



Published in final edited form as:

*Curr Urol Rep.* 2016 April ; 17(4): 29. doi:10.1007/s11934-016-0584-4.

## Novel Insights into Molecular Indicators of Response and Resistance to Modern Androgen-Axis Therapies in Prostate Cancer

John L. Silberstein<sup>#1</sup>, Maritza N. Taylor<sup>#1</sup>, and Emmanuel S. Antonarakis<sup>2</sup>

<sup>1</sup> Brady Urological Institute, Department of Urology, Johns Hopkins University School of Medicine, Baltimore, MD, USA

<sup>2</sup> Sidney Kimmel Comprehensive Cancer Center at Johns Hopkins, Department of Oncology, Johns Hopkins University School of Medicine, 1650 Orleans St, Baltimore, MD 21287, USA

# These authors contributed equally to this work.

### Abstract

While androgen ablation remains a mainstay for advanced prostate cancer therapy, nearly all patients will inevitably develop disease escape with time. Upon the development of castration-resistant prostate cancer, other androgen-axis-targeted treatments may be added in an effort to starve the disease of its androgen signaling. Nevertheless, additional androgen-pathway resistance usually develops to these novel hormonal therapies. In this review, we will discuss the resistance mechanisms to modern androgen-axis modulators and how these alterations can influence a patient's response to novel hormonal therapy. We conceptualize these resistance pathways as three broad categories: (1) reactivation of androgen/AR-signaling, (2) AR bypass pathways, and (3) androgen/AR-independent mechanisms. We highlight examples of each, as well as potential therapeutic approaches to overcome these resistance mechanisms.

### Keywords

Prostate cancer; Androgen receptor; Splice variants; Resistance; Biomarker

### Introduction

Prostate adenocarcinoma afflicts one in six American men over the course of their lifetime and is the second leading cause of cancer-related deaths in US males behind lung cancer [<sup>1</sup>]. In a cohort of 790 men with metastatic prostate cancer, a landmark trial demonstrated that

---

Emmanuel S. Antonarakis ; Email: eantona1@jhmi.edu

Compliance with Ethical Standards

**Conflict of Interest** John L. Silberstein and Maritza N. Taylor declare no potential conflicts of interest. Emmanuel S. Antonarakis has served as a paid consultant/advisor for Janssen, Astellas, Sanofi, Dendreon, Essa, and Medivation; he has received research funding from Janssen, Johnson & Johnson, Sanofi, Dendreon, Exelixis, Genentech, Novartis, and Tokai; he is a co-inventor of a technology that has been licensed to Tokai.

**Human and Animal Rights and Informed Consent** This article does not contain any studies with human or animal subjects performed by any of the authors.

the combination of hormonal therapy plus docetaxel chemotherapy increased overall survival compared to hormonal therapy alone [2]. However, even with this extended benefit from first-line chemo-hormonal treatment, most prostate cancer patients eventually progress and require treatment with additional second-line hormonal therapy.

Abiraterone acetate and enzalutamide were approved by the FDA in 2011 and 2012, respectively, for the treatment of men with CRPC. Abiraterone blocks extra-gonadal androgen biosynthesis by selectively inhibiting CYP17A1 [3], while enzalutamide directly antagonizes the AR and diminishes AR-signaling [4]. While many patients benefit from these drugs, some men exhibit primary resistance and nearly all eventually develop secondary resistance. The mechanisms underlying this resistance were poorly understood until recently and now are beginning to be elucidated. In this review, we will outline the underlying biology that determines sensitivity or resistance to these modern therapies. We would hope that this review might stimulate new therapeutic combinations to prevent cross-resistance and inspire innovation in the field.

### Rediscovering Androgen Receptor Biology

Prostate cancer is an AR-addicted disease whose development and proliferation strongly rely on adequate AR-signaling. Notwithstanding several unique distinctions, AR shares many commonalities with other steroid hormone receptors in its mechanism of action. Unliganded AR primarily resides in the cytoplasm until ligand-binding triggers its translocation to the nucleus by way of a conformational change in the receptor that liberates it from chaperone heat shock proteins [5]. AR subsequently dimerizes via N/C-terminal interactions, namely the interaction of activation function 2 (AF2) of ARLBD with the NTD [6, 7]. Agonist-bound AR also undergoes phosphorylation at several sites prior to nuclear translocation, including S81 in the NTD (the most highly phosphorylated residue on AR) and S650 in the hinge region [5, 8]. Prostate cancer cell growth is limited in the absence of S81 phosphorylation, and specific S81 phosphorylation by CDK9 regulates AR transcriptional activity [8]. Once inside the nucleus, dimerized AR recognizes and binds to AREs in the promoter or enhancer region of target genes using two zinc fingers, which then stimulates growth, survival, and differentiation of prostate cancer cells [5-7]. Additional interactions with coactivators and corepressors serve to further facilitate or inhibit transcription of these target genes. While agonist-bound AR recruits coactivators that amplify the transcription signal when AR binds to DNA, antagonist-bound AR recruits a complex of corepressors that attenuates the same signal [5].

AR amplification and gene mutations significantly contribute to the emergence of CRPC and disease progression. The AR gene is located on the human X chromosome and contains eight canonical exons: exon 1 encodes the NTD, exons 2-3 encode the DBD, and exons 4-8 encode the C-terminal LBD of the AR-FL protein [5-7]. AR-DBD comprises considerable homology with the DBD of the GR and the PR, all of which may share common gene targets [7, 9\*\*]. AR also contains a hinge region lying between its DBD and LBD, and this hinge region surrounds a nuclear localization signal with the DBD [6]. The structure of the AR-LBD strongly resembles that of many other steroid hormone receptors. The ligand-binding pocket is formed by the folding of 12 helices in the C-terminal and is subject to

conformational changes dependent on ligand-binding [5]. Point mutations in the LBD can thus alter the normal function of the AR, broadening the number of ligands that can bind the LBD [10]. The W742L/W742C mutations, for instance, are linked to bicalutamide resistance, while F877L and T878A mutations have been associated with resistance to novel androgen-directed therapies enzalutamide and abiraterone, respectively [10-13, 14, 15].

The remainder of this review will discuss specific mechanisms related to clinical androgen-pathway drug-resistance. These resistance mechanisms can be broadly divided into three classes: (1) persistent androgen/AR-signaling, (2) AR bypass pathways, and (3) androgen/AR-independent mechanisms (Fig. 1).

### Upregulation of Androgen-Synthetic Enzymes

Recent studies have shown that upregulation of enzymes in the androgen synthesis pathway contributes to castration-resistance as well as abiraterone- and enzalutamide-resistance. Extra-gonadal androgen synthesis may occur in the adrenal glands, as well as intratumorally, through upregulation of 3 $\beta$ -hydroxysteroid dehydrogenase (3 $\beta$ HSD), steroid-5 $\alpha$ -reductase (SRD5A), and 17 $\beta$ -hydroxysteroid dehydrogenase (17 $\beta$ HSD) [16, 17]. 17 $\alpha$ -hydroxylase/17,20-lyase (CYP17A1) itself, the target of abiraterone, has also been shown to be upregulated in prostate cancer patients receiving androgen-axis modulators [16]. Though Salvi et al. [18] demonstrated that AR gene gain in cfDNA had a prognostic effect on PFS in men receiving abiraterone, CYP17A1 gene gain also played a crucial role. Patients with CYP17A1 gain had a median PFS of 2.8 months on abiraterone, while those without such gene gain had a median PFS of 9.2 months [18]. These clinical data suggest that men with copy number gains in the CYP17A1 gene may be less sensitive to abiraterone.

