapeutic options for the management of pancreatic cancer. *World J Gastroenterol* 2014;20:11142-59.
 16. Kalser MH, Ellenberg SS. Pancreatic cancer. Adjuvant combined radiation and chemotherapy following curative resection. *Arch Surg* 1985;120:899-903.
 17. Neoptolemos JP, Stocken DD, Friess H, et al. A randomized trial of chemoradiotherapy and chemotherapy after resection of pancreatic cancer. *N Engl J Med* 2004;350:1200-10.
 18. Oettle H, Neuhaus P, Hochhaus A, et al. Adjuvant chemotherapy with gemcitabine and long-term outcomes among patients with resected pancreatic cancer: the CONKO-001 randomized trial. *JAMA* 2013;310:1473-81.
 19. Gillen S, Schuster T, Meyer Zum Buschenfelde C, et al. Preoperative/neoadjuvant therapy in pancreatic cancer: a systematic review and meta-analysis of response and resection percentages. *PLoS Med* 2010;7:e1000267.
 20. Lemmens VE, Bosscha K, van der Schelling G, et al. Improving outcome for patients with pancreatic cancer through centralization. *Br J Surg* 2011;98:1455-62.
 21. Takahashi H, Akita H, Gotoh K, et al. Preoperative gemcitabine-based chemoradiation therapy for pancreatic ductal adenocarcinoma of the body and tail: impact of splenic vessels involvement on operative outcome and pattern of recurrence. *Surgery* 2015;157:484-95.
 22. Nanda RH, El-Rayes B, Maithel SK, et al. Neoadjuvant modified FOLFIRINOX and chemoradiation therapy for locally advanced pancreatic cancer improves resectability. *J Surg Oncol* 2015;111:1028-34.
 23. Burris HA 3rd, Moore MJ, Andersen J, et al. Improvements in survival and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: a randomized trial. *J Clin Oncol* 1997;15:2403-13.
 24. Heinemann V, Boeck S, Hinke A, et al. Meta-analysis of randomized trials: evaluation of benefit from gemcitabine-based combination chemotherapy applied in advanced pancreatic cancer. *BMC Cancer* 2008;8:82.
 25. Conroy T, Desseigne F, Ychou M, et al. FOLFIRINOX versus gemcitabine for metastatic pancreatic cancer. *N Engl J Med* 2011;364:1817-25.
 26. Von Hoff DD, Ervin T, Arena FP, et al. Increased survival in pancreatic cancer with nab-paclitaxel plus gemcitabine. *N Engl J Med* 2013;369:1691-703.
 27. Ryan DP, Hong TS, Bardeesy N. Pancreatic adenocarcinoma. *N Engl J Med* 2014;371:2140-1.
 28. Zarogoulidis P, Pavlioglou P, Pivert PL, et al. Current and future intratumoral targeted treatment for pancreatic cancer. *Therapeutic delivery*. 2014;5:913-26.
 29. Gerber DE. Targeted therapies: a new generation of cancer treatments. *Am Fam Physician* 2008;77:311-9.
 30. Jones S, Zhang X, Parsons DW, et al. Core signaling pathways in human pancreatic cancers revealed by global genomic analyses. *Science* 2008;321:1801-6.
 31. Seicean A, Petrusel L, Seicean R. New targeted therapies in pancreatic cancer. *World J Gastroenterol* 2015;21:6127-45.
 32. Chong CR, Janne PA. The quest to overcome resistance to EGFR-targeted therapies in cancer. *Nat Med* 2013;19:1389-400.
 33. Moore MJ, Goldstein D, Hamm J, et al. Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. *J Clin Oncol* 2007;25:1960-6.
 34. Cairns RA, Harris IS, Mak TW. Regulation of cancer cell metabolism. *Nat Rev Cancer* 2011;11:85-95.
 35. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011;144:646-74.
 36. Kimmelman AC. Metabolic Dependencies in RAS-Driven Cancers. *Clin Cancer Res* 2015;21:1828-34.
 37. Bryant KL, Mancias JD, Kimmelman AC, et al. KRAS: feeding pancreatic cancer proliferation. *Trends Biochem Sci* 2014;39:91-100.
