JPP Journal of Pharmacy And Pharmacology

Hot melt extrusion as an approach to improve solubility, permeability and oral absorption of a psychoactive natural product, piperine

Eman A. Ashour^a, Soumyajit Majumdar^{a,b}, Abdulla Alsheteli^c, Sultan Alshehri^d, Bader Alsulays^c, Xin Feng^a, Andreas Gryczke^e, Karl Kolter^f, Nigel Langley^g and Michael A. Repka^{a,b}

^aDepartment of Pharmaceutics and Drug Delivery, School of Pharmacy, ^bPii Center for Pharmaceutical Technology, The University of Mississippi, University, MS 38677, USA, ^cDepartment of Pharmaceutics, College of Pharmacy, Prince Sattam Bin Abdulaziz University, AlKharj, ^dDepartment of Pharmaceutics, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia, ^eGlobal Development and Technical Marketing, ^fR&D Product Management Excipients, BASF SE, Ludwigshafen 67056, Germany and ^gBASF Corporation, 500 White Plains Road, Tarrytown, NY, USA

Keywords

hot melt extrusion; permeability; piperine; solubility

Correspondence

Michael A. Repka, Department of Pharmaceutics & Drug Delivery, Pii Center for Pharmaceutical Technology, School of Pharmacy, The University of Mississippi, University, MS 38677, USA. E-mail: marepka@olemiss.edu

Received February 5, 2016 Accepted April 30, 2016

doi: 10.1111/jphp.12579

Abstract

Objective The aims of the current research project were to investigate the efficiency of various polymers to enhance the solubility and increase the systemic absorption of piperine using hot melt extrusion technology.

Methods Piperine 10–40% w/w and Eudragit[®] EPO/Kollidon[®] VA 64 or Soluplus[®] were mixed, and the resulting blends were extruded using a twin-screw extruder (Process 11, Thermo Fisher Scientific). Drug release profiles and piperine solubility studies of the extrudates were evaluated. A non-everted intestinal sac was employed for the most promising formulation, 10% w/w piperine/Soluplus[®], and pure piperine to study the permeability characteristics.

Key Findings Dissolution studies demonstrated enhancement in piperine per cent release of 10% and 20% w/w piperine/Soluplus[®] extrudates up to 95% and 74%, respectively. The solubility of 10% and 20% piperine/Soluplus[®] increased more than 160- and 45-fold in water, respectively. Furthermore, permeability studies demonstrated the enhancement in piperine absorption of 10% w/w piperine/Soluplus[®] extrudates up to 158.9 µg/5 ml compared with pure piperine at 1.3 µg/5 ml within 20 min.

Conclusion These results demonstrated that increasing the bioavailability of piperine may be achieved as demonstrated by findings in this study.

Introduction

Oral drug delivery is considered as the simplest and easiest route of drug administration.^[1,2] Oral bioavailability is mainly affected by drug solubility, permeability^[3–5] and first pass metabolism.^[6] In fact, most of the new Active Pharmaceutical Ingredients APIs are have low water solubility with low release profiles after oral administration. The biggest challenge in the pharmaceutical industries was to enhance the solubility and the permeability of those drugs as key factors to improve their bioavailability. Various techniques have been used to improve the drug water solubility and release profile, and solid dispersions are considered to be the most successful techniques. There are two main solid dispersion manufacturing methods: the melting method, such as hot melt extrusion (HME) and solvent evaporation methods, such as spray drying.^[7] HME is one of the most commonly used technique to enhance the solubility and oral bioavailability of a poorly soluble drug as a beneficial technique for solid dispersion, which involves simple dispersion of a poorly water soluble API in an inert carrier (polymer), where the drug could exist in amorphous or crystalline state. There are many advantages of using HME due to the speed and the continuous manufacturing process. Moreover, as no solvent is required, this is considered to be a green method for enhancement of the solubility and oral bioavailability of poorly soluble drugs. Depending on the polymeric carrier, HME can be also used for other purposes such as taste masking, controlling or modifying

drug release and stabilizing the active pharmaceutical ingredient.

