Altered maternal lipid metabolism is associated with higher inflammation in obese women during late pregnancy

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Abstract

Inflammation is elevated in obese pregnant women and is associated with adverse maternal and neonatal outcomes. Maternal lipid metabolism and its relationships with maternal inflammation, insulin resistance and neonatal metabolic health are poorly understood in obese pregnant women.

18 lean (age: 26.1 ± 5.0 years, pre-pregnancy BMI: 21.5 ± 1.9 kg/m^2) and 16 obese (age: 25.0 ± 4.8 years, pre-pregnancy BMI: 36.3 ± 4.3 kg/m^2) women participated in this case-control study during the third trimester of pregnancy. Maternal plasma markers of insulin resistance (HOMA-IR) and inflammation (C-reactive protein (CRP)) were measured at rest, and lipid concentration and kinetics (lipid oxidation rate and lipolysis) were measured at rest, during a 30-minute bout of low-intensity (40% VO_{2peak}) exercise, and during a recovery period. Umbilical cord blood was collected for measurement of neonatal plasma insulin sensitivity, inflammation, and lipid concentration. Neonatal body composition was measured via air displacement plethysmography. Pregnant obese women had higher plasma CRP (9.1 ± 4.0 mg/L versus 2.3 ± 1.8 mg/L, $p<0.001$) and higher HOMA-IR (3.8 ± 1.9 versus 2.3 ± 1.5, $p=0.009$) compared to pregnant lean women. Obese women had higher lipid oxidation rates during recovery from low-intensity exercise (0.13 ± 0.03 g/min versus 0.11 ± 0.04 g/min, $p=0.02$) that was associated with higher maternal CRP ($r=0.55$, $p=0.001$). Maternal CRP was positively associated with maternal HOMA-IR ($r=0.40$, $p<0.02$) and systolic blood pressure ($r=0.40$, $p<0.02$). Maternal lipid metabolism-associated inflammation may contribute to insulin resistance and higher blood pressure in obese women during pregnancy.

Keywords

free fatty acids; inflammation; insulin resistance; lipid oxidation; obesity

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Introduction

Maternal obesity is a significant public health concern in the United States as one in three women enter pregnancy obese [1]. Maternal obesity is associated with increased risk of maternal complications including excessive gestational weight gain [2,3], gestational diabetes [4], preeclampsia and other hypertensive disorders [5–8], and long-term maternal cardiovascular morbidity [9]. Obesity during pregnancy also contributes to unfavorable offspring metabolic outcomes including excess adiposity [10,11] and insulin resistance [12].

Obese women experience metabolic dysfunction during pregnancy including higher plasma inflammation and lower insulin sensitivity [4,13,14]. Inflammation and insulin resistance are interrelated and associated with adverse pregnancy outcomes [13,15–19]. Mechanisms contributing to elevated maternal inflammation during pregnancy in obese women are unclear. Evidence suggests that non-gravid, obese adults have elevated lipid oxidation rates at rest [20] and during low-to-moderate intensity exercise [21,22], and that excessive lipid oxidation is associated with higher inflammation [23]. Elevated maternal lipid oxidation during pregnancy in obese women might contribute to increased maternal inflammation and subsequently altered maternal metabolic health. In addition, alterations in maternal lipid metabolism might impact offspring health as maternal obesity contributes to a lipotoxic placental environment that may be associated with increased *in utero* markers of inflammation and altered fetal development [24,25]. However, maternal lipid oxidation and its relationships with maternal and neonatal inflammation and metabolic health have not been previously examined.

The primary purposes of this study were to compare lipid oxidation rate and lipolysis during late pregnancy between obese and lean women, and to examine the relationships between maternal lipid metabolism, inflammation and insulin resistance. This study may identify lipid metabolism as a mechanism contributing to maternal inflammation in obese pregnant women.

Materials and methods

Subjects

Thirty-four women participated in the study (lean: n=18, obese: n=16). Women receiving prenatal care at the Women's Health Center and Women's Health Clinic at Barnes Jewish Hospital/Washington University between August 2013 and November 2014 were screened for inclusion. Approximately 350 women were screened, and 50 women who met all criteria with ongoing pregnancies were approached for participation late in their second trimester. Inclusion criteria included women ages 18–44 years, confirmed singleton viable pregnancy with no identified fetal abnormalities (as determined by routine anatomy ultrasound at 18–22 weeks), and pre-pregnancy BMI between 18.0 and 24.9 kg/m^2 for the lean group or pre-pregnancy BMI between 30 and 45 kg/m^2 for the obese group. Exclusion criteria included: 1) multiple gestation pregnancy, 2) inability to provide voluntary informed consent, 3) self-reported use of illegal drugs (cocaine, methamphetamine, opiates), 4) current smoker who did not consent to cessation, 5) current usage of daily medications by class: corticosteroids, beta-blockers (known to affect lipid metabolism) and anti-psychotics (known to alter insulin

resistance and metabolic profiles), 6) diagnosis of gestational diabetes in current pregnancy, history of gestational diabetes, pre-pregnancy diabetes or prior macrosomic (>4500 g) infant (each elevate the risk for gestational diabetes in the current pregnancy, or undiagnosed gestational diabetes), 7) history of heart disease, or 8) any other condition that would preclude exercise.

