Limited Evidence for Association of Genome-Wide Schizophrenia Risk Variants on Cortical Neuroimaging Phenotypes

Aristotle N. Voineskos^{*,1,2,5}, Daniel Felsky^{1,2,5}, Anne L. Wheeler^{1,2}, David J. Rotenberg¹, Melissa Levesque¹, Sejal Patel^{1,2}, Philip R. Szeszko^{3,4}, James L. Kennedy^{1,2}, Todd Lencz^{3,4}, and Anil K. Malhotra^{3,4}

¹Campbell Family Mental Health Institute, Centre for Addiction and Mental Health, Toronto, Ontario, Canada; ²Department of Psychiatry, University of Toronto, Toronto, Ontario, Canada; ³Zucker Hillside Hospital, Glen Oaks, NY; ⁴Center for Psychiatric Neuroscience, Feinstein Institute, Manhasset, NY

⁵These authors contributed equally to the article.

*To whom correspondence should be addressed; Kimel Family Translational Imaging-Genetics Laboratory, Research Imaging Centre, Campbell Family Mental Health Institute, Centre for Addiction and Mental Health (CAMH), 250 College Street, Toronto, Ontario M5R 1T8, Canada; tel: 416-535-8501 x33977, fax: 416-260-4162, e-mail: Aristotle.Voineskos@camh.ca

Background: There are now over 100 established genetic risk variants for schizophrenia; however, their influence on brain structure and circuitry across the human lifespan are not known. Methods: We examined healthy individuals 8-86 years of age, from the Centre for Addiction and Mental Health, the Zucker Hillside Hospital, and the Philadelphia Neurodevelopmental Cohort. Following thorough quality control procedures, we investigated associations of established genetic risk variants with heritable neuroimaging phenotypes relevant to schizophrenia, namely thickness of frontal and temporal cortical regions (n = 565) and frontotemporal and interhemispheric white matter tract fractional anisotropy (FA) (n = 530). Results: There was little evidence for association of risk variants with imaging phenotypes. No association with cortical thickness of any region was present. Only rs12148337, near a long noncoding RNA region, was associated with white matter FA (splenium of corpus callosum) following multiple comparison correction (corrected p = .012); this single nucleotide polymorphism was also associated with genu FA and superior longitudinal fasciculus FA at p < .005 (uncorrected). There was no association of polygenic risk score with white matter FA or cortical thickness. Conclusions: In sum, our findings provide limited evidence for association of schizophrenia risk variants with cortical thickness or diffusion imaging white matter phenotypes. When taken with recent lack of association of these variants with subcortical brain volumes, our results either suggest that structural neuroimaging approaches at current resolution are not sufficiently sensitive to detect effects of these risk variants or that multiple comparison correction in correlated phenotypes is too stringent, potentially "eliminating" biologically important signals.

Key words: imaging genetics/cortical thickness/DTI/polygenic risk/asymmetry

Introduction

More than 100 genetic risk loci were identified in the largest schizophrenia genome-wide association study (GWAS) to date.¹ An important next step is to identify the neural circuitry associated with these variants to help detect genetically mediated risk mechanisms of this devastating brain disorder. A recent GWAS in a large number of healthy individuals identified a small number of variants associated with subcortical brain volumes, including hippocampus and striatum; however, none of those loci were any of the 100+ schizophrenia risk loci.² While there is evidence for involvement of the hippocampus, striatum, and other subcortical structures in schizophrenia,^{3,4} there is perhaps even better evidence for alterations in cortical brain regions and related circuitry.^{5,6} Frontal and temporal cortical thickness reductions are a well-replicated finding in schizophrenia.^{6,7} In addition, diffusion anisotropy reductions in frontotemporal and interhemispheric white matter tracts are often found in people with schizophrenia compared to healthy controls,^{8,9} including in high-risk individuals.¹⁰ Therefore, a focus on cortical brain phenotypes may provide some insight into the neural mechanisms via which these genetic variants confer risk for schizophrenia.

Consistent with these proposed neural mechanisms, the first GWAS-single nucleotide polymorphism (SNP) imaging study relevant to schizophrenia demonstrated effects of a ZNF804A allele on functional frontotemporal and interhemispheric dysconnectivity phenotypes in healthy adult controls.¹¹ Others have also used the

[©] The Author 2015. Published by Oxford University Press on behalf of the Maryland Psychiatric Research Center.

All rights reserved. For permissions, please email: journals.permissions@oup.com

approach of studying healthy controls to help "validate" the potential effects of risk variants on structural and/ or functional neural circuitry, along with neurocognitive performance.^{12,13} However, not all studies demonstrate an association,¹⁴ and a recent systematic review has highlighted a number of inconsistencies in imaging genetics studies using white matter phenotypes,¹⁵ in part because phenotypes tested are often not the same.

An underutilized aspect of intermediate phenotype neuroimaging studies is the dimension of time. The use of neurodevelopmental samples, along with adult samples, can help to identify "when" in the lifespan genetic risk variants confer effects on brain structure or function, in addition to the more commonly assessed question of "where" in the brain genetic risk occurs.¹⁶ An effect observed early in life might be interpreted as more relevant for psychosis or schizophrenia disease risk while an effect seen during adulthood may be more important for disease course or progression.¹⁷

The primary objective of the present study was to assess the effects of genome-wide significant variants from the largest GWAS in schizophrenia to date¹ on intermediate phenotypes of cortical thickness and white matter fractional anisotropy (FA). The secondary objective was to assess the effects of polygenic risk score (PRS) on these same phenotypes. Three datasets (including the Philadelphia Neurodevelopmental Cohort [PNC]), that each assessed individuals at different stages of the lifespan, were used for examination of SNP-imaging phenotype associations, providing the largest known assessment of established genetic risk variants for schizophrenia on neuroimaging phenotypes of cortical thickness and white matter FA to date. Based on published individual GWAS SNP-neuroimaging phenotype findings at much smaller sample sizes, we hypothesized that a sizeable number of schizophrenia risk variants would be associated with cortical thickness and white matter FA phenotypes.