Piperine (1-piperoylpiperidine) is a crystalline pungent alkaloid isolated from black pepper (Piper nigrum), long pepper (Piper longum) and other pepper species (family Piperaceae).^[8–10] Piperine is a poor water soluble compound with a melting point at 135 °C. It has been extensively used in folk medicine in many Asian countries,^[9] and various studies have focused on investigating the pharmacological effects of piperine. Recently, it was reported as an anti-inflammatory, analgesic,^[11] antidepressant,^[12] cytoprotective, antileukaemic and antioxidant agent.^[8,13] Furthermore, piperine significantly improves spatial memory and neurodegeneration in an Alzheimer's disease animal model.^[14] Furthermore, piperine was reported to be a bioavailable enhancer.^[15] Pattanaik et al.^[16,17] reported that 20 mg of oral piperine can enhance the bioavailability of phenytoin and carbamazepine by increasing their plasma dug concentration. Atal et al.^[18] proved that piperine is a potent inhibitor of the drug metabolism. The literature reported piperine as a metabolic inhibitor that inhibits drug metabolizing enzymes, such as CYP3A4, CYP1B1, CYP1B2 and CYP2E1.^[15,19] These results allowed researchers to incorporate piperine with an anticancer agent acetyl-11-keto-ß-boswellic acid to increase its bioavailability and therapeutic efficacy.^[20] Furthermore, piperine modulates the permeability characteristics to increase the drug absorption through the cell membrane by increasing the vasodilation of the GIT membrane.^[10,21] Being a natural product, piperine had many advantages compared with other chemical entities, such as low cost due to easy availability of plant material and the extraction and isolation methods of piperine are easy and well-known.^[22] Additionally, it is safe to use.

In this study, the main goal was to enhance the solubility and permeability of piperine in order to enhance its bioavailability and therapeutic efficacy. Three polymeric carriers Soluplus[®], polyvinylpyrrolidone-co-vinylacetate 64 (Kollidon[®] VA 64) and Eudragit[®] EPO were selected via the hot melt extrusion process to investigate the solubility and permeability of piperine in order to enhance its bioavailability. The application of HME techniques to form solid dispersions of piperine to enhance solubility coupled with ex-vivo studies to demonstrate improved permeability has not been investigated.

Materials and Methods

Piperine was purchased from Sigma-Aldrich (Milwaukee, WI, USA), while Soluplus[®] and Kollidon[®] VA 64 were obtained from BASF–SE (Ludwigshafen, Germany), Eudragit[®] EPO was received as a gift sample from Evonik Industries (Parsippany, NJ, USA). All other chemicals and

reagents used in this study were of analytical grade and obtained from Fisher Scientific (Fair Lawn, NJ, USA).

Thermogravimetric Analysis

Thermogravimetric analysis (TGA) analysis were performed for piperine, Soluplus[®], Kollidon[®] VA 64 and Eudragit[®] EPO to evaluate their stability at the extrusion temperatures using a Perkin Elmer Pyris 1 TGA equipped with a Pyris manager software (PerkinElmer Life and Analytical Sciences, Shelton, CT, USA). Approximately 10–15 mg of piperine, polymers, as well as physical mixtures, was heated from 30 °C to 300 °C at a heating rate of 20 °C/min.

Differential Scanning Calorimetry

Differential scanning calorimetry (DSC) studies were obtained using Perkin Elmer Pyris 1 DSC equipped with Pyris manager software (PerkinElmer Life and Analytical Sciences). Approximately 2–4 mg of piperine, physical mixtures or extrudates was heated from 30 $^{\circ}$ C to 200 $^{\circ}$ C at a heating rate of 10 $^{\circ}$ C/min.

Hot Melt Extrusion

Piperine 10–40% w/w and Eudragit[®] EPO, Kollidon[®] VA 64, or Soluplus[®] (Table 1) were mixed using a V-shell blender (Patrtreson-Kelley Twin Shell Dry Blender) for 10 min. The resulting physical mixture blends were extruded using a twin-screw extruder (Process 11 Twin Screw Extruder; Thermo Fisher Scientific) at the screw speed of 150 rpm, at a temperature range of 100–130 °C. All extrudates were milled and sieved through an ASTM #35 mesh to obtain a uniform particle size.

