

Immune complexes in myasthenia gravis

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SUMMARY Complement components were measured and immune complexes were sought in 75 patients with myasthenia gravis. Thirty-four per cent had decreased concentrations of complement component C4, and 29% had circulating immune complexes. The greatest immunological abnormalities were found in patients with mild disease which supports recent immunoelectronmicroscopic findings.

There is compelling evidence that myasthenia gravis is an autoimmune disease. An association with other autoimmune disorders is found (Simpson, 1960); germinal centres occur in the thymic medulla in 70% of cases (Castleman, 1966); disordered T cell function and its clinical manifestations has been demonstrated (Simpson *et al.*, 1976); and both cell-mediated and humoral immunity to the acetylcholine receptor (AChR) are present (Almon *et al.*, 1974; Abramsky *et al.*, 1975; Aharanov *et al.*, 1975; Lindstrom *et al.*, 1976; Richman *et al.*, 1976).

Cell-mediated hypersensitivity to the AChR cannot be demonstrated in all patients, particularly young female myasthenics (Richman *et al.*, 1976), so that the exact role of lymphocytes in myasthenia gravis is not known. However, serum from myasthenic patients is capable of reproducing the typical clinical and electromyographic features of the disease in mice (Toyka *et al.*, 1975) which suggests a central role for humoral antibody. Antibodies to cholinergic receptor structures are found in 75% or more of patients with myasthenia gravis, and some workers have found a positive correlation between these antibodies and the severity of the disease (Lefvert *et al.*, 1978). Others have not, and indeed babies of myasthenic mothers have been described who have been perfectly healthy although they had antireceptor antibodies (Lefvert *et al.*, 1978; Barkas *et al.*, 1979). Antibody alone, however, has been shown to increase the rate of degradation of junctional and extrajunctional acetylcholine receptors in muscle (Reiness *et al.*, 1978).

A role for complement in myasthenia gravis was demonstrated by Strauss *et al.* (1960) when they described humoral antibodies to skeletal muscle A bands and pointed out that these antibodies were complement-fixing. Fluctuations in serum complement concentrations were then found (Nastuk *et al.*, 1960) in the serum of 68 patients examined serially—decreased complement levels were shown in one or more serum samples from 40 patients. When the disease remitted, 50% of these patients demonstrated an increase in serum complement to the normal or supernormal range. Immune complexes of immunoglobulin G (IgG) and complement factor 3 (C3) have been identified at the motor endplate in myasthenics (Engel *et al.*, 1977).

In the experimental model of myasthenia gravis produced by passive transfer of serum from man to mouse, a pathogenetic role for the complement system has been shown (Toyka *et al.*, 1977). Complement depleted rats cannot be immunised to develop experimental allergic myasthenia gravis (EAMG) (Lennon *et al.*, 1978). Finally, in EAMG, C3 has been shown together with IgG at the motor endplate (Sahashi *et al.*, 1978).

We, therefore, examined the serum of 75 patients with myasthenia gravis to estimate concentrations of the complement components and to try to detect evidence of circulating immune complexes.

Patients and methods

Twenty-four male and 51 female patients with typical undisputed myasthenia gravis, whose ages ranged from 2 to 66 years, were studied. All patients had antiacetylcholine receptor antibody titres which were increased, and the majority had

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antistriated muscle antibodies. The disease varied from mild ptosis in some, to complete paralysis necessitating artificial respiration in others. Patients were classified according to the severity of their disease on a scale of 1 to 3, with 3 representing the most seriously affected patients.

An attempt was made to estimate the complement components serially. From 10 patients only one sample could be obtained but from the remaining 65 at least two, usually three, and in the case of 15 patients, nine samples were evaluated, taken at intervals of at least two months. The study lasted two years.

The functional efficiency of the total complement system was assayed in terms of the total haemolytic complement activity (CH50 units) using a standard technique (Kent and Fife, 1963). The following components were measured in EDTA plasma by the single radial immunodiffusion technique, using monospecific antisera: Clq, C3, C4, and Factor B. Commercially available plates (Behringwerke) were used to estimate C4 concentrations. Concentrations of C7 were measured by the reactive lysis method using activated C5/6 (Thompson and Lachmann, 1970). Conversion products of C3 were sought by crossed antibody electrophoresis (Laurell, 1965) and of Factor B, by routine immunoelectrophoresis. The antisera used were prepared in our laboratory by standard techniques.

Immune complexes were looked for by the anti-complementary method as standardised by Mayer (1961). Serum anticomplementary activity was measured by incubating 0.1 ml of heat-inactivated test serum (56°C for 60 minutes) with 2.5 units of guinea pig complement, at 37°C for 60 minutes. The residual anticomplementary activity was then titrated. In our laboratory immune complexes are normally considered present when less than 50% lysis occurs.