 38. Daye D, Wellen KE. Metabolic reprogramming in cancer: unraveling the role of glutamine in tumorigenesis. *Semin Cell Dev Biol* 2012;23:362-9.
 39. Swierczynski J, Hebanowska A, Sledzinski T. Role of abnormal lipid metabolism in development, progression, diagnosis and therapy of pancreatic cancer. *World J Gastroenterol* 2014;20:2279-303.
 40. Ying H, Kimmelman AC, Lyssiotis CA, et al. Oncogenic Kras maintains pancreatic tumors through regulation of

- anabolic glucose metabolism. *Cell* 2012;149:656-70.
41. Son J, Lyssiotis CA, Ying H, et al. Glutamine supports pancreatic cancer growth through a KRAS-regulated metabolic pathway. *Nature* 2013;496:101-5.
 42. Cohen R, Neuzillet C, Tijeras-Raballand A, et al. Targeting cancer cell metabolism in pancreatic adenocarcinoma. *Oncotarget* 2015;6:16832-47.
 43. Rajeshkumar NV, Dutta P, Yabuuchi S, et al. Therapeutic Targeting of the Warburg Effect in Pancreatic Cancer Relies on an Absence of p53 Function. *Cancer Res* 2015;75:3355-64.
 44. Mayers JR, Vander Heiden MG. Famine versus feast: understanding the metabolism of tumors in vivo. *Trends Biochem Sci* 2015;40:130-40.
 45. Mayers JR, Wu C, Clish CB, et al. Elevation of circulating branched-chain amino acids is an early event in human pancreatic adenocarcinoma development. *Nat Med* 2014;20:1193-8.
 46. Viale A, Pettazoni P, Lyssiotis CA, et al. Oncogene ablation-resistant pancreatic cancer cells depend on mitochondrial function. *Nature* 2014;514:628-32.
 47. Yang S, Wang X, Contino G, et al. Pancreatic cancers require autophagy for tumor growth. *Genes Dev* 2011;25:717-29.
 48. Perera RM, Stoykova S, Nicolay BN, et al. Transcriptional control of autophagy-lysosome function drives pancreatic cancer metabolism. *Nature* 2015;524:361-5.
 49. Wolpin BM, Rubinson DA, Wang X, et al. Phase II and pharmacodynamic study of autophagy inhibition using hydroxychloroquine in patients with metastatic pancreatic adenocarcinoma. *Oncologist* 2014;19:637-8.
 50. Rosenfeldt MT, O'Prey J, Morton JP, et al. p53 status determines the role of autophagy in pancreatic tumour development. *Nature* 2013;504:296-300.
 51. Yang A, Kimmelman AC. Inhibition of autophagy attenuates pancreatic cancer growth independent of TP53/TRP53 status. *Autophagy* 2014;10:1683-4.
 52. Commisso C, Davidson SM, Soydaner-Azeloglu RG, et al. Macropinocytosis of protein is an amino acid supply route in Ras-transformed cells. *Nature* 2013;497:633-7.
 53. Kamphorst JJ, Nofal M, Commisso C, et al. Human pancreatic cancer tumors are nutrient poor and tumor cells actively scavenge extracellular protein. *Cancer Res* 2015;75:544-53.
 54. Kamphorst JJ, Cross JR, Fan J, et al. Hypoxic and Ras-transformed cells support growth by scavenging unsaturated fatty acids from lysophospholipids. *Proc Natl Acad Sci U S A* 2013;110:8882-7.
 55. Baenke F, Peck B, Miess H, et al. Hooked on fat: the role of lipid synthesis in cancer metabolism and tumour development. *Disease models & mechanisms*. 2013;6:1353-63.
 56. Rysman E, Brusselmans K, Scheys K, et al. De novo lipogenesis protects cancer cells from free radicals and chemotherapeutics by promoting membrane lipid saturation. *Cancer Res* 2010;70:8117-26.
 57. Guillaumond F, Bidaut G, Ouaiissi M, et al. Cholesterol uptake disruption, in association with chemotherapy, is a promising combined metabolic therapy for pancreatic adenocarcinoma. *Proc Natl Acad Sci U S A* 2015;112:2473-8.