Scanning Electron Microscopy

The morphology and physical state of the extrudates were evaluated using Scanning electron microscopy (SEM)

 Table 1
 Piperine formulation composition of hot melt extrusion (HME)

Formulation	Piperine (PIP) (%)	Eudragit [®] PEO (%)	Soluplus [®] (%)	Kollidon [®] VA 64 (%)
P ₁	10	90	_	_
P ₂	20	80	_	-
P ₃	40	60	_	_
P ₄	10	-	90	-
P ₅	20	_	80	_
P ₆	40	-	60	-
P ₇	10	_	_	90
P ₈	20	_	-	80
P ₉	40	_	_	60

analysis. Samples were mounted on adhesive carbon pads placed on aluminium and were sputter coated with gold using a Hummer[®] 6.2 sputtering system (Anatech Ltd, Springfield, VA, USA) in a high vacuum evaporator. A JEOL JSM-5600 scanning electron microscope operating (SEM) at an accelerating voltage of 10 kV was used for imaging.

Fourier Transforms Infrared Spectroscopy

Fourier transforms infrared spectroscopy (FTIR) spectra of piperine, polymeric carriers, physical mixtures, as well as extrudates, were performed using Agilent Cary 630 FTIR spectrometer equipped with a DTGS detector. The same (2-4 mg) was placed directly on ATR, the scanning range was 400–4000 cm⁻¹, and the resolution was 1 cm⁻¹.

High-Performance Liquid Chromatography

High-performance liquid chromatography (HPLC) analyses were performed on all samples to ascertain the piperine content using a Waters HPLC equipped with Empower software to analyse the data. HPLC consisted of a Waters e2695 separation Module and Waters 2489 UV/Visible detector (Waters Technologies Corporation, Milford, MA, USA). The column used was Phenomenex Luna C18 (5 μ , 250 \times 4.6 mm). The mobile phase consisted of a mixture of 0.1% ortho phosphoric acid and acetonitrile (45:55 v/v) with a flow rate of 1.2 ml/min and 20 μ L injection volume. The column temperature was 35 °C (±2). The UV detector wavelength for piperine detection was set at λ_{max} 262 nm.²³

Solubility Test

The solubility was determined at pH 1.2, 5 (water) and 6.8 for formulations P_4 – P_9 as well as pure piperine. Samples equivalent to approximately 100 mg of piperine based on the drug load of each formulation were shaken in 20-ml scintillation vials containing 10 ml tested pH solution. The samples were shaken at 80 rpm and at 25 °C using a Thermo Scientific Precision Reciprocal Shaking Bath. A sample volume of 1 ml was collected at 4, 10, 20 and 24 h. The samples were filtered and analysed by HPLC, and 1 ml of the same pH solution was added back to the scintillation vials to maintain the volume.

In-vitro Drug Release

A sample equivalent to 40 mg piperine of each extrudate as well as pure piperine was filled in gelatin capsules, and the *in-vitro* drug release profiles were run using a USP type II dissolution apparatus. The used dissolution medium was 900 ml of 0.1 N HCl maintained at 37 °C. A sample volume of 2 ml was taken at 10, 20, 30, 45, 60, 90 and 120 min, filtered and analysed using HPLC; then, 2 ml of fresh dissolution medium was added back to the dissolution vessel at each time point to maintain the volume.

Statistical Analysis

Statistical analysis was performed to compare the effects of different polymers and drug loading on piperine drug release utilizing the Kruskal–Wallis test, a nonparametric multiple comparison test followed by the Dunn's test to identify the individual differences between the formulations using SPSS software (IBM Corp. Released 2010. IBM SPSS Statistics for Windows, Version 19.0. Armonk, NY: IBM Corp.). Values of P < 0.05 were considered significant.

Ex-vivo Permeability Model (Non-Everted Intestinal Sac)

The permeability studies were performed for the most promising formulation (P_4) , and pure piperine was used as the control. Non-everted intestinal sacs of male Sprague Dawley rats, weighing approximately 200-250 g, were provided from Charles River (Wilmington, MA, USA). Sacs of 4-5 cm in length were prepared. Each sac was filled with 0.5 ml of incubated Krebs-Ringer bicarbonate phosphate buffer, pH 7.4 with piperine 5 mg/ml. Each non-everted sac was placed in a 25-ml beaker containing of 5 ml Krebs-Ringer bicarbonate phosphate buffer, pH 7.4. A sample volume of 1 ml was taken from the solution outside the sac at 0, 20, 50, 80 and 120 min. The samples were filtered and analysed for the content using HPLC. Fresh Krebs-Ringer buffer (1 ml) was added back to the beaker at each time point to maintain the volume. Cumulative dissolution profiles were calculated.

