

Diagnostic Assessment & Prognosis

Revolutionizing Alzheimer's disease and clinical trials through biomarkers

Niklas Mattsson^{a,*}, Maria C. Carrillo^b, Robert A. Dean^c, Michael D. Devous, Sr.^d,
Tania Nikolcheva^e, Pedro Pesini^f, Hugh Salter^{g,h}, William Z. Potterⁱ, Reisa S. Sperling^j,
Randall J. Bateman^k, Lisa J. Bain^l, Enchi Liu^m

^aClinical Memory Research Unit, Lund University, Sweden

^bAlzheimer's Association, Chicago, IL, USA

^cEli Lilly & Co, Inc., Indianapolis, IN, USA

^dAvid Radiopharmaceuticals, Philadelphia, PA, USA

^eF. Hoffmann-La Roche, Basel, Switzerland

^fAraclon Grifols, Zaragoza, Spain

^gAztraZeneca, Stockholm, Sweden

^hDepartment of Clinical Neuroscience, Karolinska Institutet, Sweden

ⁱFoundation for the NIH, Bethesda, MD, USA

^jBrigham and Women's Hospital, Boston, MA, USA

^kWashington University School of Medicine, St. Louis, MO, USA

^lElverson, PA, USA

^mJanssen Research and Development, LLC., San Diego, CA, USA

Abstract

The Alzheimer's Association's Research Roundtable met in May 2014 to explore recent progress in developing biomarkers to improve understanding of disease pathogenesis and expedite drug development. Although existing biomarkers have proved extremely useful for enrichment of subjects in clinical trials, there is a clear need to develop novel biomarkers that are minimally invasive and that more broadly characterize underlying pathogenic mechanisms, including neurodegeneration, neuroinflammation, and synaptic dysfunction. These may include blood-based assays and new neuropsychological testing protocols, as well as novel ligands for positron emission tomography imaging, and advanced magnetic resonance imaging methodologies. In addition, there is a need for biomarkers that can serve as theragnostic markers of response to treatment. Standardization remains a challenge, although international consortia have made substantial progress in this area and provide lessons for future standardization efforts.

© 2015 The Authors. Published by Elsevier Inc. on behalf of the Alzheimer's Association. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Keywords: Alzheimer's disease; Biomarkers; Amyloid; Tau; PET; Imaging; MRI; Cerebrospinal fluid; CSF; Blood biomarkers; Clinical trials

1. Introduction

A biomarker is a characteristic that can be objectively measured and evaluated as an indicator of a normal or path-

ologic process, or as a measure of response to therapy [1] (Biomarker Working Group 2001). Biomarker research has revolutionized the understanding of Alzheimer's disease (AD) and is in the process of transforming the design of AD clinical trials. Until recently, AD was only imprecisely diagnosed in life using clinical assessments during the dementia stage or at time of death by neuropathology. Nonetheless, substantial progress over the past decades in

*Corresponding author. Tel.: +46-(0)-40-33-50-36; Fax: +46-(0)-40-33-56-57.

E-mail address: Niklas.mattsson@med.lu.se

developing cerebrospinal fluid (CSF) and imaging biomarkers has shown that AD brain changes can be detected and used for diagnosis and prognosis of AD [2,3].

As these biomarkers have been included in observational studies of AD, better understanding of the biochemical and pathologic changes of AD has occurred. This has led to confirmation of the hypothesis [4,5] that AD is a disease progressing from preclinical to early and then late clinical stages, and which is now emphasized in novel research diagnostic criteria incorporating biomarkers [6]. Previously, drug developers focused on the dementia stage of the disease. This has now radically changed as clinical trials move toward earlier stages of AD, before extensive neurodegeneration has occurred [7–9], and even to secondary prevention before symptom onset [10–12], when disease-modifying treatments are likely to have maximal effect. Biomarkers play a key role in the design of these trials, both for inclusion of subjects with AD pathology and to track biological effects of drugs. Yet even though it is a widely held belief that AD biomarkers can be used for diagnosis, prognosis/prediction, and to monitor the effects of therapy [1,13], in the absence of an effective treatment to slow progression of AD (and the underlying pathogenic processes), the link between biomarkers and effect on disease cannot be established.

Data from many studies all over the world, including the Alzheimer's Disease Neuroimaging Initiative [14], its worldwide partners (WW-ADNI) [15], and the Dominantly Inherited Alzheimer's Network (DIAN) [16], have done much to delineate the temporal changes in biomarkers over time and clarify their relationship to cognition and function. Yet despite the field's growing acceptance of the need for biomarkers in drug development, the belief that biomarkers could improve clinical trial design and the success of those trials was shaken by recent mixed clinical trial results. The phase III bapineuzumab trial, in particular, went forward in part based on findings in phase II studies that showed modest reductions in brain amyloid [17], and CSF phosphorylated tau (P-tau) concentrations [18]. The presumption that these biomarker effects represented a clinically relevant treatment effect, however, was called into question when no clinical benefit was found in the phase III trials, despite hints that biomarkers were impacted by therapy [19]. The phase III results for solanezumab also did not provide statistically significant effects for coprimary outcomes; however, planned secondary analyses were consistent with clinical benefit of solanezumab in patients with mild AD dementia without evidence of an impact of solanezumab on brain amyloid burden, downstream neurodegeneration markers of CSF tau proteins or brain volume, but with an increase in total CSF A β_{42} and A β_{40} [20].