 58. Siegel R, Ma J, Zou Z, et al. Cancer statistics, 2014. *CA Cancer J Clin* 2014;64:9-29.
 59. Ryan DP, Hong TS, Bardeesy N. Pancreatic adenocarcinoma. *N Engl J Med* 2014;371:1039-49.
 60. Maitra A, Fukushima N, Takaori K, et al. Precursors to invasive pancreatic cancer. *Adv Anat Pathol* 2005;12:81-91.
 61. Gnani A, Licchetta A, Scarpa A, et al. Carcinogenesis of pancreatic adenocarcinoma: precursor lesions. *Int J Mol Sci* 2013;14:19731-62.
 62. Tanaka M, Fernandez-del Castillo C, Adsay V, et al. International consensus guidelines 2012 for the management of IPMN and MCN of the pancreas. *Pancreatology* 2012;12:183-97.
 63. Matthaei H, Norris AL, Tsiatis AC, et al. Clinicopathological characteristics and molecular analyses of multifocal intraductal papillary mucinous neoplasms of the pancreas. *Ann Surg* 2012;255:326-33.
 64. Poultsides GA, Reddy S, Cameron JL, et al. Histopathologic basis for the favorable survival after resection of intraductal papillary mucinous neoplasm-associated invasive adenocarcinoma of the pancreas. *Ann Surg* 2010;251:470-6.
 65. Mino-Kenudson M, Fernandez-del Castillo C, Baba Y, et al. Prognosis of invasive intraductal papillary mucinous neoplasm depends on histological and precursor epithelial subtypes. *Gut* 2011;60:1712-20.
 66. Kanda M, Matthaei H, Wu J, et al. Presence of somatic mutations in most early-stage pancreatic intraepithelial neoplasia. *Gastroenterology* 2012;142:730-3 e9.
 67. Amato E, Molin MD, Mafficini A, et al. Targeted next-generation sequencing of cancer genes dissects the molecular profiles of intraductal papillary neoplasms of the pancreas. *J Pathol* 2014;233:217-27.
 68. Schönleben F, Qiu W, Bruckman KC, et al. BRAF and KRAS gene mutations in intraductal papillary mucinous

- neoplasm/carcinoma (IPMN/IPMC) of the pancreas. *Cancer Lett* 2007;249:242-8.
69. Furukawa T, Kuboki Y, Tanji E, et al. Whole-exome sequencing uncovers frequent GNAS mutations in intraductal papillary mucinous neoplasms of the pancreas. *Sci Rep* 2011;1:161.
 70. Wu J, Jiao Y, Dal Molin M, et al. Whole-exome sequencing of neoplastic cysts of the pancreas reveals recurrent mutations in components of ubiquitin-dependent pathways. *Proc Natl Acad Sci U S A* 2011;108:21188-93.
 71. Wu J, Matthaei H, Maitra A, et al. Recurrent GNAS mutations define an unexpected pathway for pancreatic cyst development. *Sci Transl Med* 2011;3:92ra66.
 72. Xiao SY. Intraductal papillary mucinous neoplasm of the pancreas: an update. *Scientifica (Cairo)* 2012;2012:893632.
 73. von Figura G, Fukuda A, Roy N, et al. The chromatin regulator Brg1 suppresses formation of intraductal papillary mucinous neoplasm and pancreatic ductal adenocarcinoma. *Nat Cell Biol* 2014;16:255-67.
 74. Kopp JL, von Figura G, Mayes E, et al. Identification of Sox9-dependent acinar-to-ductal reprogramming as the principal mechanism for initiation of pancreatic ductal adenocarcinoma. *Cancer Cell* 2012;22:737-50.
 75. Melcher A, Parato K, Rooney CM, et al. Thunder and lightning: immunotherapy and oncolytic viruses collide. *Mol Ther* 2011;19:1008-16.
 76. Tobita K, Kijima H, Dowaki S, et al. Epidermal growth factor receptor expression in human pancreatic cancer: Significance for liver metastasis. *International journal of molecular medicine*. 2003;11:305-9.
 77. da Cunha Santos G, Dhani N, Tu D, et al. Molecular predictors of outcome in a phase 3 study of gemcitabine and erlotinib therapy in patients with advanced pancreatic cancer: National Cancer Institute of Canada Clinical Trials Group Study PA.3. *Cancer* 2010;116:5599-607.