Results and Discussion

Pre-Formulation Studies

Under all utilized processing temperatures, TGA data showed no decrease in sample weight, which indicated that all formulations in the study were stable under all applied extrusion temperatures. DSC data showed the endothermic crystalline melting peak at 135 °C for pure piperine and all the physical mixture samples (Figure 1). Alternatively, all the extrudates showed absence of crystalline melting peaks in DSC data (Figure 2).

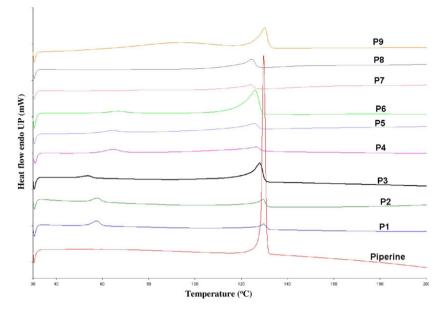


Figure 1 Differential scanning calorimetry (DSC) thermogram of 10%, 20% and 40% PIP/Eudragit[®] PEO, PIP/Soluplus[®] and PIP/Kollidon[®] VA 64 physical mixtures.

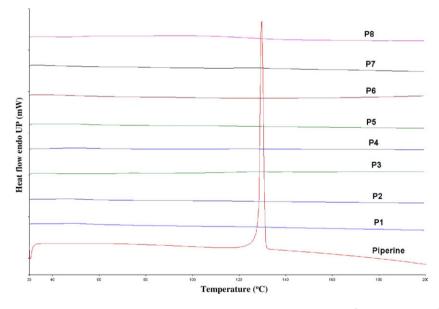


Figure 2 Differential scanning calorimetry (DSC) thermogram of 10%, 20% and 40% of PIP/Eudragit[®] PEO, PIP/Soluplus[®] and PIP/Kollidon[®] VA 64 extrudates.

Hot Melt Extrusion

Hot melt extrusion process was carried out using Process 11 Twin Screw Extruder, Thermo Fisher Scientific. The Thermo Fisher standard screw configuration was used in this study, which consists of four conveying zones and three mixing zones (Figure 3). The HME processing conditions such as extrusion temperature, screw speed and torque values are shown in Table 2. The resulting extrudates were transparent, greenish yellow and brittle, except for P_3 (40% piperine/ Eudragit[®] EPO), which was opaque. This observation can be explained by the fact that 40% of the drug load exceeded the Eudragit[®] EPO carrier capacity allowing the extrudate to mimic the appearance of the piperine only. All extrudates were milled using a coffee grinder and sieved through an ASTM #35 mesh to obtain a uniform particle size.

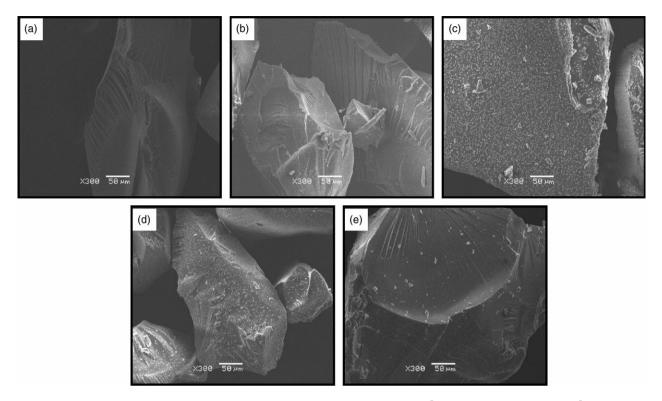


Figure 3 Scanning electron microscopy (SEM) of extrudates: (a) 10% w/w piperine/Soluplus[®], (b) 20% w/w piperine/Soluplus[®], (c) 40% w/w piperine/Soluplus[®], (d) 20% piperine/Eudragit[®] EPO, (e) 20% piperine/Kollidon[®] VA64.

