PHILOSOPHICAL TRANSACTIONS B

rstb.royalsocietypublishing.org

CrossMar

Review

Cite this article: Berridge MJ. 2016 Vitamin D, reactive oxygen species and calcium signalling in ageing and disease. *Phil. Trans. R. Soc. B* **371**: 20150434. http://dx.doi.org/10.1098/rstb.2015.0434

Accepted: 15 March 2016

One contribution of 15 to a Theo Murphy meeting issue 'Evolution brings Ca²⁺ and ATP together to control life and death'.

Subject Areas:

cellular biology, biochemistry

Keywords:

vitamin D, calcium, cardiac disease, Alzheimer's disease, klotho, inositol trisphosphate

Author for correspondence:

Michael J. Berridge

e-mail: michael.berridge@babraham.ac.uk

Vitamin D, reactive oxygen species and calcium signalling in ageing and disease

Michael J. Berridge

Babraham Institute, Babraham, Cambridge CB22 3AT, UK

Vitamin D is a hormone that maintains healthy cells. It functions by regulating the low resting levels of cell signalling components such as Ca²⁺ and reactive oxygen species (ROS). Its role in maintaining phenotypic stability of these signalling pathways depends on the ability of vitamin D to control the expression of those components that act to reduce the levels of both Ca²⁺ and ROS. This regulatory role of vitamin D is supported by both Klotho and Nrf2. A decline in the vitamin D/Klotho/Nrf2 regulatory network may enhance the ageing process, and this is well illustrated by the age-related decline in cognition in rats that can be reversed by administering vitamin D. A deficiency in vitamin D has also been linked to two of the major diseases in man: heart disease and Alzheimer's disease (AD). In cardiac cells, this deficiency alters the Ca²⁺ transients to activate the gene transcriptional events leading to cardiac hypertrophy and the failing heart. In the case of AD, it is argued that vitamin D deficiency results in the Ca²⁺ landscape that initiates amyloid formation, which then elevates the resting level of Ca²⁺ to drive the memory loss that progresses to neuronal cell death

This article is part of the themed issue 'Evolution brings Ca^{2+} and ATP together to control life and death'.

1. Introduction

A large number of cellular processes are regulated by calcium (Ca^{2+}). An important component of Ca^{2+} signalling is the $InsP_3/Ca^{2+}$ signalling pathway, which has two main operational modes. It functions either as a primary signalling pathway or it can operate as a modulatory signal. Its primary role is evident mainly in non-excitable cells where it generates the Ca^{2+} signals to control processes as diverse as fertilization, proliferation, metabolism, secretion and smooth muscle contraction. In excitable cells, the primary Ca^{2+} signal depends on the entry of Ca^{2+} through voltage-operated channels and the release of Ca^{2+} by ryanodine receptors (RYRs) on the internal stores. This primary Ca^{2+} pathway regulates processes such as contraction in the heart or memory formation in neurons. The $InsP_3/Ca^{2+}$ signalling pathway provides a modulatory signal that can induce subtle changes in the generation and function of this primary Ca^{2+} signal. In this review, I will argue that subtle changes in the nature of this modulatory role of the $InsP_3/Ca^{2+}$ signalling pathway may be responsible for the onset of two major human diseases: Alzheimer's disease (AD) and cardiovascular disease.

Both cardiac disease and AD are age related and what is remarkable is their very slow progression. Most individuals who develop these diseases are completely unaware that the disease is developing, and it is this aspect that may be explained by the subtle modulatory activity of the InsP₃/Ca²⁺ signalling pathway. It will be argued that one of the main causes of the alteration in this modulatory pathway is vitamin D deficiency that causes the small alterations in the Ca²⁺ signalling pathway responsible for the onset of these two diseases [1–3]. All this evidence raises a major question concerning what it is about vitamin D that makes it such an important component of a healthy life. Any hypothesis as to how vitamin D deficiency might contribute to disease has to take into account a possible relationship between ageing and vitamin D deficiency. There is increasing evidence that vitamin D acts by maintaining the

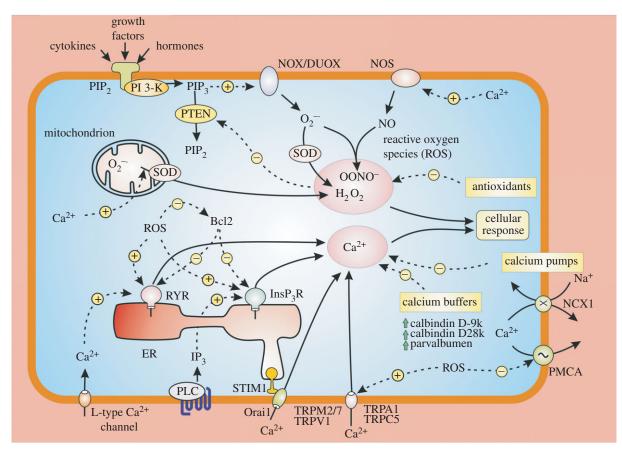


Figure 1. The main reactive oxygen species (ROS) in cells are superoxide $(0_2^{-\cdot})$, hydrogen peroxide (H_2O_2) and peroxynitrite $(0NOO^-)$. The dashed lines represent the many interactions that operate between the Ca^{2+} and redox signalling pathways. An increase in Ca^{2+} can promote ROS formation by entering the mitochondria to form $O_2^{-\cdot}$ that is converted into H_2O_2 by SOD2. Ca^{2+} can also stimulate the nitric oxide synthase (NOS) that forms NO that interacts with $O_2^{-\cdot}$ to form $ONOO^-$. In a reciprocal way, an increase in cytosolic ROS can markedly enhance Ca^{2+} signalling by either increasing the activity of various channels such as the $InsP_3Rs$ and RYRs or by inhibiting the PMCA pump.

integrity of cell signalling pathways such as those regulated by Ca^{2+} and reactive oxygen species (ROS) [1–3]. It will be argued that low vitamin D levels result in an increase in the activity of these two signalling pathways that not only act to accelerate the ageing process but may also set the stage for the onset of a large number of diseases.

2. Integrated calcium and redox signalling pathways

A large number of cellular processes are regulated by Ca²⁺ signalling pathways often operating in conjunction with the redox signalling pathway [1,2]. What is remarkable about these two signalling systems is the way they interact with each other [4] (figure 1). When Ca²⁺ builds up within the mitochondrion, it increases mitochondrial metabolism resulting in an increased formation of superoxide (O₂⁻). Another action of Ca²⁺ is to stimulate nitric oxide synthase (NOS) to increase the formation of NO that contributes to the generation of peroxynitrite (ONOO⁻). Similarly, ROS can enhance Ca²⁺ signalling. For example, ROS sensitizes both the RYRs [5-7] and the InsP₃Rs [8] to increase the release of Ca²⁺ from the internal store. The expression of Bcl-2, which regulates Ca²⁺ signalling by controlling Ca²⁺ release by the InsP₃ receptors [9] and RYRs [10,11], is reduced by ROS [12]. ROS can activate a number of TRP channels that gate Ca²⁺ (e.g. TRPM2, TRPA1 and TRPV1) [13].

