



Genome Sequence of the Autotrophic Acetogen *Clostridium magnum* DSM 2767

Ronny Uhlig,^a DAnja Poehlein,^b Ralf-Jörg Fischer, Rolf Daniel,^b Hubert Bahl^a

Institut für Biowissenschaften, Abteilung Mikrobiologie, Universität Rostock, Rostock, Germany^a; Genomic and Applied Microbiology & Göttingen Genomics Laboratory, Georg-August University Göttingen, Göttingen, Germany^b

R.U. and A.P. contributed equally to this work.

Here we report the draft genome sequence (6.6 Mbp) of the type strain *Clostridium magnum*, an acetogen with two operons coding for two separate Rnf complexes. *C. magnum* grows on a broad range of organic substrates and converts CO_2 and H_2 to acetate using the Wood-Ljungdahl pathway.

Received 18 April 2016 Accepted 21 April 2016 Published 9 June 2016

Citation Uhlig R, Poehlein A, Fischer R-J, Daniel R, Bahl H. 2016. Genome sequence of the autotrophic acetogen *Clostridium magnum* DSM 2767. Genome Announc 4(3):e00464-16. doi:10.1128/genomeA.00464-16.

Copyright © 2016 Uhlig et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Hubert Bahl, hubert.bahl@uni-rostock.de.

C*lostridium magnum* DSM 2767^T belongs to the group of Gram-positive bacteria with Gram-negative staining. It is an anaerobic, rod-shaped, spore-forming, motile bacterium, which was enriched from pasteurized freshwater sediment with 2,3 butanediol as the sole carbon and energy source by Schink in 1984 (1). In 1991, Bomar et al. (2) showed that *C. magnum* grows with H_2/CO_2 , formate, or methanol as the substrate when 0.025% (wt/ vol) yeast extract was provided in the growth medium. 2,3 Butanediol, acetoin, fructose, glucose, sucrose, xylose, malate, and citrate can serve as the substrates and are completely converted to acetate (1).

Genomic DNA of C. magnum DSM 2767 was isolated with the Tissue genomic DNA purification mini prep kit (Genaxxon, Ulm, Germany). Sequencing was done by a combined approach of single-molecule real-time (SMRT) and Illumina sequencing technologies according to the manufacturer's protocols. Genome sequencing was performed with a PacBio RSII (Pacific Biosciences, Menlo Park, CA) using P6 chemistry and an Illumina MiSeq using reagent kit v3 (2 \times 300 bp). The *de novo* assembly using the RS_ HGAP_Assembly.3" protocol included in SMRT Portal version 2.3.0 and 49,222 postfiltered reads (average read length of 12,759 bp) resulted in 21 contigs with an average coverage of 71.34-fold. The obtained Illumina reads (1,727,812) were quality filtered using Trimmomatic version 0.32 (3) and mapped onto the contigs with Bowtie2 version 2.0.6 (4) to improve the quality of the final sequence. The genome of C. magnum consists of a circular chromosome (6,600,766 bp) with an overall G+C content of 32.11%. Automatic gene prediction and identification of rRNA and tRNA genes was performed using the software tool Prokka (5). The genome contains 71 rRNA genes, 125 tRNA genes, 4,392 proteinencoding genes with predicted functions, and 1,773 genes coding for hypothetical proteins.

With a size of 6.6 Mbp, the genome of *C. magnum* is so far the largest sequenced genome of all acetogens, followed by *C. scatologenes* (5.75 Mbp, CP009933), *C. drakei* (5.64 Mbp, JIBU00000000) and *C. carboxidivorans* (5.63 Mbp). The genes encoding enzymes

of the Wood-Ljungdahl pathway revealed the same arrangement as that in all other acetogenic bacteria of the genus *Clostridium* (6). Interestingly, *C. magnum* is the first acetogen with a complete additional set of Rnf complex formation-related genes. Moreover, three more genes coding for RnfC-like proteins were found next to genes that are known to be involved in microcompartment formation as well as ethanol, ethanolamine, and 1,2 propanediol degradation (7). The genome analysis furthermore revealed genes encoding proteins for the fixation of molecular nitrogen, sporulation and formation of granulose.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at the DDBJ/EMBL/GenBank database under the accession no. LWAE00000000. The version described in this paper is version LWAE01000000.

ACKNOWLEDGMENT

We thank Kathleen Gollnow for technical support.

REFERENCES

- Schink B. 1984. Clostridium magnum sp. nov., a nonautotrophic homoacetogenic bacterium. Arch Microbiol 137:250–255. http://dx.doi.org/ 10.1007/BF00414553.
- Bomar M, Hippe H, Schink B. 1991. Lithotrophic growth and hydrogen metabolism by *Clostridium magnum*. FEMS Microbiol Lett 83:347–349.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. http://dx.doi.org/ 10.1093/bioinformatics/btu170.
- 4. Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with bowtie 2. Nat Methods 9:357–359. http://dx.doi.org/10.1038/nmeth.1923.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. http://dx.doi.org/10.1093/bioinformatics/btu153.
- Poehlein A, Cebulla M, Ilg MM, Bengelsdorf FR, Schiel-Bengelsdorf B, Whited G, Andreesen JR, Gottschalk G, Daniel R, Dürre P. 2015. The complete genome sequence of *Clostridium aceticum*: a missing link between Rnf- and cytochrome-containing autotrophic acetogens. mBio 6:e01168-15. http://dx.doi.org/10.1128/mBio.01168-15.
- Yeates TO, Crowley CS, Tanaka S. 2010. Bacterial microcompartment organelles: protein shell structure and evolution. Annu Rev Biophys 39: 185–205. http://dx.doi.org/10.1146/annurev.biophys.093008.131418.