Methods

Participant Recruitment, Assessment, Imaging and Genetics

Centre for Addiction and Mental Health Sample. A total of 112 healthy volunteers were recruited at the Centre for Addiction and Mental Health (CAMH; Toronto, Ontario, Canada). All individuals (age 18–86) completed extensive clinical assessment protocols including the Structured Clinical Interview for DSM-IV Disorders (SCID-I) and the Mini-Mental State Examination,¹⁸ to rule out the presence of any psychiatric disorder or a dementia, as previously described.^{19,20} Genotyping for all subjects was performed using the Illumina HumanOmniExpress-12v1.0 BeadChips assay. T1-weighted and diffusion-weighted magnetic resonance imaging (MRI) acquisitions were as previously described²¹; see supplementary methods for details. The study was approved by the Research

Ethics Board of CAMH, and all participants provided informed, written consent. After excluding all non-Caucasian subjects (those not matched to the HapMap 3 reference populations CEU or TSI; http://www.broadinstitute.org/~debakker/p3.html), ethnic outliers, and highly related individuals, n = 96 subjects remained for cortical thickness analyses and n = 93 for diffusion tensor imaging (DTI) analyses.

Zucker Hillside Hospital Sample. A total of 107 healthy Caucasian subjects (age 8-68) were examined from an ongoing study at the Zucker Hillside Hospital (ZHH), Glen Oaks, NY, recruited by advertisement and word of mouth. Exclusion criteria included serious medical illness and any history of psychosis or major mood disorders, as determined by structured and semistructured assessments as previously published.²²⁻²⁴ Genotyping for all subjects was performed using the Illumina HumanOmniExpress-12v1.0 BeadChips assay. T1-weighted and diffusion-weighted MRI acquisitions were as previously described^{25,26}—see supplementary methods for details. This study was approved by the Institutional Review Board of the North Shore-Long Island Jewish Health System. After excluding all non-Caucasian subjects, ethnic outliers, and highly related individuals, n = 99 subjects remained for cortical thickness analyses and n = 99 for DTI analyses.

PNC Sample. A total of 802 subjects with qualitycontrolled T1-weighted MRI and 705 subjects with diffusion-weighted MRI scans were analyzed from the PNC cohort.²⁷ Genome-wide genotyping was performed on Affymetrix (6.0 Genechip and Axiom) and Illumina (Human610, HumanHap550 v1.0, and HumanHap550 v3.0) platforms. T1-weighted and diffusion-weighted MRI acquisitions were as previously described²⁸—see supplementary methods for details. The institutional review boards of both the University of Pennsylvania and the Children's Hospital of Philadelphia approved all study procedures. After excluding all non-Caucasian subjects, ethnic outliers, and highly related individuals, n = 370 subjects remained for cortical thickness analyses and n = 338 for DTI analyses (table 1).

Image Analysis

Cortical Thickness Processing. To calculate vertex-wise cortical thickness in the 3 samples, the T1-weighted images were submitted to the CIVET pipeline (version 1.1.10, Montreal Neurological Institute at McGill University, Montreal, Québec, Canada) to generate cortical models, as described previously.²⁶ Quality control criteria for each subject were generated by examining quantitative metrics associated with the quality of image registration and surface extraction. For each subject, vertex-wise cortical thickness maps were parcellated using cortical regions

defined in the LONI Probabilistic Brain Atlas²⁹ (LPBA40; available online at http://www.loni.usc.edu/atlases/), with additional manual segmentation of the middle frontal gyri.²⁶ Average cortical thickness was computed in a total of 52 distinct cortical regions; 24 frontal and temporal regions were then chosen from these for analysis (listed in supplementary table 1).

DTIProcessing and White Matter Parcellation. Correction for head motion and eddy currents by affine registration between each diffusion weighted image and b=0 reference volume was performed using FSL (v4.1.9)³⁰ was performed. Diffusion gradient orientations were also corrected for rotational motion.^{31,32} In the CAMH dataset, the three 1.5T diffusion repetitions were averaged, to increase signal to noise, prior to FA calculation.

Tract-based spatial statistics (TBSS) is a commonly used technique to investigate white matter differences between populations.^{33,34} The TBSS pipeline provides a framework to align FA images from multiple subjects to a template FA image in MNI152-space for subsequent voxelwise analysis. Nonlinear transformations are first calculated between each FA image to the standard FA template. Each subject's registered image was manually inspected for gross anatomical inconsistencies and/or any apparent warping. These transformations are then applied to each subject FA image and an average FA is generated from these registered FA maps. The mean FA image is utilized to generate a white matter skeleton onto which the maximum FA from each of the subjects is projected.

White matter parcellation analysis was then performed using regions of interest (ROI) defined in the ENIGMA protocol.³⁵ A set of FA maps were first registered to a common space defined by the MNI152 FA template. The mean FA values were calculated over each white matter region as defined by intersection of the Mori John Hopkins white matter atlas in MNI152-space and the rigid white matter skeleton resulting from the TBSS pipeline. A total of 20 ROIs were selected for analysis (listed in supplementary table 2).

Genetics Analysis

Quality control of genetic data was performed according a standard protocol as outlined by Anderson et al (2010).³⁶ Imputation was performed for all samples and platforms separately with PLINK (v1.90b)³⁷ and IMPUTE2 (v2.3.1)^{38,39} software packages, using the 1000 Genomes Phase I integrated haplotypes (b37) reference panel (see supplementary methods). Ethnicity was determined from the genetic data using multidimensional scaling, and degree of cryptic relatedness between subjects was calculated using estimates of identity-by-descent, both using PLINK (see supplementary methods).