In this setting, the Alzheimer's Association Research Roundtable convened a meeting in May, 2014 to explore the extent to which biomarkers have furthered our understanding of the disease, supported drug development, and improved the care of patients; and more importantly, to identify what needs to be done to realize their full potential. Can

biomarkers indeed provide answers to guide future trials toward more successful outcomes? The Roundtable examined evidence to support this premise, identified unanswered questions, and explored areas of potential collaboration in precompetitive space among key stakeholders that might expedite this effort.

2. Biomarkers as enrichment tools for clinical trials

Further critical examination of the bapineuzumab and solanezumab studies suggested several possible reasons for the negative trial results. One contributing factor is that some of the enrolled trial subjects may not have AD [21]. Clinical criteria for patient inclusion in each program resulted in study populations with a significant percentage of participants without evidence of brain amyloid by positron emission tomography (PET; ~7 and 36% amyloid negative in apolipoprotein E (*APOE*) $\epsilon 4$ carriers and noncarriers, respectively) [22]. Using amyloid biomarkers to enrich for trial subjects who are amyloid-positive—and thus presumably on the AD trajectory—may improve the ability of future trials to detect a treatment effect especially for anti-amyloid therapies. Indeed, data from several studies have shown that among cognitively normal elderly, those who are amyloid-positive are at greater risk of decline compared with those who are amyloid-negative [6,23–26]. In the placebo arms of both the bapineuzumab and solanezumab studies, which enrolled subjects with mild-to-moderate AD, amyloid-positive subjects had significant decline on both cognitive and functional measures, whereas the amyloid-negative subjects did not [22]. Importantly, the effect of amyloid pathology on longitudinal memory decline may be greater in *APOE* $\epsilon 4$ carriers compared with *APOE* $\epsilon 4$ -noncarriers [27].

Disease severity may be another factor that contributed to the negative trial results. In comparison with subjects with mild disease, those with more advanced clinical disease may have far more advanced neurodegeneration. The modest impact of treatment on the underlying pathology and markers of the pathology may not be sufficient to translate to a clinical benefit. In the bapineuzumab studies, even individuals with the largest reported decrease in amyloid still had elevated values in the AD range and although significant treatment differences were observed between bapineuzumab and placebo, the change from baseline values in the bapineuzumab groups ranged ~0%–10% (with reductions in the CSF P-tau concentration and inhibited further accumulation of brain amyloid by PET) [17–19]. Finally, it is possible that the presence of copathologies (for example tau, vascular, Lewy body, or transactive response DNA binding protein 43 [TDP-43] pathology) may influence cognitive trajectories independent of amyloid pathology [28,29] and impact trial results.

Many trials currently underway or planned are therefore enrolling subjects in earlier stages of disease and using amyloid biomarkers, either amyloid PET imaging or CSF A β_{42} levels, to enrich for trial subjects thought more likely to

Table 1
Current ongoing or planned AD clinical trials sponsored by pharmaceutical industry as reported at the Research Roundtable (May 2014)

	A4 (Lilly)	AstraZeneca	Biogen Idec	Eisai	Eli Lilly	Merck	Merck pAD	Roche	Takeda (TOMMORROW)
Phase/duration	3.3 y	2.0 y	Ph1b/2.0 y	Ph2b/1.5 y	Ph3/1.5 y	Ph3/1.5 y	Ph3/1.5 y	Ph3/2.0 y	Ph3/2.0 y
Population	Amyloid+; 65–85 yo; normal cognition	Amyloid+; 55–85 yo; mild and pAD	Amyloid+; mild and pAD	Amyloid+; early AD (mild and pAD)	Amyloid+; mild AD	55–85 yo; MtM AD	Amyloid+ 50–85 yo; pAD	Amyloid+; (1) pAD 50–85 yo; (2) mild AD, 50–90 yo	Preclinical AD (asymptomatic at risk for MCI due to AD)
Sample Size	1000+	1500+	~160	Adaptive; up to 800	2100	1960	1500	(1) 700; (2) 1000	3800
Regions	USA, Canada, Australia	Global	USA	North America, EU	North America, EU, Japan	Global	Global	Global	North America, EU, Russia, Australia
Primary Biomarker Screen	Amyloid PET (Amyvid)	Amyloid PET or CSF A β ₄₂	Amyloid PET (Amyvid)	Amyloid PET	Amyloid PET (Amyvid1) or CSF A β ₄₂	None	Amyloid PET (Vizamyl)	CSF A β ₄₂	TOMM40 rs10524523, APOE, age (Risk algorithm)

Abbreviations: AD, Alzheimer's disease; MCI, mild cognitive impairment; PET, positron emission tomography; CSF, cerebrospinal fluid; y, years; yo, years old; pAD, prodromal AD; MtM, mild-to-moderate.

benefit from therapy, as reported at this Roundtable (Table 1). Only the TOMMORROW trial of the mitochondrial targeting agent pioglitazone (an approved antidiabetic prescription drug) uses a genetic enrichment strategy based on *TOMM40* and *APOE* genotype and age to identify normal individuals at risk of developing mild cognitive impairment over a 5-year period.