The reason for concentrating on these two pathways is because the expression of many of the genes responsible for regulating them is controlled by vitamin D [1,2]. Any deficiency in vitamin D will result in an alteration in how they operate, and this can have profound consequences for many different cellular processes and may be responsible for triggering a number of the diseases that have been linked to vitamin D deficiency.

3. Vitamin D regulation of the Ca²⁺ and redox signalling pathways

The active component of vitamin D is 1α ,25-dihydroxyvitamin D₃ $[1\alpha$,25(OH)₂D₃] that is formed by a series of reactions that begin in the skin where sunlight converts 7-dehydrocholesterol to vitamin D₃ (cholecalciferol) (figure 2). The latter is transferred to the liver where a hydroxyl group is added to the C-25 position by a vitamin D-25 hydroxylase (encoded by the CYP27A1 gene) to form 25-hydroxyvitamin D₃ [25(OH)D₃] that is the immediate precursor for active vitamin D. This 25(OH)D₃ is carried in the blood to enter multiple cell types where a 25(OH)D₃-1 α -hydroxylase (encoded by the CYP27B1 gene) adds another hydroxyl group to the 1 position to form the active hormone 1α ,25(OH)₂D₃, which will be referred to hereafter as vitamin D, that functions to regulate many different cellular processes [14].

Vitamin D can act in two ways. Firstly, it has non-genomic actions where it alters the activity of various signalling

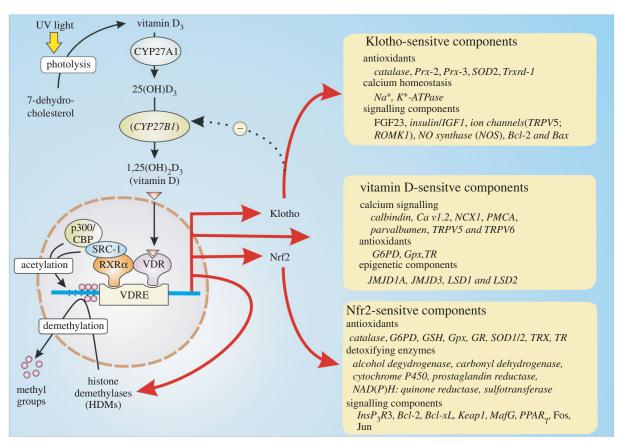


Figure 2. The vitamin D hormone 1,25-dihydroxyvitamin D_3 [1α ,25(OH) $_2D_3$] binds to the vitamin D receptor (VDR) that interacts with the retinoid X receptor (RXR) to form the VDR/RXR heterodimer that binds to the vitamin D response element (VDRE). Once in place, the VDR initiates the expression of a large number of genes located in many different cell types to express proteins that function in a number of cellular processes. Many of its actions also depend on its ability to increase the expression of both Klotho and Nrf2 that carry out many of its homeostatic functions.

pathways. Secondly, it has a genomic action that is mediated by its binding to the vitamin D receptor (VDR), which interacts with the retinoid X receptor (RXR) before binding to the vitamin D response element (VDRE) located on a large number of vitamin D-sensitive target genes (figure 2). Two of the important genes that are activated by vitamin D are *Nrf2* and the anti-ageing gene *Klotho*, both of which have multiple roles in maintaining the integrity of cellular signalling systems (figure 2). Many of the genes that are controlled by the vitamin D/Klotho/Nrf2 regulatory network function to maintain Ca²⁺ and redox homeostasis. For example, vitamin D increases the expression of Ca²⁺ pumps, exchangers and buffers to maintain low levels of Ca²⁺. In addition, vitamin D together with Klotho and Nrf2 all increase cellular antioxidants to maintain the normal reducing environment within the cell [15,16].

Vitamin D may also play a significant role in regulating the balance between autophagy and apoptosis [17,18]. This ability of vitamin D to promote autophagy over apoptosis may depend on its ability to regulate Ca^{2+} signalling that plays a significant role in controlling autophagy [19,20]. Subsequent studies revealed that the $\text{InsP}_3/\text{Ca}^{2+}$ signalling pathway plays a prominent role in regulating autophagy [21–23]. Elevation in various pathological aggregates such as amyloid, tau, α -synucleins and mutant Huntington fragments, which contribute to neuro-degenerative disease such as AD, Parkinson's disease and Huntington's disease, may accumulate because of a decline in autophagy due to an alteration in Ca^{2+} signalling that occurs when vitamin D is deficient.

Another important action of vitamin D is to control the epigenetic landscape of multiple gene promoters to maintain

the transcription activity of all the genes that function in its regulatory network [24]. Vitamin D influences the epigenetic landscape by controlling both the acetylation and methylation states of multiple gene promotor regions. The VDR/RXR dimer recruits histone acetyltransferases (HATs) such as p300/CBP and steroid receptor coactivators 1 and 2 (SRC1 and SRC2) that carry out the acetylation reactions that open up the chromatin structure to facilitate transcription so as to maintain phenotypic stability (figure 2).

Vitamin D can also regulate phenotypic stability by regulating demethylation. Many of the genes regulated by vitamin D are silenced by methylation of CpG islands located in their promotor regions [25]. For example, the decline in SERCA2a activity in cardiovascular disease may be caused by hypermethylation of its promotor region [26]. Expression of the Klotho gene, which acts together with vitamin D to regulate phenotypic stability, is silenced by methylation [27,28]. Such hypermethylation of promotor regions increases during ageing and is evident in many of the diseases such as cancer, cardiovascular and neurodegenerative diseases [29]. For example, hypermethylation of promotors in GABAergic neurons may contribute to the phenotypic remodelling responsible for schizophrenia and bipolar disorder [30]. Vitamin D modulates methylation by inducing the expression of a number of key DNA demethylases such as Jumonji domain-containing protein 1A and 3 (JMJD1A, JMJD3) and lysine-specific demethylase 1 and 2 (LSD1, LSD2) that contributes to its ability to maintain phenotypic stability [31].

This ability of vitamin D to modulate the epigenetic landscape is in keeping with its proposed role in maintaining phenotypic stability, and this may explain why vitamin D deficiency has been linked to both ageing and so many of the age-related diseases.

4. Vitamin D and ageing

There is increasing evidence that vitamin D may play an important role in the process of ageing. For example, the decline in cognition that occurs normally in older adults has been linked to vitamin D deficiency [32-35]. The ability of human skin to synthesize vitamin D declines with age [36], and this may account for the decline in the level of vitamin D and Klotho during ageing. Vitamin D and Klotho deficiency may contribute to the ageing process through dysregulation of the Ca²⁺ and redox cell signalling pathways. Nrf2 may also act to regulate longevity [37]. Dysregulation of Ca²⁺ signalling, which is closely linked to mitochondrial dysfunction and ROS formation, has been implicated in ageing [38,39]. In ageing striatal neurons, there is a marked decline in the expression of Bcl2 [38], which would contribute to the dysregulation of Ca²⁺, because one of its functions is to inhibit the InsP₃Rs [9] (figure 1).