Scanning Electron Microscopy

Scanning electron microscopy images showed absence of crystals in 10% w/w piperine/Soluplus[®] indicating that piperine was dispersed in the Soluplus[®] polymer carrier as an amorphous form. This may help to enhance the solubility and bioavailability of this formulation. However, crystals were evident in all other formulations with different ratios (Figure 3). These results are not consistent with the DSC results that showed absence of the piperine crystalline peak in the resulting extrudates. This observation be explained by the fact that all polymers used in this study, that is Eudragit® EPO, Kollidon® VA 64 and Soluplus®, have a Tg of 45 °C, 101 °C and 70 °C, respectively, which is lower than the piperine melting point. Due to this, the polymers used will soften before the melting temperature of piperine allowing the piperine to solubilize in the polymer matrix and prevent the melting peak appearance.

Fourier Transforms Infrared Spectroscopy

Fourier transforms infrared spectroscopy analysis was performed to evaluate any chemical or physical interaction between piperine and the different polymeric carriers. Figure 4a shows the FTIR spectra of Soluplus and their formulations; the physical mixture displays a sharp peak at 1623 cm⁻¹, and this peak shifted to approximately 1629 cm⁻¹ for all Soluplus formulations, indicating the presence of hydrogen bonding between piperine and Soluplus[®]. In contrast, there were no observed shifts in Eudragit[®] EPO and Kollidon[®] VA 64 formulations (Figure 4b and c). The formation of hydrogen bonding will increase the stability of the Soluplus solid dispersion formulations and prevent recrystallization of piperine.^[7,24,25]

Solubility Evaluation

Solubility study was performed for formulations P_4 , P_5 , P_6 , P_7 , P_8 and P_9 using three different solvents pH (1.2, 5 [water] and 6.8) to evaluate whether piperine solubility is pH dependent or not. The pH did not have any effect on the solubility of piperine as there were slight changes in the solubility values at pH 1.2 and 6.8 (Table 3). Furthermore, the solubility of piperine in formulations P_4 , P_5 , P_6 , P_7 , P_8 and P_9 increased more than 160-, 45-, 25-, 20-, fivefold and threefold, respectively, compared with the water solubility of piperine (Table 3). This significant enhancement in the water solubility of piperine in its amorphous state which was supported by SEM. Amorphous solid dispersion plays a significant role in increasing the solubility and dissolution rate of poorly water soluble APIs.^[26-29] In general,

Improved oral absorption of piperine via HME

Formulation	Processing temperature (°C)	Screw speed (rpm)	Torque (Nm)	Extrudate image
P ₁	110	150	6–7	
2	110	150	4–5	
P ₃	110	150	3	
P ₄	120	150	7–8	
P ₅	120	150	4–5	
P ₆	120	150	3-4	
P ₇	130	150	7–8	
P ₈	130	150	3-4	
P9	130	150	3–4	

amorphous API is more soluble than crystalline ones, because of its higher effective surface area than the crystalline form.^[30–32] However, the main problem associated with amorphous API is its instability, which can lead to recrystallization during storage as well as during dissolution.^[33] In contrast, amorphous solid dispersions are stable as polymeric carriers prevent API recrystallization.^[34]

In-Vitro Release Profiles

Dissolution studies demonstrated improvement in piperine drug release of 10% and 20% w/w piperine/Soluplus[®] extrudates up to 95% and 74%, respectively (Figure 5). However, no effect on piperine drug release profiles for other formulations was noted due to the remaining crystalline piperine. These results confirmed that the dispersion of piperine in the amorphous state plays a big role in enhancing the solubility^[35] and drug release. Additionally, it can be concluded that the maximum drug loading capacity of piperine in Soluplus to form stabilized amorphous solid dispersion is approximately 10%. Soluplus[®] polyvinyl caprolactam–polyvinyl acetate–polyethylene glycol graft copolymer is a relatively new polymer designed mainly for use in solid dispersion preparation APIs.^[36] It acts as a polymeric stabilizer and solubilizing agent to improve the solubility and bioavailability of poor water