Study procedures

All study procedures were performed at the Washington University School of Medicine Institute for Clinical and Translational Sciences Clinical Research Unit (CRU). All pregnant women participated in two maternal visits between 32 and 37 weeks gestation. Approval for this study was granted by the Institutional Review Board at Washington University (IRB ID: 201306109, NCT: NCT02039414).

Maternal visit 1

Body composition was measured using skinfold anthropometry in order to determine maternal percent body fat. Body fat percentage was determined by pressing folds of the skin at seven sites with a caliper (Harpenden Skinfolds Caliper, Baty International, United Kingdom), recording skin thickness, and entering the data into a standardized equation that accounts for age as previously described [26]. Participants also completed the YMCA submaximal cycle test as previously performed in order to predict cardiorespiratory fitness levels [27]. National Institutes of Health's Dietary History Questionnaire II was completed by each participant to determine potential differences in maternal diet in the month leading up to study visits [28]. Previous literature demonstrates that dietary history questionnaires are valid and reproducible among pregnant populations [29].

Maternal visit 2

Approximately one week after Visit 1, subjects were admitted to the CRU the morning after an overnight fast. The night prior, subjects were instructed to consume a standardized meal consisting of 50% carbohydrates, 30% fats, and 20% protein. Upon admission to the CRU, height, weight, and vital signs were obtained. A catheter (IV) was placed in a hand vein and heated to 55°C by using a thermostatically controlled box in order to obtain arterialized blood samples as previously described [30]. Participants kept their hand in the box throughout the entire study visit. Participants rested for approximately 30 minutes prior to measuring lipid oxidation rate using indirect calorimetry (True One 2400, Parvomedics, Sandy, UT). Participants laid supine while a canopy was placed over their head for 15 minutes to measure oxygen consumption and carbon dioxide production in order to determine lipid oxidation rate [31]. After the initial indirect calorimetry measurement, a baseline blood collection was obtained. Participants then exercised continuously at approximately 40% of their predicted VO_{2peak} (based on the YMCA submaximal cycle test) for 30 minutes. Lipid metabolism was examined via indirect calorimetry and plasma analysis during and after exercise because low-to-moderate intensity exercise increases both adipose tissue breakdown (*i.e.* lipolysis) and lipid oxidation, and low-to-moderate intensity exercise might mimic their daily activity levels (~3–5 METS, e.g. household chores, caring for other children, walking). Exercise indirect calorimetry was performed and blood was drawn at the 10, 20, and 30 minute time points of submaximal exercise. After exercise

termination, participants returned to supine and blood was drawn 10, 30, and 60 minutes-post cessation of exercise (*i.e.* recovery). Indirect calorimetry was performed 30 minutes post-exercise for 15 minutes as described for the baseline measurement.

Sample analyses and calculations

All samples were immediately placed on ice and plasma was separated by centrifugation within 30 minutes of collection. Plasma samples were stored at −80°C until final analyses were performed. Blood samples for glucose were collected in heparinized tubes and analyzed immediately with an automated glucose analyzer (Yellow Springs Instruments Co, Yellow Springs, OH). Plasma insulin concentration was measured by electrochemiluminescence technology (Elecsys 2010, Roche Diagnostics, Indianapolis, IN). Insulin and glucose levels were used to calculate the homeostatic model assessment-insulin resistance (HOMA-IR) [32]. The HOMA-IR is an index of insulin resistance that reflects fasting glucose concentration measured at the fasting insulin concentration. Inflammation was examined through high-sensitivity C-reactive protein (CRP) and measured by immunoturbidimetric assay (Roach Diagnostics, Indianapolis, IN). Blood samples used to determine plasma free fatty acids were collected in in tubes containing EDTA. Plasma free fatty acid concentrations were determined by enzymatic colorimetric assay (Wako Pure Chemical Industries, Osaka, Japan). Lipolysis was calculated by the area under the curve (AUC) [33,34] for free fatty acids from baseline to the end of the study period. A summary score to represent total lipid oxidation rate throughout the study period was determined by calculating the AUC using lipid oxidation rates from baseline, exercise, and recovery time points. Clinical lipid profiles including triglycerides, total cholesterol, low-density lipoprotein, and high-density lipoprotein were also obtained at the CRU.