Statistical Analysis

Power Analysis. Due to the uncommonly large sample size analyzed in the Ripke et al $(2014)^1$ paper, several SNPs found to be genome-wide significant for schizophrenia have low minor allele frequencies (MAFs; 9 SNPs have MAF < 0.1); this results in inadequate statistical power when analyzed in relatively smaller samples, especially when considering continuous traits and additive genetic models (for a SNP with MAF = 0.1, in sample of n = 1000, only 10 subjects would be expected to possess the rare homozygous genotype). Therefore, Quanto software (v1.2.4, http://biostats.usc.edu/Quanto.html) was used to determine the MAF threshold for inclusion in our study for cortical thickness and white matter microstructure analyses separately. For cortical thickness, in order to detect small effect sizes (Cohen's D = 0.2) in the combined sample (CAMH/ZHH/PNC) of n = 565, SNPs with MAF <0.22 were excluded from analysis. For white matter microstructure, in the combined sample of N = 530, the corresponding MAF threshold was 0.27.

Analyses. Cortical Thickness Statistical Primary analysis was conducted using R software $(v3.1.1)^{40}$ with Frank Harrel's "rms" statistical package.⁴¹ After excluding SNPs with MAF < 0.22, 60 of the 91 SNPs remained. Of these 60 SNPs, 2 (rs77502336 and rs6434928) showed slight deviations from Hardy-Weinberg equilibrium (HWE), with chi-squared p = .009 and p = .045, respectively, and were excluded from further analyses (final number of SNPs = 58). We conducted combined-sample analyses in 2 steps for each SNP/cortical ROI combination: first, between-sample variance in thickness was accounted for using ordinary least squares regression, by modeling average thickness as a product of study site. Second, to identify the effects of each SNP, residuals from this first model were entered into a second linear regression model that included additive genotype, age, sex, and handedness as predictors (supplementary methods). In total, 1392 SNP \times ROI combinations were tested, resulting in a Bonferroni multiple-correction significance threshold of $p = 3.59 \times 10^{-5}$ (2 sided). Corresponding models including age-by-SNP interaction terms were also tested in the combined sample. For models where the combined-sample genetic effect had a *p* value below this threshold, post hoc tests of dominant and recessive models were tested within each subsample separately. To test robustness of our findings, linear mixed-effects regression⁴² was also performed, with study site modeled as a random effect and genotype, sex, age, and handedness modeled as fixed effects, for all primary analyses. Results according to this method were not different (supplementary methods).

White Matter Microstructure The same 2-step modeling approach was used for analyzing the effect of each SNP on average FA for each white matter ROI. After excluding SNPs with MAF <0.27 and HWE p <.05, 43 remained, resulting in a total of 860 tests and a Bonferroni multiple-correction significance threshold of $p = 5.81 \times 10^{-5}$. Due to significant observed nonlinear effects of age on all average FA measures, as well as the known trajectory of FA change across the lifespan,^{43,44} age was modeled using a second degree polynomial. Again, corresponding models including age-by-SNP interaction terms were also tested in the combined sample.

Secondary Analyses. Polygene Risk Score In order to assess the additive effect of genome-wide significant SNPs from the Ripke et al (2014) paper,¹ we calculated a single PRS for each subject in all 3 study samples and evaluated the effect of this score on cortical thickness and white matter microstructure. The polygene score for each subject was calculated by multiplying the number of risk alleles possessed at each SNP by the natural logarithm of its respective combined-sample OR (for schizophrenia risk, from Ripke et al [2014]¹) and summing the products. Association of the PRS with each imaging phenotype was tested in the combined sample using the same 2-step regression technique as for the individual SNP analyses.

Hemispheric Laterality To explore potential genetic effects on hemispheric asymmetry (a potential neuroimaging phenotype for schizophrenia⁴⁵⁻⁴⁹) in our measures of cortical thickness and FA, we calculated a laterality index⁵⁰ for each bilateral ROI using the following formula:

$$laterality index = \frac{left \ value - right \ value}{(left \ value + right \ value)/2}$$

These indices were then used as outcomes in regression models as specified above.

Results

Primary Analyses

Associations of Genetic Risk Variants for Schizophrenia With Cortical Thickness. No SNP was significantly associated with cortical thickness of any region tested, after correction for 1392 tests (58 SNPs \times 24 cortical ROIs). Each tested cortical thickness ROI and top-associated SNP is presented in supplementary table 1. No age-by-SNP interaction was significant after correction.

Associations of Genetic Risk Variants for Schizophrenia With White Matter FA. In the combined sample of 530 subjects, the T allele of rs12148337 was significantly associated with an increase in FA of the splenium, after correction for 860 tests (additive t = 4.40, $p_{\text{unadjusted}} = 1.31 \times 10^{-5}$, $p_{\text{Bon-}}_{\text{ferroni}} = .011$). While the allelic direction of effect was consistent across subsamples, post hoc testing of the genotype effect within each subsample separately yielded significant effects at p < .05 in the CAMH (n = 93, additive t = 2.47, p = .015) and PNC (n = 338, additive t = 3.07, p = .0023) datasets only (figure 1). The direction of effect between homozygote groups in the ZHH sample was consistent with the other samples (ie, T/T subjects had greater thickness than C/C subjects), however, the heterozygote group showed the greatest cortical thickness, which is inconsistent with an additive model; the additive effect was not statistically significant (n = 99, t = 1.04, p = .30). The rs12148337 SNP also showed an association with a number of other white matter tracts (at uncorrected p < .005) including genu of the corpus callosum, and the left and right superior longitudinal fasciculus among others. Only one other SNP (rs10791097) was associated with more than one white matter tract at uncorrected p < .005. Each tested FA ROI and top-associated SNP is presented in supplementary table 2. No age-by-SNP interaction was significant after correction.

Secondary Analyses

PRS Analysis. The additive PRS, derived from 91 SNPs imputed with high quality, was normally distributed in both the cortical thickness (n = 565, Shapiro-Wilk p = .43) and FA (n = 530, Shapiro-Wilk p = .63) analysis. After correction for multiple testing, PRS was not associated with any neuroimaging phenotype in the combined sample (all p > .05), and no within-subsample associations survived multiple correction independently (all $p_{\text{Bonferroni}} > .05$) (see supplementary figure 1).