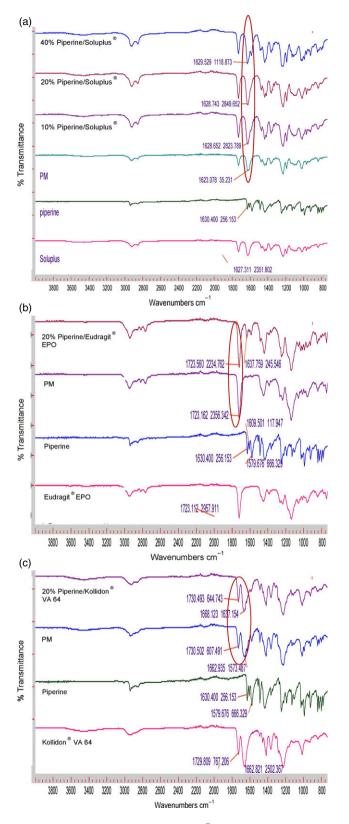


Figure 4 (a) Fourier transforms infrared spectroscopy (FTIR) spectra of Soluplus[®], piperine, physical mixture, 10% piperine/Soluplus[®], 20% piperine/Soluplus[®] and 40% piperine/Soluplus[®]. (b) FTIR spectra of Eudragit[®] EPO, piperine, 20% piperine/Eudragit[®] EPO physical mixture and extrudates. (c) FTIR spectra of Kollidon[®] VA 64, piperine, 20% piperine/Kollidon[®] VA 64 physical mixture and extrudates.

© 2016 Royal Pharmaceutical Society, Journal of Pharmacy and Pharmacology, 68 (2016), pp. 989–998

soluble APIs.^[37] It is a good polymeric carrier candidate to prepare amorphous solid dispersions of many poor water soluble APIs such as clotrimazole, carbamazepine, griseofulvin and itraconazole by improving their dissolution profiles as well as their intestinal absorption and bioavailability.^[37] In the recent study, Soluplus[®] was

Table 3 Solubility of piperine and piperine formulation in water, pH 1.2 and 6.8 $\,$

Formulation	Solubility in water (mg/l)	Solubility in pH 1.2 (mg/l)	Solubility in pH 6.8 (mg/l)
Piperine	1	0.9	0.9
P ₄	163	117	117
P ₅	47	39	45
P ₆	27	21	27
P ₇	20	18	17
P ₈	6	6	6
P ₉	3	3	3

successfully used to prepare 10% piperine amorphous solid dispersion with a significant increase (P < 0.05) in the piperine release profile.

Ex-vivo Permeability Model (Non-Everted Intestinal Sac)

There are many *in-vivo*, *ex-vivo* and *in-vitro* assays used to evaluate the intestinal drug permeability.^[38] These assays include perfusion, diffusion chambers, gut sac, cultured cell and artificial membrane.^[39] The results of a research investigation showed that there was a good correlation between rat and human oral absorption. There are two types of rat intestinal sac models that are used to evaluate the drug permeability: everted and non-everted sacs. Some studies have been performed to compare the permeability values of some APIs through both everted and non-everted sacs, and no significant difference was observed in the permeability results. In this study, the non-everted rat intestinal sac

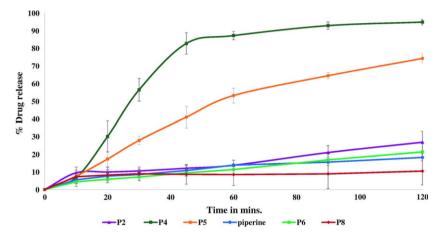


Figure 5 *In-vitro* release profiles of 10% w/w piperine/Soluplus[®], 20% w/w piperine/Soluplus[®], 40% w/w piperine/Soluplus[®], 20% piperine/Eudragit[®] EPO, 20% piperine/Kollidon[®] VA64 and pure piperine.

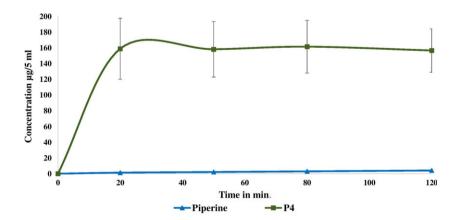


Figure 6 Piperine absorption characteristics using jejunum non-everted sac under normal physiological conditions (Krebs–Ringer bicarbonate phosphate buffer, pH 7.4).

model was used to study piperine absorption. The permeability study results demonstrated the enhancement in piperine absorption of 10% w/w piperine/Soluplus[®] up to 158.9 μ g/5 ml compared with 1.4 μ g/5 ml in the case of pure piperine within 20 min (Figure 6) as a direct result of the solubility improvement (over 112-fold increase in absorption).