Another androgen-synthetic enzyme that has been implicated in novel hormonal therapy resistance is aldo-keto reductase 1C3 (AKR1C3). This enzyme is responsible for the conversion of 4-androstene-3,17-dione to testosterone, and 5 $\alpha$ -androstenedione to DHT [19]. Intratumoral upregulation of AKR1C3 has been shown to mediate resistance to both enzalutamide and abiraterone in prostate cancer patients, with this enzyme's ability to convert low levels of circulating androgens into potent AR agonists [20, 21]. Although indomethacin is known to be a weak inhibitor of AKR1C3, the search is on to identify additional more specific AKR1C3 inhibitors for clinical development. In addition, a gain-of-function mutation in the 3 $\beta$ HSD1 gene (N367T) has recently been discovered [22]. This mutation renders 3 $\beta$ HSD1 resistant to ubiquitination and degradation, leading to profound intracellular accumulation of the enzyme and increased levels of DHT [22]. Increased 3 $\beta$ HSD1 activity has been associated with resistance to first-line androgen deprivation therapy, and may even be a mechanism of abiraterone resistance [17, 23].

Notably, there may be promise in new compounds that can antagonize multiple parts of the androgen-axis. A metabolite of abiraterone, called 4-abiraterone (D4A), was recently shown to be more potent than abiraterone in vitro [16]. With a chemical structure similar to testosterone, D4A can antagonize the AR directly, in addition to inhibiting CYP17A1, 3 $\beta$ HSD, and SRD5A [16]. Whether D4A will be developed into a clinical drug entity is unclear at this time, but remains of great interest. Galeterone, another CYP17A1 inhibitor, has moved from the lab into the clinic and is currently under investigation in a phase III

clinical trial (NCT02438007; Table 1). This compound may have activity against both AR and AR-Vs due to its ability to trigger E3 ubiquitin ligase function and degrade all forms of the AR protein [24]. An analog of galeterone, VNPT55, may also prove more effective in targeting the multitude of androgen-axis resistance pathways [24].

### Androgen Receptor Amplification

Prostate cancer cells remain responsive to and dependent on androgen-axis signaling, even in the castration-resistant setting [5]. While a temporary halt in disease progression is often observed when these cells are deprived of androgen, AR-signaling adaptations inevitably ensue in response to the selective pressures [5, 25]. One such adaptation is AR gene (and protein) amplification to increase sensitivity to circulating and intratumoral androgens. This heightened sensitivity enables prostate cancer cells to thrive under otherwise limiting conditions [5]. In a study designed to assess the prevalence of AR amplification, Zhang et al. [25] obtained tumor biopsy specimens from 37 patients developing castration-resistant disease, determined AR amplification by FISH analysis, and confirmed the link between AR gene amplification and prostate cancer progression. A clinically significant finding of this study is that patients with AR gene amplification also exhibited increased levels of AR protein, and PFS in these patients was shorter than in patients with normal AR copy numbers, illustrating the significance of AR gene amplification and overexpression in the development of CRPC [25].

AR amplification can also be linked with resistance to enzalutamide and abiraterone. Though not associated with HSPC, AR copy number gain often indicates disease progression in patients with CRPC and is correlated with poor prognosis [14, 15, 18]. Patients with AR amplification prior to initiating enzalutamide have worse clinical outcomes, exhibited by significantly lower PSA response rates and shorter PFS [14]. Azad et al. [14] analyzed cfDNA from 62 metastatic-CRPC (mCRPC) patients progressing on systemic therapy, which revealed that AR amplification occurred at a significantly higher frequency in patients progressing on enzalutamide than on any other agent (53 vs. 21 %). Interestingly, however, a previous study exhibited that bone marrow-specific nuclear AR expression, as well as CYP17A1 expression, was correlated with benefit to enzalutamide therapy [26]. More recently, enzalutamide was shown to trigger a subcellular nuclear-to-cytoplasmic shift of AR, thereby suppressing AR-signaling, but also inducing an adaptive feedback mechanism in which testosterone levels are increased in the bone marrow [26]. Combined therapy that directly and indirectly inhibits AR-signaling through AR antagonism and androgen biosynthesis inhibition could evade this feedback mechanism and improve therapeutic efficacy in some CRPC patients.

In the setting of abiraterone, Carreira et al. [10] observed AR copy number gain in 35 and 37 % of CRPC liquid and tumor biopsy samples, respectively, compared to only in 6 % of precastration samples. Moreover, AR copy number gain was associated with resistance to abiraterone [10]. Similarly, Salvi et al. [18] evaluated AR copy number variations (CNVs) in cfDNA from 53 CRPC patients prior to receiving abiraterone, 16 of which exhibited AR gain. Ten (62.5 %) of those 16 patients exhibited early progression on abiraterone, also suggesting that AR CNVs may predict abiraterone resistance [18]. Likewise, abiraterone-

naïve patients with AR gain have been reported to be 4.9 times less likely to have a 50 % PSA reduction and also exhibit significantly worse PFS and OS than patients without CNVs [15]. Salvi et al. [18] concordantly found that AR amplification remained predictive of both PFS and OS, as patients with AR gene gain had PFS 3.4 times shorter and OS more than fourfold shorter than patients expressing normal AR copy numbers. Notably, a recent study showed that AR copy number at progression on abiraterone remains relatively unchanged from baseline, suggesting that abiraterone resistance in patients expressing normal AR copy numbers may be explained by other mechanisms [15].

### Androgen Receptor Point Mutations

Another way the AR-signaling axis can be rescued in prostate cancer is via point mutations in the LBD that develop in response to treatment with specific antiandrogens. These mutations alter the steric and chemical properties of the ligand-binding pocket, thus conferring AR agonistic activity to alternative ligands and former AR antagonists [5, 27, 28]. Numerous studies have confirmed the importance of these mutations in patients receiving novel hormonal therapies. For example, the L702H point mutation converts AR to a glucocorticoid-activated phenotype, while T878A renders AR progesterone-responsive [10, 27, 28]. L702H often mediates resistance to abiraterone treatment due to the fact that abiraterone is given together with corticosteroids, which can agonize this mutated AR [10, 15]. Similarly, T878A allows for abiraterone resistance since abiraterone inhibits CYP17A1, resulting in an increase in upstream steroids (i.e., progestins) [29].

F877L also appears to mediate resistance to enzalutamide and apalutamide (ARN-509) [10, 12, 13, 14, 30]. This mutation transforms enzalutamide and apalutamide into AR agonists, and also maintains the AR's sensitivity to androgens. F877L is probably more relevant as an acquired (rather than a primary) mechanism of resistance to enzalutamide/apalutamide and is found in <10 % of patients at the time of progression on these agents [30].

A novel antiandrogen, ODM-201, has been shown to antagonize AR even with the F877L and T878A mutations in pre-clinical studies [31]. These data, combined with the drug's promising phase II clinical results [32], have led to a phase III study using this compound in non-metastatic castration-resistant men (NCT02200614; Table 1). ODM-201's lower risk of seizures and activity against mutant AR may help improve its chances for FDA approval in an ever-crowded antiandrogen market.