Hemispheric Laterality. For cortical thickness ROIs, there were no significant associations of laterality index found with any SNP in the combined sample. For tractwise FA, one SNP showed significant association with laterality index after correction for multiple testing: the A allele of rs11139497 was associated with increased laterality index (greater left vs right asymmetry) of uncinate fasciculus FA in the combined sample, controlling for handedness (n = 528, additive t = 4.0, $p_{unadjusted} = 6.9 \times 10^{-5}$, $p_{Bonferroni} = .047$). The directionality of this effect was consistent across study subsamples (CAMH: t = 3.01, p = .003; ZHH: t = 2.0, p = .049; PNC: t = 1.6, p = .11).

Discussion

We examined the association of recently established genome-wide significant SNPs for schizophrenia with frontotemporal and interhemispheric cortical thickness and white matter FA neuroimaging phenotypes in healthy individuals across the lifespan. No SNP was associated with cortical thickness following multiple comparison correction. The rs12148337 SNP was associated with FA of the splenium of the corpus callosum following multiple comparison correction, while the rs11139497 SNP was associated with the laterality index of FA of the uncinate fasciculus. PRS was also not associated with cortical thickness or white matter FA,



Fig. 1. Significant effect of rs12148337 on splenium fractional anisotropy in (A) combined sample of N = 530 ($p = 1.31 \times 10^{-5}$) and subsamples: (B) CAMH (p = .015), (C) ZHH (p = .30), and (D) PNC (p = .0023). Model residuals in panel (A) are derived from a different model than in (B), (C), and (D), as they are from regression predicting site-normalized residuals (residuals from final model were manually verified as being approximately normally distributed). The curves in (A) were drawn based on predicted values from the model for each genotypic group (Levene's test for equal variance among genotype groups p = .14). Residuals plotted in (B), (C), and (D) are from regression models predicting raw splenium FA, covarying for nonlinear effects of age, as well as sex. Center lines of boxplots (panels B, C, and D) are medians, and error bars extend $\pm 1.5 \times$ interquartile range; p values are 2 sided.

following correction for multiple comparisons. When taken together, our findings provide limited evidence for association of GWAS-significant SNPs for schizophrenia with neuroimaging phenotypes of cortical thickness and white matter FA.^{51,52}

We began with all 114 SNPs from the PGC2 study, not including insertion/deletion variants. After imputation across the 3 samples, 91 remained. Based on our power

calculations, we proceeded to assess 60 SNPs in relation to neuroimaging phenotypes. With primarily negative results, our findings stand in contrast to the predominantly positive findings in imaging genetics studies of GWAS-significant SNPs for schizophrenia and cortical neuroimaging phenotypes,^{12,53–56} including recent studies assessing PRS on cortical phenotypes, typically in much smaller samples.^{57–59} Our results are surprising, because a

Demographic	CAMH (<i>n</i> = 96)	ZHH (<i>n</i> = 99)	PNC (<i>n</i> = 370)	Diff. (<i>p</i>)*	Combined $(n = 565)$
Cortical thickness analysis					
Age, mean (v) (SD)	46 (18.8)	33 (16)	15 (3.6)	<.0001	23 (16.4)
Sex	50 M. 46 F	53 M. 47 F	178 M. 193 F	.58	281 M. 286 F
Handedness	89 R. 7 L	89R. 10 L	328 R. 42 L	.57	506 R. 59 L
IQ, mean score (SD) ^a	118 (8.5) ^b	106 (9.6) ^c	106 (13.2) ^d	_	
	CAMH (<i>n</i> = 93)	ZHH (<i>n</i> = 99)	PNC (<i>n</i> = 338)	Diff. (p)*	Combined $(n = 530)$
Fractional anisotropy anal	vsis				
Age, mean (v) (SD)	46 (18.7)	32 (16)	15 (3.5)	<.0001	23 (16.4)
Sex	48 M, 45 F	52 M, 47 F	164 M, 178 F	.66	264 M, 270 F
Handedness	87 R, 6 L	88R, 10L, 1 NA	303 R, 38 L, 1 NA	.44	478 R, 54 L, 2 NA
IQ, mean score (SD) ^a	118 (8.5) ^e	106 (9.6) ^f	107 (13.2) ^g		

 Table 1. Sample Demographics Summary

Note: CAMH, Centre for Addiction and Mental Health; IQ, Intelligence Quotient; NA, data not available; PNC, Philadelphia Neurodevelopmental Cohort; ZHH, Zucker Hillside Hospital. R, right handed; L, left handed; M, male; F, female.

^aIQ was evaluated using different tests in each sample subset; for CAMH sample, the Wechsler Test of Adult Reading (WTAR); for ZHH, the Wide-Range Achievement Test v3 (WRAT-3); and for PNC, the Wide-Range Achievement Test v4 (WRAT-4). Therefore, no cross-sample or combined-sample statistical comparison of IQ score was performed.

IQ data were only available for a subset of each sample: ${}^{b}n = 85$, ${}^{c}n = 88$, ${}^{d}n = 370$, ${}^{e}n = 84$, ${}^{f}n = 87$, ${}^{g}n = 337$.

**p* values for sample differences are 2 sided and were calculated using ANOVA for continuous data (age) and Fisher's Exact Test for factor data (sex, handedness).

number of aspects of our study design might have biased us toward positive association. First, it might be expected that our decision to focus on frontal and temporal cortical thickness, and frontotemporal and interhemispheric white matter FA would have biased the likelihood toward positive association based on the extant literature. Our selected neuroimaging phenotypes (eg, cortical thickness) are separable from other cortical phenotypes (surface area) in their genetic underpinnings.⁵² There is also a wealth of evidence for alterations in frontal and temporal brain gene expression⁶⁰⁻⁶² and brain structure^{3,63,64} and function^{65,66} in schizophrenia. Second, the collective sample size of our study was larger than most published imaging genetics studies, particularly those examining cortical thickness and white matter FA and those that have employed polygenic risk approaches.^{58,67,68} Third, the sampling of participants across the lifespan provided an opportunity to assess SNPneuroimaging associations with the dimension of time, which might have revealed effects of SNPs on neuroimaging phenotypes at a given phase of the lifespan.

