

Synergistic effect of antioxidant system and osmolyte in hydrogen sulfide and salicylic acid crosstalk-induced heat tolerance in maize (*Zea mays* L.) seedlings

Zhong-Guang Li*

School of Life Sciences; Engineering Research Center of Sustainable Development and Utilization of Biomass Energy; Ministry of Education; Key Laboratory of Biomass Energy and Environmental Biotechnology; Yunnan Normal University; Kunming, PR China

Keywords: antioxidant system, heat tolerance, maize seedlings, osmolyte, salicylic acid, signal crosstalk, sodium hydrosulfide

Salicylic acid (SA), is a plant hormone with multifunction that is involved in plant growth, development and the acquisition of stress tolerance. Hydrogen sulfide (H₂S) is emerging similar functions, but crosstalk between SA and H₂S in the acquisition of heat tolerance is not clear. Our recent study firstly reported that SA treatment enhanced the activity of L-cysteine desulphydrase (L-DES), a key enzyme in H₂S biosynthesis, followed by induced endogenous H₂S accumulation, which in turn improved the heat tolerance of maize seedlings.¹ In addition, NaHS, a H₂S donor, enhanced SA-induced heat tolerance, while its biosynthesis inhibitor DL-propargylglycine (PAG) and scavenger hydroxylamine (HT) weakened SA-induced heat tolerance. Also, NaHS had no significant effect on SA accumulation and its biosynthesis enzymes phenylalanine ammonia lyase (PAL) and benzoic-acid-2-hydroxylase (BA2H) activities, as well as significant difference was not observed in NaHS-induced heat tolerance of maize seedlings by SA biosynthesis inhibitors paclobutrazol (PAC) and 2-aminoindan-2-phosphonic acid (AIP) treatment.¹ Further study displayed that SA induced osmolytes (proline, betaine and trehalose) accumulation and enhancement in activity of antioxidant system in maize seedlings. These results showed that antioxidant system and osmolyte play a synergistic role in SA and H₂S crosstalk-induced heat tolerance of maize seedlings.

Salicylic acid (SA), a plant hormone with multifunction that is involved in plant growth, development and the acquisition of stress tolerance, is mainly synthesized through the phenylalanine (Phe) route localized in the cytoplasm, phenylalanine ammonia lyase (PAL) and benzoic-acid-2-hydroxylase (BA2H) are key enzymes in SA biosynthesis in plants.²⁻⁵ SA has long been considered as an important endogenous immune signal in the induction of disease resistance response in plants.^{3,6}

In addition, SA has also functions in responses to abiotic stress factors such as heavy metal toxicity, salt, drought, chilling and heat.³⁻⁴ Spraying potato plants with an acetyl SA can improve the resistance of plants to heat stress in a concentration manner.² In addition, treatment with SA led to an increase in the level of endogenous H₂O₂ by reduction in catalase activity, similar to heat hardening at 45°C for 1 h.⁷ In maize, treatment of seeds with SA not only could markedly enhanced chilling tolerance, but also heat tolerance, and the acquisition of this tolerance is closely relative to enhancement in the activity of antioxidant system.⁸ Our previous work firstly found that exogenous SA enhanced the activity of L-cysteine desulphydrase (L-DES), a key enzyme in H₂S biosynthesis, followed by induced endogenous

H₂S accumulation, which in turn improved the heat tolerance of maize seedlings, NaHS (H₂S donor) enhanced SA-induced heat tolerance, indicated that H₂S might be a downstream signal molecule in SA-induced heat tolerance of maize seedlings,¹ but the mechanism of SA and H₂S interaction-induced heat tolerance is not completely clear.

Hydrogen sulfide (H₂S) has recently been identified as a third endogenous gaseous transmitter after nitric oxide (NO) and reactive oxygen species (ROS).⁹ In plants, L-cysteine desulphydrase (L-DES, E.C. 4.4.1.1.) is considered to be a key enzyme in H₂S biosynthesis.^{9,10} Recently, many positive effects of H₂S is being emerged in multiple physiological processes, including seed germination, organogenesis, stomata movement, osmotic stress, salt stress, chilling stress, oxidative stress and heavy metal stress.¹¹⁻¹⁷ Our previous research results also showed that NaHS treatment can improve heat tolerance of tobacco cells, maize and wheat seedlings.^{1,18-24}

Enhancement in antioxidant defense system and osmotic adjustment are common mechanisms of plants adapt to adverse environments including heat stress by scavenging reactive oxygen species (ROS); buffering redox; stabilizing protein, nucleic acid and biomembrane; regulating acidity in cytoplasm.^{25,26} Further