### Androgen Receptor Splice Variants

Constitutively active AR-Vs contribute yet another mechanism of resistance to novel hormonal therapies, as these truncated molecules have been shown to regulate transcription in an androgen-independent fashion [33]. Upward of 20 AR-Vs have been identified to date [34••], some whose function has been well elucidated, others whose role in advanced prostate cancer remains obscure. AR-Vs usually result from cryptic exon insertions downstream of DBD coding regions or (much more rarely in humans) from deletion of LBD-coding regions [33]. ADT enhances the rate of AR gene transcription and recruitment of critical splicing factors to AR pre-mRNAs, which can result in increased expression of AR-V mRNA and protein [35]. An inverse relationship between canonical AR-FL signaling

and AR-V expression has previously been shown [36]: inhibition of AR-FL signaling corresponded with increased expression of AR-Vs in vitro, whereas AR-Vs were not overexpressed during normal AR-FL signaling. Compared to levels in HSPC, AR-V1 and AR-V7 levels have been shown to be increased >20-fold in CRPC [33]. These AR-Vs may serve as transcription factors for enhanced expression of cell-cycle genes, while AR-FL signaling may preferentially activate transcription of genes involved in biosynthesis, cellular metabolism, and differentiation [36]. Most notably, Hu et al. [36] demonstrated a correlation between AR-V7 (and AR<sup>V567es</sup>) overexpression and upregulation of the cell-cycle gene UBE2C in CRPC patient specimens. Other studies have shown that AR<sup>V567es</sup> is upregulated in the castration setting, promotes expression and activity of AR-FL, activates a gene set distinct from AR-FL, and is associated with increased nuclear localization of AR and shorter survival [37, 38].

AR-V7 in particular has emerged as an important biomarker, yielding insights into disease prognosis and response to novel hormonal agents. Retrospective studies of human bone and prostate biopsies suggest inferior clinical outcomes for CRPC patients overexpressing AR-V7 [38, 39]. Another study showed that AR-V7 was upregulated in 429 prostate cancer biopsy specimens, and increased nuclear expression of ARV7 was associated with higher risk of disease recurrence following radical prostatectomy [40]. To support this idea of ARV7 driven progression, knockdown of AR-V7 resulted in a weakened propensity for prostate cancer cell growth [40].

Recent prospective studies have shown that using AR-V7 as a prognostic marker is feasible in the setting of novel hormonal therapy. A myriad of data bolster AR-V7 status as an independent predictive factor in disease development and progression, and as a marker of resistance to enzalutamide and abiraterone. In an early study collecting bone marrow specimens from men embarking on enzalutamide treatment, patients with high levels of pre-treatment androgen signaling demonstrated clinical benefit, while AR-V7 positivity (using immunohistochemistry) was associated with primary resistance [26]. These findings were corroborated by a study from Antonarakis et al. [41••] in 62 mCRPC patients (31 treated with enzalutamide, 31 treated with abiraterone) analyzing baseline CTC-derived AR-V7 status as a predictor of response or resistance to these therapies. Notably, >50 % of patients pre-treated with abiraterone and/or enzalutamide were ARV7-positive as indicated by the CTC-based RT-PCR assay, whereas <15 % of abiraterone-naïve and enzalutamide-naïve patients were found to express AR-V7 [41••]. Men receiving abiraterone or enzalutamide who were positive for AR-V7 had no PSA responses, shorter clinical or radiographic PFS, and shorter OS than their AR-V7 negative counterparts [41••]. Interestingly, AR-V7 negative to positive conversions were noted in four patients receiving enzalutamide and in two patients receiving abiraterone: these men had intermediate clinical outcomes [41••]. In a third study, Steinestel et al. [42] showed that AR-V7 positivity in CTCs most likely emerges under selective therapeutic pressures and is associated with the absolute number of prior hormonal therapies received. That study also showed inferior clinical outcomes to abiraterone and enzalutamide in AR-V7-positive compared to AR-V7-negative men [42]. Of note, no ADT-naïve patient had detectable levels of AR-V7 in their CTCs, supporting the idea that AR-V7 expression may be an adaptive response to first-line and novel androgen-axis therapies.

AR-V7 status has also become an important biomarker in the setting of chemotherapy. Though AR-V7 positivity in CTCs is linked to enzalutamide and abiraterone resistance, taxane chemotherapy may remain an effective therapeutic alternative in these patients. PSA responses have been observed in taxane-treated patients irrespective of AR-V7 status and prior treatment with second-line androgen-axis therapies [43–45], and treatment with cabazitaxel in particular does not appear to be influenced by AR-V7 status [44]. In post hoc analyses, AR-V7-positive men treated with taxane chemotherapy exhibit superior clinical outcomes to those treated with enzalutamide or abiraterone [43]. Interestingly, the authors reported that only one AR-V7-negative patient converted to AR-V7-positive while receiving taxane chemotherapy, whereas nearly 50 % of AR-V7-positive patients became ARV7-negative during taxane treatment [43]. This phenomenon may be due to conversion from CTC-positive to CTC-negative. Nakazawa et al. [45] also observed conversions from AR-V7-negative to positive status in men treated with AR-directed therapies and taxane chemotherapies; whereas, ARV7-positive to negative reversions were observed solely with taxane chemotherapies [45]. Conversions may reflect adaptive induction of and reliance on AR-V7 to maintain AR-axis signaling, while reversions may reflect some disinhibition of canonical AR-signaling and thereby reduced pressure for ARV7 expression [45]. AR-V7 reversions after taxane chemotherapy may present a unique opportunity for benefit from re-treatment with enzalutamide or abiraterone, although confirmation of this hypothesis is awaited. At this time, the clinical significance of AR-V7 transitions remains unclear.

Some potential therapeutic strategies to overcome AR-V-mediated resistance are listed in Table 1. These include drugs that target and degrade all AR protein (including AR-Vs; e.g., galeterone), agents that inhibit the AR-NTD (e.g., EPI-506), and epigenetic therapies that interfere with AR transcriptional activity (e.g., bromodomain/BET inhibitors). Moreover, combinatorial immunotherapy strategies (e.g., ipilimumab plus nivolumab) may be a fruitful approach in AR-V-expressing patients.

### Glucocorticoid Receptor Induction

Another proposed mechanism of resistance to androgen-axis therapies involves upregulation of the GR. The GR may be able to substitute for the AR in various circumstances, binding to AREs and other promoter elements to sustain cell survival [9••]. These two steroid receptors have overlapping transcriptomes, potentially allowing proliferation signals to bypass the AR in patients being treated with enzalutamide or apalutamide [9••]. Of the tissues analyzed in one study, GR mRNA levels were 27-fold higher in enzalutamide- and apalutamide-resistant tumors compared to control [9••]. In vivo, cells expressing high levels of GR were resistant to treatment with enzalutamide and grew aggressively; whereas, LNCaP cells (which express low levels of GR) showed minimal growth [9••]. By knocking down the GR with shRNA, tumor growth was significantly delayed in this previously drug-resistant mouse model [9••]. Concordant results were also seen when assessing human bone marrow biopsies by immunohistochemistry in enzalutamide-treated patients, with GR-positive patients being less likely to have a durable response to therapy [9••]. These data might also imply that enzalutamide should not be given together with corticosteroids, which may be capable of further agonizing an upregulated GR in this setting. Intriguingly, a hypothesis-generating post hoc analysis of the AFFIRM study (which allowed treating physicians to use concurrent

steroids, if desired, together with enzalutamide) suggested that enzalutamide produced inferior PFS and OS when combined with steroids [46].