Our results were not entirely negative, however. The association of the rs12148337 SNP with FA of the splenium of the corpus callosum that survived Bonferroni correction (for 900 comparisons) was consistent in direction across all 3 samples. The splenium of the corpus callosum provides interhemispheric temporal, parietal, and occipital connections. A meta-analysis showed consistent alterations in FA in the splenium of the corpus callosum in people with schizophrenia compared to healthy controls, supporting a role for this structure in schizophrenia.⁶⁹ The trajectory of splenium "maturation" as

indexed by FA across our 3 samples is consistent with the literature, where a peak is reached at approximately 25 years of age.⁴⁴ The effect of this SNP on splenium FA appears to be consistent across the lifespan, supporting an "early hit" model. As shown in supplementary table 2, the allelic direction of effect indicates that the rs12148337 T risk allele from Ripke et al (2014)¹ is associated with higher FA in our study samples. Association at an uncorrected p <.005 of this same SNP with FA of the genu of the corpus callosum, along with left and right superior longitudinal fasciculus, which are all tracts strongly implicated in schizophrenia,⁵ provides added confidence for our primary finding, given that association with the splenium alone might be due to registration error, or some other spurious effect.

The rs12148337 SNP is located on chromosome 15q23 approximately 25kb 5' of the long noncoding RNA gene RP11-543G18.1 (HaploReg v2⁷⁰). The SNP is also near genes MIR629, a microRNA gene and TLE3, transducinlike enhancer of split 3 which functions in the Notch signaling pathway that regulates determination of cell fate during development. Rs12148337 is in high linkage disequilibrium $(r^2 = .93, D' = 0.98;$ HaploReg v2⁷⁰) with SNP rs1971791 which has been associated with the neurodegenerative disorder, amyotrophic lateral sclerosis.⁷¹ Long noncoding RNAs play regulatory roles during transcription and translation, as well as in epigenetic regulation, and are known to play an important role in brain development, neuron function and maintenance, and neuropsychiatric disorders.⁷² Less is known about the rs11139497 SNP, which was associated with laterality of uncinate fasciculus FA, following multiple comparison correction. Although less typically a focus of neuroimaging investigations, good evidence for altered laterality of brain structure in schizophrenia exists.^{45,73} The rs11139497 SNP is located on chromosome 9, approximately 63kb 5' of the *FAM75B* (also *SPATA31B1P* or *C9orf36B*) gene within an uncharacterized locus (LOC105376107, ncRNA). The NCBI Gene database lists 2 Gene Ontology biological process terms for this pseudogene: cell differentiation and spermatogenesis, although there are no supporting publications (evidence inferred from electronic annotation).

Some potential limitations deserve consideration. It is possible that some of the SNPs examined might be associated with parietal or occipital brain structure, given reports of alterations in these brain regions⁷⁴⁻⁷⁶ or of more anatomically widespread alterations in brain structure⁷⁷ in schizophrenia. We did not study patients with schizophrenia: effects of genetic variants in patient populations may be different than those of controls. A previous study¹⁷ by our group found no effect of the MIR137 GWASsignificant SNP on these same neuroimaging phenotypes in healthy individuals but found a prominent effect in patients with schizophrenia or schizoaffective disorder, in addition to effects on age at onset. Different scanners were used at each site. Despite rigorous statistical modeling that took potential study site effects into account, it remains possible that there was an effect of site or an effect of the scanner at that site which may have influenced our results. Ours is not the first study, however, to consider imaging genetics data from multiple sites.⁷⁸ The main advantage of a multisite analysis, which is also true of the present study, is the increase in sample size and accompanying power. Some have argued that with the appropriate methodological approaches, sample sizes much smaller than those used in the present study should be able to detect effects of genetic variation on brain structure or function.79,80 In contrast, others have argued that much larger sample sizes are needed. However, 2 studies that included >10000 participants^{2,81} revealed no significant association of any schizophrenia GWAS-significant variant with subcortical brain volumes, which suggests that an especially large sample size may not be the answer. The SNPs that we were underpowered to examine might have effect on our neuroimaging phenotypes of interest, especially since more rare variants may have disproportionately large effect sizes.

When taken together, our findings provide limited evidence for effects of genome-wide significant risk variants for schizophrenia or for PRS on well-established frontotemporal and interhemispheric neuroimaging phenotypes of cortical thickness and white matter FA. Even larger sample sizes may not be the answer for discovery of neural mechanisms of established schizophrenia risk variants, although such studies would need to be conducted to be certain. Since schizophrenia is a brain disorder, it may be that structural neuroimaging phenotypes at current resolution⁸² are not sufficiently sensitive to reliably detect associations with these risk variants in healthy individuals. Notably, most PGC-derived susceptibility SNPs are thought to be regulatory in nature. It may be that variants which play a direct role in protein conformation (eg, BDNF Val66Met^{21,83,84} and APOE ε4^{20,85,86} important in neuropsychiatric disorders) may have greater impact on brain, which is detectable using current structural MRI approaches. Alternatively, the statistical threshold for detection in studies with many comparisons may be too stringent, "drowning out" important biological clues and mechanisms, particularly given that thickness of brain regions tend to covary, as does FA of different white matter tracts,⁸ and as such may not be truly "independent." Relaxing Bonferroni thresholds for imaging genetics studies, possibly in combination with plausible in vitro or complementary analyses, may provide a clearer window into novel genetically mediated risk mechanisms for schizophrenia and therapeutic opportunities for this debilitating disorder.

Supplementary Material

Supplementary material is available at http://schizophreniabulletin.oxfordjournals.org.

Funding

Canadian Institutes of Health Research Vanier Canada Graduate Scholarship and the Peterborough K.M. Hunter Graduate Scholarship to D.F.; the Canadian Institutes of Health Research, the CAMH Foundation, Ontario Ministry of Research and Innovation, Brain and Behavior Research Foundation, and Canada Foundation for Innovation to A.N.V.; and the National Institute of Mental Health (grant numbers R01MH099167 and R01MH102324 to A.N.V., R01MH079800 and P50MH080173 to A.K.M.). No sponsor or funder played any role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; and preparation, review, or approval of the manuscript.