 78. Cascinu S, Berardi R, Labianca R, et al. Cetuximab plus gemcitabine and cisplatin compared with gemcitabine and cisplatin alone in patients with advanced pancreatic cancer: a randomised, multicentre, phase II trial. *Lancet Oncol* 2008;9:39-44.
 79. Philip PA, Benedetti J, Corless CL, et al. Phase III study comparing gemcitabine plus cetuximab versus gemcitabine in patients with advanced pancreatic adenocarcinoma: Southwest Oncology Group-directed intergroup trial S0205. *J Clin Oncol* 2010;28:3605-10.
 80. Fountzilias G, Bobos M, Kalogera-Fountzila A, et al. Gemcitabine combined with gefitinib in patients with inoperable or metastatic pancreatic cancer: a phase II Study of the Hellenic Cooperative Oncology Group with biomarker evaluation. *Cancer Invest* 2008;26:784-93.
 81. Kimura K, Sawada T, Komatsu M, et al. Antitumor effect of trastuzumab for pancreatic cancer with high HER-2 expression and enhancement of effect by combined therapy with gemcitabine. *Clin Cancer Res* 2006;12:4925-32.
 82. Safran H, Iannitti D, Ramanathan R, et al. Herceptin and gemcitabine for metastatic pancreatic cancers that overexpress HER-2/neu. *Cancer Invest* 2004;22:706-12.
 83. Harder J, Ihorst G, Heinemann V, et al. Multicentre phase II trial of trastuzumab and capecitabine in patients with HER2 overexpressing metastatic pancreatic cancer. *Br J Cancer* 2012;106:1033-8.
 84. Safran H, Miner T, Resnick M, et al. Lapatinib/gemcitabine and lapatinib/gemcitabine/oxaliplatin: a phase I study for advanced pancreaticobiliary cancer. *Am J Clin Oncol* 2008;31:140-4.
 85. Su D, Jiao SC, Wang LJ, et al. Efficacy of nimotuzumab plus gemcitabine usage as first-line treatment in patients with advanced pancreatic cancer. *Tumour Biol* 2014;35:2313-8.
 86. Strumberg D, Schultheis B, Scheulen ME, et al. Phase II study of nimotuzumab, a humanized monoclonal anti-epidermal growth factor receptor (EGFR) antibody, in patients with locally advanced or metastatic pancreatic cancer. *Invest New Drugs* 2012;30:1138-43.
 87. Ioannou N, Dalglish AG, Seddon AM, et al. Anti-tumour activity of afatinib, an irreversible ErbB family blocker, in human pancreatic tumour cells. *Br J Cancer* 2011;105:1554-62.
 88. PD Dr. med. Volker Heinemann. Afatinib as Cancer Therapy for Exocrine Pancreatic Tumours (ACCEPT). In: *ClinicalTrials.gov* [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01728818>, NLM Identifier: NCT01728818.
 89. di Magliano MP, Logsdon CD. Roles for KRAS in pancreatic tumor development and progression. *Gastroenterology* 2013;144:1220-9.
 90. Takashima A, Faller DV. Targeting the RAS oncogene. *Expert Opin Ther Targets* 2013;17:507-31.
 91. Pylayeva-Gupta Y, Grabocka E, Bar-Sagi D. RAS oncogenes: weaving a tumorigenic web. *Nat Rev Cancer* 2011;11:761-74.
 92. Van Cutsem E, van de Velde H, Karasek P, et al. Phase III trial of gemcitabine plus tipifarnib compared with gemcitabine plus placebo in advanced pancreatic cancer. *J Clin Oncol* 2004;22:1430-8.

93. Bodoky G, Timcheva C, Spigel DR, et al. A phase II open-label randomized study to assess the efficacy and safety of selumetinib (AZD6244 [ARRY-142886]) versus capecitabine in patients with advanced or metastatic pancreatic cancer who have failed first-line gemcitabine therapy. *Invest New Drugs* 2012;30:1216-23.
