

Incorporating Biomarkers Into Cancer and Aging Research

Joleen M. Hubbard, Harvey J. Cohen, and Hyman B. Muss

Joleen M. Hubbard, Mayo Clinic, Rochester, MN; Harvey J. Cohen, Duke University Medical Center, Durham; and Hyman B. Muss, University of North Carolina, Chapel Hill, NC.

Published online ahead of print at www.jco.org on July 28, 2014.

Authors' disclosures of potential conflicts of interest and author contributions are found at the end of this article.

Corresponding author: Joleen M. Hubbard, MD, Mayo Clinic, 200 First St SW, Rochester MN 55905; e-mail: hubbard.joleen@mayo.edu.

© 2014 by American Society of Clinical Oncology

0732-183X/14/3224w-2611w/\$20.00

DOI: 10.1200/JCO.2014.55.4261

A B S T R A C T

The challenge in treating the older adult with cancer is accurately accounting for and adapting management to the heterogeneity in health status of the individual patient. Many oncologists recognize that chronological age alone should not be the determinant when deciding on a treatment regimen. Easily measurable markers that provide an assessment of functional age would be ideal to assess frailty, which may predispose the patient to complications from cancer treatment, including increased toxicity, functional decline, decreased quality of life, and poorer survival. Several categories of potential markers, including chronic inflammatory markers, markers of cellular senescence, and imaging to assess muscle mass to detect sarcopenia, may provide insight into the likelihood of treatment-related complications. This article discusses candidate markers and strategies to evaluate these markers in cancer treatment trials, with the aim of developing a method to assess risk of oncologic outcomes and guide management decisions for both the physician and patient.

J Clin Oncol 32:2611-2616. © 2014 by American Society of Clinical Oncology

INTRODUCTION

There is great heterogeneity in the ability of older adults to tolerate cancer treatment. Older adults are at risk for increased toxicity from cancer therapy, but standard methods to accurately determine this risk are lacking. Clinical factors routinely collected during the cancer assessment such as age, performance status (PS), and comorbidities are not reliable predictors of toxicity.¹⁻⁴ Out of concern for poor tolerability, chemotherapy is often withheld from older patients on the basis of age alone, despite evidence that some older adults can derive benefit from treatment similar to that derived by younger patients.^{4,5} As a result, older adults with cancer are often undertreated, make up a minority of patients enrolled onto clinical trials,⁶ and are not gaining the benefits of cancer therapeutic advances as much as younger patients.⁷

To individualize treatment for the older patient, more data are needed beyond their chronological age and comorbidities. An accurate sense of their functional age with more objective measures that are easily measured, easily reproduced, and predictive of outcome are needed. Some consider frailty a reflection of functional age because it gives a measure of physiological age not necessarily in proportion to chronological age.⁸ Frailty can be defined as the inability of an individual to return to their baseline physical status after an insult to the body, or a measure of resilience. Fried et al⁹ described a phenotype of frailty as having three of the following: 10-

pound unintentional weight loss, poor grip strength, exhaustion, slow gait, and low physical activity level. The degree of resilience varies among older individuals as commonly seen by the differing degrees to which older patients tolerate treatment in terms of adverse effects.

The ideal marker would reflect the degree of a patient's functional reserve and predict tolerance to cancer treatment. Some measures of this already exist. The comprehensive geriatric assessment can assess multiple aspects of a patient's life, including physical function, physical and mental health, cognition, and socioeconomic circumstances. The application of the geriatric assessment in oncology is discussed elsewhere in this issue. But, likely because of lack of time, resources, and expertise, the comprehensive geriatric assessment is not widely used in clinical practice. In addition, one could imagine that it may be difficult to measure repeatedly in patients who are likely fatigued from the cancer treatment itself. There are several proposed biologic markers of aging with various amounts of data on their ability to correlate with physical function or predict functional decline and/or mortality. Another largely unexplored area is the use of imaging studies for assessment of the ratio of muscle mass or fat to muscle. This assessment could be done on computed tomography scans commonly used for tumor staging evaluations that could potentially give more information on functional reserve and/or predict a decline in physical function.