*Correspondence to: Zhong-Guang Li; Email: zhongguang_li@163.com

Submitted: 04/20/2015; Accepted: 05/11/2015

<http://dx.doi.org/10.1080/15592324.2015.1051278>

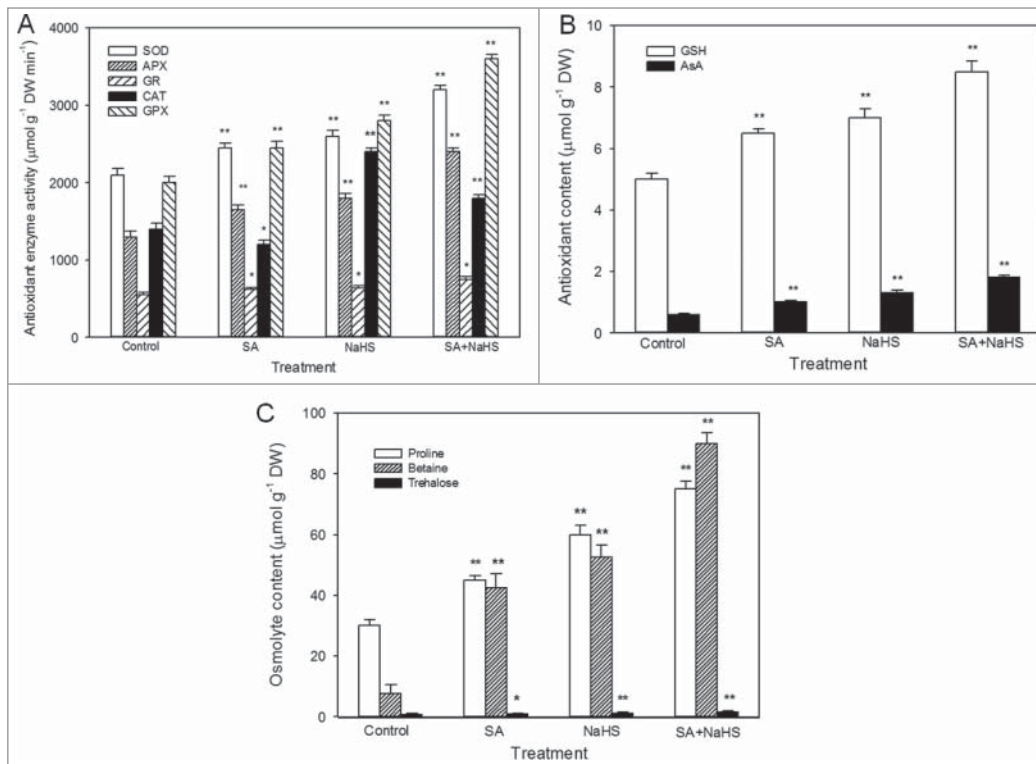


Figure 1. Effects of SA pretreatment on the accumulation of osmolyte (A), antioxidant enzyme activity (B) and antioxidant content (C) in maize seedlings under normal culture conditions. Error bars represent standard error and each data in figure represents the mean \pm SE of 3 experiments, and asterisk and double asterisks indicate significant difference ($P < 0.05$) and very significant difference ($P < 0.01$) from the control without SA or NaHS treatment, respectively.

study showed that SA and NaHS treatment alone or in combination significantly increased the activity of antioxidant enzymes superoxide dismutase (SOD), ascorbate peroxidase (APX), glutathione reductase (GR), catalase (CAT, SA treatment alone inhibited CT activity) and guaiacol peroxidase (GPX), and the content of antioxidants ascorbic acid (AsA) and glutathione (GSH), as well as the accumulation of endogenous osmolytes such as proline, betaine and trehalose in maize seedlings (Fig. 1). In addition, these effects induced by SA were enhanced by NaHS treatment (Fig. 1). These data indicated that antioxidant system and osmolyte exert a very important role in SA and H₂S interaction-induced heat tolerance of maize seedlings.

In conclusion, pretreatment of maize seedlings with SA significantly stimulated an increase in L-DES activity

in maize seedlings, followed by induced endogenous H₂S accumulation, which in turn increased the heat tolerance of maize seedlings (Fig. 2).¹ In addition, SA-induced heat tolerance was enhanced by NaHS, weakened by PAG and HT (Fig. 2),¹ suggesting that crosstalk between SA and H₂S exist in the heat tolerance of maize seedlings. Further research showed that SA and H₂S interaction induced an increase in the activity of antioxidant system and the accumulation of osmolyte in maize seedlings, implied that antioxidant system and osmolyte play a synergistic role in SA and H₂S crosstalk-induced heat tolerance of maize seedlings (Fig. 2). However, plant heat tolerance is involved in crosstalk among Ca²⁺, H₂O₂, NO, H₂S and plant hormones, which needs to be further investigated in the future.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

We appreciate the reviewers and editors for their exceptionally helpful comments on the article.