There has long been an interest in the possibility that alterations in the cellular redox balance [40,41] and Ca²⁺ signalling [42] might be responsible for ageing [43]. The way in which vitamin D deficiency and a concomitant decline in both Klotho and Nrf2 function contributes to many diseases may be explained through the ability of these custodial systems to maintain the stability of the redox and Ca2+ signalling systems described earlier [2]. For example, during ageing, there is a decline in the capacity of cells to maintain NAD(P)H levels in neurons [44,45], and this accounts for a decline in the levels of glutathione (GSH), which is essential to maintain low redox levels [46]. Such a decline in GSH results in a selective decline in the activity of GABAergic neurons in the hippocampus and could contribute to schizophrenia [47]. Vitamin D acts to maintain the expression of the Nrf2 antioxidant pathway [48]. There is a marked decline in the level of Nrf2 in the AD brain compared with age-matched controls [49]. Genetic ablation of the VDR results in premature ageing in mice suggesting that vitamin D can maintain normal physiological ageing [50].

Some of the most convincing evidence that vitamin D deficiency contributes to the ageing process has emerged from studies on the decline of memory in ageing rats. When considering memory mechanisms, it is important to point out that the ageing process does not affect long-term memories, but it induces a slow and progressive deterioration in the formation and retention of new memories [51]. This initial age-related decline in working memory is very subtle and has been linked to small changes in both the Ca2+ and redox signalling pathways [51-53]. An alteration in Ca²⁺ signalling has been linked to ageing in the brain [54-58]. The early loss of memory is caused by a number of subtle changes such as an elevation in the resting level of Ca²⁺ [56] and an increase in the expression of the Ca_V1.2 L-type Ca²⁺ channel [58], which is one of the proteins that is normally down-regulated by vitamin D (figure 2). Such changes may also depend on a decrease in the neuronal Ca²⁺ buffers and a decline in the mechanisms responsible for extruding Ca²⁺ from the cytoplasm [59]. Enhancing the intracellular buffering capacity markedly enhanced the learning capacity of aged rats [60].

At the electrophysiological level, the loss of memory during ageing has been linked to the progressive increase in the amplitude of the slow after hyperpolarization (sAHP) [52,61]. This sAHP is caused by increased fluxes of Ca²⁺ through Ca_V1.2 L-type voltage-gated channels, which are known to be elevated during ageing [58], and the RYRs resulting in abnormally high Ca²⁺ transients that activate SK potassium channels to hyperpolarize the membrane [51,55,57]. This sAHP reduces working memory in two ways. Firstly, the hyperpolarization reduces the spiking activity necessary for memory formation through long-term potentiation (LTP). Secondly, the increase in Ca²⁺ stimulates calcineurin to induce the long-term depolarization (LTD) that erases memories [52].

One of the interesting aspects of this dysregulation is that the relatively subtle elevation in the sensitivity of the Ca²⁺ signalling pathway appears to be driven by an increase in the oxidative state of the neurons [62]. In ageing mice, there is a marked increase in oxidative stress that contributes to a reduction in memory formation [63]. The enhanced ROS levels may increase sAHP by sensitizing the RYRs (figure 1). This would seem to be the case because the sAHP can be reversed by treating neurons with dithiothreitol (DTT) [64]. Similarly, a decrease in ROS could also contribute to the increase in cognition observed in ageing rats following treatment with the anti-inflammatory drug montelukast that is used normally to treat asthma [65]. The dysregulation of both Ca²⁺ and ROS signalling that is responsible for development of the sAHP during ageing seems to depend on vitamin D deficiency. The vitamin D/Klotho/NRF2 regulatory system can prevent the dysregulation of the Ca²⁺ and ROS signalling responsible for the sAHP through multiple mechanisms. For example, vitamin D suppresses the expression of the Ca_V1.2 L-type Ca²⁺ channel [66] that initiates the Ca²⁺ signal that induces the sAHP, and it also maintains the expression of PMCA and NCX1, which extrude Ca²⁺ from the cell. Klotho acts to stimulate the Na⁺/K⁺-ATPase responsible for maintaining the Na⁺ gradient necessary for Ca²⁺ extrusion by NCX1. Finally, NRF2 increases the expression of many antioxidants that ensure that ROS levels are kept low, which will prevent the sensitization of the RYRs that are triggering the sAHP and memory erasure.

Such a conclusion is strongly supported by the observation that vitamin D can reverse the Ca2+-dependent processes responsible for the age-related decline in memory [67]. What is more significant is that vitamin D can enhance hippocampal synaptic function, and more significantly, it could prevent the decline in cognition [68].

The fact that vitamin D deficiency brings about a dysregulation of both the Ca²⁺ and redox signalling pathways during ageing has raised an interesting possibility that it could also contribute to the initiation of age-related diseases [2].

5. The vitamin D/Klotho/Nrf2 regulatory network and disease

While most attention has been focused on establishing a link between vitamin D deficiency and disease, there is little information as to what the mechanism might be. To answer this question, I have developed a phenotypic stability hypothesis that is based on the idea that vitamin D may play an essential role through its ability to maintain both the redox and Ca²⁺

signalling pathways as described earlier [1,2]. A decline in the activity of the vitamin D/klotho/Nrf2 regulatory network has been linked to many diseases. Roselli & Caroni [69] have emphasized the importance of studying the early preclinical phases of neurodegenerative diseases. AD is a case in point in that the preclinical phase can last for many years before the disease is diagnosed. The following conceptual framework attempts to explain what might drive the early preclinical disease development and how this may be related to the ageing process. The basic idea is that there is a slow but progressive dysregulation of the Ca²⁺ and redox signalling pathways resulting from a deficiency in vitamin D [1,2]. It will be argued that this dysregulation results in an alteration in the modulatory activity of the InsP₃/Ca²⁺ signalling pathway, and this creates subtle alterations in the normal cellular signalling pathway resulting in the onset of disease. To understand why such subtle alterations occur can lead to various disease states, it is important to consider the way the Ca²⁺ signalling system is organized in each specific cell type to provide either primary or modulatory signals.

(a) Cardiovascular disease

Vitamin D deficiency has been linked to hypertension and cardiovascular disease [70-76]. The ability of vitamin D to protect the cardiovascular system may depend on its ability to maintain the stability of the ROS and Ca²⁺ signalling systems, which are known to be dysregulated in hypertension, cardiac hypertrophy, congestive heart failure (CHF) and atrial arrhythmias.

One of the main causes of cardiac hypertrophy and CHF is hypertension. The renin-angiotensin system (RAS) plays a major role in regulating blood pressure. One of the primary actions of vitamin D is to curb RAS to prevent the hypertension that is a major risk factor for heart disease [77]. Vitamin D regulates the secretion of renin by renin-producing granular cells, which is controlled by the cyclic AMP signalling pathway. Vitamin D acts by preventing the cyclic AMP response element-binding protein (CREB) from binding to the renin gene promotor [78]. In mice, deletion of either the enzyme 25(OH)D 1α hydroxylase or the VDR resulted in an increase in the renin-angiotensin system, hypertension and the onset of cardiac hypertrophy [79-81]

In patients with type 2 diabetes, the associated hypertension was improved following vitamin D supplementation [82]. The excessive release of renin and the resulting increase in angiotensin II can have multiple effects on some of the key components of the cardiovascular system. One of the actions of angiotensin II is to increase the formation of endothelin-1 (ET-1), which is a potent vasoconstrictor and thus contributes to angiotensin II-induced hypertension [83,84].