Table 1. Summary of Proposed Markers of Functional Age

Marker	Source	Test	Association With Frailty and/or Function	Association With Mortality	References
Chronic inflammatory markers	Serum or plasma	ELISA	Yes (CRP, IL-6, TNF- α , D-dimer, IL-1RA)	Yes (CRP, IL-6, D-dimer, sVCAM)	Cesari et al ¹⁰ Ferrucci et al ¹¹ Hubbard et al ¹² Leng et al ¹³ Walston et al ¹⁴ Cohen et al ¹⁵ de Saint-Hubert et al ¹⁶ Huffman et al ¹⁷ Puts et al ¹⁸ Reuben et al ¹⁹ Rønning et al ²⁰
Telomere length	Leukocyte DNA	q-PCR or Southern blot	Yes	Yes	Cawthon et al ²¹ Epel et al ²² Farzaneh-Far et al ²³ Risques et al ²⁴
p16 ^{INK4a}	T-lymphocyte RNA	qRT-PCR	No	No	Krishnamurthy et al ²⁵ Liu et al ²⁶ Song et al ²⁷
Sarcopenia	CT scan	Commercially available software for body composition analysis	Yes	Yes	Baumgartner et al ²⁸ Heymsfield et al ²⁹ Janssen et al ³⁰ Metter et al ³¹

Abbreviations: CRP, C-reactive protein; CT, computed tomography; ELISA, enzyme-linked immunosorbent assay; IL-6, interleukin-6; IL-1RA, IL-1 receptor antagonist; q-PCR, quantitative polymerase chain reaction; qRT-PCR, quantitative real-time PCR; sVCAM, soluble vascular cell adhesion molecule; TNF- α , tumor necrosis factor- α .

This review will discuss potential markers of functional age to complement clinical geriatric assessment as well as their potential incorporation into clinical trials to assess their value. Validation will be necessary before any marker can be routinely used in practice to better inform patients and physicians of the potential harms and/or risks associated with treatment and to guide clinical management decisions. A summary of the proposed markers is listed in Table 1.

POTENTIAL BIOMARKERS THAT WARRANT FURTHER STUDY

Markers of Systemic Inflammation

Markers of chronic inflammation are potential biomarkers of frailty and functional reserve that have been studied most in terms of their correlation with clinical measures of frailty, functional decline, and mortality. Prothrombotic factors have also been noted to be increased with chronic inflammatory markers, likely because of a costimulatory effect between the two processes. For instance, inflammatory cytokines such as tumor necrosis factor α (TNF- α) and interleukin-6 (IL-6) can stimulate production of prothrombotic factors such as plasminogen activator inhibitor-1 (PAI-1) and fibrinogen.³² The synthesis of cytokines IL-1B, IL-6, and PAI-1 is also induced by D-dimer, a marker of the activated coagulation system.³³ Likewise, when vascular cell adhesion molecule (VCAM) is exposed to inflammatory markers TNF- α and IL-1B, it is cleaved to soluble vascular cell adhesion molecule (sVCAM), which has been shown to be increased in patients with age-related diseases such as rheumatoid arthritis.³⁴ For the purpose of this discussion, markers of the coagulation system such as D-dimer and sVCAM will be included with chronic inflammatory markers. In general, proinflammatory mediators such as IL-6, TNF- α , D-dimer, and PAI-1, increase with age, even among healthy individuals.³⁶⁻³⁸ These markers are proposed to accelerate the aging process and exacerbate multiple age-related diseases.³⁹⁻⁴¹

Multiple studies have shown that these markers correlate with clinical measures of frailty and are increased to a greater degree in frail patients than in age-matched, nonfrail controls.^{10-14,42,43} One study of 110 patients older than age 75 years evaluated a combination of inflammatory markers (TNF- α , IL-6, C-reactive protein [CRP]) and low albumin and their relationship to several different clinical measures of frailty.¹² The degree of clinical frailty independently correlated with increased inflammatory marker levels and lower albumin levels, adjusting for multiple factors, including age, sex, body mass index, smoking status, number of comorbidities, and number of medications. Therefore, these markers might be useful as a quick measure of the patient's degree of frailty or biologic age and might provide insight into an individual's tolerance to cancer treatment.

Decline in physical function and loss of independence is of great concern for older adults undergoing cancer treatment, and several studies in the general geriatric population have shown that increased chronic inflammatory and procoagulant markers predict functional decline.^{15-19,45,46} In an analysis of moderately to severely disabled women age 65 years and older on the Women's Health and Aging Study, higher baseline IL-6 levels were associated with significantly higher levels of functional decline, including decreased mobility, activities of daily living deficits, increased walking limitations, and decreased walking speed, compared with women with lower IL-6 levels.⁴⁶ Elevated inflammatory cytokines and procoagulant marker levels also correlate with postoperative complications and functional decline after oncologic surgery.^{16,20} Whether chronic inflammatory markers can independently predict functional decline associated with cancer treatment warrants further study.

Elevated inflammatory markers are also associated with mortality risk in the elderly.^{15,17,19,44} In a study of community dwelling adults (mean age, 78 years), higher levels of sVCAM were independently

associated with poorer baseline functional status and mortality at 4 years (hazard ratio, 1.2; $P = .002$).¹⁷ Higher sVCAM, D-dimer, and IL-6 were independently related to 4-year mortality, adjusting for functional status, demographics, and comorbidities.