Acknowledgments

A.K.M. is a consultant to Forum Pharmaceuticals and Genomind Inc. J.L.K. has received honoraria from Roche, Novartis, and Eli Lilly Co. He also serves in an unpaid role on the advisory board to Assure Rx. A.N.V., D.F., A.L.W., D.J.R., M.L., S.P., P.R.S., and T.L. reported no biomedical financial interests or potential conflicts of interest.

References

- Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophreniaassociated genetic loci. *Nature*. 2014;511:421–427.
- Hibar DP, Stein JL, Renteria ME, et al.; Alzheimer's Disease Neuroimaging Initiative; CHARGE Consortium; EPIGEN; IMAGEN; SYS. Common genetic variants influence human subcortical brain structures. *Nature*. 2015;520:224–229.

- Shenton ME, Dickey CC, Frumin M, McCarley RW. A review of MRI findings in schizophrenia. *Schizophr Res.* 2001;49:1–52.
- 4. Adriano F, Caltagirone C, Spalletta G. Hippocampal volume reduction in first-episode and chronic schizophrenia: a review and meta-analysis. *Neuroscientist*. 2012;18:180–200.
- Wheeler AL, Voineskos AN. A review of structural neuroimaging in schizophrenia: from connectivity to connectomics. *Front Hum Neurosci.* 2014;8:653.
- Voineskos AN, Foussias G, Lerch J, et al. Neuroimaging evidence for the deficit subtype of schizophrenia. JAMA Psychiatry. 2013;70:472–480.
- Rimol LM, Nesvåg R, Hagler DJ Jr, et al. Cortical volume, surface area, and thickness in schizophrenia and bipolar disorder. *Biol Psychiatry*. 2012;71:552–560.
- Voineskos AN, Lobaugh NJ, Bouix S, et al. Diffusion tensor tractography findings in schizophrenia across the adult lifespan. *Brain*. 2010;133:1494–1504.
- Kubicki M, Styner M, Bouix S, et al. Reduced interhemispheric connectivity in schizophrenia-tractography based segmentation of the corpus callosum. *Schizophr Res.* 2008;106:125–131.
- von Hohenberg CC, Pasternak O, Kubicki M, et al. White matter microstructure in individuals at clinical high risk of psychosis: a whole-brain diffusion tensor imaging study. *Schizophr Bull*. 2014;40:895–903.
- 11. Esslinger C, Walter H, Kirsch P, et al. Neural mechanisms of a genome-wide supported psychosis variant. *Science*. 2009;324:605.
- Voineskos AN, Lerch JP, Felsky D, et al. The ZNF804A gene: characterization of a novel neural risk mechanism for the major psychoses. *Neuropsychopharmacology*. 2011;36:1871–1878.
- Bigos KL, Mattay VS, Callicott JH, et al. Genetic variation in CACNA1C affects brain circuitries related to mental illness. *Arch Gen Psychiatry*. 2010;67:939–945.
- Cousijn H, Eissing M, Fernández G, et al. No effect of schizophrenia risk genes MIR137, TCF4, and ZNF804A on macroscopic brain structure. *Schizophr Res.* 2014;159:329–332.
- 15. Gurung R, Prata DP. What is the impact of genome-wide supported risk variants for schizophrenia and bipolar disorder on brain structure and function? A systematic review. *Psychol Med.* 2015;45:2461–2480.
- Felsky D, Szeszko P, Yu L, et al. The SORL1 gene and convergent neural risk for Alzheimer's disease across the human lifespan. *Mol Psychiatry*. 2014;19:1125–1132.
- Lett TA, Chakravarty MM, Chakavarty MM, et al. The genome-wide supported microRNA-137 variant predicts phenotypic heterogeneity within schizophrenia. *Mol Psychiatry*. 2013;18:443–450.
- Folstein MF, Folstein SE, McHugh PR. "Mini-mental state". A practical method for grading the cognitive state of patients for the clinician. J Psychiatr Res. 1975;12:189–198.
- Voineskos AN, O'Donnell LJ, Lobaugh NJ, et al. Quantitative examination of a novel clustering method using magnetic resonance diffusion tensor tractography. *Neuroimage*. 2009;45:370–376.
- 20. Felsky D, Voineskos AN. APOE ϵ 4, aging, and effects on white matter across the adult life span. *JAMA Psychiatry*. 2013;70:646–647.
- Voineskos AN, Lerch JP, Felsky D, et al. The brain-derived neurotrophic factor Val66Met polymorphism and prediction of neural risk for Alzheimer disease. *Arch Gen Psychiatry*. 2011;68:198–206.