Several considerations are raised with the use of amyloid PET imaging or CSF A β ₄₂ as a screening tool and enrichment criterion to be addressed for trials of anti-amyloid therapies:

- How practical is it to require amyloid positivity by PET or CSF as an inclusion criterion in a large (global) trial?
- Is there an advantage of one amyloid test over the other (CSF vs. PET)?
- Is it possible to establish standard cutoffs for PET or CSF amyloid assessments to differentiate between normal and pathologic state?
- Will standard cutoffs differ based on the stage of disease, for example as discussed in Lim et al. (2015) [30]?
- Are quantitative PET reads required, or is a visual read sufficient?
- Are the different radiotracers interchangeable, or is it necessary for all screens to be performed using a single radiotracer?
- Are different CSF assays interchangeable? Can novel development of reference standard procedures and materials overcome variability problems?
- How can between-site variability for PET scans or CSF biomarker measurements be overcome?
- If a subject has had a previous amyloid biomarker measurement that indicates amyloid-positivity, can this historical data be used to satisfy the inclusion criterion?

- What are the labeling implications of having amyloid positivity as an eligibility criterion? Will the amyloid tests be considered companion diagnostics?

3. Supporting a disease modification claim with biomarkers

In the US Food and Drug Administration (FDA) draft guidance on developing drugs for early stage disease, disease modification is defined as a “direct effect on the underlying disease pathophysiology.” The guidance goes on to consider the possibility “that a claim of disease modification could be supported by evidence of a meaningful effect on a biomarker in combination with a clinical benefit” [31]. In its Guideline on Medicinal Products for the Treatment of Alzheimer's Disease and Other Dementias, the European Medicines Agency defines disease modification as “slowing or arrest of symptom progression of the dementing process,” and like FDA, suggests that “Ideally proof of a disease modifying effect would require demonstration of clinically relevant changes... [and] supportive evidence for a change in the underlying disease process based on biological markers,” [32].

Based on research from the past several decades and more recent data from larger observational studies such as ADNI, Australian Imaging, Biomarker & Lifestyle Flagship Study of Ageing (AIBL), and DIAN, a consensus is emerging that AD begins decades before dementia onset [5,33] and is heralded by successive changes in markers of the underlying disease processes [4,34]. Several biomarkers are known to be relevant in AD and have been studied intensely [35–38]. Hippocampal atrophy rates show high correlation with cognitive change [39]. However, by its very nature, magnetic resonance imaging (MRI) does not

measure a single molecular entity, thus atrophy could reflect not only neuron loss but other neurodegeneration-associated events including loss of axonal tracts/dendrites, loss of other cell types (e.g. astrocytes, microglia), inflammatory processes, and associated alterations of interstitial fluid volume [37,40].

Other available biomarkers that reflect pathogenic processes of the disease include CSF levels of total tau (T-tau) and P-tau, which are related to cortical axonal degeneration and tangle pathology [41,42]; CSF A β_{42} and amyloid PET imaging, which correlate negatively and positively, respectively, with amyloid plaque load at autopsy [43]; and fludeoxyglucose (FDG)-PET, a measure of glucose metabolism, which may partly reflect synaptic dysfunction [44].

However, even if a biomarker has been shown to be relevant to AD, its utility as a theragnostic biomarker is not guaranteed. For example, although biomarkers such as CSF T-tau/P-tau and volumetric MRI are clearly affected in observational studies of AD, the effect of a disease-modifying therapy on these biomarkers may be influenced by multiple factors, such as the mechanism of action of the therapeutic or related disease processes. Thus, each candidate biomarker must be evaluated empirically for response to therapy.

For example, although both bapineuzumab and solanezumab are monoclonal antibodies against the A β_{42} peptide, there were differences in which AD relevant biomarkers responded to treatment. These differences may be due to differences in binding properties of the antibodies, i.e., soluble versus aggregated amyloid [45–47], and illustrate the fact that not all AD relevant biomarkers may serve as theragnostic biomarkers in all trials and that multiple factors will influence their utility. An additional point is that the observed biomarker changes in response to therapy were small to modest and not accompanied by a slowing of decline in cognition and, thus, do not support a disease-modifying effect. To date, it is unknown to what degree treatment-induced change in a biomarker would be sufficient to test the underlying therapeutic hypothesis. Thus, it is premature to judge the theragnostic value of any biomarker that has, to date, only been modestly perturbed by any treatment.

Furthermore, differentiation between utility as biomarkers of target engagement versus theragnostic response may be needed. For an anti-amyloid treatment, an amyloid marker may be used to assess target engagement, but use of a downstream neurodegenerative marker, such as a marker of tau biology, may be required for therapeutic monitoring. Similarly, for a therapeutic agent targeting tau pathology, measurement of CSF tau levels or tau PET imaging might prove useful for trial enrichment and/or target engagement, but other markers might be required to characterize treatment effect on the downstream pathology. Thus, novel biomarkers need to be discovered and/or developed that can characterize other aspects of the neurodegenerative process. Importantly, when analyzing biomarkers, whether new or old, longitudinal studies are needed that provide evidence of change over time.

4. Standardization

For biomarkers to be used in clinical trials, standardization of sample and data collection protocols and analytical methods is essential. Particularly, the ability to combine data across time and study sites is crucial. Perhaps more importantly, the ability to reduce or control variability introduced by the measurement methodology improves the potential to detect and measure biological changes resulting from therapeutic intervention. Standardization can promote comparison across studies and ease the transition from research use to use in a regulatory environment (pivotal clinical trials) and in clinical practice. Here, we will review standardization efforts for CSF and imaging biomarkers.