Several chronic inflammatory markers, (IL-6, D-dimer, and CRP) also have the ability to predict both decline in function and mortality, even after controlling for age, comorbidities, and physical function,^{15,19,42} and may have greater predictive ability among patients without baseline functional impairments, suggesting they may identify prefrail patients that may not otherwise have been identified without extensive geriatric assessment testing. The majority of studies evaluating chronic inflammatory markers in the general geriatric population are mainly epidemiologic observations. The role of these markers in the management of general geriatric patients is being studied in ongoing geriatric research.

The direct link between increased inflammatory markers and functional decline has not been established. It has been postulated that interconnections between inflammatory cytokines and the CNS, endocrine systems, and musculoskeletal system could result in sarcopenia and/or bone loss leading to decreases in function. Several animal model studies suggest a role in the catabolic effects contributing to muscle wasting,⁴⁷⁻⁴⁹ but whether this is a direct effect of the inflammatory markers or mediated by other circulating factors remains unknown.

The benefits of using chronic inflammatory markers include the ease of measurement. They can be collected during blood draws routinely done for cancer management. They can be measured with enzyme-linked immunosorbent assays, and a panel of inflammatory markers can be included in a multiplex enzyme-linked immunosorbent assay. The problem with assessing chronic inflammatory markers is that they reflect and may be produced by the cancer itself. Experts in the biomarkers of aging development process have recommended that “. . . an aging marker should both correlate with functional age but also be independent from specific pathologic conditions.”⁵⁰ Although chronic inflammatory markers may not technically be true aging biomarkers, they are potentially useful in reflecting correlated processes that may predict oncologic outcomes.

An argument for measuring cytokines in the setting of an ongoing malignancy is that they are associated with poorer PS and quality of life (QOL), as well as higher levels of fatigue,^{51,52} that may affect a patient's ability to tolerate treatment. There is evidence that cytokine levels may provide prognostic information beyond PS. In a study of 377 patients with acute myeloid leukemia, cytokine expression (IL-6, IL-1, IL-2, TNF- α , and CRP) was evaluated in 58 patients identified at the extremes of survival (median survival, 4.9 months and 46.3 months in patients with poor and good prognosis, respectively).⁵³ There was no statistically significant difference in survival when evaluated by PS (Eastern Cooperative Oncology Group 0 to 1 v 2). However, when categorized by cytokine expression levels (low, intermediate, high) and PS, there was a large difference in survival between low-, intermediate-, and high-risk groups (56.1 months, 9.85 months, and 8.35 months, respectively; $P = .02$).

Markers of Cellular Senescence

If a cell does not enter the apoptotic pathway in response to DNA damage, the cell can activate a DNA damage response pathway leading to permanent cell cycle arrest termed cellular senescence.^{54,55} Cellular senescence, a term for the mitotic arrest of a cell, is induced by DNA

damage through several mechanisms, including cell division and subsequent telomere shortening, as well as forms of cellular stress, including activation of oncogenes and oxidative stress.⁵⁵ The assumption is that as a person ages, one accumulates more and more senescent cells.

There may be a connection between senescent cells and the increasing levels of chronic inflammatory markers seen in older individuals. Cellular division has ceased in senescent cells, but they actively secrete proinflammatory proteins collectively known as the senescence-associated secretory phenotype (SASP).⁵⁶ SASP factors are proposed to increase inflammation in surrounding tissues and in the circulation and contribute to the aging process. Factors within the SASP are similar to the inflammatory and coagulation markers associated with frailty and mortality in the elderly population.^{44,57}

Because there are no established circulating markers of cellular senescence, markers associated with senescence, including telomere length and p16^{INK4a}, have been explored as potential biologic markers of aging.

Telomere length. Telomeres are proteins at the ends of chromosomes that shorten with each cell division, eventually leading to mitotic arrest termed “replicative senescence,” which has been proposed to contribute to the aging process.⁵⁸ Because telomere length decreases with increasing age, telomere length has been evaluated as a biomarker of aging. In a study of 143 patients older than 60 years, Cawthon et al²¹ demonstrated a correlation between shorter telomere length by age and increased mortality. Since that time, multiple studies have attempted to replicate this finding with varying results, but because of the correlation with aging as well as age-related disease, telomere length continues to be a marker of interest in the search for markers of biologic age.⁵⁹

Interestingly, telomere length may also have a role in predicting cancer prognosis. Shorter telomere length has been associated with poorer prognosis among patients in retrospective studies of colorectal cancer, soft tissue sarcomas, breast cancer, and lung cancer.⁶⁰ Other studies on the association between telomere length and cancer-related mortality have yielded conflicting results. Telomere length has also been associated with functional status or mortality in several studies.^{21,22-24}