The changes in Ca²⁺ signalling in ventricular cardiac cells, which result in hypertrophy and CHF, are relatively minor. There is a small increase in the amplitude of the Ca²⁺ transient that occurs during each heartbeat. This amplification of each transient is caused by an increase in the activity of InsP₃/Ca²⁺ modulatory signalling pathway, which is driven by the increased levels of angiotensin II and ET-1 [85]. In the presence of these two hormones, there are subtle changes in the spatial properties of the individual Ca²⁺ transients. It was proposed that the increase in InsP₃ acts on perinuclear InsP₃R2s to create a nuclear Ca²⁺ signal

responsible for driving the transcriptional processes that initiate hypertrophy [86] (figure 3). There is now considerable experimental evidence to show that activation of InsP₃Rs can indeed function to induce the nuclear Ca2+ transients that activate the transcriptional events responsible for the onset of hypertrophy [87-93]. One of the genes that is activated is ITPR2 that codes for the InsP₃R2 that is responsible for the nuclear Ca²⁺ signal that drives hypertrophy [94]. In cardiomyoctes, miR-133a acts to inhibit the expression of InsP₃R2 [95]. Down-regulation of miR-133a accounts for an increase in the level of the InsP₃R2s, and this is a major contributory factor for the onset of cardiac hypertrophy.

Vitamin D deficiency contributes to the onset of hypertrophy by increasing the Ca²⁺ and redox signalling pathways. For example, there is a decrease in the expression of both SERCA and phospholamban (PLN) that contributes to an increased Ca²⁺ transient amplitude and a decline in the recovery phase [96]. Vitamin D deficiency will also result in an increase in ROS levels that then enhances the Ca²⁺ signalling events that initiate the processes of hypertrophy that results in CHF [97,98]. The angiotensin II and ET-1 not only act to increase Ca2+, but they also increase ROS levels by stimulating NOX at the plasma membrane [99-101] (figure 3). ROS acts by increasing the activity of the ion channels (Na_V1.5 sodium channel, Ca_V1.2 channels and RYR2) and pumps (SERCA) that contribute to the Ca²⁺cycling events that occur during each heartbeat. In addition, ROS can also act indirectly by increasing the activity of protein kinases such as PKA and CaMKIIδ_c that act normally to regulate cardiac activity [101]. These increased ROS and Ca²⁺ signalling processes contribute to the alterations in gene transcription that result in hypertrophy [98]. The cardiac hypertrophy in spontaneously hypertensive rats is reduced by vitamin D [102], and vitamin D supplementation can also markedly improve the outcome of patients suffering from heart failure [74,103].

(b) Alzheimer's disease

AD is another example of a major human disease where the initial change is so subtle that it can go undiagnosed for long periods. The initial symptoms are a decline in working memory, which closely resemble those that occur in ageing as described earlier. The onset of AD depends on the accumulation of extracellular β -amyloid (A β) deposits that disrupt neuronal signalling pathways to reduce cognition. The Ca²⁺ hypothesis considers that the loss of memory depends on an up-regulation of neuronal Ca²⁺ signalling [104–109]. When Ca²⁺ is measured in the spines and dendrites of cortical pyramidal neurons of transgenic mice, there was a higher than normal resting level in those neurons located close to amyloid deposits [110]. Similarly, the resting level of Ca^{2+} in the cortical neurons of 3xTg-AD animals was 247 nmol l⁻¹, which was twice that found in the non-Tg controls (110 nmol l^{-1}) [111]. Such evidence of a persistent elevation in the resting level of Ca²⁺ led to the suggestion that it may continuously activate LTD to explain why memories are erased shortly after they are formed [112,113].

The relatively small elevation in the resting level of Ca²⁺ does not alter the overall function of the brain. Information from the sensory organs can still be processed, new memories can be formed, but they are not retained because the persistent elevation in Ca²⁺ erases them shortly after they are formed. A number of mechanisms have been proposed to explain the

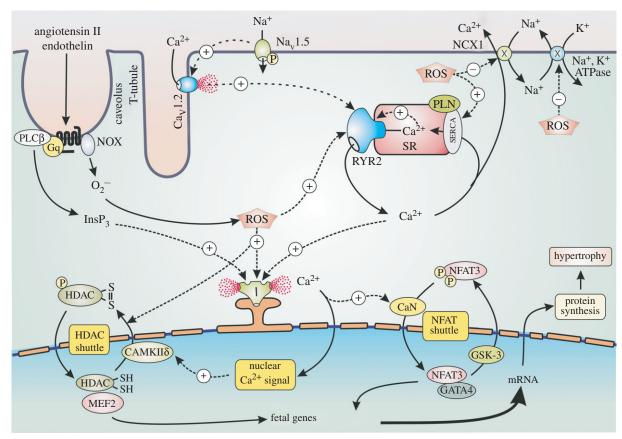


Figure 3. The role of enhanced Ca^{2+} and ROS levels in cardiac hypertrophy. A number of signalling pathways have been implicated in the activation of hypertrophy. A major pathway is induced by angiotensin II and endothelin that stimulate the formation of $InsP_3$ that triggers a nuclear Ca^{2+} signal that activates the HDAC and NFAT shuttles to stimulate the transcription factors responsible for switching on the fetal genes that induce hypertrophy. These hormones also activate NOX to form reactive oxygen species (ROS) that contributes to hypertrophy by enhancing the sensitivity of $InsP_3R$ (I) and ryanodine receptor 2 (RYR2).

elevation of intracellular Ca^{2+} levels by the $A\beta$ protein [114,115]. Many of these mechanisms depend on the InsP₃/ Ca^{2+} signalling pathway [116–119]. A β can bind to the cellular prion protein (PrPC), which is coupled to mGluR5 that increases InsP₃ formation and Ca²⁺ release [117] (figure 4). The formation of InsP₃ is also increased by Aβ acting on the calcium-sensing receptor (CaSR) [118]. Phospholipase Cη1 (PLC η 1), which is activated by Ca²⁺, may contribute to the dysregulation of Ca2+ signalling by amplifying these Aβ-dependent elevations in Ca^{2+} [119]. Activation of the mGluR5 receptor by the $A\beta$ protein has been shown to enhance the process of LTD responsible for memory loss [120]. The significance of InsP₃R activation in the pathogenesis of AD has also emerged from studies on the effects of presenilin mutations. In familial Alzheimer's disease (FAD), presenilin mutations enhance the activity of InsP₃Rs resulting in an increase in Ca²⁺ signalling in both human cells and mouse neurons [121,122]. In a mouse model of AD, which had mutations in presenilin, the AD symptoms were reversed following a reduction in the expression of the InsP₃R, thus supporting the notion that the InsP₃/Ca²⁺ signalling pathway plays a significant role in disease pathogenesis [122].