Currently there are several commercially available assays to measure CSF biomarkers (A β_{42} , P-tau, and T-tau), some of which have Conformité Européenne (CE)-marking in Europe, but none of which have been cleared by FDA. Although performance of the different assays to differentiate between normal and pathologic is similar, the absolute values reported by them for any particular analyte are different [48]. There are several different possible sources of preanalytical and analytical variability of CSF biomarker measurements [49]. Standardization efforts for CSF biomarkers to address preanalytical and analytical issues have been initiated by the Alzheimer's Association (AA) under the Global Biomarkers Standardization Consortium [50]. To date, the lack of standardization across available assays has required use of assay/laboratory-specific cut points to convert numerical data from each method into clinically meaningful categorical results. The standardization programs described previously aim to enable a universally acceptable cut point to differentiate normal from pathologic states using CSF biomarker results generated across time, assays, and laboratories.

Quantitative analyses from amyloid PET images acquired from multicenter studies require standardization of protocols to reduce measurement variability and error with attention and emphasis on different aspects when conducting cross-sectional or longitudinal comparisons. Standardization efforts involve three main areas of focus: scan acquisition, image processing, and image analyses. A comprehensive review of these topics is recently detailed [51]. With the introduction of multiple tracers, each of which with slightly different characteristics, comparison of results across studies has become even more challenging. The Centiloid Project proposes to calibrate all amyloid imaging tracers according to a unified and standardized numerical scale [52] and would theoretically allow results from studies using different tracers to be combined and compared.

Standardization of methods for structural MRI measures of hippocampal volume has been undertaken by the European Alzheimer's Disease Consortium (EADC)-ADNI Hippocampal Harmonization Protocol project, with funding from the AA and six medical device, diagnostics, and pharmaceutical companies. This project has developed a harmonized protocol for estimating hippocampal volume through

manual segmentation [53], which addresses issues relating to acquisition of images (e.g., different manufacturers, sequences, positioning of the patient), variability in algorithms, readers, and patient characteristics.

5. Pattern and time course of biomarker changes in AD

The hypothetical model of dynamic biomarkers proposed by Jack et al. [4], in which a gradual, ordered, and successive alteration in AD biomarkers, beginning with amyloid pathology, then tauopathy, and later neurodegeneration, preceding clinical symptoms, and eventual dementia, recapitulates the key tenets of the amyloid cascade hypothesis [54,55]. Although recent data support many aspects of this model [2,16,26,56–58], some do not and the model has been updated accordingly [34]. The main impetus for the change is due to the evidence that medial temporal lobe tau pathology occurs in normal aging independent of amyloid [59,60]. Furthermore, data support that amyloidosis and neurodegeneration (tauopathy) may initiate independently but that abnormal (high levels) amyloid and tau interact with amyloid accelerating the downstream neurodegeneration [61]. Thus, although the model provides a framework for the general trend of biomarker changes at a population level, more work is needed to understand possible individual differences in the ordering of biomarker changes [34].

6. Novel biomarkers

Roundtable participants agree on the need for additional novel biomarkers for other aspects of AD pathophysiology, including neuroinflammation, neurodegeneration, synaptic dysfunction, and other associated neuropathologies (e.g., α -synuclein) and comorbid conditions. Although CSF biomarkers provide the most direct biochemical access to the brain with few confounds from other organs [62], blood-based biomarkers are desired for large-scale screening. Novel imaging biomarkers, including tau imaging [63–67], advanced MRI methodologies [68,69], and new neuropsychological testing protocols, have also shown promise in providing a more comprehensive picture of the progression of dementing diseases [70–73].

A number of novel CSF biomarkers have been identified for staging and potentially tracking AD, including visinin-like protein (VILIP)-1 [74,75], YKL-40 [76] and heart-type fatty acid binding protein (HFABP) [77], among others. Novel biomarkers may also be helpful in differentiating AD patients with copathologies.

Ideally, to enrich studies for subjects who are likely to have AD pathology, low cost and minimally invasive biomarkers optimized for sensitivity may be part of a screening funnel that can reduce the number of subjects to be evaluated by a more costly/invasive amyloid test (PET or CSF). Blood-based assays are currently in development for a variety of analytes, including various species of A β and tau, α -synuclein,

and TDP-43. Furthermore, proteomic studies have identified hundreds of potential markers of inflammation, oxidative stress, mitochondrial dysfunction, neuronal and microvascular injury, and metabolic processes. Investigators have constructed several multianalyte panels to detect, classify, and predict disease progression, although these findings require replication [78–80]. Other approaches such as microRNA and plasma exosome analyses are also being investigated for utility in identifying individuals likely to have AD pathology [81–83]. An international working group of experts in the field has been formed to address the challenges that have hampered development of blood-based biomarkers [84].

7. Conclusion and steps forward

The Roundtable concluded that more longitudinal studies on biomarker trajectories, specifically evaluating change in individuals, are needed to confirm or modify the findings of cross-sectional studies that have dominated the field to this point. In addition, there was widespread support for increased effort and focus on studies linking neuropathology to biomarkers, and a suggestion that additional public-private funding may be needed to achieve this. Specifically, it would be valuable to assess and compare postmortem samples from patients involved in different clinical trials. Finally, novel biomarkers that reflect other disease processes downstream of the initiating AD pathologies would not only increase our understanding of AD but can potentially provide needed tools for next-generation AD therapies in development.