Telomere length can be measured in peripheral blood leukocytes under the presumption that it represents telomere length in other tissues. This marker could be obtained in conjunction with routine blood draws, but it requires laboratory methods that have technical limitations. Telomere length can be measured by estimating the size of cleaved telomere fragments (by using the terminal restriction fragment method⁶¹), shortest telomere length by fluorescent in situ hybridization, or by quantitative real-time polymerase chain reaction.⁵⁹ A potential disadvantage to using telomere length as a biomarker is that it could be affected by many processes other than age, including genetic factors, environmental exposure, and dietary intake.⁵⁹ As with chronic inflammatory markers, a correlation of frailty and telomere length may be important for predicting outcomes of interest, regardless of whether it is considered a pure aging biomarker.

Dysfunctional telomeres. Dysfunctional telomeres may also have a role as biomarkers of aging. In a preclinical study, Jiang et al⁶² identified markers secreted by bone marrow cells with telomere dysfunction (stathmin, CRAMP, EF1 α , and chitinase 3). These markers were found to distinguish old from young adults and healthy older adults versus older adults with age-related diseases. The benefit to

these markers is that they can be detected in the serum, making their measurement much easier than assessments of telomere length.

p16^{INK4a}. The activation of oncogenes or the loss of tumor suppressor function can also induce senescence, primarily controlled by the p53 pathway and its downstream mediators.^{63,64} *p16^{INK4a}* is involved in an alternate pathway involved in permanent cell-cycle arrest that is now recognized as a marker of cellular senescence in animal models.²⁵ It has been demonstrated that *p16^{INK4a}* levels increase with age in mammalian models and humans, and its expression has also been correlated with higher levels of IL-6.²⁵⁻²⁷

The role of *p16* levels in the geriatric oncology population is currently being evaluated in an ongoing clinical trial (NCT00849758). Interestingly, *p16^{INK4a}* levels have recently been found to increase in patients receiving chemotherapy for breast cancer, making this a potential marker for studying the effects of chemotherapy on the aging process.⁶⁵ Levels of *p16^{INK4a}* can be obtained from the blood via quantitative real-time polymerase chain reaction on T-lymphocyte RNA, although the assay is complicated and not generally available at this time.⁶⁰

Imaging for Sarcopenia

Another modality of evaluation that may predict underlying frailty and/or functional age is imaging for sarcopenia. Sarcopenia can be defined as muscle mass two standard deviations below that of a healthy adult.⁶⁶ Sarcopenia is more prevalent in older adults and is a hallmark of frailty and subsequent disability.⁶⁷ In addition, circulating inflammatory mediators likely contribute to the development and progression of sarcopenia.^{68,69}

Beyond sarcopenia, there is currently major interest in body composition and how it correlates with sarcopenia; for example, obese patients can have a decrease in muscle mass termed sarcopenic obesity. A study of 250 patients with body mass index more than 30 evaluated muscle mass by using whole-body dual-energy x-ray absorptiometry and computed tomography scans.⁷⁰ Patients with sarcopenic obesity had poorer response to chemotherapy, poorer PS, and a 10-month decrement in median survival after adjusting for standard predictors of cancer-related mortality.

The advantages to using radiographic technology to assess for sarcopenia is that this information could be collected at the time of routine imaging studies done for tumor evaluation and/or restaging.⁷¹ Sarcopenia may be present before the onset of disability,⁶⁷ and sarcopenia may be disguised by obesity. Therefore, imaging studies for sarcopenia may identify the prefrail or frail patient who otherwise may not have been detected. There is also evidence that body composition may be related to toxicity from cytotoxic chemotherapy.⁷² A disadvantage of imaging evaluation for sarcopenia is the cost of the computerized imaging technology and the expertise required to calculate the degree of muscle mass in each patient. It is uncertain whether radiographic evaluation for sarcopenia will add more to geriatric assessment than standard measures such as grip strength and timed up and go. One possibility would be to use the aforementioned measures as a screening tool, which might prompt further evaluation for sarcopenia.

Other Potential Aging Markers

There are other aging biomarker candidates that may prove to be helpful predictors of oncologic outcomes. These include genes associated with longevity, single nucleotide polymorphisms associated with

aging, lymphopenia, and oxidative stress markers.^{50,60} Data for these potential markers in relation to clinical measures of frailty and mortality prediction is currently scant, but future studies may provide more insight into their use in geriatric oncology.

MOVING FORWARD: INCORPORATING BIOMARKERS INTO ONCOLOGY CLINICAL TRIALS

The need for more accurate assessment of the functional age of patients with older cancer is clear, and clinical trials are beginning to incorporate some of the aforementioned markers. The European Organisation for Research and Treatment of Cancer is including collection of aging biomarkers such as markers of chronic inflammation, aging genes, and markers of cellular senescence in patients enrolled onto clinical trials. The ideal biomarker of functional age or frailty would provide additional information beyond what is collected during routine oncology evaluation and would be easily measured and easily repeated throughout treatment without causing increased burden to the patient.