There are an increasing number of studies indicating that a deficiency in vitamin D may contribute to the onset of AD [123–128]. The level of vitamin D in AD patients is lower than that in controls [129]. Enhanced dietary vitamin D intake lowered the risk of developing AD in a study of older women [130]. VDR polymorphisms have been associated with age-related decline in cognition and are also a

risk factor for AD [126,131,132]. Since AD seems to be caused by abnormal elevations in Ca^{2+} , it is reasonable to propose that the deleterious effect of vitamin D deficiency may be explained by a decrease in its normal role as a custodian of Ca^{2+} and ROS homeostasis. Similarly, a decrease in ROS could also contribute to the increase in cognition observed in ageing rats following treatment with the anti-inflammatory drug montelukast that is used normally to treat asthma [65].

Vitamin D may prevent the onset of AD by regulating a number of processes. Firstly, vitamin D can increase the expression of the multidrug resistance protein 1 (MDR1) gene that codes for the P-glycoprotein (P-gp), which is an efflux transporter that acts to reduce the accumulation of Aβ [133]. Secondly, vitamin D may act to control the expression of those toolkit components responsible for maintaining low ROS and Ca²⁺ levels. For example, vitamin D stimulates the expression of Ca2+ pumps and exchangers (PMCA and NCX) and Ca²⁺ buffers such as calbindin (CB) and parvalbumin (figure 4). The expression of neuronal CB is known to be reduced in AD [134]. Mice expressing mutant APP also display a decline in the level of CB especially in the dentate gyrus region of the hippocampus, which functions in learning and memory [135]. Vitamin D can curb the influx of external Ca2+ by reducing the expression of L-type voltage-sensitive channels, which are markedly elevated in rat hippocampal neurons [66].

Many of the deleterious effects of vitamin D deficiency in AD may depend on a decline in the expression of

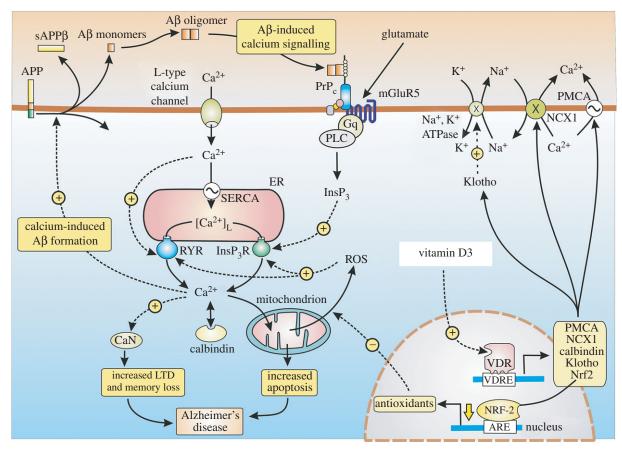


Figure 4. Dysregulation of Ca^{2+} and redox signalling in Alzheimer's disease (AD). The calcium hypothesis of AD suggests that the formation of Aβ oligomers brings about an overall increase in Ca^{2+} signalling that results in a permanent elevation in the resting level of Ca^{2+} to 300 nM that then erases memories soon after they are formed by activating calcineurin (CaN) inducing long-term depression (LTD). An elevation of Ca^{2+} sets up a positive feedback loop by acting to stimulate the hydrolysis of the amyloid precursor protein (APP) to generate the Aβ oligomers that bind to the cellular prion protein (PrP^{C}) that then activates the $Prace{InsP_3/Ca^{2+}}$ signalling pathway. Vitamin D reduces the risk of AD by acting to maintain $Prace{InsP_3/Ca^{2+}}$ and redox signalling at their normal low resting levels.

its two collaborators Nrf2 and klotho. Nrf2 levels are markedly reduced in the brain of patient with AD [49]. Vector-mediated expression of Nrf2 in the hippocampus of AD transgenic mice resulted in a marked improvement in cognition [135]. One of the main functions of Nrf2 is to maintain the cellular level of the redox buffer GSH [136], which is a critical factor in preventing AD [44]. The level of Bcl-2, which inhibits the ability of InsP₃ to activate the InsP₃ receptors [9] and the RYRs [10,11] (figure 1), is maintained by Nrf2 and Klotho, thereby reducing the level of Ca²⁺. The ability of Bcl-2 to reduce the symptoms of AD in transgenic mice [137,138] may be explained by this reduction in the activity of both the InsP₃R and RYRs. Klotho may also play a role in AD, because its levels in the CSF of patients with AD are lower than those in age-matched controls [139]. In the senescence-accelerated mouse prone-8 (SAMP8) mouse, a decline in the expression of klotho has been linked to symptoms of AD, including a decline in cognition and an accumulation of amyloid- β_{1-42} [140].

It is clear that dysregulation of the vitamin D/klotho/Nrf2 regulatory network results in a decline in cell signalling stability that results in the elevated neuronal Ca²⁺ and ROS levels that seem to responsible for the onset of AD. Such a mechanism suggests an interesting explanation for the sporadic nature of AD. Despite it being referred to as an age-related disease, not everyone who ages develops AD. So what is it that triggers the onset of sporadic AD? One possibility is that it is induced in those individuals who are

deficient in vitamin D and thus have abnormally elevated levels of Ca^{2+} that may initiate the formation of the pathological $A\beta$ oligomers [2,3]. This possibility is supported by the fact that Ca^{2+} acts to stimulate the formation of $A\beta$ (figure 4) [57,107,141–145]. Inhibiting the RYR2 with dantrolene that reduces their release of Ca^{2+} was found to markedly reduce the formation of $A\beta$ [145]. Such Ca^{2+} -induced increases in amyloid formation then initiates a positive feedback loop, because it is followed by $A\beta$ -induced Ca^{2+} signalling and it is this $A\beta/Ca^{2+}$ positive feedback loop that may be responsible for the onset of AD [146]. Such a scenario is entirely consistent with the fact the vitamin D deficiency is such a strong risk factor for AD.

6. Conclusion

The phenotypic stability of the interacting Ca²⁺ and ROS signalling pathways is maintained by vitamin D. It is argued that a deficiency in vitamin D results in an elevation in both the ROS and Ca²⁺ signalling pathways that may contribute to the process of ageing. An example of this is the age-related decline in the cognition of rats that can be reversed by administering vitamin D. Such deficiencies in vitamin D may also set the stage for the onset of both heart disease and AD.

Competing interests. I have no competing interests. Funding. I received no funding for this study.