Another hypothesis suggests that GR is negatively regulated by AR, meaning that AR inhibition automatically leads to GR upregulation. ChIP analysis in one study confirmed that an ARE regulates the expression of GR, leading to more questions regarding the high levels of GR in enzalutamide-resistant cells [47]. However, clinical studies have shown PSA reductions in response to dexamethasone and other steroids (GR agonists) [48-51]. This responsiveness may also indicate that PSA levels are not regulated by the GR. To further clarify the GR landscape, a study using dexamethasone in patients developing acquired resistance to enzalutamide is currently underway, and will allow re-treatment with enzalutamide after a period of dexamethasone therapy (NCT02491411; Table 1). Conversely, another trial is analyzing the use of a GR antagonist, mifepristone, in combination with enzalutamide, to block both signaling pathways concurrently (NCT02012296; Table 1). Further studies must be done to show the importance of the GR in maintaining tumor cell populations and inducing proliferation in CRPC. If shown to be a driving force behind drug-resistance to enzalutamide or other novel hormonal agents, the GR has the potential to be therapeutically targeted in combination with other AR-directed strategies.

### Progesterone Receptor Activation

The PR also shares significant homology with the AR, especially in the DBD (>80 % identity in this region). Therefore, it is also possible that PR (like GR) may become induced during androgen ablation, thereby restoring expression of AR-regulated genes [52, 53]. In one study, PR was detected using immunohistochemical staining in about 30 % of mCRPC biopsy specimens [54], and preliminary clinical data from another study suggest that activated (nuclear) PR may be associated with development of castration-resistance [55]. While the role of PR in castration-resistant progression admittedly remains unclear, a phase I/II study is currently being conducted using the oral PR antagonist, onapristone, in men with abiraterone- or enzalutamide-refractory CRPC (NCT02049190; Table 1) [56]. In that study, all patients will undergo baseline tumor biopsies to evaluate activated PR using an analytically validated immunohistochemical assay.

### Androgen/AR-Independent Mechanisms

While the androgen/AR pathways are important in understanding progression on novel hormonal therapies, many other resistance mechanisms have also been implicated (Fig. 1; Table 2). With the addition of modern androgen/AR-directed therapies to the treatment regimen, refractory prostate cancer can develop increasingly more mutations and lead to an aggressive and lethal phenotype. For example, neuroendocrine differentiation can result from the loss or mutation of tumor suppressors like Rb and/or p53 (Table 2) [34••, 57]. PTEN is also a commonly lost tumor suppressor gene in prostate cancer, and its loss has been shown to drive metastasis (Table 2) [34••, 58, 59]. Recent studies have linked both canonical and non-canonical Wnt signaling to therapeutic resistance and disease progression (Fig. 1) [34••, 60•, 61]. Similarly, amplification of two oncogenes, N-Myc (MYCN) and Aurora kinase A (AURKA), can lead to neuroendocrine differentiation (Table 2) [62].



MYCN appears to be upregulated in 40 % of neuroendocrine prostate cancer cases and approximately 5 % of prostate adenocarcinomas [63]. Each of these pathways presents a potential therapeutic target for the future, and ongoing clinical trials will determine their clinical relevance moving forward. For a more detailed review of androgen/AR-independent mechanisms of escape, we refer the reader to several excellent recent reviews [64, 65].

DNA repair pathways have also emerged recently as a clinically relevant and exploitable avenue for therapeutic manipulation, and it is becoming increasingly understood that PARP enzymes may have a dual role in DNA damage repair and AR transcriptional regulation [66]. Since prostate cancer cells are rapidly multiplying, they need to upregulate DNA repair enzymes due to the constant stress of proliferation. A recent study showed that patients with defects in certain DNA repair enzymes (including BRCA1/2, ATM, Fanconi's anemia genes, and CHEK2) had an 88 % response rate to the PARP inhibitor olaparib [67\*\*]. These results may lead to biomarker-driven precision medicine trials using biopsies to determine treatment selection. This study has led to a subsequent phase II trial in mCRPC patients who have failed taxane chemotherapy (NCT01682772; Table 1). As we begin to have a better understanding of this disease's underlying biology, we can develop more rational therapeutics and pragmatic clinical trials.

## Conclusions

The resistance mechanisms behind androgen-axis therapies are numerous and our knowledge of them is ever expanding. New agents are entering clinical trials that hope to block AR-signaling even further than the compounds currently approved by the FDA. EPI-506, with its ability to bind the NTD of AR-FL and AR-Vs in the preclinical setting, is currently being tested in a phase I clinical trial (NCT02606123; Table 1). Galeterone has entered a pivotal phase III clinical trial in AR-V7-positive patients and may prove superior to enzalutamide in that population (NCT02438007; Table 1). In regard to the current therapies on the market, our best strategy may be to use biomarkers to determine a patient's ideal opportunity for responsiveness. However, even those patients with response to these therapies will develop progressive disease.

With this in mind, one new approach is to target pathways that are less variable and cannot be mutated in response to therapy, perhaps using immune-directed strategies. To this end, a biomarker-driven immunotherapy trial in prostate cancer is going to test ipilimumab (an anti-CTLA-4 antibody) combined with nivolumab (an anti-PD-1 antibody) in AR-V7-positive CRPC patients (NCT02601014; Table 1). The hypothesis is that the cancers from these patients will have higher mutational burden and will respond better to therapies that help the immune system recognize self from non-self (i.e., cancer). If successful, this trial could change the prostate cancer landscape from that of novel hormonally targeted therapies to one of immunotherapies. Immunotherapy has been successful in many other cancer types, and remains the only systemic therapy that can produce lasting responses after treatment has been completed.

In conclusion, treatment selection using androgen-axis modulators should ideally be guided by biomarkers, including AR-Vs and AR mutations as well as others. The use of such

biomarkers (whether derived from biopsies, CTCs, or circulating nucleic acids) promises to play a critical role in determining which patients will respond best to certain therapies. Only by further prospectively validating these biomarkers in the clinic and developing new ones can we create a superior experience for prostate cancer patients.