When evaluating potential biomarkers in clinical trials, we need end points that will accurately predict treatment tolerance and outcomes. Outcomes should include the development of adverse events, functional decline, QOL, and survival, which would enable patients to make better-informed decisions regarding their treatment. Such markers would also help clinicians identify the most appropriate treatment regimen for the patient (ie, full-dose chemotherapy, modified doses, best supportive care) as well as identify patients in need of additional supportive care during treatment.

When assessing biomarkers in the context of cancer and cancer treatment, it will be important to assess the biomarkers at various time points to evaluate the impact of aging markers at different time points in the cancer evaluation and treatment trajectory. The measurement of biomarkers over time could be used to address four important questions.

First, how much does tumor burden and treatment (recent surgery, radiation therapy, and chemotherapy) affect the physical function (degree of frailty) for the individual patient? Starting with adjuvant therapy trials will likely remove the confounding factor of the underlying malignant process contributing to the biomarker levels and potentially clinical measures of frailty and QOL. Consider measuring markers before tumor removal, and at various time points after surgery because surgery itself would be expected to temporarily elevate these markers and contribute to a temporary decline in physical function, depending on the impact of the surgery on the individual patient. The most appropriate time point after surgery to measure biomarkers is not known, but circulating acute phase reactants from the surgery itself should be resolved by 6 to 8 weeks after surgery.^{73,74}

Second, do the increased circulating markers correlate with clinical measures of frailty? And if so, even if the increased biomarkers were the result of the underlying tumor burden, will they still have an impact on treatment tolerance? Will they decrease with treatment or increase? If they decrease, would this improve treatment tolerance to the point that the dose of chemotherapeutic drugs in the regimen could potentially be increased?

Third, will chemotherapy and/or radiation therapy contribute to an increase in senescent cell burden? There are suggestions that chemotherapy contributes to the development of cellular senescence. Would this result in an increase in circulating SASP factors and contribute to a decline in physical function or health-related QOL greater

than that for a patient who had not received chemotherapy? Could a resulting increase in SASP factors contribute to accelerated aging? In studies done on survivors of childhood cancers, investigators have found an increase in age-related pathologies such as atherosclerotic disease.^{75,76} Is this a result of the development of senescent cells in response to treatment resulting in premature aging?

Fourth, if senescent cells contribute to local tissue inflammation (destruction?) resulting in changes such as epithelial to mesenchymal transition, would this result in decreased disease-free survival, progression-free survival, or overall survival?

Future Directions

In the development of biomarkers, it is anticipated that there will be a combination or panel of markers that will have the best predictive power of end points such as toxicity, functional decline, QOL, and survival. As with the geriatric assessment, patients would potentially be placed into frailty and/or aging categories (ie, low-, intermediate-, or high-risk groups) to predict risk for the specific end point. This panel of markers should then be validated in older patients enrolled onto clinical trials. If validated, the panel could then be evaluated as a decision tool in a prospective trial, which could be used to stratify patients and assign them into different management categories such as modified treatment regimens.

Ultimately, research on biomarkers of aging may provide us with more accurate assessments of risk and may also identify biologic targets for interventions to ease the burden of cancer treatment for older patients. Future studies using a biomarker of aging panel could be incorporated into interventional studies to assess whether interventions in patients identified as at risk for poorer outcomes (toxicity or decreased survival or decline in physical function) can be modified or prevented by focused interventions. Potential interventions could include pharmacologic agents targeting cachexia/sarcopenia, chronic inflammation, and/or the burden of senescent cells. Other interventions may include physical rehabilitation, nutritional supplementation, or psychosocial methods.

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

The author(s) indicated no potential conflicts of interest.