References

- Berridge MJ. 2015 Vitamin D: a custodian of cell signalling stability in health and disease. *Biochem.* Soc. Trans. 43, 349 – 358. (doi:10.1042/ BST20140279)
- Berridge MJ. 2015 Vitamin D cell signalling in health and disease. *Bioch. Biophys. Res. Commun.* 460, 53–71. (doi.org/10.1016/j.bbrc.2015.01.008)
- Berridge MJ. 2015 Vitamin D, cell signalling phenotypic stability and Alzheimer's disease. Austin J. Clin. Neurol. 2, 1033–1036.
- Hidalgo C, Donoso P. 2008 Crosstalk between calcium and redox signaling: from molecular mechanisms to health implications. *Antioxid. Redox Signal.* 10, 1275 – 1312. (doi:10.1089/ars. 2007.1886)
- Donoso P, Sanchez G, Bull R, Hidalgo C. 2011 Modulation of cardiac ryanodine receptor activity by ROS and RNS. Front. Biosci. 16, 553-567. (doi:10. 2741/3705)
- Prosser BL, Khairallah RJ, Ziman AP, Ward CW, Lederer WJ. 2013 X-ROS signaling in the heart and skeletal muscle: stretch-dependent local ROS regulates [Ca²⁺](i). *J. Mol. Cell Cardiol.* 58, 172–181. (doi:10.1016/j.yjmcc.2012.11.011)
- Hidalgo C. 2005 Cross talk between Ca²⁺ and redox signalling cascades in muscle and neurons through the combined activation of ryanodine receptors/ Ca²⁺ release channels. *Phil. Trans. R. Soc. B* 360, 2237 – 2246. (doi:10.1098/rstb.2005.1759)
- Bánsághi S, Golenár T, Madesh M, Csordás G, RamachandraRao S, Sharma K, Yule DI, Joseph SK, Hajnóczky G. 2014 Isoform- and species-specific control of inositol 1,4,5-trisphosphate (IP₃) receptors by reactive oxygen species. J. Biol. Chem. 289, 8170–8181. (doi:10.1074/jbc.M113.504159)
- Rong YP, Distelhorst CW. 2008 Bcl-2 protein family: versatile regulators of calcium signaling in cell survival and apoptosis. *Annu. Rev. Physiol.* 70, 73 – 91. (doi:10.1146/annurev.physiol.70.021507.105852)
- Vervliet T et al. 2014 Bcl-2 binds to and inhibits ryanodine receptors. J. Cell Sci. 127, 2782 – 2792. (doi:10.1242/jcs.150011)
- Vervliet T, Parys JB, Bultynck G. 2015 Bcl-2 and FKBP12 bind to IP3 and ryanodine receptors at overlapping sites: the complexity of protein-protein interactions for channel regulation. *Biochem. Soc. Trans.* 43, 396–404. (doi:10.1042/BST20140298)
- Hildeman DA, Mitchell T, Aronow B, Wojciechowski S, Kappler J, Marrack P. 2003 Control of Bcl-2 expression by reactive oxygen species. *Proc. Natl Acad. Sci. USA* 100, 15 035 – 15 040. (doi:10.1073/pnas.1936213100)
- Kozai D, Ogawa N, Mori Y. 2014 Redox regulation of transient receptor potential channels. *Antioxid*. *Redox Signal*. 21, 971–986. (doi:10.1089/ars. 2013.5616)
- Dusso AS. 2014 Update on the biologic role of vitamin D on the endocrine system. *Curr. Vasc. Pharmacol.* 12, 272–277. (doi:10.2174/15701611113119990026)

- Kaspar JW, Niture SK, Jaiswal AK. 2009 Nrf2:INrf2 (Keap1) signaling in oxidative stress. Free Radic. Biol. Med. 47, 1304–1309. (doi:10.1016/j. freeradbiomed.2009.07.035)
- Hayes JD, Dinkova-Kostova AT. 2014 The Nrf2 regulatory network provides an interface between redox and intermediary metabolism. *Trends Biochem. Sci.* 39, 199–218. (doi:10.1016/j.tibs. 2014.02.002)
- Høyer-Hansen M, Nordbrandt SP, Jäättelä M. 2010 Autophagy as a basis for the health-promoting effects of vitamin D. *Trends Mol. Med.* 16, 295 – 302. (doi:10.1016/j.molmed.2010.04.005)
- Uberti F, Lattuada D, Morsanuto V, Nava U, Bolis G, Vacca G, Squarzanti DF, Cisari C, Molinari C. 2014 Vitamin D protects human endothelial cells from oxidative stress through the autophagic and survival pathways. J. Clin. Endocrinol. Metab. 99, 1367 – 1374. (doi:10.1210/jc.2013-2103)
- Galluzzi L, Pietrocola F, Levine B, Kroemer G.
 2014 Metabolic control of autophagy. *Cell* **159**, 1263 – 1276. (doi:10.1016/j.cell.2014.11.006)
- Høyer-Hansen M et al. 2007 Control of macroautophagy by calcium, calmodulin-dependent kinase kinase-beta, and Bcl-2. Mol. Cell. 25, 193 – 205. (doi:10.1016/j.molcel.2006.12.009)
- Cárdenas C. 2010 Essential regulation of cell bioenergetics by constitutive InsP3 receptor Ca²⁺ transfer to mitochondria. *Cell* 142, 270–283. (doi:10.1016/j.cell.2010.06.007)
- Harr MW, Distelhorst CW. 2010 Apoptosis and autophagy: decoding calcium signals that mediate life or death. *Cold Spring Harb. Perspect. Biol.* 2, a005579. (doi:10.1101/cshperspect.a005579)
- Decuypere JP, Bultynck G, Parys JB. 2011 A dual role for Ca²⁺ in autophagy regulation. *Cell Calcium* 50, 242 – 250. (doi:10.1016/j.ceca.2011.04.001)
- 24. Fetahu IS, Höbaus J, Kállay E. 2014 Vitamin D and the epigenome. *Front. Physiol.* **5**, 164. (doi:10.3389/fphys.2014.00164)
- Saccone D, Asani F, Bornman L. 2015 Regulation of the vitamin D receptor gene by environment, genetics and epigenetics. *Gene* 561, 171–180. (doi:10.1016/j.gene.2015.02.024)
- Kao YH, Cheng CC, Chen YC, Chung CC, Lee TI, Chen SA, Chen YJ. 2011 Hydralazine-induced promoter demethylation enhances sarcoplasmic reticulum Ca²⁺-ATPase and calcium homeostasis in cardiac myocytes. *Lab. Invest.* 91, 1291–1297. (doi:10. 1038/labinvest.2011.92)
- King GD, Rosene DL, Abraham CR. 2011 Promoter methylation and age-related downregulation of Klotho in rhesus monkey. *Age* 34, 1405 – 1419. (doi:10.1007/s11357-011-9315-4)
- Lee J et al. 2010 Theranti-aging gene KLOTHO is a novel target for epigenetic silencing in human cervical carcinoma. Mol. Cancer 9, 109 – 119. (doi:10.1186/1476-4598-9-109)
- 29. van Otterdijk SD, Mathers LC, Strathdee G. 2013 Do age-related changes in DNA methylation play a role