Neonatal measurements

At parturition, neonatal birth weight was obtained. In addition, 44 mL of umbilical vein blood was collected, centrifuged within 30 minutes of parturition, and placed in a −80°C freezer for further analysis. Umbilical cord blood was used to determine neonatal HOMA-IR (insulin and glucose levels), free fatty acid concentration, and CRP.

Within 48 hours of delivery, neonatal anthropometrics were measured in the CRU. Neonatal length (Pediatric Length Board, Ellard Instrumentation LTD, Monroe, WA) and head circumference (Gulick II Tape Measure, model 67020, Country Technology Inc., Gays Mills, WI) were measured. Body composition (fat and lean mass) was measured by skin fold thickness at four different sites (triceps, subscapular, ilium, and thigh, measured by one recorder) and by air displacement plethysmography (Pea Pod, Life Measurement, Inc., Concord, CA). All neonates were full-term (37 weeks gestation) at the time of delivery except for one in the obese group. This neonate's data were excluded from the analysis.

Statistical analysis

Normality of the distribution for each variable was tested using Kolmogorov-Smirnov tests student's Independent T-Tests for normally distributed variables and Mann-Whitney U tests for non-normally distributed variables were used to compare metabolic outcomes between lean and obese groups. Pearson product-moment correlation coefficients for normally

distributed variables or Spearman's rank-order correlation coefficient for non-normally distributed variables were used to assess the degree of the relationship between variables. Two-way repeated-measures ANOVAs (group × time) were used with Tukey post hoc analyses when comparing baseline, exercise, and recovery conditions. Study data were collected and managed using REDCap electronic data capture tools hosted at Washington University School of Medicine [35]. All data analyses were conducted using IBM SPSS Statistics, Version 22 (Armonk, New York).

Based on previous data comparing lipid metabolism between lean and obese females at rest and during exercise [22], we estimated an effect size of 1.1. Therefore, with an alpha level of 0.05, 16 women per group were required to power the study at β=0.80.

Results

Maternal demographic characteristics

Obese women had significantly higher pre-pregnancy body mass indexes and body fat percentages than the lean group (Table 1). Age, parity, income level, race, and gestation age during study visits were similar between groups (Table 1).

Maternal baseline metabolic characteristics

Maternal baseline metabolic characteristics are presented in Table 1. Obese pregnant women had significantly higher plasma insulin concentration than lean pregnant women. HOMA-IR was higher in obese women compared to lean pregnant women. Obese pregnant women had higher plasma CRP concentration compared to lean pregnant women. Maternal systolic blood pressure was higher in obese pregnant women compared to lean pregnant women across all time points (F=3.90, p=0.03). CRP, HOMA-IR, and systolic blood pressure data are shown in Figure 1. Obese pregnant women had significantly higher resting energy expenditure compared to lean pregnant women (Table 1).

Dietary composition was similar between groups (Table 2). Three women (two lean, one obese) did not complete the dietary survey.

Maternal lipid metabolism

Pregnant obese women had a higher post-exercise lipid oxidation rate compared to their lean counterparts (Recovery: 0.11 ± 0.04 g/min *vs.* 0.13 ± 0.03 g/min, p=0.02) and tended to have higher lipid oxidation rates during baseline and exercise conditions (baseline- lean: 0.10 ± 0.04 g/min *vs.* obese: 0.12 ± 0.03 g/min, p=0.13; exercise- lean: 0.16 ± 0.09 g/min *vs.* obese: 0.22 ± 0.09 g/min, p=0.09) (Figure 2A). Total lipid oxidation was significantly higher in the obese group (lean: 11.8 ± 5.4 g *vs.* obese: 15.3 ± 4.4 g, p=0.05) (Figure 2B). Lipolysis was similar between groups throughout the baseline, exercise, and resting conditions.

Correlations between maternal metabolic characteristics

CRP was positively associated with lipid oxidation rate at baseline (r=0.42, p=0.02) (Figure 3A) and during the 1-hour recovery period (r=.55, p=0.001) (Figure 3B). Maternal CRP was also related to total lipid oxidation (r=0.42, p=0.01) (Figure 3C). Similarly, baseline systolic

blood pressure was positively correlated with lipid oxidation rate at baseline (r=0.34, p=0.05) and during the 1-hour recovery period (r=0.39, p=0.02). However, when accounting for CRP, the relationships between lipid oxidation rate and systolic blood pressure did not exist. Maternal CRP was positively correlated with maternal circulating insulin levels (r= 0.44, p=0.01), HOMA-IR (r=0.40, p=0.02) (Figure 4A), and systolic blood pressure (r=0.40, p=0.02) (Figure 4B).