AUTHOR CONTRIBUTIONS

Conception and design: Joleen M. Hubbard, Harvey J. Cohen, Hyman B. Muss

Manuscript writing: All authors

Final approval of manuscript: All authors

REFERENCES

- Gross CP, McAvay GJ, Guo Z, et al: The impact of chronic illnesses on the use and effectiveness of adjuvant chemotherapy for colon cancer. *Cancer* 109:2410-2419, 2007
- Jatoi A, Hillman S, Stella PJ, et al: Daily activities: Exploring their spectrum and prognostic impact in older, chemotherapy-treated lung cancer patients. *Support Care Cancer* 11:460-464, 2003
- Muss HB, Woolf S, Berry D, et al: Adjuvant chemotherapy in older and younger women with lymph node-positive breast cancer. *JAMA* 293:1073-1081, 2005
- Sargent DJ, Goldberg RM, Jacobson SD, et al: A pooled analysis of adjuvant chemotherapy for resected colon cancer in elderly patients. *N Engl J Med* 345:1091-1097, 2001
- Kornblith AB, Kemeny M, Peterson BL, et al: Survey of oncologists' perceptions of barriers to accrual of older patients with breast carcinoma to clinical trials. *Cancer* 95:989-996, 2002
- Jessup JM, Stewart A, Greene FL, et al: Adjuvant chemotherapy for stage III colon cancer: Implications of race/ethnicity, age, and differentiation. *JAMA* 294:2703-2711, 2005
- Smith BD, Jiang J, McLaughlin SS, et al: Improvement in breast cancer outcomes over time: Are older women missing out? *J Clin Oncol* 29:4647-4653, 2011
- Mitnitski AB, Graham JE, Mogilner AJ, et al: Frailty, fitness and late-life mortality in relation to chronological and biological age. *BMC Geriatr* 2:1, 2002
- Fried LP, Tangen CM, Walston J, et al: Frailty in older adults: Evidence for a phenotype. *J Gerontol A Biol Sci Med Sci* 56:M146-M156, 2001
- Cesari M, Penninx BW, Pahor M, et al: Inflammatory markers and physical performance in older persons: The InCHIANTI study. *J Gerontol A Biol Sci Med Sci* 59:242-248, 2004
- Ferrucci L, Cavazzini C, Corsi A, et al: Biomarkers of frailty in older persons. *J Endocrinol Invest* 25:10-15, 2002
- Hubbard RE, O'Mahony MS, Savva GM, et al: Inflammation and frailty measures in older people. *J Cell Mol Med* 13:3103-3109, 2009
- Leng SX, Xue QL, Tian J, et al: Inflammation and frailty in older women. *J Am Geriatr Soc* 55:864-871, 2007
- Walston J, McBurnie MA, Newman A, et al: Frailty and activation of the inflammation and coagulation systems with and without clinical comorbidities: Results from the Cardiovascular Health Study. *Arch Intern Med* 162:2333-2341, 2002
- Cohen HJ, Harris T, Pieper CF: Coagulation and activation of inflammatory pathways in the development of functional decline and mortality in the elderly. *Am J Med* 114:180-187, 2003
- de Saint-Hubert M, Jamart J, Morrhaye G, et al: Serum IL-6 and IGF-1 improve clinical prediction of functional decline after hospitalization in older patients. *Aging Clin Exp Res* 23:106-111, 2011
- Huffman KM, Pieper CF, Kraus VB, et al: Relations of a marker of endothelial activation (s-VCAM) to function and mortality in community-dwelling older adults. *J Gerontol A Biol Sci Med Sci* 66:1369-1375, 2011
- Puts MT, Visser M, Twisk JW, et al: Endocrine and inflammatory markers as predictors of frailty. *Clin Endocrinol (Oxf)* 63:403-411, 2005
- Reuben DB, Cheh AI, Harris TB, et al: Peripheral blood markers of inflammation predict mortality and functional decline in high-functioning community-dwelling older persons. *J Am Geriatr Soc* 50:638-644, 2002
- Rønning B, Wyller TB, Seljeflot I, et al: Frailty measures, inflammatory biomarkers and post-operative complications in older surgical patients. *Age Ageing* 39:758-761, 2010
- Cawthon RM, Smith KR, O'Brien E, et al: Association between telomere length in blood and mortality in people aged 60 years or older. *Lancet* 361:393-395, 2003
- Epel ES, Merkin SS, Cawthon R, et al: The rate of leukocyte telomere shortening predicts mortality from cardiovascular disease in elderly men. *Aging (Albany NY)* 1:81-88, 2008
- Farzaneh-Far R, Cawthon RM, Na B, et al: Prognostic value of leukocyte telomere length in patients with stable coronary artery disease: Data from the Heart and Soul Study. *Arterioscler Thromb Vasc Biol* 28:1379-1384, 2008
- Risques RA, Arbeeve KG, Yashin AI, et al: Leukocyte telomere length is associated with disability in older U.S. population. *J Am Geriatr Soc* 58:1289-1298, 2010
- Krishnamurthy J, Torrice C, Ramsey MR, et al: Ink4a/Arf expression is a biomarker of aging. *J Clin Invest* 114:1299-1307, 2004
- Liu Y, Sanoff HK, Cho H, et al: Expression of p16(INK4a) in peripheral blood T-cells is a biomarker of human aging. *Aging Cell* 8:439-448, 2009
- Song Z, von Figura G, Liu Y, et al: Lifestyle impacts on the aging-associated expression of biomarkers of DNA damage and telomere dysfunction in human blood. *Aging Cell* 9:607-615, 2010
- Baumgartner RN, Koehler KM, Gallagher D, et al: Epidemiology of sarcopenia among the elderly in New Mexico. *Am J Epidemiol* 147:755-763, 1998
- Heymsfield SB, Wang Z, Baumgartner RN, et al: Human body composition: Advances in models and methods. *Annu Rev Nutr* 17:527-558, 1997
- Janssen I, Heymsfield SB, Ross R: Low relative skeletal muscle mass (sarcopenia) in older persons is associated with functional impairment and physical disability. *J Am Geriatr Soc* 50:889-896, 2002
- Metter EJ, Talbot LA, Schrager M, et al: Skeletal muscle strength as a predictor of all-cause mortality in healthy men. *J Gerontol A Biol Sci Med Sci* 57:B359-B365, 2002