## Abbreviations

<b>ADT</b>	androgen deprivation therapy
<b>AR</b>	androgen receptor
<b>ARE</b>	androgen response element
<b>AR-FL</b>	full-length androgen receptor
<b>AR-V</b>	androgen receptor splice variant
<b>cfDNA</b>	cell-free DNA
<b>CRPC</b>	castration-resistant prostate cancer
<b>CTC</b>	circulating tumor cell
<b>CYP17A1</b>	cytochrome P450 17A1
<b>DBD</b>	DNA-binding domain
<b>DHT</b>	dihydrotestosterone
<b>GR</b>	glucocorticoid receptor
<b>HSPC</b>	hormone-sensitive prostate cancer
<b>LBD</b>	ligand-binding domain
<b>NTD</b>	N-terminal domain
<b>PFS</b>	progression-free survival
<b>PR</b>	progesterone receptor
<b>OS</b>	overall survival

## References

Papers of particular interest, published recently, have been highlighted as:

• Of importance

•• Of major importance

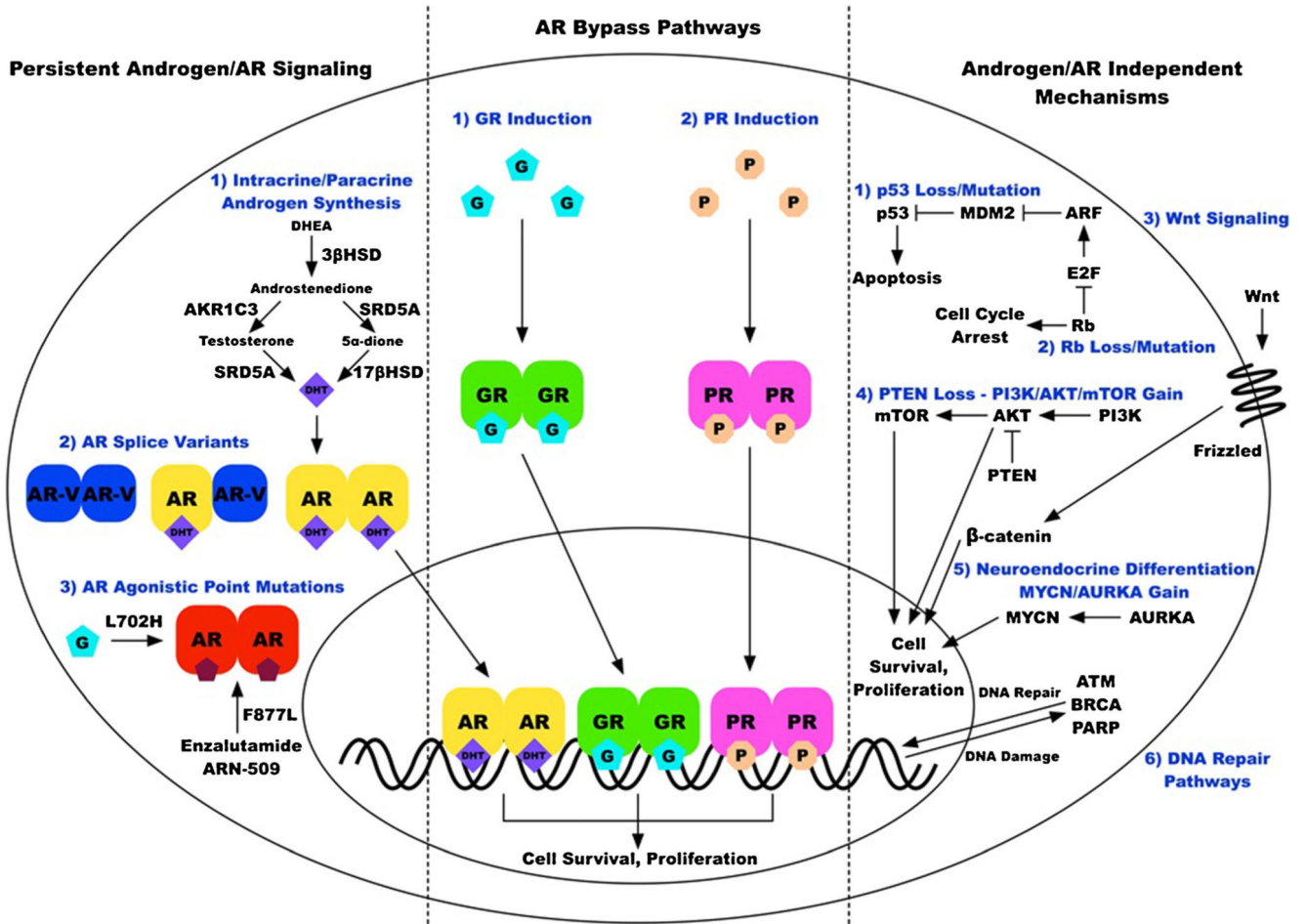
1. Howlander, N.; Noone, AM.; Krapcho, M.; Garshell, J.; Miller, D.; Altekruse, SF., et al. SEER Cancer Statistics Review, 1975–2012. 2015. Available from: [http://seer.cancer.gov/csr/1975\\_2012/](http://seer.cancer.gov/csr/1975_2012/)
2. Sweeney CJ, Chen YH, Carducci M, Liu G, Jarrard DF, Eisenberger M, et al. Chemohormonal therapy in metastatic hormone-sensitive prostate cancer. *N Engl J Med.* 2015; 373:737–46. [PubMed: 26244877]
3. Fizazi K, Scher HI, Molina A, Logothetis CJ, Chi KN, Jones RJ, et al. Abiraterone acetate for treatment of metastatic castration-resistant prostate cancer: final overall survival analysis of the

- COU-AA-301 randomised, double-blind, placebo-controlled phase 3 study. *Lancet Oncol.* 2012; 13:983–92. [PubMed: 22995653]
4. Scher HI, Fizazi K, Saad F, Taplin ME, Sternberg CN, Miller K, et al. Increased survival with enzalutamide in prostate cancer after chemotherapy. *N Engl J Med.* 2012; 367:1187–97. [PubMed: 22894553]
  5. Gelmann EP. Molecular biology of the androgen receptor. *J Clin Oncol.* 2002; 20:3001–15. [PubMed: 12089231]
  6. Gao W, Bohl CE, Dalton JT. Chemistry and structural biology of androgen receptor. *Chem Rev.* 2005; 105:3352–70. [PubMed: 16159155]
  7. Egan A, Dong Y, Zhang H, Qi Y, Balk SP, Sartor O. Castration-resistant prostate cancer: adaptive responses in the androgen axis. *Cancer Treat Rev.* 2014; 40:426–33. [PubMed: 24139549]
  8. Gordon V, Bhadel S, Wunderlich W, Zhang J, Ficarro SB, Mollah SA, et al. CDK9 regulates AR promoter selectivity and cell growth through serine 81 phosphorylation. *Mol Endocrinol.* 2010; 24:2267–80. [PubMed: 20980437]
  - 9••. Arora VK, Schenkein E, Murali R, Subudhi SK, Wongvipat J, Balbas MD, et al. Glucocorticoid receptor confers resistance to antiandrogens by bypassing androgen receptor blockade. *Cell.* 2013; 155:1309–22. [PubMed: 24315100] [This article is one of the first studies to show the clinical relevance of GR in prostate cancer, especially in the setting of enzalutamide resistance.]
  10. Carreira S, Romanel A, Goodall J, Grist E, Ferraldeschi R, Miranda S, et al. Tumor clone dynamics in lethal prostate cancer. *Sci Transl Med.* 2014; 6:254ra125.
  11. Korpai M, Korn JM, Gao X, Rakiec DP, Ruddy DA, Doshi S, et al. An F876L mutation in androgen receptor confers genetic and phenotypic resistance to MDV3100 (enzalutamide). *Cancer Discov.* 2013; 3:1030–43. [PubMed: 23842682]
  12. Joseph JD, Lu N, Qian J, Sensintaffar J, Shao G, Brigham D, et al. A clinically relevant androgen receptor mutation confers resistance to second-generation antiandrogens enzalutamide and ARN-509. *Cancer Discov.* 2013; 3:1020–9. [PubMed: 23779130]
  13. Balbas MD, Evans MJ, Hosfield DJ, Wongvipat J, Arora VK, Watson PA, et al. Overcoming mutation-based resistance to antiandrogens with rational drug design. *eLife.* 2013; 2:e00499. [PubMed: 23580326]
  - 14•. Azad AA, Volik SV, Wyatt AW, Haegert A, Le Bihan S, Bell RH, et al. Androgen receptor gene aberrations in circulating cell-free DNA: biomarkers of therapeutic resistance in castration-resistant prostate cancer. *Clin Cancer Res.* 2015; 21:2315–24. [PubMed: 25712683] [This study showed the feasibility and prognostic impact of using cfDNA as a biomarker to analyze androgen receptor copy number variations and mutations.]
  15. Romanel A, Tandefelt DG, Conteduca V, Jayaram A, Casiraghi N, Wetterskog D, et al. Plasma AR and abiraterone-resistant prostate cancer. *Sci Transl Med.* 2015; 7:312re10.
  16. Li Z, Bishop AC, Alyamani M, Garcia JA, Dreicer R, Bunch D, et al. Conversion of abiraterone to D4A drives anti-tumour activity in prostate cancer. *Nature.* 2015; 523:347–51. [PubMed: 26030522]
  17. Li R, Evaul K, Sharma KK, Chang KH, Yoshimoto J, Liu J, et al. Abiraterone inhibits 3beta-hydroxysteroid dehydrogenase: a rationale for increasing drug exposure in castration-resistant prostate cancer. *Clin Cancer Res.* 2012; 18:3571–9. [PubMed: 22753664]
  18. Salvi S, Casadio V, Conteduca V, Burgio SL, Menna C, Bianchi E, et al. Circulating cell-free AR and CYP17A1 copy number variations may associate with outcome of metastatic castration-resistant prostate cancer patients treated with abiraterone. *Br J Cancer.* 2015; 112:1717–24. [PubMed: 25897673]
  19. Lin HK, Jez JM, Schlegel BP, Peehl DM, Pachter JA, Penning TM. Expression and characterization of recombinant type 2 3 alpha-hydroxysteroid dehydrogenase (HSD) from human prostate: demonstration of bifunctional 3 alpha/17 beta-HSD activity and cellular distribution. *Mol Endocrinol.* 1997; 11:1971–84. [PubMed: 9415401]
  20. Liu C, Lou W, Zhu Y, Yang JC, Nadiminty N, Gaikwad NW, et al. Intracrine androgens and AKR1C3 activation confer resistance to enzalutamide in prostate cancer. *Cancer Res.* 2015; 75:1413–22. [PubMed: 25649766]