Neonatal anthropometric and metabolic outcomes

Air displacement plethysmography was not performed on three neonates due to time of delivery relative to discharge, but all other neonatal anthropometrics were obtained. Cord blood could not be obtained from one lean and two obese participants. Neonatal anthropometrics and metabolic outcomes were similar between groups (Table 3).

Discussion

Our primary novel findings from the study were: 1) maternal lipid oxidation was higher in obese pregnant women compared to lean pregnant women, particularly following an acute bout of low-intensity exercise, 2) maternal lipid oxidation rate was significantly associated with maternal inflammation, and 3) maternal inflammation was related to maternal insulin resistance and systolic blood pressure. These results suggest that elevated maternal lipid oxidation might contribute to increased inflammation and subsequent reductions in insulin sensitivity during late pregnancy in obese women. Importantly, we believe the finding of elevated lipid oxidation rates in obese pregnant women following a mild bout of physical activity is clinically meaningful as this metabolic environment would be representative of women who recently participated in typical daily physical activities (e.g. running errands, cleaning the house, or taking care of another child). Lipid oxidation rate in non-gravid adults is known to increase over resting values during low-to-moderate physical activity and remained elevated for 2–3 hours post-exercise [36,37].

In the current study, higher rates of maternal lipid oxidation during rest and post-exercise recovery were associated with greater maternal inflammation. In non-gravid adults, lipid oxidation is intricately related to inflammation and insulin resistance [23,38]. Specifically, lipid oxidation by-products including reactive oxygen species have been shown to signal pathways that initiate an inflammatory response [23,39,40]. The current study also found that higher maternal inflammation was associated with higher insulin resistance, suggesting that maternal inflammation, possibly the result of increased lipid oxidation by-products, might contribute to higher maternal insulin resistance in obese pregnant women. Our results are consistent with Retnakaran *et al.* who demonstrated obesity during pregnancy elicits an inflammatory response with possible downstream metabolic sequelae including insulin resistance [41]. Similarly, Korkmazer *et al.* found higher inflammation among insulin resistant pregnant women with and without a classification of gestational diabetes [42]. The clinical implications of this could be substantial as the combination of obesity and insulin resistance during pregnancy can lead to gestational diabetes [13]. Consequently, offspring of obese, insulin resistant women are at increased risk of preterm delivery and associated neonatal morbidity, as well as childhood metabolic dysfunction which may potentiate the

vicious cycle of obesity and insulin resistance [13]. Taken together, these data suggest a relationship between maternal lipid metabolism, inflammation, and insulin resistance in obese women during pregnancy.

In the current study, lipid profiles were similar between lean and obese pregnant women (triglycerides, total cholesterol, low-density lipoprotein, high-density lipoprotein). While maternal obesity is associated with increased higher plasma concentrations of triglycerides and low-density lipoproteins early in pregnancy [43], lean and obese women reach similar concentrations during late pregnancy [43]. Our data, measured during late pregnancy, are reflective of the literature demonstrating similar lipid profiles between lean and obese pregnant women. Similarly, maternal and neonatal free fatty acid concentrations in the current study were consistent with Catalano *et al.* who reported free fatty acid concentrations in obese women and their fetuses were not different than lean pregnant women [43,44].

Although not a primary focus of the study, we also found a relationship between maternal inflammation and resting systolic blood pressure. Maternal systemic inflammation is a known feature of preeclampsia, a pregnancy-specific condition characterized by high blood pressure [45]. In fact, Redman *et al.* suggest that preeclampsia is ultimately an excessive maternal inflammatory response to pregnancy [46]. High blood pressure disorders during pregnancy, including preeclampsia, are an important cause of morbidity, long-term disability, and even death among pregnant women and their offspring [47]. Inflammation has been identified as a key contributor to pregnancy-specific high blood pressure [46,48] and our findings are consistent with this.

Oxidative stress could provide a link connecting increased maternal lipid oxidation with inflammation and subsequent insulin resistance and hypertension in obese pregnant women. In non-gravid adults, oxidative stress is elevated in obesity [49] and is associated with inflammation, vascular cell wall damage, high blood pressure, cardiovascular disease, poor metabolic function, and insulin resistance [50–58]. It is possible that elevated lipid oxidation rates observed in obese pregnant women generate partially-oxidized byproducts (*i.e.* reactive oxygen species); however, this is speculative and requires further study. Figure 5 depicts proposed pathway for the role of maternal on lipid metabolism and long-term maternal and neonatal outcomes.