Conclusion

Hot melt extrusion was successfully used to enhance the solubility of the psychoactive natural product piperine. The drug release profiles of 10% and 20% w/w piperine/Soluplus[®] hot melt extrudates were significantly enhanced due to piperine in its amorphous state. In addition, the formation of hydrogen bonds between piperine and Soluplus[®] may inhibit drug recrystallization and assist in the maintenance of the drug in its amorphous form, thereby enabling

References

- 1. Youn YS *et al.* Improved intestinal delivery of salmon calcitonin by Lys 18-amine specific PEGylation: stability, permeability, pharmacokinetic behavior and in vivo hypocalcemic efficacy. *J Control Release* 2006; 114: 334–342.
- Guggi D *et al.* Systemic peptide delivery via the stomach: in vivo evaluation of an oral dosage form for salmon calcitonin. *J Control Release* 2003; 92: 125–135.
- Pade V, Stavchansky S. Link between drug absorption solubility and permeability measurements in Caco-2 cells. *J Pharm Sci* 1998; 87: 1604–1607.
- Hörter D, Dressman J. Influence of physicochemical properties on dissolution of drugs in the gastrointestinal tract. *Adv Drug Deliv Rev* 2001; 46: 75–87.
- Martinez MN, Amidon GL. A mechanistic approach to understanding the factors affecting drug absorption: a review of fundamentals. J Clin Pharmacol 2002; 42: 620–643.
- 6. Wu Y *et al.* The role of biopharmaceutics in the development of a clinical nanoparticle formulation of MK-0869: a Beagle dog model predicts improved bioavailability and diminished food effect on absorption in

human. Int J Pharm 2004; 285: 135-

 Vasconcelos T *et al.* Solid dispersions as strategy to improve oral bioavailability of poor water soluble drugs. *Drug Discov Today* 2007; 12: 1068– 1075.

146.

- Chonpathompikunlert P *et al.* Piperine, the main alkaloid of Thai black pepper, protects against neurodegeneration and cognitive impairment in animal model of cognitive deficit like condition of Alzheimer's disease. *Food Chem Toxicol* 2010; 48: 798–802.
- 9. Wattanathorn J *et al.* Piperine, the potential functional food for mood and cognitive disorders. *Food Chem Toxicol* 2008; 46(9): 3106–3110.
- Khajuria A *et al.* Piperine modulates permeability characteristics of intestine by inducing alterations in membrane dynamics: influence on brush border membrane fluidity, ultrastructure and enzyme kinetics. *Phytomedicine* 2002; 9: 224–231.
- 11. Gupta S *et al.* Comparative anti-nociceptive, anti-inflammatory and toxicity profile of nimesulide vs nimesulide and piperine combination. *Pharmacol Res* 2000; 41(6): 657–662.
- Kontush A, Schekatolina S, Vitamin E in neurodegenerative disorders: Alzheimer's disease. Ann N Y Acad Sci, 2004. 1031: 249–262.

good formulation stability. Furthermore, a significant improvement in piperine permeability was confirmed using a non-everted rat intestinal sac. These results demonstrate that an increase in the absorption and bioavailability of piperine are possibilities.

Conflict of Interest

The authors report no conflict of interests to disclose.

Acknowledgements and Funding

This work was partially supported by Grant Number P20GM104932 from the National Institute of General Medical Sciences (NIGMS), a component of NIH. The authors also thank the Pii Center for Pharmaceutical Technology for contributions in this project.

- Selvendiran K *et al.* Cytoprotective effect of piperine against benzo [a] pyrene induced lung cancer with reference to lipid peroxidation and antioxidant system in Swiss albino mice. *Fitoterapia* 2003; 74: 109–115.
- Borre YE *et al.* Neuroprotective and cognitive enhancing effects of a multitargeted food intervention in an animal model of neurodegeneration and depression. *Neuropharmacology* 2014; 79: 738–749.
- Kesarwani K, Gupta R. Bioavailability enhancers of herbal origin: an overview. Asian Pac J Trop Biomed 2013; 3: 253–266.
- Pattanaik S *et al.* Effect of piperine on the steady-state pharmacokinetics of phenytoin in patients with epilepsy. *Phytother Res* 2006; 20(8): 683–686.
- Pattanaik S *et al.* Pharmacokinetic interaction of single dose of piperine with steady-state carbamazepine in epilepsy patients. *Phytother Res* 2009; 23: 1281–1286.
- Atal C *et al.* Biochemical basis of enhanced drug bioavailability by piperine: evidence that piperine is a potent inhibitor of drug metabolism. *J Pharmacol Exp Ther* 1985; 232(1): 258–262.
- Majeed M et al., Use of piperine as a bioavailability enhancer, 1999, Google Patents.