94. Infante JR, Somer BG, Park JO, et al. A randomised, double-blind, placebo-controlled trial of trametinib, an oral MEK inhibitor, in combination with gemcitabine for patients with untreated metastatic adenocarcinoma of the pancreas. *Eur J Cancer* 2014;50:2072-81.
95. Scott AJ, O'Neil BH, Gomes C, et al. A phase II/III randomized study to compare the efficacy and safety of rigosertib plus gemcitabine versus gemcitabine alone in patients with previously untreated metastatic pancreatic cancer. *J Clin Oncol* 2015;33:abstr 342.
96. Sachdev D, Yee D. Disrupting insulin-like growth factor signaling as a potential cancer therapy. *Mol Cancer Ther* 2007;6:1-12.
97. Philip PA, Goldman B, Ramanathan RK, et al. Dual blockade of epidermal growth factor receptor and insulin-like growth factor receptor-1 signaling in metastatic pancreatic cancer: phase Ib and randomized phase II trial of gemcitabine, erlotinib, and cixutumumab versus gemcitabine plus erlotinib (SWOG S0727). *Cancer* 2014;120:2980-5.
98. Fuchs CS, Azevedo S, Okusaka T, et al. A phase 3 randomized, double-blind, placebo-controlled trial of ganitumab or placebo in combination with gemcitabine as first-line therapy for metastatic adenocarcinoma of the pancreas: the GAMMA trial. *Ann Oncol* 2015;26:921-7.
99. Kindler HL, Niedzwiecki D, Hollis D, et al. Gemcitabine plus bevacizumab compared with gemcitabine plus placebo in patients with advanced pancreatic cancer: phase III trial of the Cancer and Leukemia Group B (CALGB 80303). *J Clin Oncol* 2010;28:3617-22.
100. Van Cutsem E, Vervenne WL, Bennouna J, et al. Phase III trial of bevacizumab in combination with gemcitabine and erlotinib in patients with metastatic pancreatic cancer. *J Clin Oncol* 2009;27:2231-7.
101. Kindler HL, Ioka T, Richel DJ, et al. Axitinib plus gemcitabine versus placebo plus gemcitabine in patients with advanced pancreatic adenocarcinoma: a double-blind randomised phase 3 study. *Lancet Oncol* 2011;12:256-62.
102. Rougier P, Riess H, Manges R, et al. Randomised, placebo-controlled, double-blind, parallel-group phase III study evaluating aflibercept in patients receiving first-line treatment with gemcitabine for metastatic pancreatic cancer. *Eur J Cancer* 2013;49:2633-42.
103. Chiorean EG, Schneider BP, Akisik FM, et al. Phase 1 pharmacogenetic and pharmacodynamic study of sorafenib with concurrent radiation therapy and gemcitabine in locally advanced unresectable pancreatic cancer. *Int J Radiat Oncol Biol Phys* 2014;89:284-91.
104. Kindler HL, Wroblewski K, Wallace JA, et al. Gemcitabine plus sorafenib in patients with advanced pancreatic cancer: a phase II trial of the University of Chicago Phase II Consortium. *Invest New Drugs* 2012;30:382-6.
105. Ltd. TP. Investigator's Initiated Phase II Study for Pancreatic Cancer Patients. In: *ClinicalTrials.gov* [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01659502>, NLM Identifier: NCT01659502.
106. Momenta Pharmaceuticals I. M402 in Combination With Nab-Paclitaxel and Gemcitabine in Pancreatic Cancer In: *ClinicalTrials.gov* [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01621243>, NLM Identifier: NCT01621243.
107. Di Marco M, Macchini M, Vecchiarelli S, et al. Hedgehog signaling: from the cuirass to the heart of pancreatic cancer. *Pancreatol* 2012;12:388-93.
108. Bailey JM, Swanson BJ, Hamada T, et al. Sonic hedgehog promotes desmoplasia in pancreatic cancer. *Clin Cancer Res* 2008;14:5995-6004.
109. Olive KP, Jacobetz MA, Davidson CJ, et al. Inhibition of Hedgehog signaling enhances delivery of chemotherapy in a mouse model of pancreatic cancer. *Science* 2009;324:1457-61.