Neonates of pregnant obese woman had similar body composition, insulin sensitivity, and inflammatory markers as lean pregnant women. These results suggest that despite the metabolic abnormalities associated with obesity during pregnancy, the fetus may be protected from some of these detrimental effects, at least at 24–48 hours post-parturition. However, the "fetal origins hypothesis" suggests the origins of adult health diseases such as obesity, cardiovascular disease, and diabetes may be caused by in utero exposures [59,60]. Therefore, it is plausible that metabolic abnormalities may be epigenetically programmed, but are not apparent or measurable until later in life. For example, Liebowitz *et al.* found that offspring of obese women had higher plasma inflammation at 12 years of age [61]. Offspring of obese women with elevated plasma inflammation during pregnancy may develop inflammation as they age as a result of fetal programming; however, these changes may not be detectable at delivery when umbilical cord blood is obtained. Similarly, Whitaker

concluded that obesity during pregnancy more than doubles the risk of obesity in the offspring between ages two and four [43,62]. Although neonatal body composition in our study was not different between lean and obese pregnant women, it is possible that differences might emerge during the preschool and childhood years. It is also plausible that our sample size was not large enough to detect a difference in birth weight or body composition as it is well-accepted that obese women have larger babies [10,11]. Based on our findings and the current literature, longitudinal studies of offspring born to obese women are needed.

A limitation of the present study was the observational study design, thus, cause-and-effect relationships could not be determined from these data. Our results should be cautiously interpreted as this pilot study was powered based on our primary outcome of lipid oxidation rate. We are not adequately powered to investigate many other maternal and neonatal outcomes. Bias may have been introduced because measurements and assessments were not blinded (it was obvious who was an obese "case" versus a lean "control"), but uniform data collection procedures were followed for all participants to minimize bias.

In conclusion we found that obese pregnant women have higher lipid oxidation rate, particularly after an acute bout of low-intensity exercise during late pregnancy. In addition, higher maternal lipid oxidation rates were associated with higher maternal inflammation. Maternal inflammation was related to insulin resistance and systolic blood pressure. We did not find any differences in neonatal metabolic outcomes between lean and obese women. Future studies investigating the role of maternal lipid oxidation-produced oxidative stress and interventions targeting lipid metabolism and inflammation to improve maternal and neonatal health in obese pregnant women are warranted.

Acknowledgments

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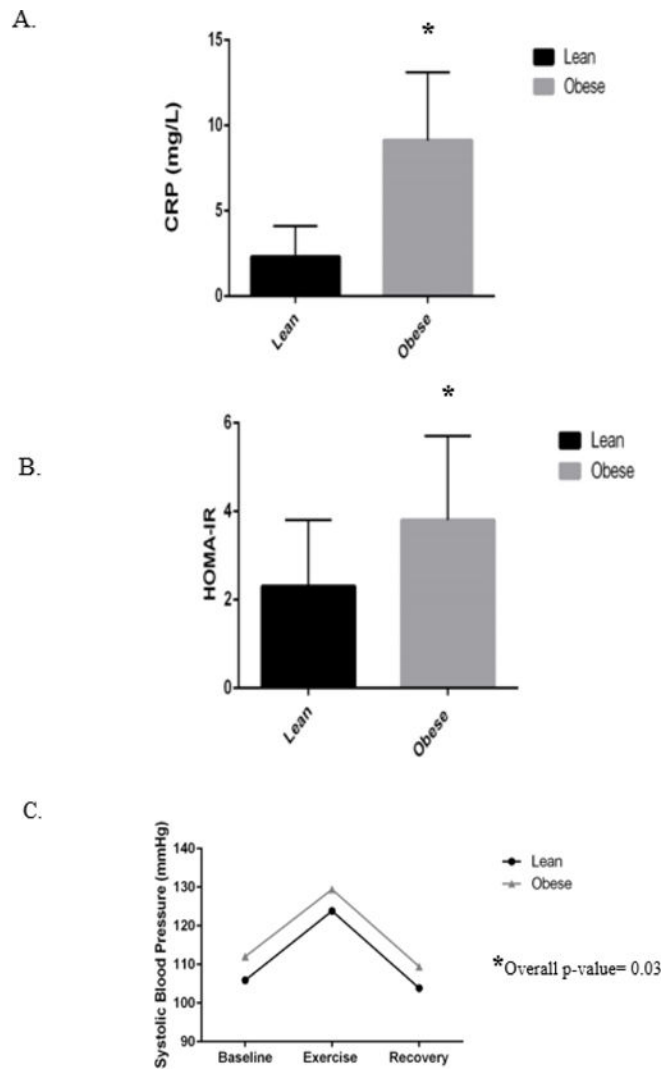
A.

B.

C.