21. Tamae D, Mostaghel E, Montgomery B, Nelson PS, Balk SP, Kantoff PW, et al. The DHEA-sulfate depot following P450c17 inhibition supports the case for AKR1C3 inhibition in high risk localized and advanced castration resistant prostate cancer. *Chem Biol Interact.* 2015; 234:332–8. [PubMed: 25514466]
22. Chang KH, Li R, Kuri B, Lotan Y, Roehrborn CG, Liu J, et al. A gain-of-function mutation in DHT synthesis in castration-resistant prostate cancer. *Cell.* 2013; 154:1074–84. [PubMed: 23993097]
23. Hearn JWD, AbuAli G, Magi-Galluzzi C, Reddy CA, Chang KH, Klein EA, et al. HSD3B1 and resistance to androgen deprivation therapy in prostate cancer. *J Clin Oncol.* 2015; 33(suppl 7) abstr 156.
24. Kwegyir-Afful AK, Senthilmurugan R, Purushottamachar P, Ramamurthy VP, Njar VC. Galeterone and VNPT55 induce proteasomal degradation of AR/AR-V7, induce significant apoptosis via cytochrome c release and suppress growth of castration resistant prostate cancer xenografts in vivo. *Oncotarget.* 2015; 6:27440–60. [PubMed: 26196320]
25. Zhang X, Hong SZ, Lin EJ, Wang DY, Li ZJ, Chen LI. Amplification and protein expression of androgen receptor gene in prostate cancer cells: fluorescence hybridization analysis. *Oncol Lett.* 2015; 9:2617–22. [PubMed: 26137116]
26. Efstathiou E, Titus M, Wen S, Hoang A, Karlou M, Ashe R, et al. Molecular characterization of enzalutamide-treated bone metastatic castration-resistant prostate cancer. *Eur Urol.* 2015; 67:53–60. [PubMed: 24882673]
27. Zhao XY, Malloy PJ, Krishnan AV, Swami S, Navone NM, Peehl DM, et al. Glucocorticoids can promote androgen-independent growth of prostate cancer cells through a mutated androgen receptor. *Nat Med.* 2000; 6:703–6. [PubMed: 10835690]
28. Steketee K, Timmerman L, Ziel-van der Made AC, Doesburg P, Brinkmann AO, Trapman J. Broadened ligand responsiveness of androgen receptor mutants obtained by random amino acid substitution of H874 and mutation hot spot T877 in prostate cancer. *Int J Cancer.* 2002; 100:309–17. [PubMed: 12115546]
29. Chen EJ, Sowalsky AG, Gao S, Cai C, Voznesensky O, Schaefer R, et al. Abiraterone treatment in castration-resistant prostate cancer selects for progesterone responsive mutant androgen receptors. *Clin Cancer Res.* 2015; 21:1273–80. [PubMed: 25320358]
30. Rathkopf, DE.; Smith, MR.; Antonarakis, ES.; Ryan, CJ.; Berry, WR.; Shore, ND., et al. AACR Annual Meeting. Vol. 75. *Cancer Res*; Philadelphia, PA: 2015. Androgen receptor mutations in patients with castration-resistant prostate cancer with and without prior abiraterone acetate treatment.. abstr CT134
31. Moilanen AM, Riikonen R, Oksala R, Ravanti L, Aho E, Wohlfahrt G, et al. Discovery of ODM-201, a new-generation androgen receptor inhibitor targeting resistance mechanisms to androgen signaling-directed prostate cancer therapies. *Sci Rep.* 2015; 5:12007. [PubMed: 26137992]
32. Fizazi K, Massard C, Bono P, Jones R, Kataja V, James N, et al. Activity and safety of ODM-201 in patients with progressive metastatic castration-resistant prostate cancer (ARADES): an open-label phase 1 dose-escalation and randomised phase 2 dose expansion trial. *Lancet Oncol.* 2014; 15:975–85. [PubMed: 24974051]
33. Hu R, Dunn TA, Wei S, Isharwal S, Veltri RW, Humphreys E, et al. Ligand-independent androgen receptor variants derived from splicing of cryptic exons signify hormone-refractory prostate cancer. *Cancer Res.* 2009; 69:16–22. [PubMed: 19117982]
- 34••. Robinson D, Van Allen EM, Wu YM, Schultz N, Lonigro RJ, Mosquera JM, et al. Integrative clinical genomics of advanced prostate cancer. *Cell.* 2015; 161:1215–28. [PubMed: 26000489] [This study is one of the most comprehensive genomic analyses of castration-resistant prostate cancer.]
35. Liu LL, Xie N, Sun S, Plymate S, Mostaghel E, Dong X. Mechanisms of the androgen receptor splicing in prostate cancer cells. *Oncogene.* 2014; 33:3140–50. [PubMed: 23851510]
36. Hu R, Lu C, Mostaghel EA, Yegnasubramanian S, Gurel M, Tannahill C, et al. Distinct transcriptional programs mediated by the ligand-dependent full-length androgen receptor and its splice variants in castration-resistant prostate cancer. *Cancer Res.* 2012; 72:3457–62. [PubMed: 22710436]