RESEARCH IN CONTEXT

1. Systematic review: There have been many reviews on Alzheimer's disease (AD) biomarkers in the last 5 years, but this article is unique that the Alzheimer's Association Research Roundtable provides an unparalleled forum in which the leading experts on Alzheimer's disease from pharmaceutical industry, nonprofit organizations, and governmental organizations can collaborate on moving the field forward by identifying areas of research that are most critical to achieve a successful next generation AD therapeutic.
2. Interpretation: This article summarizes the foremost areas of work in the view of industry drug developers to achieve a successful next generation AD therapeutic.
3. Future directions: Continued work in the Alzheimer's Association Research Roundtable (AARR) precompetitive space will allow optimization of resource use across different sectors.

References

- [1] Biomarkers Definitions Working G. Biomarkers and surrogate endpoints: Preferred definitions and conceptual framework. *Clin Pharmacol Ther* 2001;69:89–95.
- [2] Fagan AM, Xiong C, Jasielec MS, Bateman RJ, Goate AM, Benzinger TL, et al. Longitudinal change in CSF biomarkers in autosomal-dominant Alzheimer's disease. *Sci Transl Med* 2014;6:226ra30.
- [3] Shaw LM, Vanderstichele H, Knapik-Czajka M, Clark CM, Aisen PS, Petersen RC, et al. Cerebrospinal fluid biomarker signature in Alzheimer's disease neuroimaging initiative subjects. *Ann Neurol* 2009;65:403–13.
- [4] Jack CR Jr, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, et al. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. *Lancet Neurol* 2010;9:119–28.
- [5] Sperling RA, Aisen PS, Beckett LA, Bennett DA, Craft S, Fagan AM, et al. Toward defining the preclinical stages of Alzheimer's disease: Recommendations from the National Institute on Aging and the Alzheimer's Association workgroup. *Alzheimers Dement* 2011;7:280–92.
- [6] Morris JC, Roe CM, Grant EA, Head D, Storandt M, Goate AM, et al. Pittsburgh compound B imaging and prediction of progression from cognitive normality to symptomatic Alzheimer disease. *Arch Neurol* 2009;66:1469–75.
- [7] Schneider LS, Mangialasche F, Andreasen N, Feldman H, Giacomini E, Jones R, et al. Clinical trials and late-stage drug development for Alzheimer's disease: An appraisal from 1984 to 2014. *J Intern Med* 2014;275:251–83.
- [8] Gomez-Isla T, Price JL, McKeel DW Jr, Morris JC, Growdon JH, Hyman BT. Profound loss of layer II entorhinal cortex neurons occurs in very mild Alzheimer's disease. *J Neurosci* 1996;16:4491–500.
- [9] Price JL, Ko AI, Wade MJ, Tsou SK, McKeel DW, Morris JC. Neuron number in the entorhinal cortex and CA1 in preclinical Alzheimer disease. *Arch Neurol* 2001;58:1395–402.
- [10] Sperling RA, Rentz DM, Johnson KA, Karlawish J, Donohue M, Salmon DP, et al. The A4 study: Stopping AD before symptoms begin? *Sci Transl Med* 2014;6:228f13.
- [11] Bateman RJ, Aisen PS, De Strooper B, Fox NC, Lemere CA, Ringman JM, et al. Autosomal-dominant Alzheimer's disease: A review and proposal for the prevention of Alzheimer's disease. *Alzheimers Res Ther* 2011;3:1.
- [12] Reiman EM, Langbaum JB, Fleisher AS, Caselli RJ, Chen K, Ayutyanont N, et al. Alzheimer's Prevention Initiative: A plan to accelerate the evaluation of presymptomatic treatments. *J Alzheimers Dis* 2011;26(Suppl 3):321–9.
- [13] Bateman RJ, Klunk WE. Measuring target effect of proposed disease-modifying therapies in Alzheimer's disease. *Neurotherapeutics* 2008;5:381–90.
- [14] Weiner MW, Veitch DP, Aisen PS, Beckett LA, Cairns NJ, Green RC, et al. The Alzheimer's Disease Neuroimaging Initiative: A review of papers published since its inception. *Alzheimers Dement* 2013;9:e111–94.
- [15] Carrillo MC, Bain LJ, Frisoni GB, Weiner MW. Worldwide Alzheimer's disease neuroimaging initiative. *Alzheimers Dement* 2012;8:337–42.
- [16] Bateman RJ, Xiong C, Benzinger TLS, Fagan AM, Goate A, Fox NC, et al. Clinical, cognitive, and biomarker changes in the Dominantly Inherited Alzheimer Network. *N Engl J Med* 2012;367:795–804.
- [17] Rinne JO, Brooks DJ, Rossor MN, Fox NC, Bullock R, Klunk WE, et al. 11C-PiB PET assessment of change in fibrillar amyloid-beta load in patients with Alzheimer's disease treated with bapineuzumab: A phase 2, double-blind, placebo-controlled, ascending-dose study. *Lancet Neurol* 2010;9:363–72.
- [18] Blennow K, Zetterberg H, Rinne JO, Salloway S, Wei J, Black R, et al. Effect of immunotherapy with bapineuzumab on cerebrospinal fluid biomarker levels in patients with mild to moderate Alzheimer disease. *Arch Neurol* 2012;69:1002–10.
- [19] Salloway S, Sperling R, Fox NC, Blennow K, Klunk W, Raskind M, et al. Two phase 3 trials of bapineuzumab in mild-to-moderate Alzheimer's disease. *N Engl J Med* 2014;370:322–33.
- [20] Doody RS, Thomas RG, Farlow M, Iwatsubo T, Vellas B, Joffe S, et al. Phase 3 trials of solanezumab for mild-to-moderate Alzheimer's disease. *N Engl J Med* 2014;370:311–21.
- [21] Karran E, Hardy J. Anti-amyloid therapy for Alzheimer's disease—are we on the right road? *N Engl J Med* 2014;370:377–8.
- [22] Salloway S, Sperling R, Gregg K, Yu P, Joshi A, Lu M, et al. Incidence and clinical progression of placebo-treated amyloid-negative subjects with mild-to-moderate Alzheimer's disease (AD): Results from the phase III PET substudies of bapineuzumab and solanezumab. (P4-417). *Alzheimers Dement* 2013;9(4 Suppl):P889.
- [23] Knopman DS, Jack CR Jr, Wiste HJ, Weigand SD, Vemuri P, Lowe V, et al. Short-term clinical outcomes for stages of NIA-AA preclinical Alzheimer disease. *Neurology* 2012;78:1576–82.
- [24] Lim YY, Maruff P, Pietrzak RH, Ellis KA, Darby D, Ames D, et al. Abeta and cognitive change: Examining the preclinical and prodromal stages of Alzheimer's disease. *Alzheimers Dement* 2014;10:743–751.
- [25] Lim YY, Pietrzak RH, Ellis KA, Jaeger J, Harrington K, Ashwood T, et al. Rapid decline in episodic memory in healthy older adults with high amyloid-beta. *J Alzheimers Dis* 2013;33:675–9.
- [26] Vos SJ, Xiong C, Visser PJ, Jasielec MS, Hassenstab J, Grant EA, et al. Preclinical Alzheimer's disease and its outcome: A longitudinal cohort study. *Lancet Neurol* 2013;12:957–65.
- [27] Lim YY, Laws SM, Villemagne VL, Snyder PJ, Ames D, Martins RN, et al. Abeta-related memory decline in preclinical AD is delayed in APOE e4 non-carriers. *AAIC* 2015. 2015.
- [28] Schneider JA, Arvanitakis Z, Leurgans SE, Bennett DA. The neuropathology of probable Alzheimer disease and mild cognitive impairment. *Ann Neurol* 2009;66:200–8.
- [29] Toledo JB, Arnold SE, Raible K, Brettschneider J, Xie SX, Grossman M, et al. Contribution of cerebrovascular disease in autopsy confirmed neurodegenerative disease cases in the National Alzheimer's Coordinating Centre. *Brain* 2013;136(Pt 9):2697–706.
- [30] Lim YY, Maruff P, Schindler R, Ott BR, Salloway S, Yoo DC, et al. Disruption of cholinergic neurotransmission exacerbates Abeta-related cognitive impairment in preclinical Alzheimer's disease. *Neurobiol Aging* 2015;36:2709–15.
- [31] Food and Drug Administration. Draft Guidance for Industry. Alzheimer's disease: Developing drugs for the treatment of early stage disease. Available at: <http://www.fda.gov/downloads/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/UCM338287.pdf>. Accessed February 24, 2013.
- [32] European Medicines Agency Committee for Medicinal Products for Human Use (CHMP). Guideline on medicinal products for the treatment of Alzheimer's disease and other dementias. 2008.
- [33] Dubois B, Feldman HH, Jacova C, Dekosky ST, Barberger-Gateau P, Cummings J, et al. Research criteria for the diagnosis of Alzheimer's disease: Revising the NINCDS-ADRDA criteria. *Lancet Neurol* 2007;6:734–46.
- [34] Jack CR Jr, Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, et al. Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol* 2013;12:207–16.
- [35] Hampel H, Frank R, Broich K, Teipel SJ, Katz RG, Hardy J, et al. Biomarkers for Alzheimer's disease: Academic, industry and regulatory perspectives. *Nat Rev Drug Discov* 2010;9:560–74.
- [36] Morris JC, Selkoe DJ, Holtzman DM, Mayeux R, Schupf N, Klunk WE, et al. Emerging consensus for Alzheimer's biomarkers in clinical trials. *Neurobiol Aging* 2011;32:S1–66.