^{© 2016} Royal Pharmaceutical Society, Journal of Pharmacy and Pharmacology, 68 (2016), pp. 989–998

- 20. Brough C et al., Pharmaceutical formulations of acetyl-11-keto-b-boswellic acid, diindolylmethane, and curcumin for pharmaceutical applications, 2012, Google Patents.
- Khajuria A *et al.* Permeability characteristics of piperine on oral absorption–an active alkaloid from peppers and a bioavailability enhancer. *Indian J Exp Biol* 1998; 36: 46–50.
- Ribeiro TS *et al.* Toxic effects of natural piperine and its derivatives on epimastigotes and amastigotes of Trypanosoma cruzi. *Bioorg Med Chem Lett* 2004; 14: 3555–3558.
- Moorthi C *et al.* Application of validated RP–HPLC–PDA method for the simultaneous estimation of curcumin and piperine in Eudragit E 100 nanoparticles. *J Pharm Res* 2013; 7: 224–229.
- Vasanthavada M *et al.* Phase behavior of amorphous molecular dispersions II: role of hydrogen bonding in solid solubility and phase separation kinetics. *Pharm Res* 2005; 22: 440–448.
- 25. Schachter DM *et al.* Solid state NMR perspective of drug–polymer solid solutions: a model system based on poly (ethylene oxide). *Int J Pharm* 2004; 281: 89–101.
- 26. Van den Mooter G. The use of amorphous solid dispersions: a for-

mulation strategy to overcome poor solubility and dissolution rate. *Drug Discov Today Technol* 2012; 9: e79– e85.

- 27. Blagden N *et al.* Crystal engineering of active pharmaceutical ingredients to improve solubility and dissolution rates. *Adv Drug Deliv Rev* 2007; 59: 617–630.
- Lakshman JP *et al.* Application of melt extrusion in the development of a physically and chemically stable high-energy amorphous solid dispersion of a poorly water-soluble drug. *Mol Pharm* 2008; 5: 994– 1002.
- Kennedy M et al. Enhanced bioavailability of a poorly soluble VR1 antagonist using an amorphous solid dispersion approach: a case study. Mol Pharm 2008; 5: 981–993.
- 30. Pignatello R *et al.* Preparation, characterisation and photosensitivity studies of solid dispersions of diflunisal and Eudragit RS100[®] and RL100[®]. *Int J Pharm* 2001; 218: 27–42.
- Craig DQ. The mechanisms of drug release from solid dispersions in water-soluble polymers. *Int J Pharm* 2002; 231: 131–144.
- 32. Kawabata Y *et al.* Formulation design for poorly water-soluble drugs based on biopharmaceutics classification

system: basic approaches and practical applications. *Int J Pharm* 2011; 420: 1–10.

- 33. Alonzo DE *et al.* Understanding the behavior of amorphous pharmaceutical systems during dissolution. *Pharm Res* 2010; 27: 608–618.
- 34. Overhoff KA *et al.* Solid dispersions of itraconazole and enteric polymers made by ultra-rapid freezing. *Int J Pharm* 2007; 336: 122–132.
- 35. Shah S *et al.* Melt extrusion with poorly soluble drugs. *Int J Pharm* 2013; 453: 233–252.
- 36. Shamma RN, Basha M. Soluplus[®]: a novel polymeric solubilizer for optimization of Carvedilol solid dispersions: formulation design and effect of method of preparation. *Powder Technol* 2013; 237: 406–414.
- Hardung H et al. Combining HME & solubilization: Soluplus[®]—the solid solution. Drug Deliv Technol 2010; 10: 20–27.
- Ruan L-P *et al.* Prediction of human absorption of natural compounds by the non-everted rat intestinal sac model. *Eur J Med Chem* 2006; 41: 605–610.
- Volpe DA. Application of method suitability for drug permeability classification. AAPS J 2010; 12: 670– 678.