- in the development of age-related diseases? *Biochem. Soc. Trans.* **41**, 803 807. (doi:10.1042/BST20120358)
- Guidotti AJ et al. 2011 Epigenetic GABAergic targets in schizophrenia and bipolar disorder. Neuropharmacology 60, 1007 – 1016. (doi:10.1016/j. neuropharm.2010.10.021)
- Pereira F, Barbáchano A, Singh PK, Campbell MJ, Muñoz A, Larriba MJ. 2012 Vitamin D has wide regulatory effects on histone demethylase genes. *Cell Cycle* 11, 1081–1089. (doi:10.4161/ cc.11.6.19508)
- Przybelski RJ, Binkley NC. 2007 Is vitamin D important for preserving cognition? A positive correlation of serum 25-hydroxyvitamin D concentration with cognitive function. *Arch. Biochem. Biophys.* 460, 202–205. (doi:10.1016/j. abb.2006.12.018)
- Kuningas M, Mooijaart SP, Jolles J, Slagboom PE, Westendorp RG, van Heemst D. 2009 VDR gene variants associate with cognitive function and depressive symptoms in old age. *Neurobiol. Aging*. 30, 466–473. (doi:10.1016/j.neurobiolaging.2007. 07.001)
- Beydoun MA, Ding EL, Beydoun HA, Tanaka T, Ferrucci L, Zonderman AB. 2012 Vitamin D receptor and megalin gene polymorphisms and their associations with longitudinal cognitive change in US adults. Am. J. Clin. Nutr. 95, 163 – 178. (doi:10. 3945/ajcn.111.017137)
- Schlögl M, Holick MF. 2014 Vitamin D and neurocognitive function. *Clin. Interv. Aging* 9, 559–568. (doi:10.2147/CIA.S51785)
- MacLaughlin J, Holick MF. 1985 Aging decreases the capacity of human skin to produce vitamin D3.
 J. Clin. Invest. 76, 1536 – 1538. (doi:10.1172/ JCl112134)
- Lewis KN, Mele J, Hayes JD, Buffenstein R. 2010
 Nrf2, a guardian of healthspan and gatekeeper of species longevity. *Integr. Comp. Biol.* 50, 829 – 843. (doi:10.1093/icb/icq034)
- Ureshino RP, Bertoncini CR, Fernandes MJ, Abdalla FM, Porto CS, Hsu YT, Lopes GS, Smaili SS. 2010 Alterations in calcium signaling and a decrease in Bcl-2 expression: possible correlation with apoptosis in aged striatum. *J. Neurosci. Res.* 88, 438 – 447. (doi:10.1002/jnr.22214)
- Decuypere J-P, Monaco G, Missiaen L, De Smedt H, Parys JB, Bultynck G. 2011 IP₃ receptors, mitochondria, and Ca²⁺ signaling: implications for aging. J. Aging Res. 2011, 920178. (doi:10.4061/ 2011/920178)
- Harman D. 1956 Aging: a theory based on free radical and radiation chemistry. *J. Gerontol.* 11, 298–300. (doi:10.1093/qeronj/11.3.298)
- 41. Finkel T, Holbrook NJ. 2000 Oxidants, oxidative stress and the biology of ageing. *Nature* **408**, 239–247. (doi:10.1038/35041687)
- 42. Khachaturian ZS. 1989 The role of calcium regulation in brain aging: reexamination of a

- hypothesis. *Aging (Milano)* **1**, 17 34. (doi:10.1007/BF03323872)
- Ureshino RP, Rocha KK, Lopes GS, Trindade CB, Smaili SS. 2014 Calcium signaling alterations, oxidative stress, and autophagy in aging. *Antioxid. Redox Signal.* 21, 123 – 137. (doi:10.1089/ ars.2013.5777)
- 44. Ghosh D, LeVault KR, Brewer GJ. 2014 Dual-energy precursor and nuclear erythroid-related factor 2 activator treatment additively improve redox glutathione levels and neuron survival in aging and Alzheimer mouse neurons upstream of reactive oxygen species. *Neurobiol. Aging* 35, 179–190. (doi:10.1016/j.neurobiolaging.2013.06.023)
- Ghosh D, LeVault KR, Brewer GJ. 2014 Relative importance of redox buffers GSH and NAD(P)H in agerelated neurodegeneration and Alzheimer disease-like mouse neurons. *Aging Cell* 13, 631–640. (doi:10.1111/acel.12216)
- Ghosh D, LeVault KR, Barnett AJ, Brewer GJ. 2012 A reversible early oxidized redox state that precedes macromolecular ROS damage in aging nontransgenic and 3xTg-AD mouse neurons.
 J. Neurosci. 32, 5821 5832. (doi:10.1523/JNEUROSCI.6192-11.2012)
- Steullet P, Cabungcal JH, Kulak A, Kraftsik R, Chen Y, Dalton TP, Cuenod M, Do KQ. 2010 Redox dysregulation affects the ventral but not dorsal hippocampus: impairment of parvalbumin neurons, gamma oscillations, and related behaviors. J. Neurosci. 30, 2547 – 2558. (doi:10.1523/ JNEUROSCI.3857-09.2010)
- 48. Nakai K *et al.* 2014 Vitamin D activates the Nrf2-Keap1 antioxidant pathway and ameliorates nephropathy in diabetic rats. *Am. J. Hypertens.* **27**, 586–595. (doi:10.1093/ajh/hpt160)
- Ramsey CP, Glass CA, Montgomery MB, Lindl KA, Ritson GP, Chia LA, Hamilton RL, Chu CT, Jordan-Sciutto KL. 2007 Expression of Nrf2 in neurodegenerative diseases. *J. Neuropathol. Exp.* Neurol. 66, 75–85. (doi:10.1097/nen. 0b013e31802d6da9)
- Keisala TMA, Lou YR, Zou J, Kalueff AV, Pyykkö I, Tuohimaa P. 2009 Premature aging in vitamin D receptor mutant mice. *J. Steroid Biochem. Mol. Biol.* 115, 91–97. (doi:10.1016/j.jsbmb.2009.03.007)
- Toescu EC, Verkhratsky A. 2007 The importance of being subtle: small changes in calcium homeostasis control cognitive decline in normal aging. *Aging Cell* 6, 267 – 273. (doi:10.1111/j.1474-9726.2007.00296)
- 52. Foster TC. 2007 Calcium homeostasis and modulation of synaptic plasticity in the aged brain. *Aging Cell* **6**, 319–325. (doi:10.1111/j.1474-9726. 2007.00283.x)
- 53. Mattson MP. 2007 Calcium and neurodegeneration. *Aging Cell* **6**, 337 350. (doi:10.1111/j.1474-9726. 2007.00275)
- 54. Gibson GE, Peterson C. 1987 Calcium and the aging nervous system. *Neurobiol. Aging* **8**, 329 343. (doi:10.1016/0197-4580(87)90072-8)
- Landfield PW. 1987 'Increased calcium-current' hypothesis of brain aging. *Neurobiol. Aging* 8, 346–347. (doi:10.1016/0197-4580(87)90074-1)