- [37] Hampel H, Wilcock G, Andrieu S, Aisen P, Blennow K, Broich K, et al. Biomarkers for Alzheimer's disease therapeutic trials. *Prog Neurobiol* 2011;95:579–93.
- [38] Morris JC, Selkoe DJ. Recommendations for the incorporation of biomarkers into Alzheimer clinical trials: An overview. *Neurobiol Aging* 2011;32(Suppl 1):S1–3.
- [39] Lo RY, Hubbard AE, Shaw LM, Trojanowski JQ, Petersen RC, Aisen PS, et al. Longitudinal change of biomarkers in cognitive decline. *Arch Neurol* 2011;68:1257–66.
- [40] Jack CR Jr. Alliance for aging research AD biomarkers work group: Structural MRI. *Neurobiol Aging* 2011;32(Suppl 1):S48–57.
- [41] Blennow K, Hampel H, Weiner M, Zetterberg H. Cerebrospinal fluid and plasma biomarkers in Alzheimer disease. *Nat Rev Neurol* 2010;6:131–44.
- [42] Seppala TT, Nerg O, Koivisto AM, Rummukainen J, Puli L, Zetterberg H, et al. CSF biomarkers for Alzheimer disease correlate with cortical brain biopsy findings. *Neurology* 2012;78:1568–75.
- [43] Clark CM, Schneider JA, Bedell BJ, Beach TG, Bilker WB, Mintun MA, et al. Use of florbetapir-PET for imaging beta-amyloid pathology. *JAMA* 2011;305:275–83.
- [44] Mosconi L, Pupi A, De Leon MJ. Brain glucose hypometabolism and oxidative stress in preclinical Alzheimer's disease. *Ann N Y Acad Sci* 2008;1147:180–95.
- [45] Wagner M, Wolf S, Reischies FM, Daerr M, Wolfsgruber S, Jessen F, et al. Biomarker validation of a cued recall memory deficit in prodromal Alzheimer disease. *Neurology* 2012;78:379–86.
- [46] Dodart JC, Bales KR, Gannon KS, Greene SJ, DeMattos RB, Mathis C, et al. Immunization reverses memory deficits without reducing brain A β burden in Alzheimer's disease model. *Nat Neurosci* 2002;5:452–7.
- [47] DeMattos RB, Bales KR, Cummins DJ, Dodart JC, Paul SM, Holtzman DM. Peripheral anti-A β antibody alters CNS and plasma A β clearance and decreases brain A β burden in a mouse model of Alzheimer's disease. *Proc Natl Acad Sci U S A* 2001;98:8850–5.
- [48] Mattsson N, Andreasson U, Persson S, Carrillo MC, Collins S, Chalbot S, et al. CSF biomarker variability in the Alzheimer's Association quality control program. *Alzheimers Dement* 2013;9:251–61.
- [49] Paterson RW, Toombs J, Chapman MD, Nicholas JM, Heslegrave AJ, Slattery CF, et al. Do cerebrospinal fluid transfer methods affect measured amyloid beta-42, total tau, and phosphorylated tau in clinical practice. *Alzheimers Dement*. 2015;in press.
- [50] Carrillo MC, Blennow K, Soares H, Lewczuk P, Mattsson N, Oberoi P, et al. Global standardization measurement of cerebral spinal fluid for Alzheimer's disease: An update from the Alzheimer's Association Global Biomarkers Consortium. *Alzheimers Dement* 2013;9:137–40.
- [51] Schmidt ME, Chiao P, Klein G, Matthews D, Thurfjell L, Cole PE, et al. The influence of biological and technical factors on quantitative analysis of amyloid PET: Points to consider and recommendations for controlling variability in longitudinal data. *Alzheimers Dement* 2015;11:1050–68.
- [52] Klunk WE, Koeppe RA, Price JC, Benzinger TL, Devous MD Sr, Jagust WJ, et al. The Centiloid Project: Standardizing quantitative amyloid plaque estimation by PET. *Alzheimers Dement* 2015;11:1–1514.
- [53] Frisoni GB, Jack CR. Harmonization of magnetic resonance-based manual hippocampal segmentation: A mandatory step for wide clinical use. *Alzheimers Dement* 2011;7:171–4.
- [54] Hardy J, Allsop D. Amyloid deposition as the central event in the aetiology of Alzheimer's disease. *Trends Pharmacol Sci* 1991;12:383–8.
- [55] Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science* 2002;297:353–6.
- [56] Buchhave P, Minthon L, Zetterberg H, Wallin AK, Blennow K, Hansson O. Cerebrospinal fluid levels of beta-amyloid 1-42, but not of tau, are fully changed already 5 to 10 years before the onset of Alzheimer dementia. *Arch Gen Psychiatry* 2012;69:98–106.
- [57] Perrin RJ, Fagan AM, Holtzman DM. Multimodal techniques for diagnosis and prognosis of Alzheimer's disease. *Nature* 2009;461:916–22.