110. Singh BN, Fu J, Srivastava RK, et al. Hedgehog signaling antagonist GDC-0449 (Vismodegib) inhibits pancreatic cancer stem cell characteristics: molecular mechanisms. *PloS One* 2011;6:e27306.
111. Catenacci DV, Junttila MR, Karrison T, et al. Randomized Phase Ib/II Study of Gemcitabine Plus Placebo or Vismodegib, a Hedgehog Pathway Inhibitor, in Patients With Metastatic Pancreatic Cancer. *J Clin Oncol* 2015;33:4284-92.
112. National Cancer Institute (NCI). Gemcitabine Hydrochloride With or Without Vismodegib in Treating Patients With Recurrent or Metastatic Pancreatic Cancer In: *ClinicalTrials.gov* [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01064622>, NLM Identifier: NCT01064622.
113. Center. SKCC. Pancreas, Hedgehog Inhibitors for

- Metastatic Adenocarcinoma of the Pancreas.
114. Bax L. Hedgehog Inhibition for Pancreatic Ductal Adenocarcinoma (PDAC) in the Preoperative Setting (HIPPOs). In: ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01096732>, NLM Identifier: NCT01096732.
 115. Infinity Reports Update from Phase 2 Study of Saridegib Plus Gemcitabine in Patients with Metastatic Pancreatic Cancer.
 116. Sidney Kimmel Comprehensive Cancer Center. Gemcitabine + Nab-paclitaxel With LDE-225 (Hedgehog Inhibitor) as Neoadjuvant Therapy for Pancreatic Adenocarcinoma. In: ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01431794>, NLM Identifier: NCT01431794.
 117. Sjolund J, Manetopoulos C, Stockhausen MT, et al. The Notch pathway in cancer: differentiation gone awry. *Eur J Cancer* 2005;41:2620-9.
 118. Ristorcelli E, Lombardo D. Targeting Notch signaling in pancreatic cancer. *Expert Opin Ther Targets* 2010;14:541-52.
 119. Plentz R, Park JS, Rhim AD, et al. Inhibition of gamma-secretase activity inhibits tumor progression in a mouse model of pancreatic ductal adenocarcinoma. *Gastroenterology* 2009;136:1741-9 e6.
 120. De Jesus-Acosta A, Laheru D, Maitra A, et al. A phase II study of the gamma secretase inhibitor RO4929097 in patients with previously treated metastatic pancreatic adenocarcinoma. *Invest New Drugs* 2014;32:739-45.
 121. Hidalgo M, Cooray P, Jameson MB, et al. A phase Ib study of the anti-cancer stem cell agent demcizumab (DEM) & gemcitabine (GEM) +/- paclitaxel protein bound particles (nab-paclitaxel) in pts with pancreatic cancer. *J Clin Oncol* 2015;33:abstr 4118.
 122. Waddell N, Pajic M, Patch AM, et al. Whole genomes redefine the mutational landscape of pancreatic cancer. *Nature* 2015;518:495-501.
 123. Tangutoori S, Baldwin P, Sridhar S. PARP inhibitors: A new era of targeted therapy. *Maturitas* 2015;81:5-9.
 124. AstraZeneca. Study to Assess the Safety & Tolerability of a PARP Inhibitor in Combination With Gemcitabine in Pancreatic Cancer. In: ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT00515866>, NLM Identifier: NCT00515866.
 125. National Cancer Institute (NCI). Gemcitabine Hydrochloride and Cisplatin With or Without Veliparib or Veliparib Alone in Patients With Locally Advanced or Metastatic Pancreatic Cancer. Available online: <https://clinicaltrials.gov/ct2/show/NCT01585805>, NLM Identifier: NCT01585805.
 126. Hezel AF, Kimmelman AC, Stanger BZ, et al. Genetics and biology of pancreatic ductal adenocarcinoma. *Genes Dev* 2006;20:1218-49.
 127. Ito D, Fujimoto K, Mori T, et al. In vivo antitumor effect of the mTOR inhibitor CCI-779 and gemcitabine in xenograft models of human pancreatic cancer. *Int J Cancer* 2006;118:2337-43.
 128. Center ULCC. BKM120 + mFOLFOX6 in Advanced Solid Tumors With Expansion Cohort Pancreatic Cancer.: In: ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01571024>, NLM Identifier: NCT01571024.