- 22. Peters BD, Szeszko PR, Radua J, et al. White matter development in adolescence: diffusion tensor imaging and metaanalytic results. *Schizophr Bull.* 2012;38:1308–1317.
- 23. Kaufman J, Birmaher B, Brent D, et al. Schedule for Affective Disorders and Schizophrenia for School-Age Children-Present and Lifetime Version (K-SADS-PL): initial reliability and validity data. J Am Acad Child Adolesc Psychiatry. 1997;36:980–988.
- First MB, Spitzer RL, Gibbon M, Williams JBW. Structured Clinical Interview for DSM-IV-TR Axis I Disorders, Research Version, Non-Patient Edition. (SCID-I/NP). 2002.
- 25. Toteja N, Guvenek-Cokol P, Ikuta T, et al. Age-associated alterations in corpus callosum white matter integrity in bipolar disorder assessed using probabilistic tractography. *Bipolar Disord*. 2015;17:381–391.
- Wheeler AL, Wessa M, Szeszko PR, et al. Further neuroimaging evidence for the deficit subtype of schizophrenia: a cortical connectomics analysis. *JAMA Psychiatry*. 2015;72:446–455.
- 27. Satterthwaite TD, Connolly JJ, Ruparel K, et al. The Philadelphia Neurodevelopmental Cohort: a publicly available resource for the study of normal and abnormal brain development in youth. *Neuroimage*. 2016;124:1115–1119.
- Satterthwaite TD, Elliott MA, Ruparel K, et al. Neuroimaging of the Philadelphia Neurodevelopmental Cohort. *Neuroimage*. 2014;86:544–553.
- Shattuck DW, Mirza M, Adisetiyo V, et al. Construction of a 3D probabilistic atlas of human cortical structures. *Neuroimage*. 2008;39:1064–1080.
- Jenkinson M, Beckmann CF, Behrens TE, Woolrich MW, Smith SM. FSL. *Neuroimage*. 2012;62:782–790.
- Jeurissen B, Leemans A, Sijbers J. Automated correction of improperly rotated diffusion gradient orientations in diffusion weighted MRI. *Med Image Anal.* 2014;18:953–962.
- Leemans A, Jones DK. The B-matrix must be rotated when correcting for subject motion in DTI data. *Magn Reson Med.* 2009;61:1336–1349.
- Smith SM, Jenkinson M, Johansen-Berg H, et al. Tract-based spatial statistics: voxelwise analysis of multi-subject diffusion data. *Neuroimage*. 2006;31:1487–1505.
- Smith SM, Jenkinson M, Woolrich MW, et al. Advances in functional and structural MR image analysis and implementation as FSL. *Neuroimage*. 2004;23(suppl 1):S208–S219.
- 35. Jahanshad N, Kochunov PV, Sprooten E, et al. Multi-site genetic analysis of diffusion images and voxelwise heritability analysis: a pilot project of the ENIGMA-DTI working group. *Neuroimage*. 2013;81:455–469.
- Anderson CA, Pettersson FH, Clarke GM, Cardon LR, Morris AP, Zondervan KT. Data quality control in genetic case-control association studies. *Nat Protoc*. 2010;5:1564–1573.
- Purcell S, Neale B, Todd-Brown K, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet*. 2007;81:559–575.
- Howie B, Fuchsberger C, Stephens M, Marchini J, Abecasis GR. Fast and accurate genotype imputation in genomewide association studies through pre-phasing. *Nat Genet*. 2012;44:955–959.
- Marchini J, Howie B, Myers S, McVean G, Donnelly P. A new multipoint method for genome-wide association studies by imputation of genotypes. *Nat Genet*. 2007;39:906–913.
- 40. R Core Team. R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation For Statistical Computing; 2014.

- 41. Harrel F Jr. Regression modeling strategies, "rms" package for R Statistical Program (v4.2-1). 2014.
- 42. Bates D, Machler M, Bolker B, et al. Linear mixed-effects models using eigen and S4, "lme4" package for R Statistical Program (v1.1-7). 2014.
- Westlye LT, Walhovd KB, Dale AM, et al. Life-span changes of the human brain white matter: diffusion tensor imaging (DTI) and volumetry. *Cereb Cortex*. 2010;20:2055–2068.
- Lebel C, Gee M, Camicioli R, Wieler M, Martin W, Beaulieu C. Diffusion tensor imaging of white matter tract evolution over the lifespan. *Neuroimage*. 2012;60:340–352.
- 45. Ribolsi M, Daskalakis ZJ, Siracusano A, Koch G. Abnormal asymmetry of brain connectivity in schizophrenia. *Front Hum Neurosci.* 2014;8:1010.
- Niznikiewicz M, Donnino R, McCarley RW, et al. Abnormal angular gyrus asymmetry in schizophrenia. *Am J Psychiatry*. 2000;157:428–437.
- Oertel-Knöchel V, Linden DE. Cerebral asymmetry in schizophrenia. *Neuroscientist*. 2011;17:456–467.
- Sommer IE, Ramsey NF, Kahn RS. Language lateralization in schizophrenia, an fMRI study. *Schizophr Res.* 2001;52:57–67.
- Gur RE, Chin S. Laterality in functional brain imaging studies of schizophrenia. *Schizophr Bull*. 1999;25:141–156.
- Catani M, Allin MP, Husain M, et al. Symmetries in human brain language pathways correlate with verbal recall. *Proc Natl Acad Sci USA*. 2007;104:17163–17168.
- 51. Kochunov P, Jahanshad N, Marcus D, et al. Heritability of fractional anisotropy in human white matter: a comparison of Human Connectome Project and ENIGMA-DTI data. *Neuroimage*. 2015;111:300–311.
- 52. Eyler LT, Chen CH, Panizzon MS, et al. A comparison of heritability maps of cortical surface area and thickness and the influence of adjustment for whole brain measures: a magnetic resonance imaging twin study. *Twin Res Hum Genet*. 2012;15:304–314.
- Wassink TH, Epping EA, Rudd D, et al. Influence of ZNF804a on brain structure volumes and symptom severity in individuals with schizophrenia. *Arch Gen Psychiatry*. 2012;69:885–892.
- Schultz CC, Nenadic I, Riley B, et al. ZNF804A and cortical structure in schizophrenia: in vivo and postmortem studies. *Schizophr Bull*. 2014;40:532–541.
- 55. Rose EJ, Hargreaves A, Morris D, et al. Effects of a novel schizophrenia risk variant rs7914558 at CNNM2 on brain structure and attributional style. *Br J Psychiatry*. 2014;204:115–121.
- Thong JY, Qiu A, Sum MY, et al. Effects of the neurogranin variant rs12807809 on thalamocortical morphology in schizophrenia. *PLoS One*. 2013;8:e85603.
- Papiol S, Mitjans M, Assogna F, et al. Polygenic determinants of white matter volume derived from GWAS lack reproducibility in a replicate sample. *Transl Psychiatry*. 2014;4:e362.
- Walton E, Turner J, Gollub RL, et al. Cumulative genetic risk and prefrontal activity in patients with schizophrenia. *Schizophr Bull*. 2013;39:703–711.
- Terwisscha van Scheltinga AF, Bakker SC, van Haren NE, et al.; Psychiatric Genome-wide Association Study Consortium. Genetic schizophrenia risk variants jointly modulate total brain and white matter volume. *Biol Psychiatry*. 2013;73:525–531.
- Harrison PJ, Weinberger DR. Schizophrenia genes, gene expression, and neuropathology: on the matter of their convergence. *Mol Psychiatry*. 2005;10:40–68; image 5.