- [58] Roe CM, Fagan AM, Grant EA, Holtzman DM, Morris JC. CSF biomarkers of Alzheimer disease: "nongenerative" outcomes. *Neurology* 2013;81:2028–31.
- [59] Price JL, Morris JC. Tangles and plaques in nondemented aging and "preclinical" Alzheimer's disease. *Ann Neurol* 1999;45:358–68.
- [60] Braak H, Braak E. Frequency of stages of Alzheimer-related lesions in different age categories. *Neurobiol Aging* 1997;18:351–7.
- [61] Knopman DS. β -Amyloidosis and neurodegeneration in Alzheimer disease: Who's on first? *Neurology* 2014;82:1756–7.
- [62] Fagan AM, Perrin RJ. Upcoming candidate cerebrospinal fluid biomarkers of Alzheimer's disease. *Biomark Med* 2012;6:455–76.
- [63] Chien DT, Szardenings AK, Bahri S, Walsh JC, Mu F, Xia C, et al. Early clinical PET imaging results with the novel PHF-tau radioligand [F18]-T808. *J Alzheimers Dis* 2014;38:171–84.
- [64] Maruyama M, Shimada H, Suhara R, Shinotoh H, Ji B, Maeda J, et al. Imaging of tau pathology in a tauopathy mouse model and in Alzheimer patients compared to normal controls. *Neuron* 2013;79:1094–108.
- [65] Okamura N, Furumoto S, Fodero-Tavoletti MT, Mulligan RS, Harada R, Yates P, et al. Non-invasive assessment of Alzheimer's disease neurofibrillary pathology using 18F-THK5105 PET. *Brain* 2014;137(Pt 6):1762–71.
- [66] Villemagne VL, Furumoto S, Fodero-Tavoletti MT, Mulligan RS, Hodges J, Harada R, et al. In vivo evaluation of a novel tau imaging tracer for Alzheimer's disease. *Eur J Nucl Med Mol Imaging* 2014;41:816–26.
- [67] Xia CF, Arteaga J, Chen G, Gangadharmath U, Gomez LF, Kasi D, et al. [(18)F]T807, a novel tau positron emission tomography imaging agent for Alzheimer's disease. *Alzheimers Dement* 2013;9:666–76.
- [68] Kerchner GA. Ultra-high field 7T MRI: A new tool for studying Alzheimer's disease. *J Alzheimers Dis* 2011;26(Suppl 3):91–5.
- [69] Nir TM, Jahanshad N, Villalon-Reina JE, Toga AW, Jack CR, Weiner MW, et al. Effectiveness of regional DTI measures in distinguishing Alzheimer's disease, MCI, and normal aging. *Neuroimage Clin* 2013;3:180–95.
- [70] Ayutyanont N, Langbaum JB, Hendrix SB, Chen K, Fleisher AS, Friesenhahn M, et al. The Alzheimer's Prevention Initiative composite cognitive test score: Sample size estimates for the evaluation of preclinical Alzheimer's disease treatments in presenilin 1 E280A mutation carriers. *J Clin Psychiatry* 2014;75:652–60.
- [71] Donohue MC, Sperling RA, Salmon DP, Rentz DM, Raman R, Thomas RG, et al. The preclinical Alzheimer cognitive composite: Measuring amyloid-related decline. *JAMA Neurol* 2014;71:961–70.
- [72] Insel PS, Mattsson N, Mackin RS, Kornak J, Nosheny R, Tosun-Turgut D, et al. Biomarkers and cognitive endpoints to optimize trials in Alzheimer's disease. *Ann Clin Transl Neurol* 2015;2:534–47.
- [73] Langbaum JB, Hendrix SB, Ayutyanont N, Chen K, Fleisher AS, Shah RC, et al. An empirically derived composite cognitive endpoint with improved power to track and evaluate treatments for preclinical Alzheimer's disease. *Alzheimers Dement* 2014;10:666–74.
- [74] Tarawneh R, D'Angelo G, Macy E, Xiong C, Carter D, Cairns NJ, et al. Visinin-like protein-1: Diagnostic and prognostic biomarker in Alzheimer disease. *Ann Neurol* 2011;70:274–85.
- [75] Tarawneh R, Lee JM, Ladenson JH, Morris JC, Holtzman DM. CSF VILIP-1 predicts rates of cognitive decline in early Alzheimer disease. *Neurology* 2012;78:709–19.
- [76] Craig-Schapiro R, Perrin RJ, Roe CM, Xiong C, Carter D, Cairns NJ, et al. YKL-40: A novel prognostic fluid biomarker for preclinical Alzheimer's disease. *Biol Psychiatry* 2010;68:903–12.
- [77] Chiasserini D, Parnetti L, Andreasson U, Zetterberg H, Giannandrea D, Calabresi P, et al. CSF levels of heart fatty acid binding protein are altered during early phases of Alzheimer's disease. *J Alzheimers Dis* 2010;22:1281–8.
- [78] Henriksen K, O'Bryant SE, Hampel H, Trojanowski JQ, Montine TJ, Jeromin A, et al. The future of blood-based biomarkers for Alzheimer's disease. *Alzheimers Dement* 2014;10:115–31.