Funding

This research is supported by National Natural Science Foundation of China (31360057).

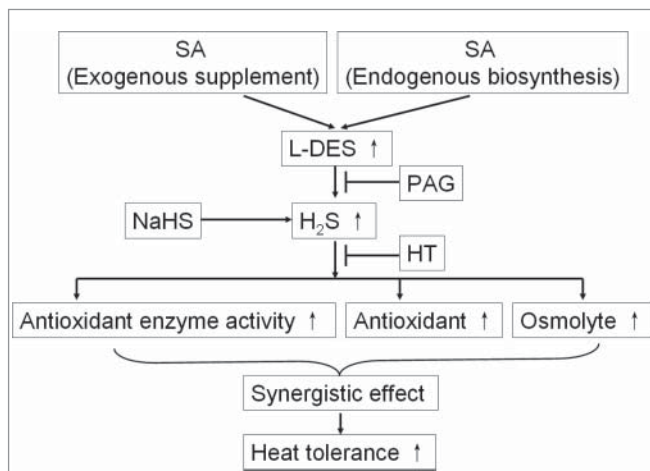


Figure 2. A possible model of crosstalk between SA and H₂S in the acquisition of heat tolerance in plants. Arrows (→) indicate positive effects, arrows (↑) represent increase in index, blunt line (⊥) indicates negative effect. H₂S, hydrogen sulfide; HT, hydroxylamine; L-DES, L-cysteine desulfhydrase; PAG, DL-propargylglycine; SA, salicylic acid (adapted from Li et al.¹).