Results

Positive anticomplementary assays were obtained in 22 of the 75 patients. Serial testing of the patients revealed occasional reversal to give a negative result. The results are shown in Fig. 1, from which it can be seen that most of the positive assays were found in group 1, where 60% of the less severely affected patients had positive assays, compared with 15% of those with moderate myasthenia and 25% of those with incapacitating disease. Thus the positive assays tended to correlate with mild disease.

Estimation of the plasma complement components revealed conspicuous abnormalities in

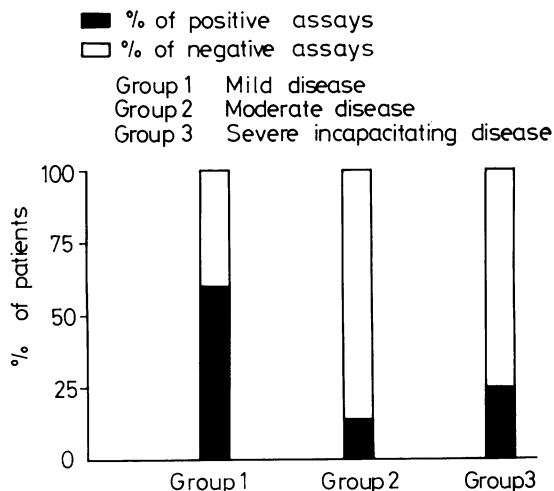


Fig. 1 Percentage of positive anticomplementary assays in 75 patients with myasthenia gravis.

component C4. Decreased concentrations of C4 were found in 26 of the 75 patients and increased concentrations in 18. Normal results were obtained in 31 patients. The normal plasma range in our laboratory is 31–55 mg/dl, and the low levels found were from 13.5 to 28 mg/dl while the high levels were from 56 to 72 mg/dl. In Fig. 2, the C4 concentrations are shown for each group, with the percentage of patients showing decreased, normal or increased C4 concentrations indicated. There was a conspicuous association between decreased C4 concentrations and mild or moderate

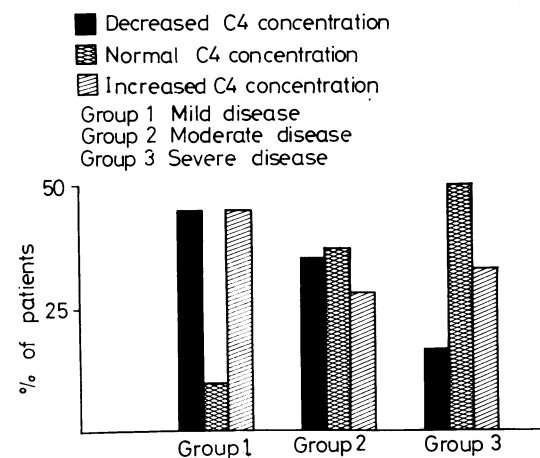


Fig. 2 Plasma C4 concentrations in 75 patients with myasthenia gravis.

disease—45% of patients in group 1 had low levels of C4 as did 35% of patients in group 2. In patients with severe, incapacitating myasthenia gravis the percentage fell to 17%.

Estimations of Clq, C3, and Factor B were normal, although the Clq and C3 concentrations tended to be in the low normal range. Estimations of C7 showed a good deal of variation. The CH50 unit estimations correlated well with the C4 results. Occasional C3 conversion products were detected in patients with decreased C4 concentrations but no conversion of Factor B was found.

In each group some patients showed increased C4 concentration above the normal range—45% in group 1, 28% in group 2, and 33% in group 3.

During the period of study, where serial measurements were made, the results were essentially similar on each occasion.

Discussion

There has long been circumstantial evidence suggesting complement involvement in the pathogenesis of myasthenia gravis. Early studies showed that sera from myasthenic patients were capable of lysing the surface cells of a preparation of frog sartorius muscle (Nastuk *et al.*, 1959). The active principle in these sera was heat-labile and destroyed on heating at 56°C for one hour. Involvement of complement was suspected, and indeed an inverse correlation between serum complement concentration and disease activity in myasthenic subjects was then reported in a preliminary study (Nastuk *et al.*, 1956). A large group of myasthenic patients was next investigated with serial estimations of complement concentrations. A close correlation was shown with decreases in complement paralleling disease activity. Associated virus or bacterial infections were ruled out (Nastuk *et al.*, 1960).

In 1973 it was shown that rabbits immunised with acetylcholine receptor (AChR) provided an experimental model for myasthenia gravis (Patrick and Lindstrom, 1973). Further studies revealed a factor in the serum of myasthenic subjects capable of binding to purified AChR (Almon *et al.*, 1974; Bender *et al.*, 1975). A role for complement was suggested by experiments in which myasthenic serum passively transferred to mice produced clinical signs in the recipients which were significantly less if the mice were depleted of C3 (Toyka *et al.*, 1977). In addition, EAMG could not be induced in rats depleted of complement by intraperitoneal inoculation of cobra venom factor (Lennon *et al.*, 1978). Thus, complement appeared to be essential for AChR antibodies to be pathogenic.