37. Sun S, Sprenger CC, Vessella RL, Haugk K, Soriano K, Mostaghel EA, et al. Castration resistance in human prostate cancer is conferred by a frequently occurring androgen receptor splice variant. *J Clin Invest.* 2010; 120:2715–30. [PubMed: 20644256]
38. Hornberg E, Ylitalo EB, Crnalic S, Antti H, Stattin P, Widmark A, et al. Expression of androgen receptor splice variants in prostate cancer bone metastases is associated with castration-resistance and short survival. *PLoS One.* 2011; 6:e19059. [PubMed: 21552559]
39. Qu Y, Dai B, Ye D, Kong Y, Chang K, Jia Z, et al. Constitutively active AR-V7 plays an essential role in the development and progression of castration-resistant prostate cancer. *Sci Rep.* 2015; 5:7654. [PubMed: 25563505]
40. Guo Z, Yang X, Sun F, Jiang R, Linn DE, Chen H, et al. A novel androgen receptor splice variant is up-regulated during prostate cancer progression and promotes androgen depletion-resistant growth. *Cancer Res.* 2009; 69:2305–13. [PubMed: 19244107]
- 41••. Antonarakis ES, Lu C, Wang H, Luber B, Nakazawa M, Roeser JC, et al. AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. *N Engl J Med.* 2014; 371:1028–38. [PubMed: 25184630] [This study showed that AR-V7 might be a clinically relevant biomarker for enzalutamide and abiraterone resistance that can be tested from CTCs through non-invasive means.]
42. Steinestel J, Luedeke M, Arndt A, Schnoeller TJ, Lennerz JK, Wurm C, et al. Detecting predictive androgen receptor modifications in circulating prostate cancer cells. *Oncotarget.* 2015
43. Antonarakis ES, Lu C, Luber B, Wang H, Chen Y, Nakazawa M, et al. Androgen receptor splice variant 7 and efficacy of taxane chemotherapy in patients with metastatic castration-resistant prostate cancer. *JAMA Oncol.* 2015; 1:582–91. [PubMed: 26181238]
44. Onstenk W, Sieuwerts AM, Kraan J, Van M, Nieuweboer AJ, Mathijssen RH, et al. Efficacy of cabazitaxel in castration-resistant prostate cancer is independent of the presence of AR-V7 in circulating tumor cells. *Eur Urol.* 2015; 68:939–45. [PubMed: 26188394]
45. Nakazawa M, Lu C, Chen Y, Paller CJ, Carducci MA, Eisenberger MA, et al. Serial blood-based analysis of AR-V7 in men with advanced prostate cancer. *Ann Oncol.* 2015; 26:1859–65. [PubMed: 26117829]
46. Scher HI, Fizazi K, Saad F, Chi KN, Taplin ME, Sternberg CN, et al. Impact of on-study corticosteroid use on efficacy and safety in the phase III AFFIRM study of enzalutamide, an androgen receptor inhibitor. *J Clin Oncol.* 2013; 31(suppl 6) abstr 6.
47. Xie N, Cheng H, Lin D, Liu L, Yang O, Jia L, et al. The expression of glucocorticoid receptor is negatively regulated by active androgen receptor signaling in prostate tumors. *Int J Cancer.* 2015; 136:E27–38. [PubMed: 25138562]
48. Storlie JA, Buckner JC, Wiseman GA, Burch PA, Hartmann LC, Richardson RL. Prostate specific antigen levels and clinical response to low dose dexamethasone for hormone-refractory meta-static prostate carcinoma. *Cancer.* 1995; 76:96–100. [PubMed: 8630883]
49. Nishimura K, Nonomura N, Yasunaga Y, Takaha N, Inoue H, Sugao H, et al. Low doses of oral dexamethasone for hormone-refractory prostate carcinoma. *Cancer.* 2000; 89:2570–6. [PubMed: 11135218]
50. Shamash J, Powles T, Sarker SJ, Protheroe A, Mithal N, Mills R, et al. A multi-centre randomised phase III trial of Dexamethasone vs Dexamethasone and diethylstilbestrol in castration-resistant prostate cancer: immediate vs deferred Diethylstilbestrol. *Br J Cancer.* 2011; 104:620–8. [PubMed: 21285990]
51. Venkitaraman R, Thomas K, Huddart RA, Horwich A, Dearnaley DP, Parker CC. Efficacy of low-dose dexamethasone in castration-refractory prostate cancer. *BJU Int.* 2008; 101:440–3. [PubMed: 17941935]
52. Miyahira AK, Simons JW, Soule HR. The 20th Annual Prostate Cancer Foundation Scientific Retreat report. *Prostate.* 2014; 74:811–9. [PubMed: 24719035]
53. Grindstad T, Andersen S, Al-Saad S, Donnem T, Kiselev Y, Nordahl Melbo-Jorgensen C, et al. High progesterone receptor expression in prostate cancer is associated with clinical failure. *PLoS One.* 2015; 10:e0116691. [PubMed: 25723513]

