

Lost & found: C9ORF72 and the autophagy pathway in ALS/FTD

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C9ORF72 expression is reduced in a substantial number of patients with amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD), which may contribute to disease pathogenesis. However, its normal molecular function remains unknown. In this issue of *The EMBO Journal*, Sellier *et al* (2016) identified a novel protein complex consisting of C9ORF72, WDR41, and SMCR8 that acts as a GDP-GTP exchange factor (GEF) for RAB8a and RAB39b and is regulated by TBK1, whose partial loss of function also causes ALS and FTD. They further reveal a potential modulatory role for this novel complex in macroautophagy (autophagy), especially in the context of ataxin-2 toxicity.

See also: C Sellier *et al* (June 2016)

When GGGGCC (G₄C₂) repeat expansions in the first intron of *C9ORF72* were identified as the most common genetic cause of ALS and FTD (DeJesus-Hernandez *et al*, 2011; Renton *et al*, 2011), nothing was known about the molecular functions of *C9ORF72*. Since then, breathtaking progress has been made in understanding the effects of this mutation, largely focusing on the toxic products derived from expanded G₄C₂ repeats (Gitler & Tsuiji, 2016). For instance, the expanded repeats in nuclear RNA foci might misregulate RNA processing by sequestering some RNA-binding proteins. Moreover, abnormal translation of the sense and antisense repeat RNAs through a repeat-associated non-AUG (RAN) translation mechanism gives rise to toxic dipeptide repeat (DPR) proteins. However, expression of *C9ORF72* mRNA is also reduced in ALS/FTD patients with

C9ORF72 repeat expansions—raising the possibility that compromised *C9ORF72* function contributes to disease pathogenesis as well (DeJesus-Hernandez *et al*, 2011).

To understand the normal function of *C9ORF72*, Sellier *et al* (2016) set out to identify proteins that interact with it directly. Immunoprecipitation from N2A mouse cells followed by tandem mass spectrometry analysis revealed a novel protein complex containing Smcr8 (a DENN domain-containing protein predicted to function as a GEF), Wdr41 (a WD40 repeat-containing protein of unknown function), and Rab8a and Rab39b (two small GTPases involved in membrane trafficking), and transfected *C9ORF72* (Fig 1). Additional biochemical analysis confirmed the existence of such a protein complex endogenously in mouse brain. Interestingly, RAB8 directly interacts with optineurin and TBK1, mutations in which are implicated in ALS/FTD (Bettencourt & Houlden, 2015). Moreover, in a *Drosophila* model of FTD linked to chromosome 3, Rab8 is a strong modifier of mutant CHMP2B toxicity (West *et al*, 2015). Thus, RAB8 may be a signaling hub for several disease proteins in ALS/FTD.

What molecular functions might *C9ORF72* fulfill in this novel protein complex? Sellier *et al* (2016) found that the complex composed of *C9ORF72*, SMCR8, and WDR41 serves as a GEF for RAB8a and RAB39b but not for some other RAB proteins, suggesting a certain degree of specificity. However, *C9ORF72* alone does not have any GEF activity even though it has a DENN domain. This complex also interacts with autophagy adaptors p62 and optineurin; however, this interaction seems to be weak and its functional significance needs to be further confirmed.

The authors went on to examine the cellular functions of the *C9ORF72*-containing complex. Knockdown of *C9orf72* partially blocked the formation of autophagosomes after activation by torin or rapamycin in cultured primary mouse cortical neurons, consistent with a suggested role for *C9ORF72* in autophagy (Farg *et al*, 2014). This function required RAB39b, since a constitutively active form of RAB39b, but not RAB8a or other RAB proteins, rescued autophagy defects caused by *C9orf72* depletion. Accordingly, knockdown of *C9orf72* or its binding partners such as Smcr8 or Wdr41 resulted in the accumulation of p62 in rodent neurons. This accumulation was rescued by the expression of the long isoform of *C9ORF72*, demonstrating that p62 accumulation is indeed a direct consequence of *C9orf72* knockdown. Interestingly, the short isoform of *C9ORF72* failed to do so, suggesting a distinct function. Moreover, cytoplasmic aggregates of TDP-43 were observed in some cell types with *C9orf72* depletion.

These results in cultured neurons raise several intriguing issues that need to be addressed. First, it seems that *C9ORF72* is not essential for autophagy induction but may play a modulatory role in some cell types. Exactly how *C9ORF72* and RAB39b do so at the molecular level is unclear. Second, the functional significance of the *C9ORF72*–RAB8a interaction remains to be further investigated. Third, elevated p62 and a potential autophagy defect were observed in human cortical neurons differentiated from induced pluripotent stem cells of FTD patients with *C9ORF72* repeat expansions (Almeida *et al*, 2013). It will be informative to examine how much of this defect in human neurons results from partial

