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## Addendum: Independent optical excitation of distinct neural populations

A trafficking variant of the Chrimson molecule (**Addendum Fig. 1c**) was used for the *Drosophila* experiments in the original version of the paper (i.e., **Fig. 3, Supplementary Figs. 14–16, and the Supplementary Videos**). This trafficking variant, called CsChrimson-KGC-GFP-ER2, is a CsChR-Chrimson chimera, replacing the Chrimson N-terminus with the CsChR N-terminus (**Addendum Fig. 1a, Addendum Supplementary Fig. 1**), with appended KGC and ER2 trafficking sequences (**Addendum Fig. 1c**).

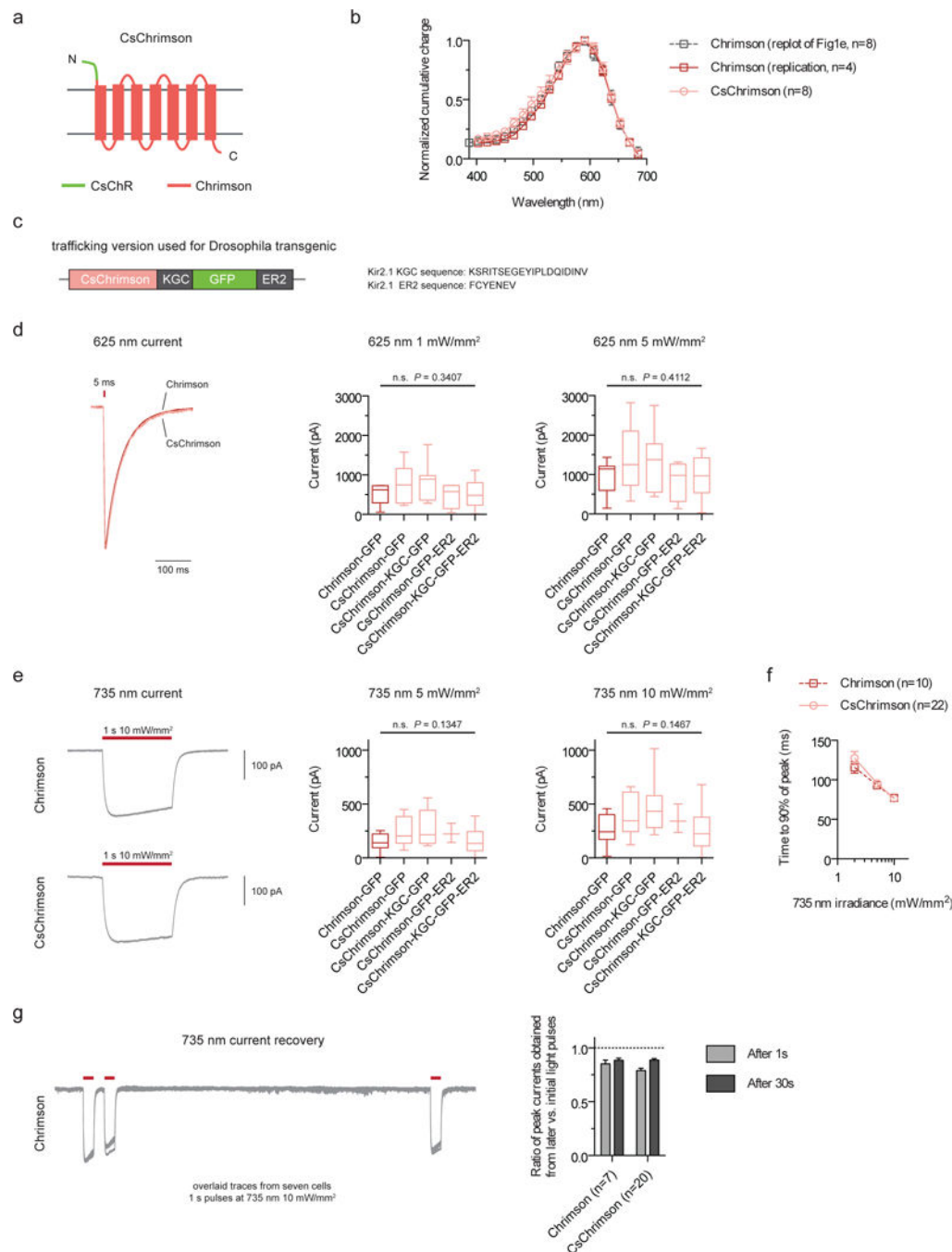
In the original paper, we found CsChR to have high membrane expression levels (**Supplementary Figs. 5–6**). We therefore attempted to boost Chrimson expression by swapping the Chrimson N-terminus with that of the CsChR N-terminus. As no transmembrane regions were modified, we unsurprisingly found that CsChrimson has the same spectral and kinetic properties as Chrimson in murine cultured neurons (**Addendum Figs. 1b, d, f, g**). We additionally compared CsChrimson with and without KGC and/or ER2 trafficking sequences and found all variants to have similar photocurrents in cultured neurons (**Addendum Figs. 1d, e**). However, we observed more cytosolic aggregates with the KGC version and a reduction of aggregates with the ER2 version (**Addendum Supplementary Fig. 2**). It is therefore likely that CsChrimson will be of use with the ER2 trafficking sequence in some biological contexts.

### Methods

Whole-cell patch clamp recordings were made using a Multiclamp 700B amplifier and a Digidata 1550 digitizer (Molecular Devices, Sunnyvale, CA). All other experimental conditions are the same as previously described.

### Supplementary Material

Refer to Web version on PubMed Central for supplementary material.



**Figure 1. CsChrimson characterization in cultured cells**

(a) Schematic of CsChrimson chimera. (b) Action spectra for Chrimson and CsChrimson, as well as the Chrimson spectrum data from the original manuscript (HEK293 cells; measured using equal photon fluxes of  $\sim 2.5 \times 10^{21}$  photons/s/m<sup>2</sup>). (c) Schematic of trafficking sequences used to generate the CsChrimson *Drosophila* transgenics. (d–e) Maximum photocurrents in response to red (625-nm) and far-red (735-nm) light as measured in cultured neurons. (f–g) Turn-on (f) and recovery kinetics (g) in response to 735-nm light. CsChrimson kinetic data were pooled from all trafficking versions. All constructs in this

panel were expressed under CaMKII promoter and selected based solely on the presence of co-transfected cytosolic tdTomato expression. Illumination conditions are as labeled in each panel. Box plot whiskers represent minimum and maximum data points. Box limits represent 25<sup>th</sup> percentile, median, and 75<sup>th</sup> percentile. *n* values: Chromson-GFP (*n* = 9 cells in **d**, *n* = 12 cells in **e**), CsChrimson-GFP (*n* = 7 cells in **d**, *n* = 8 cells in **e**), CsChrimson-KGC-GFP (*n* = 7 cells in **d**, **e**), CsChrimson-GFP-ER2 (*n* = 4 cells in **d**, *n* = 3 cells in **e**), and CsChrimson-KGC-GFP-ER2 (*n* = 10 cells in **d**, *n* = 11 cells in **e**). Plotted data are mean ± s.e.m. in **b**, **f**, and **g**. ANOVA with Dunnett's *post hoc* test with Chromson-GFP as reference in **d**, **e**.