54. Mateo, J.; Nowakowska, K.; Jayaram, A.; Rodrigues, DN.; Riisnaes, R.; Zukiwski, A., et al. Prostate Cancer Foundation Scientific Retreat. Carlsbad, CA: 2014. Phase 1 study of onapristone, a progesterone receptor (PR) antagonist, in castration-resistant prostate cancer.. abstract 60
55. Zukiwski A, Bosq J, Gilles EM, Beldegrun A. Progesterone receptor (PR), a potential mechanism of resistance and target in AIPC. Prostate Cancer Foundation Scientific Retreat. 2014 abstract 47.
56. Mateo, J.; Rodrigues, DN.; Lopez, RP.; Flohr, P.; Riisnaes, R.; Lokiec, FM., et al. A phase 1–2 study of the type I progesterone receptor (PR) antagonist, onapristone, in patients with advanced castration-resistant prostate cancer.. *J Clin Oncol*; ASCO Annual Meeting; 2014; abstract TPS5097
57. Tan HL, Sood A, Rahimi HA, Wang W, Gupta N, Hicks J, et al. Rb loss is characteristic of prostatic small cell neuroendocrine carcinoma. *Clin Cancer Res*. 2014; 20:890–903. [PubMed: 24323898]
58. Mithal P, Allott E, Gerber L, Reid J, Welbourn W, Tikishvili E, et al. PTEN loss in biopsy tissue predicts poor clinical outcomes in prostate cancer. *Int J Urol*. 2014; 21:1209–14. [PubMed: 25099119]
59. Mulholland DJ, Kobayashi N, Ruscetti M, Zhi A, Tran LM, Huang J, et al. Pten loss and RAS/ MAPK activation cooperate to promote EMT and metastasis initiated from prostate cancer stem/progenitor cells. *Cancer Res*. 2012; 72:1878–89. [PubMed: 22350410]
60. Miyamoto DT, Zheng Y, Wittner BS, Lee RJ, Zhu H, Broderick KT, et al. RNA-Seq of single prostate CTCs implicates noncanonical Wnt signaling in antiandrogen resistance. *Science*. 2015; 349:1351–6. [PubMed: 26383955] [This study was the first to accomplish single-cell RNA sequencing and uncovered a new potential mechanism of enzalutamide resistance related to non-canonical Wnt signaling.]
61. Sun Y, Campisi J, Higano C, Beer TM, Porter P, Coleman I, et al. Treatment-induced damage to the tumor microenvironment promotes prostate cancer therapy resistance through WNT16B. *Nat Med*. 2012; 18:1359–68. [PubMed: 22863786]
62. Mosquera JM, Beltran H, Park K, MacDonald TY, Robinson BD, Tagawa ST, et al. Concurrent AURKA and MYCN gene amplifications are harbingers of lethal treatment-related neuroendocrine prostate cancer. *Neoplasia*. 2013; 15:1–10. [PubMed: 23358695]
63. Beltran H. The N-myc oncogene: maximizing its targets, regulation, and therapeutic potential. *Mol Cancer Res*. 2014; 12:815–22. [PubMed: 24589438]
64. Karantanos T, Evans CP, Tombal B, Thompson TC, Montironi R, Isaacs WB. Understanding the mechanisms of androgen deprivation resistance in prostate cancer at the molecular level. *Eur Urol*. 2015; 67:470–9. [PubMed: 25306226]
65. Beltran H, Tomlins S, Aparicio A, Arora V, Rickman D, Ayala G, et al. Aggressive variants of castration-resistant prostate cancer. *Clin Cancer Res*. 2014; 20:2846–50. [PubMed: 24727321]
66. Feng FY, de Bono JS, Rubin MA, Knudsen KE. Chromatin to clinic: the molecular rationale for PARP1 inhibitor function. *Mol Cell*. 2015; 58:925–34. [PubMed: 26091341]
67. Mateo J, Carreira S, Sandhu S, Miranda S, Mossop H, Perez-Lopez R, et al. DNA-repair defects and olaparib in metastatic prostate cancer. *N Engl J Med*. 2015; 373:1697–708. [PubMed: 26510020] [This study showed the first potential genetic signature predicting response to the PARP inhibitor, olaparib, in men with CRPC.]



**Fig. 1.** Signaling pathways implicated in resistance to novel androgen/AR-directed therapies. These resistance mechanisms are conceptualized in three broad categories: (1) reactivation of androgen/AR-signaling leading to persistent AR-signaling, (2) AR bypass pathways leading to activation of androgen-regulated genes by alternative steroid receptors, and (3) a large number of androgen/AR-independent pathways

**Table 1**

## Clinical trials attempting to address divergent mechanisms of androgen/AR resistance

Upregulation of androgen-synthetic enzymes				
Dutasteride	Phase II	Enzalutamide & Dutasteride as 1st Line Treatment for Patients 65 Years Old With Prostate Cancer		NCT02213107
Abiraterone Acetate	Phase II	A Phase II Study of Increased-Dose Abiraterone Acetate in Patients With Castration Resistant Prostate Cancer		NCT01637402
Androgen receptor amplification				
Apalutamide/ARN-509	Phase III	A Study of ARN-509 in Men With Non-Metastatic Castration-Resistant Prostate Cancer (SPARTAN)		NCT01946204
Androgen receptor point mutations				
ODM-201	Phase III	Efficacy and Safety Study of BAY1841788 (ODM-201) in Men With High-risk Non-metastatic Castration-resistant Prostate Cancer (ARAMIS)		NCT02200614
VT-464	Phase II	Once-daily Oral VT-464 in Patients With Castration-Resistant Prostate Cancer Progressing on Enzalutamide or Abiraterone		NCT02445976
Androgen receptor splice variants				
Testosterone	Phase II	RE-sensitizing With Supraphysiologic Testosterone to Overcome REsistant (The RESTORE Study)		NCT02090114
Galeterone	Phase III	A Study of Galeterone Compared to Enzalutamide In Men Expressing Androgen Receptor Splice Variant-7 mRNA (AR-V7) Metastatic CRPC (ARMOR3-SV)		NCT02438007
EPI-506	Phase I/II	Safety and Anti-Tumor Study of Oral EPI-506 for Patients With Metastatic Castration-Resistant Prostate Cancer		NCT02606123
GS-5829	Phase I/II	Safety, Tolerability, Pharmacokinetics, and Pharmacodynamics of GS-5829 as a Single Agent and In Combination With Enzalutamide in Participants With Metastatic Castrate-Resistant Prostate Cancer		NCT02607228
Niclosamide	Phase I	Niclosamide and Enzalutamide in Treating Patients With Androgen Receptor Splice Variant-Positive, Castration-Resistant, Metastatic Prostate Cancer		NCT02532114
Glucocorticoid receptor induction				
Dexamethasone	Phase II	Dexamethasone Prior to Re-treatment With Enzalutamide in Treating Patients With Metastatic Hormone-Resistant Prostate Cancer Previously Treated With Enzalutamide and Docetaxel (DEXTER)		NCT02491411
Mifepristone	Phase I/II	Enzalutamide and Mifepristone in Treating Patients With Metastatic Hormone Resistant Prostate Cancer		NCT02012296
Progesterone receptor activation				
Onapristone	Phase I/II	Phase 1-2 Study of Onapristone in Patients With Advanced Castration-resistant Prostate Cancer		NCT02049190
Androgen/AR-independent mechanisms				
Ipilimumab + Nivolumab	Phase II	Biomarker-Driven Therapy With Nivolumab and Ipilimumab in Treating Patients With Metastatic Hormone-Resistant Prostate Cancer Expressing AR-V7 (STARVE-PC)		NCT02601014
LY3023414	Phase II	A Study of Enzalutamide and LY3023414 in Men With Prostate Cancer		NCT02407054
Alisertib	Phase I/II	Alisertib, Abiraterone Acetate and Prednisone in Treating Patients With Hormone-Resistant Prostate Cancer		NCT01848067
Olaparib	Phase II	TOPARP: A Phase II Trial of Olaparib in Patients With Advanced Castration Resistant Prostate Cancer		NCT01682772
Niraparib	Phase I	Enzalutamide and Niraparib in the Treatment of Metastatic Castrate-Resistant Prostate Cancer		NCT02500901



**Table 2**

## Androgen/AR-independent mechanisms of resistance

Substrate	Mechanism of resistance in CRPC
Src-1	Stimulation of MAPK signaling
IL-6	Stimulation of MAPK signaling; Resistance to bicalutamide via upregulation of TIF2
HER2/HER3	Stabilization of AR and increased binding to AREs
PTEN loss	Activation of PI3K/Akt signaling pathway
Akt/PI3K	Inhibition of AR degradation via increased interaction of AR with p300
EZH2	Epigenetic silencing of tumor suppressor genes
STAT3	Resistance to enzalutamide; Promotion of PCa stemlike cells
c-Met	Enhanced PCa cell proliferation, motility, and invasion
RB1 deletion	Increased cell growth via disinhibition of cell-cycle progression
TP53 deletion or mutation	Dysregulation of cell division
MYCN gain	Dysregulation of some cell proliferation genes
AURKA gain	Dysregulation of cell-cycle progression
PARP1 overexpression	Disruption of proper DNA damage repair; Disruption of transcriptional regulation
IGF1 and FGF	Increased cell growth and proliferation/inhibition of apoptosis via Akt pathway