Figure 1.
Maternal (A) inflammation (B) insulin resistance (C) systolic blood pressure between lean and obese pregnant women. Obese women have higher plasma CRP concentrations (A) higher insulin resistance (HOMA-IR) (B) and higher systolic blood pressure (C) than lean women during late pregnancy. All data are mean ± SD. *$p<0.05$.
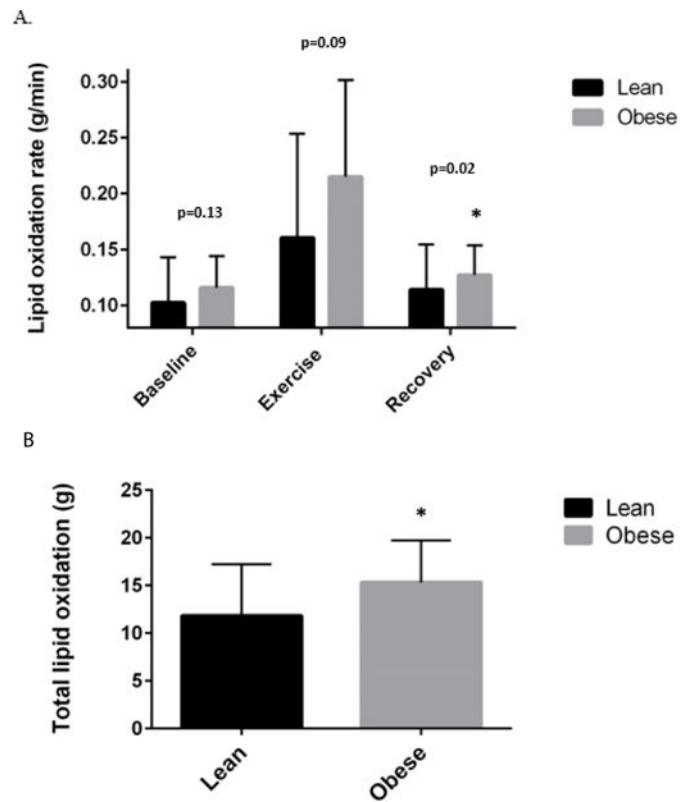
Figure 2.
(A) Lipid oxidation rates between lean and obese groups at baseline, during exercise, and during recovery from exercise (B). Total lipid oxidation measured by AUC across all timepoints between lean and obese pregnant women. Lipid oxidation rates are higher in obese pregnant women, particularly during post-exercise recovery (A), and a total lipid oxidation is higher in obese pregnant women compared to lean pregnant women (B). All data are mean ± SD. *$p<0.05$.