- Mirnics K, Middleton FA, Marquez A, Lewis DA, Levitt P. Molecular characterization of schizophrenia viewed by microarray analysis of gene expression in prefrontal cortex. *Neuron*. 2000;28:53–67.
- 62. Guillozet-Bongaarts AL, Hyde TM, Dalley RA, et al. Altered gene expression in the dorsolateral prefrontal cortex of individuals with schizophrenia. *Mol Psychiatry*. 2014;19:478–485.
- 63. Hulshoff Pol HE, Kahn RS. What happens after the first episode? A review of progressive brain changes in chronically ill patients with schizophrenia. *Schizophr Bull*. 2008;34:354–366.
- 64. Kasai K, Shenton ME, Salisbury DF, et al. Progressive decrease of left superior temporal gyrus gray matter volume in patients with first-episode schizophrenia. *Am J Psychiatry*. 2003;160:156–164.
- 65. Spoletini I, Cherubini A, Di Paola M, et al. Reduced frontotemporal connectivity is associated with frontal gray matter density reduction and neuropsychological deficit in schizophrenia. *Schizophr Res.* 2009;108:57–68.
- 66. Gur RE, Gur RC. Functional magnetic resonance imaging in schizophrenia. *Dialogues Clin Neurosci.* 2010;12:333–343.
- 67. Whalley HC, Sprooten E, Hackett S, et al. Polygenic risk and white matter integrity in individuals at high risk of mood disorder. *Biol Psychiatry*. 2013;74:280–286.
- Oertel-Knöchel V, Lancaster TM, Knöchel C, et al. Schizophrenia risk variants modulate white matter volume across the psychosis spectrum: evidence from two independent cohorts. *Neuroimage Clin.* 2015;7:764–770.
- Patel S, Mahon K, Wellington R, Zhang J, Chaplin W, Szeszko PR. A meta-analysis of diffusion tensor imaging studies of the corpus callosum in schizophrenia. *Schizophr Res.* 2011;129:149–155.
- Ward LD, Kellis M. HaploReg: a resource for exploring chromatin states, conservation, and regulatory motif alterations within sets of genetically linked variants. *Nucleic Acids Res.* 2011:40:D930–D934.
- Ahmeti KB, Ajroud-Driss S, Al-Chalabi A, et al. Age of onset of amyotrophic lateral sclerosis is modulated by a locus on 1p34.1. *Neurobiol Aging*. 2013;34:357.e7–357.e19.
- Wu P, Zuo X, Deng H, Liu X, Liu L, Ji A. Roles of long noncoding RNAs in brain development, functional diversification and neurodegenerative diseases. *Brain Res Bull.* 2013;97:69–80.
- Honea R, Crow TJ, Passingham D, Mackay CE. Regional deficits in brain volume in schizophrenia: a meta-analysis of voxel-based morphometry studies. *Am J Psychiatry*. 2005;162:2233–2245.
- Kuperberg GR, Broome MR, McGuire PK, et al. Regionally localized thinning of the cerebral cortex in schizophrenia. *Arch Gen Psychiatry*. 2003;60:878–888.
- Narr KL, Toga AW, Szeszko P, et al. Cortical thinning in cingulate and occipital cortices in first episode schizophrenia. *Biol Psychiatry*. 2005;58:32–40.
- Cheung V, Chiu CP, Law CW, et al. Positive symptoms and white matter microstructure in never-medicated first episode schizophrenia. *Psychol Med.* 2011;41:1709–1719.
- Anderson VM, Goldstein ME, Kydd RR, Russell BR. Extensive gray matter volume reduction in treatmentresistant schizophrenia. *Int J Neuropsychopharmacol.* 2015;18:pyv016.
- 78. Thompson PM, Stein JL, Medland SE, et al.; Alzheimer's Disease Neuroimaging Initiative, EPIGEN Consortium, IMAGEN Consortium, Saguenay Youth Study (SYS) Group. The ENIGMA Consortium: large-scale collaborative

analyses of neuroimaging and genetic data. *Brain Imaging Behav.* 2014;8:153–182.

- Meyer-Lindenberg A, Nicodemus KK, Egan MF, Callicott JH, Mattay V, Weinberger DR. False positives in imaging genetics. *Neuroimage*. 2008;40:655–661.
- Silver M, Montana G, Nichols TE; Alzheimer's Disease Neuroimaging Initiative. False positives in neuroimaging genetics using voxel-based morphometry data. *Neuroimage*. 2011;54:992–1000.
- 81. Stein JL, Medland SE, Vasquez AA, et al.; Alzheimer's Disease Neuroimaging Initiative; EPIGEN Consortium; IMAGEN Consortium; Saguenay Youth Study Group; Cohorts for Heart and Aging Research in Genomic Epidemiology Consortium; Enhancing Neuro Imaging Genetics through Meta-Analysis Consortium. Identification of common variants associated with human hippocampal and intracranial volumes. *Nat Genet*. 2012;44:552–561.
- Weinberger DR, Radulescu E. Finding the elusive psychiatric "lesion" with 21st-century neuroanatomy: a note of caution [published online ahead of print August 28, 2015]. *Am J Psychiatry*. doi: appiajp201515060753.
- Toro R, Chupin M, Garnero L, et al. Brain volumes and Val66Met polymorphism of the BDNF gene: local or global effects? *Brain Struct Funct*. 2009;213:501–509.
- Yang X, Liu P, Sun J, et al. Impact of brain-derived neurotrophic factor Val66Met polymorphism on cortical thickness and voxel-based morphometry in healthy Chinese young adults. *PLoS One*. 2012;7:e37777.
- Madhyastha T, Borghesani P, Aylward E, et al. Effect of the APOE-ε4 allele on longitudinal changes in cortical thickness in normal aging. *Alzheimers Dement J Alzheimers Assoc.* 2012;8:P4.
- Nyberg L, Salami A. The APOE ε4 allele in relation to brain white-matter microstructure in adulthood and aging. *Scand J Psychol.* 2014;55:263–267.