Recently there have been elegant demonstrations of immune complexes (IgG and C3) at the motor endplate in myasthenic patients. The deposits of IgG and C3 were localised to the segments of the postsynaptic membrane where acetylcholine is known to be distributed, and were also found on degenerating junctional folds in the synaptic space (Engel *et al.*, 1977). Immune complexes were much more sparse and difficult to find in the severe myasthenics. In EAMG, similar studies have been made of the motor endplates, and similar complexes of IgG and C3 have been identified on the terminal expansions of the junctional folds and on detached fragments of these folds in the synaptic space. Similarly, complexes were present in decreased amounts in the more severe cases of EAMG (Sahashi *et al.*, 1978). These findings were attributed to the fact that in severe myasthenia gravis or EAMG a smaller quantity of AChR remains at the endplates. The length of postsynaptic membrane on which immune complex deposits were found correlated with the miniature endplate potential (mepp) amplitude, as would be expected, since it had been shown previously that there was a linear relationship between the mepp amplitude and the amount of postsynaptic membrane reacting for AChR (Engel *et al.*, 1977). All these results suggest that the reduction in AChR is secondary to complement-mediated damage.

In the *in vitro* situation, using EAMG rat sera and innervated rat diaphragm in organ culture, AChR antibody has been shown to have two effects: to bind to AChR and, more importantly, to increase the rate of degradation of AChR (Heinemann *et al.*, 1978; Reiness *et al.*, 1978). The latter effect of antigen modulation appears to be complement independent in this model. *In vivo*, however, the involvement of complement has been clearly shown (Engel *et al.*, 1977).

Our finding of a decreased serum concentration of CH50 units in patients with myasthenia gravis agrees with that of previous workers (Strauss *et al.*, 1960). We used a more accurate and standardised microtechnique (Mayer, 1961) and found the best correlation with the plasma C4 concentrations. The Clq, C3, and Factor B estimations fell within the normal range; C7 concentrations showed wide variation. In the cases with low C4 concentrations, some conversion of C3 was frequently detectable. Conversion of Factor B was not found. Thus, the findings confirm that there is classical pathway utilisation of complement and that it is the early components of the cascade which are most involved (Toyka *et al.*, 1977).

In the original study (Strauss *et al.*, 1960) 60%

of patients showed decreased serum complement. We found that, overall, 34% of myasthenic patients had reduced C4 concentrations but, as can be seen in Fig. 2, the percentage of patients showing this reduction reached 45% in those least affected, was 35% in those with moderate disease, and fell to 17% in patients with severe myasthenia. It is of great interest that the depression of C4 occurred most frequently in the least affected patients, because this may be explained by previous observations on deposition of immune complexes at the muscle endplate (Engel *et al.*, 1977). We also confirm the finding of Strauss *et al.* (1960) of wide variations in serum complement. Increased C4 concentrations were detected in 24% of the patients. These results suggest that complement metabolism should be investigated in myasthenic patients.

The anticomplementary assays were positive in 30% of our cases, suggesting the presence of circulating immune complexes. There was a much higher percentage of positive results (60%) in the less severely affected myasthenics than in those with incapacitating myasthenia gravis (30%). Circulating immune complexes have been sought in small groups of patients with myasthenia gravis by other investigators. Casali *et al.* (1976) detected complexes in seven of eight patients, using the Clq-binding assay. In five of the eight patients there was an associated decrease in the serum C4 concentration. On the other hand, Tachovsky *et al.* (1976) found no serum antigen-antibody complexes in five patients with myasthenia gravis when they employed the Raji cell radioimmune assay. Thus in each of these studies a method different from the one used here, was utilised.

At present all tests for immune complexes in myasthenia gravis are "antigen-nonspecific". Results depend on differing size, concentrations, and biological properties of the complexes. The Clq-binding assay is highly sensitive and similar to the anticomplementary assay in that it depends on the complex interacting with complement by reacting with a receptor on the free molecule. Thus, our results and those of Casali *et al.* (1976) are comparable.

The Raji cell radioimmune assay, however, depends on interaction of the complex with complement receptors on the Raji cells, and detects IgG complexes only. IgM antigen-antibody complexes (which certainly may be present in these patients) will not be identified. It is likely that complexes of more than one type are present in patients with myasthenia gravis, as has been shown in, for instance, patients with dermatitis herpetiformis and coeliac disease (Mohammed *et al.*,

1976). Therefore, in future it would be wise to employ several different assays for their detection.

The identity of the antigen in the immune complexes demonstrated in myasthenia gravis is unknown but in view of the fact that AChR is lost from the postsynaptic membrane, and that antibody-coated segments of receptor are shed into the postsynaptic space in both myasthenia gravis and EAMG, these complexes may represent antibody-bound receptor. Their nature awaits elucidation.

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