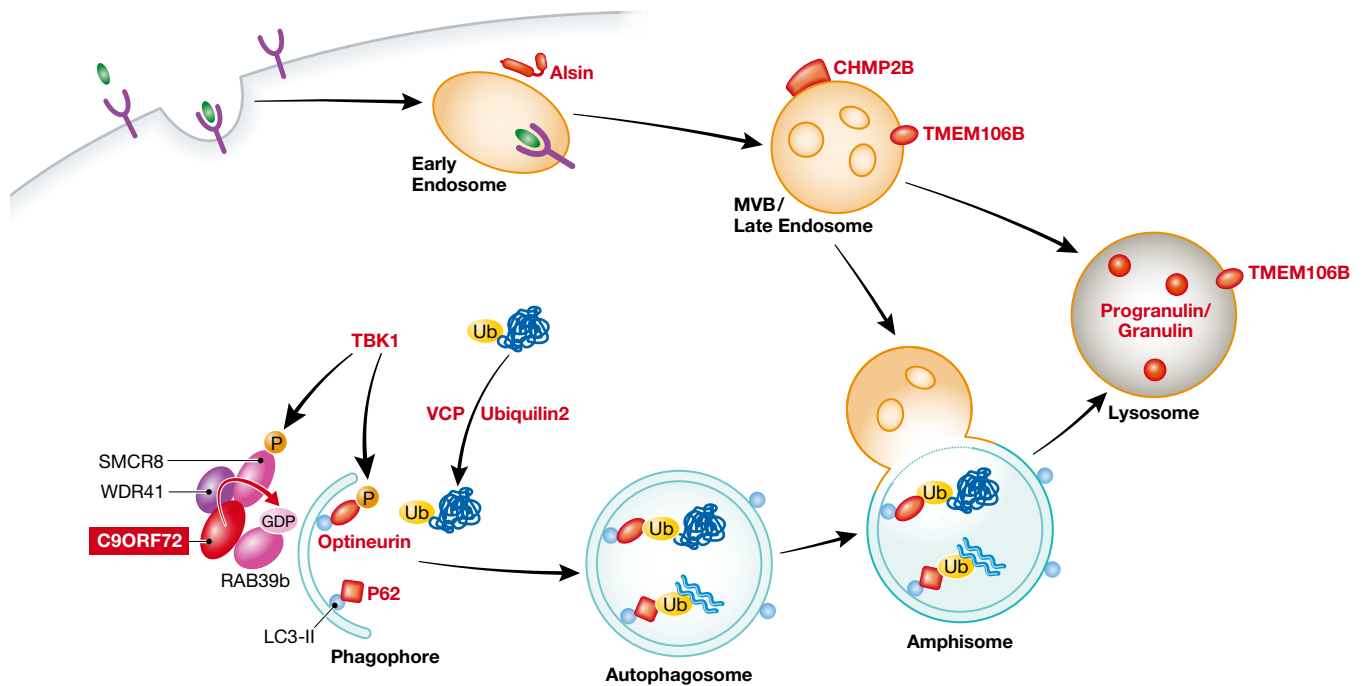


Figure 1. Schematic overview of ALS/FTD disease proteins known to play a role in the autophagy and endosomal–lysosomal pathways.

Sellier *et al* (2016) identified a novel complex consisting of C9ORF72, WDR41, and SMCR8 that serves as a GEF for RAB39b. This complex may play a modulatory role in the initiation of autophagy in certain cell types, but detailed molecular mechanisms remain unclear. Key functions of many other ALS/FTD disease proteins (in red) in some steps of the autophagy and endosomal–lysosomal pathways are also highlighted. MVB, multivesicular body.

loss of C9ORF72 activity or from DPR protein toxicity. Fourth, p62 accumulation and TDP-43 aggregates were not reported in *C9orf72*-knockout mice (Atanasio *et al*, 2016; O'Rourke *et al*, 2016). Thus, compensatory mechanisms may exist when C9orf72 is absent throughout development *in vivo*.

Another interesting aspect of this study is the exploration of interactions between the C9ORF72-containing complex and other ALS/FTD disease proteins, such as TBK1, whose haploinsufficiency causes a subset of ALS/FTD cases (Bettencourt & Houlden, 2015). Sellier *et al* (2016) showed that SMCR8 is a target of TBK1, which also phosphorylates several other targets that function in the autophagy pathway. A form of SMCR8 that mimics a constitutive TBK1 phosphorylation rescued autophagy defects caused by depletion of SMCR8 or TBK1, demonstrating the importance of TBK1-mediated SMCR8 phosphorylation in mediating the function of the C9ORF72/SMCR8/WDR41 complex in autophagy.

The authors also examined the gene encoding ataxin-2, in which intermediate expansion of polyglutamine repeats (27–33) increases risk for ALS and FTD (Elden *et al*, 2010). *C9orf72* depletion and simultaneous

expression of ataxin-2 with 30 glutamines, but not each manipulation alone, induced neurodegeneration in cultured rodent neurons and in a zebrafish model. This synergy seems to be specific to ataxin-2, since *C9orf72* depletion did not enhance the toxicity of mutant SOD1 or other disease proteins. The cause of this specificity remains to be further examined. Reducing *C9orf72* expression did not significantly affect neuronal viability and had only a partial effect on basal autophagy, raising the possibility that this complex plays a modulatory role. It will be interesting to examine whether loss of C9orf72 also affects the accumulation of DPR proteins, especially in mouse models.

C9orf72-knockout mice do not show signs of neurodegeneration in the brain (Atanasio *et al*, 2016; O'Rourke *et al*, 2016). Remarkably, however, they instead develop progressive splenomegaly and lymphadenopathy, a finding reminiscent of the increased incidence of lymphomas and other tumors caused by partial loss of function of beclin 1, a key factor in autophagy initiation (Yue *et al*, 2003). More importantly, loss of C9orf72 led to the accumulation of lysosomes and to abnormal immune responses

in microglia (O'Rourke *et al*, 2016), raising the intriguing possibility that partial loss of function of *C9ORF72* contributes to neurodegeneration in ALS/FTD patients through a non-cell-autonomous mechanism. Future studies are warranted to investigate the cell type-specific roles of the newly identified C9ORF72/SMCR8/WDR41 complex in the endosomal–lysosomal pathway, which is so closely linked to autophagy and also affected by mutations in several other disease genes such as *CHMP2B* and *GRN*.

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