32. Kanapuru B, Ershler WB: Inflammation, coagulation, and the pathway to frailty. *Am J Med* 122:605-613, 2009
33. Robson SC, Shephard EG, Kirsch RE: Fibrin degradation product D-dimer induces the synthesis and release of biologically active IL-1 beta, IL-6 and plasminogen activator inhibitors from monocytes in vitro. *Br J Haematol* 86:322-326, 1994
34. Carter RA, Wicks IP: Vascular cell adhesion molecule 1 (CD106): A multifaceted regulator of joint inflammation. *Arthritis Rheum* 44:985-994, 2001
35. Ershler WB, Sun WH, Binkley N, et al: Interleukin-6 and aging: Blood levels and mononuclear cell production increase with advancing age and in vitro production is modifiable by dietary restriction. *Lymphokine Cytokine Res* 12:225-230, 1993
36. Sindermann J, Kruse A, Frercks HJ, et al: Investigations of the lymphokine system in elderly individuals. *Mech Ageing Dev* 70:149-159, 1993
37. Fagiolo U, Cossarizza A, Scala E, et al: Increased cytokine production in mononuclear cells of healthy elderly people. *Eur J Immunol* 23:2375-2378, 1993
38. Cushman M, Psaty BM, Macy E, et al: Correlates of thrombin markers in an elderly cohort free of clinical cardiovascular disease. *Arterioscler Thromb Vasc Biol* 16:1163-1169, 1996
39. Bruunsgaard H, Pedersen M, Pedersen BK: Aging and proinflammatory cytokines. *Curr Opin Hematol* 8:131-136, 2001
40. Franceschi C, Capri M, Monti D, et al: Inflammaging and anti-inflammaging: A systemic perspective on aging and longevity emerged from studies in humans. *Mech Ageing Dev* 128:92-105, 2007
41. Vasto S, Candore G, Balistreri CR, et al: Inflammatory networks in ageing, age-related diseases and longevity. *Mech Ageing Dev* 128:83-91, 2007
42. Pieper CF, Rao KM, Currie MS, et al: Age, functional status, and racial differences in plasma D-dimer levels in community-dwelling elderly persons. *J Gerontol A Biol Sci Med Sci* 55:M649-M657, 2000
43. Yao X, Li H, Leng SX: Inflammation and immune system alterations in frailty. *Clin Geriatr Med* 27:79-87, 2011
44. De Martinis M, Franceschi C, Monti D, et al: Inflammation markers predicting frailty and mortality in the elderly. *Exp Mol Pathol* 80:219-227, 2006
45. Ferrucci L, Harris TB, Guralnik JM, et al: Serum IL-6 level and the development of disability in older persons. *J Am Geriatr Soc* 47:639-646, 1999
46. Ferrucci L, Penninx BW, Volpato S, et al: Change in muscle strength explains accelerated decline of physical function in older women with high interleukin-6 serum levels. *J Am Geriatr Soc* 50:1947-1954, 2002
47. Goodman MN: Interleukin-6 induces skeletal muscle protein breakdown in rats. *Proc Soc Exp Biol Med* 205:182-185, 1994
48. Todorov P, Cariuk P, McDevitt T, et al: Characterization of a cancer cachectic factor. *Nature* 379:739-742, 1996
49. Mitch WE, Goldberg AL: Mechanisms of muscle wasting: The role of the ubiquitin-proteasome pathway. *N Engl J Med* 335:1897-1905, 1996
50. Falandry C, Gilson E, Rudolph KL: Are aging biomarkers clinically relevant in oncogeriatrics? *Crit Rev Oncol Hematol* 85:257-265, 2013
51. Panju AH, Danesh A, Minden MD, et al: Associations between quality of life, fatigue, and cytokine levels in patients aged 50+ with acute myeloid leukemia. *Support Care Cancer* 17:539-546, 2009
52. Rich T, Innominato PF, Boerner J, et al: Elevated serum cytokines correlated with altered behavior, serum cortisol rhythm, and dampened 24-hour rest-activity patterns in patients with metastatic colorectal cancer. *Clin Cancer Res* 11:1757-1764, 2005
53. Rao AV, Valk PJ, Metzeler KH, et al: Age-specific differences in oncogenic pathway dysregulation and anthracycline sensitivity in patients with acute myeloid leukemia. *J Clin Oncol* 27:5580-5586, 2009