- [79] Kiddle SJ, Sattlecker M, Proitsi P, Simmons A, Westman E, Bazenec C, et al. Candidate blood proteome markers of Alzheimer's disease onset and progression: A systematic review and replication study. *J Alzheimers Dis* 2014;38:515–31.
- [80] Snyder HM, Carrillo MC, Grodstein F, Henriksen K, Jeromin A, Lovestone S, et al. Developing novel blood-based biomarkers for Alzheimer's disease. *Alzheimers Dement* 2014;10:109–14.
- [81] Cheng L, Quek CY, Sun X, Bellingham SA, Hill AF. The detection of microRNA associated with Alzheimer's disease in biological fluids using next-generation sequencing technologies. *Front Genet* 2013;4:150.
- [82] Sheinerman KS, Umansky SR. Circulating cell-free microRNA as biomarkers for screening, diagnosis and monitoring of neurodegenerative diseases and other neurologic pathologies. *Front Cell Neurosci* 2013;7:150.
- [83] Tsilioni I, Panagiotidou S, Theoharides TC. Exosomes in neurologic and psychiatric disorders. *Clin Ther* 2014;36:882–8.
- [84] O'Bryant SE, Gupta V, Henriksen K, Edwards M, Jeromin A, Lista S, et al. Guidelines for the standardization of preanalytic variables for blood-based biomarker studies in Alzheimer's disease research. *Alzheimers Dement* 2015;11:549–60.