 129. Rexahn Pharmaceuticals I. A Safety and Efficacy Study of RX-0201 Plus Gemcitabine in Metastatic Pancreatic Cancer.: In: ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01028495>, NLM Identifier: NCT01028495.
 130. Pharmaceuticals. N. Safety, Pharmacokinetics and Pharmacodynamics of BEZ235 Plus MEK162 in Selected Advanced Solid Tumor Patients. In: ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01337765>, NLM Identifier: NCT01337765.
 131. Wolpin BM, Hezel AF, Abrams T, et al. Oral mTOR inhibitor everolimus in patients with gemcitabine-refractory metastatic pancreatic cancer. *J Clin Oncol* 2009;27:193-8.
 132. Kordes S, Klumpen HJ, Weterman MJ, et al. Phase II study of capecitabine and the oral mTOR inhibitor everolimus in patients with advanced pancreatic cancer. *Cancer Chemother Pharmacol* 2015;75:1135-41.
 133. Pharmaceuticals. N. Treatment of Patients Suffering From a Progressive Pancreas Carcinoma With Everolimus (RAD001) and Gemcitabine. In: ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT00560963>, NLM Identifier: NCT00560963.
 134. (NCI). NCI. CCI-779 in Treating Patients With Locally Advanced or Metastatic Pancreatic Cancer. In: ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://>

- clinicaltrials.gov/ct2/show/NCT00075647, NLM Identifier: NCT00075647.
135. Heinemann V, Reni M, Ychou M, et al. Tumour-stroma interactions in pancreatic ductal adenocarcinoma: rationale and current evidence for new therapeutic strategies. *Cancer Treat Rev* 2014;40:118-28.
 136. Provenzano PP, Cuevas C, Chang AE, et al. Enzymatic targeting of the stroma ablates physical barriers to treatment of pancreatic ductal adenocarcinoma. *Cancer Cell* 2012;21:418-29.
 137. Therapeutics H. PEGPH20 Plus Nab-Paclitaxel Plus Gemcitabine Compared With Nab-Paclitaxel Plus Gemcitabine in Subjects With Stage IV Untreated Pancreatic Cancer (HALO-109-202). In: *ClinicalTrials.gov* [Internet]. Bethesda (MD): National Library of Medicine (US). Available online: <https://clinicaltrials.gov/ct2/show/NCT01839487>, NLM Identifier: NCT01839487.
 138. Hwang RF, Yokoi K, Bucana CD, et al. Inhibition of platelet-derived growth factor receptor phosphorylation by STI571 (Gleevec) reduces growth and metastasis of human pancreatic carcinoma in an orthotopic nude mouse model. *Clin Cancer Res* 2003;9:6534-44.
 139. Institute RPC. Dovitinib Lactate, Gemcitabine Hydrochloride, and Capecitabine in Treating Patients With Advanced or Metastatic Solid Tumors, Pancreatic Cancer and Biliary Cancers. In: *ClinicalTrials.gov* [Internet]. Bethesda (MD): National Library of Medicine (US) Available online: <https://clinicaltrials.gov/ct2/show/NCT01497392>, NLM Identifier: NCT01497392.
 140. Bramhall SR, Schulz J, Nemunaitis J, et al. A double-blind placebo-controlled, randomised study comparing gemcitabine and marimastat with gemcitabine and placebo as first line therapy in patients with advanced pancreatic cancer. *Br J Cancer* 2002;87:161-7.
 141. Lichty BD, Breitbach CJ, Stojdl DF, et al. Going viral with cancer immunotherapy. *Nat Rev Cancer* 2014;14:559-67.
 142. Kelly E, Russell SJ. History of oncolytic viruses: genesis to genetic engineering. *Mol Ther* 2007;15:651-9.
 143. Naik S, Russell SJ. Engineering oncolytic viruses to exploit tumor specific defects in innate immune signaling pathways. *Expert Opin Biol Ther* 2009;9:1163-76.
 144. Chiocca EA, Rabkin SD. Oncolytic viruses and their application to cancer immunotherapy. *Cancer Immunol Res* 2014;2:295-300.