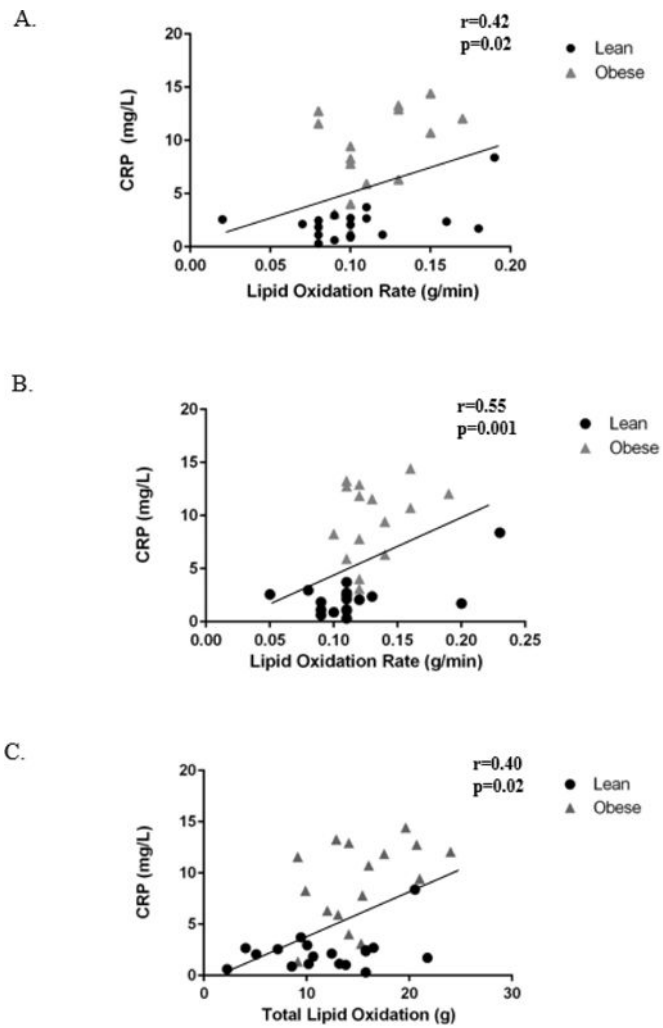
Figure 3.
Relationships between CRP and (A) baseline lipid oxidation rate, (B) post-exercise recovery lipid oxidation rate, and (C) total lipid oxidation. Maternal plasma CRP concentration is significantly correlated to lipid oxidation rate at baseline (A), during recovery (B), and to total lipid oxidation (C). *p<0.05.
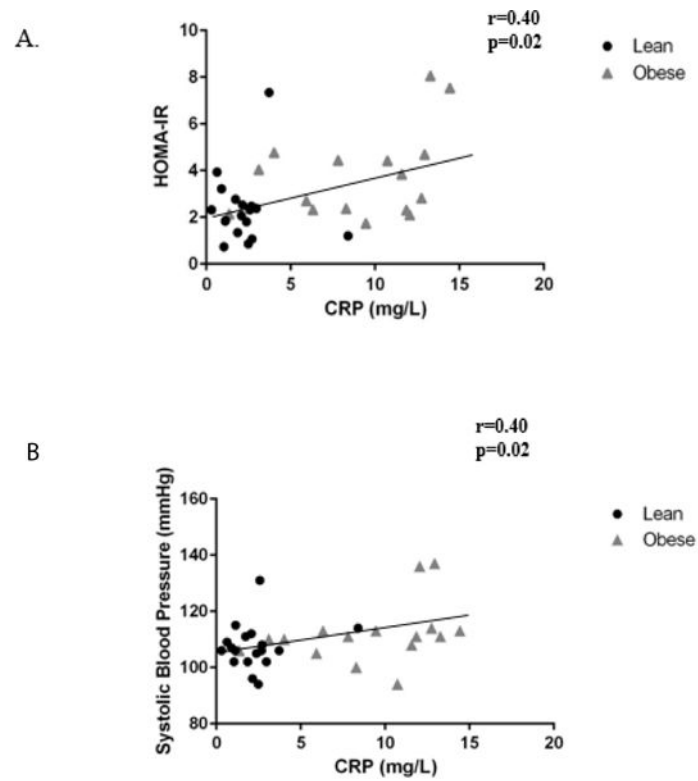
Figure 4.
Relationships between baseline maternal inflammation and (A) insulin resistance (B) systolic blood pressure. Maternal plasma CRP concentration is significantly correlated to maternal insulin resistance (A) and maternal systolic blood pressure (B). *p<0.05.
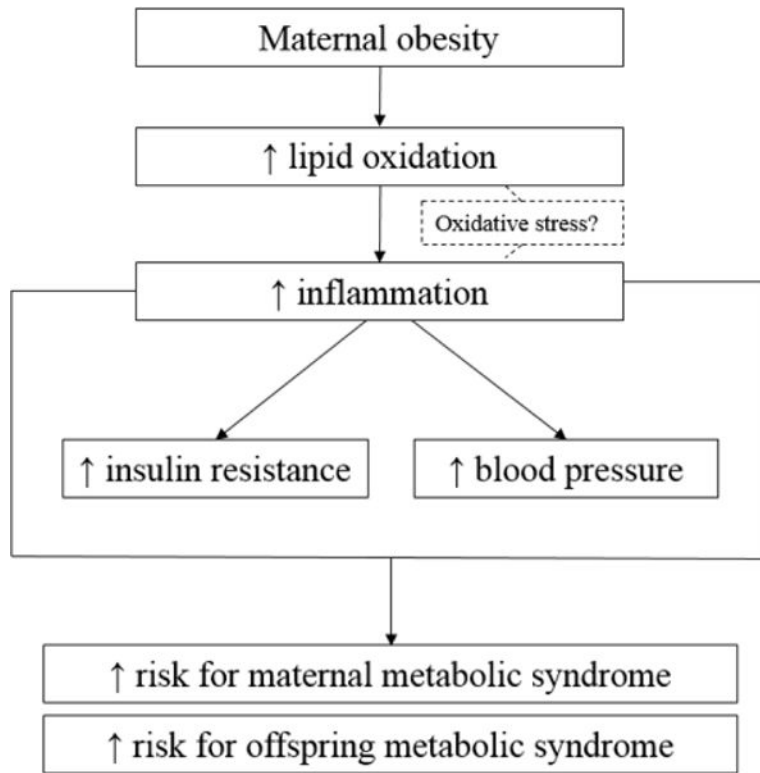
Figure 5.
Proposed pathway for maternal lipid metabolism and long-term maternal and neonatal outcomes.