References

- Li Z-G, Xie L-R, Li X-J. Hydrogen sulfide acts as a downstream signal molecule in salicylic acid-induced heat tolerance in maize (*Zea mays* L.) seedlings. *J Plant Physiol* 2015; 177:121-7; PMID:25727780; <http://dx.doi.org/10.1016/j.jplph.2014.12.018>
- Szalai G, Horgosi S, Soós V, Majláth I, Balázs E, Janda T. Salicylic acid treatment of pea seeds induces its de novo synthesis. *J Plant Physiol* 2011; 168:213-9; PMID:20933297; <http://dx.doi.org/10.1016/j.jplph.2010.07.029>
- Pál M, Szalai G, Kovács V, Gondor OK, Janda T. Salicylic acid-mediated abiotic stress tolerance. In: Salicylic acid. Hayat S, Ahmad A, Alyemeni MN, editors. LLC: Springer Science+Business Media; 2013; pp183-247.
- Janda M, Ruelland E. Magical mystery tour: Salicylic acid signaling. *Environ Exp Bot* 2014; 114:117-28; <http://dx.doi.org/10.1016/j.envexpbot.2014.07.003>
- Miura K, Tada Y. Regulation of water, salinity, and cold stress responses by salicylic acid. *Front Plant Sci* 2014; 5:4; PMID:24478784; <http://dx.doi.org/10.3389/fpls.2014.00004>
- Kumar D. Salicylic acid signaling in disease resistance. *Plant Sci* 2014; 228:127-34; PMID:25438793; <http://dx.doi.org/10.1016/j.plantsci.2014.04.014>
- Dat JF, Foyer CH, Scott IM. Changes in salicylic acid and antioxidants during induced thermotolerance in mustard seedlings. *Plant Physiol* 1998; 118:455-61; PMID:9847121; <http://dx.doi.org/10.1104/pp.118.4.1455>
- Du CK, Li ZG, Gong M. The adaptations to heat and chilling stresses and relation to antioxidant enzymes of maize seedlings induced by salicylic acid. *Plant Physiol Commun* 2005; 41:19-22
- García-Mata C, Lamattina L. Gasotransmitters are emerging as new guard cell signaling molecules and regulators of leaf gas exchange. *Plant Sci* 2013; 201:202:66-73; PMID:23352403; <http://dx.doi.org/10.1016/j.plantsci.2012.11.007>
- Lisjak M, Teklic T, Wilson ID, Whiteman M, Hancock JT. Hydrogen sulfide: Environmental factor or signalling molecule? *Plant Cell Environ* 2013; 36:1607-16; PMID:23347018; <http://dx.doi.org/10.1111/pce.12073>
- Li ZG, Gong M, Liu P. Hydrogen sulfide is a mediator in H₂O₂-induced seed germination in *Jatropha Curcas*. *Acta Physiol Plant* 2012; 34:2207-13; <http://dx.doi.org/10.1007/s11738-012-1021-z>
- Fang T, Cao ZY, Li JL, Shen WB, Huang LQ. Auxin-induced hydrogen sulfide generation is involved in lateral root formation in tomato. *Plant Physiol Biochem* 2014; 76:44-51; PMID:24463534; <http://dx.doi.org/10.1016/j.plaphy.2013.12.024>
- Lisjak M, Teklic T, Wilson ID, Wood M, Whiteman M, Hancock JT. Hydrogen sulfide effects on stomatal apertures. *Plant Signal Behav* 2011; 6:1444-6; PMID:21904118; <http://dx.doi.org/10.4161/psb.6.10.17104>
- Christou A, Manganaris GA, Papadopoulos I, Fotopoulos V. Hydrogen sulfide induces systemic tolerance to salinity and non-ionic osmotic stress in strawberry plants through modification of reactive species biosynthesis and transcriptional regulation of multiple defence pathways. *J Exp Bot* 2013; 64:1953-66; PMID:23567865; <http://dx.doi.org/10.1093/jxb/ert055>
- Fu PN, Wang WJ, Hou LX, Liu X. Hydrogen sulfide is involved in the chilling stress response in *Vitis vinifera* L. *Acta Soc Bot Pol* 2013; 82:295-302; <http://dx.doi.org/10.5586/asbp.2013.031>
- Zhang H, Hua SL, Zhang ZJ, Hua LY, Jiang CX, Wei ZJ, Liu JA, Wang HL, Jiang ST. Hydrogen sulfide acts as a regulator of flower senescence in plants. *Postharv Biol Technol* 2011; 60:251-7; <http://dx.doi.org/10.1016/j.postharvbio.2011.01.006>
- Chen J, Wang WH, Wu FH, You CY, Liu WT, Dong XK, He JX, Zheng HL. Hydrogen sulfide alleviates aluminum toxicity in barley seedlings. *Plant Soil* 2013; 362:301-18; <http://dx.doi.org/10.1007/s11104-012-1275-7>
- Li Z-G, Gong M, Xie H, Yang L, Li J. Hydrogen sulfide donor sodium hydrosulfide-induced heat tolerance in tobacco (*Nicotiana tabacum* L.) suspension cultured cells and involvement of Ca²⁺ and calmodulin. *Plant Sci* 2012; 185/186:185-9; PMID:22325880; <http://dx.doi.org/10.1016/j.plantsci.2011.10.006>
- Li Z-G, Ding X-J, Du P-F. Hydrogen sulfide donor sodium hydrosulfide-improved heat tolerance in maize and involvement of proline. *J Plant Physiol* 2013; 170:741-7; PMID:23523123; <http://dx.doi.org/10.1016/j.jplph.2012.12.018>
- Li Z-G, Yang S-Z, Long W-B, Yang G-X, Shen Z-Z. Hydrogen sulfide may be a novel downstream signal molecule in nitric oxide-induced heat tolerance of maize (*Zea mays* L.) seedlings. *Plant Cell Environ* 2013; 36:1564-72; PMID:23489239; <http://dx.doi.org/10.1111/pce.12092>
- Li Z-G, Luo L-J, Zhu L-P. Involvement of trehalose in hydrogen sulfide donor sodium hydrosulfide-induced the acquisition of heat tolerance in maize (*Zea mays* L.) seedlings. *Bot Stud* 2014; 55:20; <http://dx.doi.org/10.1186/1999-3110-55-20>
- Li Z-G, Yi X-Y, Li Y-T. Effect of pretreatment with hydrogen sulfide donor sodium hydrosulfide on heat tolerance in relation to antioxidant system in maize (*Zea mays*) seedlings. *Biologia* 2014; 69:1001-9
- Wu DH, Li YL, Xia X, Pu ZP, Liao JM, Huang K, Li ZG. Hydrogen sulfide donor sodium hydrosulfide pretreatment improved multiple resistance abilities of wheat to high temperature and drought stress. *J Yunnan Normal Univ* 2013; 33:29-35
- Li Z-G, Zhu L-J. Hydrogen sulfide donor sodium hydrosulfide-induced accumulation of betaine is involved in the acquisition of heat tolerance in maize seedlings. *Braz J Bot* 2015; 38(1):31-8; <http://dx.doi.org/10.1007/s40415-014-0106-x>
- Foyer CH, Noctor G. Redox regulation in photosynthetic organisms: signaling, acclimation, and practical implications. *Antioxid Redox Signal* 2009; 11:861-905; PMID:19239350; <http://dx.doi.org/10.1089/ars.2008.2177>
- Li Z-G, Yuan L-X, Wang Q-L, Ding Z-L, Dong C-Y. Combined action of antioxidant defense system and osmolytes in chilling shock-induced chilling tolerance in *Jatropha curcas* seedlings. *Acta Physiol Plant* 2013; 35:2127-36; <http://dx.doi.org/10.1007/s11738-013-1249-2>