 145. Wong HH, Lemoine NR, Wang Y. Oncolytic Viruses for Cancer Therapy: Overcoming the Obstacles. *Viruses* 2010;2:78-106.
 146. Martuza RL, Malick A, Markert JM, et al. Experimental therapy of human glioma by means of a genetically engineered virus mutant. *Science* 1991;252:854-6.
 147. Thorne SH, Hwang TH, O'Gorman WE, et al. Rational strain selection and engineering creates a broad-spectrum, systemically effective oncolytic poxvirus, JX-963. *J Clin Invest* 2007;117:3350-8.
 148. Pan W, Bodempudi V, Esfandyari T, et al. Utilizing ras signaling pathway to direct selective replication of herpes simplex virus-1. *PLoS One* 2009;4:e6514.
 149. Doloff JC, Waxman DJ, Jounaidi Y. Human telomerase reverse transcriptase promoter-driven oncolytic adenovirus with E1B-19 kDa and E1B-55 kDa gene deletions. *Hum Gene Ther* 2008;19:1383-400.
 150. Nishimoto T, Yoshida K, Miura Y, et al. Oncolytic virus therapy for pancreatic cancer using the adenovirus library displaying random peptides on the fiber knob. *Gene Ther* 2009;16:669-80.
 151. Conner J, Braidwood L, Brown SM. A strategy for systemic delivery of the oncolytic herpes virus HSV1716: redirected tropism by antibody-binding sites incorporated on the virion surface as a glycoprotein D fusion protein. *Gene Ther* 2008;15:1579-92.
 152. Heinemann MK. Buy one - get one free! *Thorac Cardiovasc Surg* 2014;62:1-2.
 153. Sobol PT, Boudreau JE, Stephenson K, et al. Adaptive antiviral immunity is a determinant of the therapeutic success of oncolytic virotherapy. *Mol Ther* 2011;19:335-44.
 154. Dai MH, Zamarin D, Gao SP, et al. Synergistic action of oncolytic herpes simplex virus and radiotherapy in pancreatic cancer cell lines. *Br J Surg* 2010;97:1385-94.
 155. Pandha HS, Heinemann L, Simpson GR, et al. Synergistic effects of oncolytic reovirus and cisplatin chemotherapy in murine malignant melanoma. *Clin Cancer Res* 2009;15:6158-66.
 156. Nishizaki M, Meyn RE, Levy LB, et al. Synergistic inhibition of human lung cancer cell growth by adenovirus-mediated wild-type p53 gene transfer in combination with docetaxel and radiation therapeutics in vitro and in vivo. *Clin Cancer Res* 2001;7:2887-97.
 157. Yu W, Fang H. Clinical trials with oncolytic adenovirus in China. *Curr Cancer Drug Targets* 2007;7:141-8.
 158. Green NK, Seymour LW. Adenoviral vectors: systemic delivery and tumor targeting. *Cancer Gene Ther* 2002;9:1036-42.
 159. O'Shea CC, Johnson L, Bagus B, et al. Late viral RNA export, rather than p53 inactivation, determines ONYX-015 tumor selectivity. *Cancer Cell* 2004;6:611-23.

160. ClinicalTrials.gov. ClinicalTrials.gov [Internet]. Bethesda (MD): National Library of Medicine (US); [2015]. Available online: <http://www.clinicaltrials.gov>.
161. Chantrill LA, Nagrial AM, Watson C, et al. Precision Medicine for Advanced Pancreas Cancer: The Individualized Molecular Pancreatic Cancer Therapy

- (IMPACT) Trial. Clin Cancer Res 2015;21:2029-37.
162. Ahn J, Miura Y, Yamada N, et al. Antibody fragment-conjugated polymeric micelles incorporating platinum drugs for targeted therapy of pancreatic cancer. Biomaterials 2015;39:23-30.

Cite this article as: Kosmidis C, Sapalidis K, Kotidis E, Mixalopoulos N, Zarogoulidis P, Tsavlis D, Baka S, Man YG, Kanellos J. Pancreatic cancer from bench to bedside: molecular pathways and treatment options. Ann Transl Med 2016;4(9):165. doi: 10.21037/atm.2016.05.11