Table 1

Maternal demographic and metabolic characteristics.

	Lean (n=18) (Mean ± SD)	Obese (n=16) (Mean ± SD)	p-value
Age (yrs)	26.1 ± 5.0	25.0 ± 4.8	0.52
Pre-pregnancy BMI (kg/m²) *	21.5 ± 1.9	36.3 ± 4.3	<0.001
Body fat percentage (%) *	20.7 ± 4.0	37.7 ± 3.5	<0.001
Gestation age at visit 2 (weeks)	35.2 ± 1.0	34.7 ± 1.4	0.25
Gestational weight gain (kg)	14.5 ± 4.5	10.3 ± 9.2	0.12
Heart rate (bpm)	84.1 ± 9.1	90.0 ± 10.8	0.08
Systolic blood pressure (mmHg)	107.3 ± 8.1	112.0 ± 10.9	0.16
Diastolic blood pressure (mmHg)	69.3 ± 7.6	71.5 ± 4.3	0.31
Glucose (mg/dL)	76.9 ± 5.5	80.3 ± 7.9	0.24
Insulin (uU/mL) *	12.3 ± 8.2	18.7 ± 8.4	0.01
HOMA-IR *	2.3 ± 1.5	3.8 ± 1.9	0.01
C-reactive protein (mg/L) *	2.3 ± 1.8	9.1 ± 4.0	<0.001
Total cholesterol (mg/dL)	228.4 ± 36.3	207.8 ± 35.9	0.11
HDL (mg/dL)	67.9 ± 18.2	65.1 ± 12.3	0.61
LDL (mg/dL)	130.2 ± 35.0	110.6 ± 34.0	0.24
Triglycerides (mg/dL)	151.8 ± 41.8	160.0 ± 60.9	0.65
Free fatty acids (meq/L)	0.47 ± 0.12	0.49 ± 0.15	0.64
Resting energy expenditure (kcal/day) *	1795 ± 225	2166 ± 291	<0.001
	# of women (%)	# of women (%)	p-value (χ^2-test)
Parity			
Nulliparous	11 (61%)	10 (63%)	0.93
Multiparous	7 (39%)	6 (37%)	
Income			
Low income	10 (56%)	8 (56%)	0.75
Mod/high income	8 (44%)	8 (43%)	
Race			
Caucasian	8 (44%)	5 (31%)	0.73
African-American	9 (50%)	10 (63%)	
Other	1 (6%)	1 (6%)	

*
p<0.05

Table 2

Average maternal daily dietary composition.

	Lean (n=16) mean ± SD	Obese (n=15) mean ± SD	p-value
Energy intake (kcal/day)	2554.7 ± 1574	2058.7 ± 845	0.29
Fat (g)	94.6 ± 50.5	69.1 ± 31.2	0.10
Carbohydrate (g)	353.5 ± 256.2	295.4 ± 137.3	0.44
Protein (g)	84.1 ± 39.0	73.3 ± 35.9	0.43

Table 3

Neonatal anthropometric and metabolic outcomes.

Neonatal demographic characteristics	Lean (n=18) mean ± SD	Obese (n=16) mean ± SD	p-value
Gestational age at delivery (weeks)	39.1 ± 0.9	39.5 ± 1.4	0.33
Birth weight (g)	3221.6 ± 377	3344 ± 428	0.57
Length (cm)	49.5 ± 2.2	49.9 ± 2.0	0.88
Head circumference (cm)	33.7 ± 1.7	34.4 ± 1.2	0.22
Fat mass (%)	11.1 ± 4.3	11.2 ± 3.6	0.98
Skinfolds (cm)			
Triceps	4.9 ± 0.8	5.1 ± 1.1	0.61
Subscapular	4.4 ± 0.8	4.4 ± 0.9	0.97
Ilium	4.7 ± 1.3	5.2 ± 1.4	0.56
Thigh	6.4 ± 1.3	6.9 ± 1.9	0.45
	# of women (%)	**# of women (%)**	
Mode of Delivery			
Vaginal	14 (78%)	9 (56%)	0.27
Cesarean	4 (22%)	7 (44%)	
Gender			
Male	7 (39%)	10 (63%)	0.30
Female	11 (61%)	6 (37%)	
Neonatal cord blood values	**Lean (n=17) mean ± SD**	**Obese (n=14) mean ± SD**	**p-value**
Glucose (mg/dL)	88.4 ± 14.0	80.7 ± 12.8	0.13
Insulin (uU/mL)	7.9 ± 6.2	7.5 ± 4.9	0.98
HOMA-IR	1.7 ± 1.5	1.6 ± 1.2	0.68
Free fatty acids (meq/L)	0.19 ± 0.08	0.16 ±0.06	0.28
C-reactive protein (mg/L)	0.20 ± 0.10	0.24 ± 0.21	0.55

*
p<0.05