54. Campisi J: Senescent cells, tumor suppression, and organismal aging: Good citizens, bad neighbors. *Cell* 120:513-522, 2005
55. Collado M, Blasco MA, Serrano M: Cellular senescence in cancer and aging. *Cell* 130:223-233, 2007
56. Coppé JP, Patil CK, Rodier F, et al: Senescence-associated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor. *PLoS Biol* 6:2853-2868, 2008
57. Freund A, Orjalo AV, Desprez PY, et al: Inflammatory networks during cellular senescence: Causes and consequences. *Trends Mol Med* 16:238-246, 2010
58. Hemann MT, Strong MA, Hao LY, et al: The shortest telomere, not average telomere length, is critical for cell viability and chromosome stability. *Cell* 107:67-77, 2001
59. Mather KA, Jorm AF, Parslow RA, et al: Is telomere length a biomarker of aging? A review. *J Gerontol A Biol Sci Med Sci* 66:202-213, 2011
60. Pallis AG, Hatse S, Brouwers B, et al: Evaluating the physiological reserves of older patients with cancer: The value of potential biomarkers of aging? *J Geriatr Oncol* 5:204-218, 2014
61. Harley CB, Futcher AB, Greider CW: Telomeres shorten during ageing of human fibroblasts. *Nature* 345:458-460, 1990
62. Jiang H, Schiffer E, Song Z, et al: Proteins induced by telomere dysfunction and DNA damage represent biomarkers of human aging and disease. *Proc Natl Acad Sci U S A* 105:11299-11304, 2008
63. Campisi J: Cellular senescence as a tumor-suppressor mechanism. *Trends Cell Biol* 11:S27-S31, 2001
64. Brown JP, Wei W, Sedivy JM: Bypass of senescence after disruption of p21CIP1/WAF1 gene in normal diploid human fibroblasts. *Science* 277:831-834, 1997
65. Muss HB, Krishnamurthy J, Alston SM, et al: P16INK4a expression after chemotherapy in older women with early-stage breast cancer. *J Clin Oncol* 29:550s, 2011 (suppl; abstr 9002)
66. Roubenoff R: Sarcopenia: Effects on body composition and function. *J Gerontol A Biol Sci Med Sci* 58:1012-1017, 2003
67. Baumgartner RN, Wayne SJ, Waters DL, et al: Sarcopenic obesity predicts instrumental activities of daily living disability in the elderly. *Obes Res* 12:1995-2004, 2004
68. Zoico E, Roubenoff R: The role of cytokines in regulating protein metabolism and muscle function. *Nutr Rev* 60:39-51, 2002
69. Schragger MA, Metter EJ, Simonsick E, et al: Sarcopenic obesity and inflammation in the InCHIANTI study. *J Appl Physiol* 102:919-925, 2007
70. Prado CM, Lieffers JR, McCargar LJ, et al: Prevalence and clinical implications of sarcopenic obesity in patients with solid tumours of the respiratory and gastrointestinal tracts: A population-based study. *Lancet Oncol* 9:629-635, 2008
71. Prado CM, Birdsell LA, Baracos VE: The emerging role of computerized tomography in assessing cancer cachexia. *Curr Opin Support Palliat Care* 3:269-275, 2009
72. Prado CM, Baracos VE, McCargar LJ, et al: Body composition as an independent determinant of 5-fluorouracil-based chemotherapy toxicity. *Clin Cancer Res* 13:3264-3268, 2007
73. Baigrie RJ, Lamont PM, Kwiatkowski D, et al: Systemic cytokine response after major surgery. *Br J Surg* 79:757-760, 1992
74. Wirtz DC, Heller KD, Miltner O, et al: Interleukin-6: A potential inflammatory marker after total joint replacement. *Int Orthop* 24:194-196, 2000
75. Mueller S, Fullerton HJ, Stratton K, et al: Radiation, atherosclerotic risk factors, and stroke risk in survivors of pediatric cancer: A report from the Childhood Cancer Survivor Study. *Int J Radiat Oncol Biol Phys* 86:649-655, 2013
76. Armstrong GT, Liu Q, Yasui Y, et al: Late mortality among 5-year survivors of childhood cancer: A summary from the Childhood Cancer Survivor Study. *J Clin Oncol* 27:2328-2338, 2009