- Kirischuk S, Verkhratsky A. 1996 Calcium homeostasis in aged neurones. *Life Sci.* 59, 451–459. (doi:10.1016/0024-3205(96)00324-4)
- 57. Thibault O, Gant JC, Landfield PW. 2007 Expansion of the calcium hypothesis of brain aging and Alzheimer's disease: minding the store. *Aging Cell* **6**, 307 317. (doi:10.1111/j.1474-9726.2007.00295.x)
- Zanos P, Bhat S, Terrillion CE, Smith RJ, Tonelli LH, Gould TD. 2015 Sex-dependent modulation of agerelated cognitive decline by the L-type calcium channel gene Cacna1c (Cav 1.2). Eur. J. Neurosci. 42, 2499 – 2507. (doi:10.1111/ejn.12952)
- Zaidi A, Gao J, Squier TC, Michaelis ML. 1998 Agerelated decrease in brain synaptic membrane Ca²⁺-ATPase in F344/BNF1 rats. *Neurobiol. Aging* 19, 487 – 495. (doi:10.1016/S0197-4580(98)00078-5)
- Tonkikh A, Janus C, El-Beheiry H, Pennefather PS, Samoilova M, McDonald P, Ouanounou A, Carlen PL. 2006 Calcium chelation improves spatial learning and synaptic plasticity in aged rats. *Exp. Neurol.* 197, 291–300. (doi:10.1016/j.expneurol. 2005.06.014)
- Disterhoft J, Oh M. 2007 Alterations in intrinsic neuronal excitability during normal aging. *Aging Cell* 6, 327–336. (doi:10.1111/j.1474-9726.2007. 00297.x)
- 62. Serrano F, Klann E. 2004 Reactive oxygen species and synaptic plasticity in the aging hippocampus. *Ageing Res. Rev.* **3**, 431–443. (doi:10.1016/j.arr. 2004.05.002)
- Watson JB, Arnold MM, Ho YS, O'Dell TJ. 2006 Agedependent modulation of hippocampal long-term potentiation by antioxidant enzymes. *J. Neurosci.* Res. 84, 1564–1574. (doi:10.1002/jnr.21040)
- Bodhinathan K, Kumar A, Foster TC. 2010 Redox sensitive calcium stores underlie enhanced after hyperpolarization of aged neurons: role for ryanodine receptor mediated calcium signaling. *J. Neurophysiol.* 104, 2586–2593. (doi:10.1152/jn. 00577.2010)
- Marschallinger J et al. 2015 Structural and functional rejuvenation of the aged brain by an approved anti-asthmatic drug. Nat. Commun. 6, 8466. (doi:10.1038/ncomms9466)
- 66. Brewer LD, Thibault V, Chen KC, Langub MC, Landfield PW, Porter NM. 2001 Vitamin D hormone confers neuroprotection in parallel with downregulation of L-type Ca²⁺ channel expression in hippocampal neurons. *J. Neurosci*. 21, 98–108.
- Brewer LD, Porter NM, Kerr DS, Landfield PW, Thibault 0. 2006 Chronic 1α,25-(0H)₂vitamin D₃ treatment reduces Ca²⁺-mediated hippocampal biomarkers of aging. *Cell Calcium* 40, 277 – 286. (doi:10.1016/j.ceca.2006.04.001)
- Latimer CS et al. 2014 Vitamin D prevents cognitive decline and enhances hippocampal synaptic function in aging rats. Proc. Natl Acad. Sci. USA 111, E4359 – E4366. (doi:10.1073/pnas.1404477111)
- Roselli F, Caroni P. 2015 From intrinsic firing properties to selective neuronal vulnerability in neurodegenerative diseases. *Neuron* 85, 901–910. (doi:10.1016/j.neuron.2014.12.063)

- Scragg R, Jackso R, Holdaway IM, Lim T, Beaglehole R. 1990 Myocardial infarction is inversely associated with plasma 25-hydroxyvitamin D3 levels: a community-based study. *Int. J. Epidemiol.* 19, 559–563. (doi:10.1093/ije/19.3.559)
- Heaney RH. 2008 Vitamin D in Health and Disease. *Clin. J. Am. Soc. Nephrol.* 3, 1535 – 1541. (doi:10. 2215/CJN.01160308)
- Kim DH, Sabour S, Sagar UN, Adams S, Whellan DJ. 2008 Prevalence of hypovitaminosis D in cardiovascular diseases (from the National Health and Nutrition Examination Survey 2001 to 2004).
 Am. J. Cardiol. 102, 1540 – 1544. (doi:10.1016/j. amjcard.2008.06.067)
- Anderson JL, May HT, Horne BD, Bair TL, Hall NL, Carlquist JF, Lappé DL, Muhlestein JB. 2010 Relation of vitamin D deficiency to cardiovascular risk factors, disease status, and incident events in a general healthcare population. *Am. J. Cardiol.* 106, 963 – 968. (doi:10.1016/j.amjcard.2010.05.027)
- Vacek JL, Vanga SR, Good M, Lai SM, Lakkireddy D, Howard PA. 2012 Vitamin D deficiency and supplementation and relation to cardiovascular health. Am. J. Cardiol. 109, 359 – 363. (doi:10.1016/ j.amjcard.2011.09.020)
- Hossein-nezhad A, Holick MF. 2013 Vitamin D for health: a global perspective. *Mayo Clin. Proc.* 88, 720–755. (doi:10.1016/j.mayocp. 2013.05.011)
- 76. Dong J, Lau CW, Wong SL, Huang Y. 2014 Cardiovascular benefits of vitamin D. *Acta Physiologica Sinica* **66**, 30–36. (doi:10.13294/ j.aps.2014.0005)
- Weng S, Sprague JE, Oh J, Riek AE, Chin K, Garcia M, Bernal-Mizrachi C. 2013 Vitamin D deficiency induces high blood pressure and accelerates atherosclerosis in mice. *PLoS ONE* 8, e54625. (doi:10.1371/journal.pone.0054625)
- Yuan W et al. 2007 1,25-Dihydroxyvitamin D3 suppresses renin gene transcription by blocking the activity of the cyclic AMP response element in the renin gene promoter. J. Biol. Chem. 282, 29 821— 29 830. (doi:10.1074/jbc.M705495200)
- Li YC, Kong J, Wei M, Chen Z-F, Liu SQ, Cao L-P. 2002 1,25-Dihydroxyvitamin D(3) is a negative endocrine regulator of the renin—angiotensin system. J. Clin. Invest. 110, 229—238. (doi:10.1172/ JCI15219)
- 80. Xiang W *et al.* 2005 Cardiac hypertrophy in vitamin D receptor knockout mice: role of the systemic and cardiac renin—angiotensin systems. *Am. J. Physiol. Endocrinol. Metab.* **288**, E125—E132. (doi:10.1152/ajpendo.00224.2004)
- 81. Zhou C, Lu F, Cao K, Xu D, Goltzman D, Miao D. 2008 Calcium-independent and 1,25(0H)2D3-dependent regulation of the renin-angiotensin system in 1alpha-hydroxylase knockout mice. *Kidney Int.* **74**, 170 179. (doi:10.1038/ki.2008.101)
- 82. Nasri H, Behradmanesh S, Ahmadi A, Rafieian-Kopaei M. 2014 Impact of oral vitamin D (cholecalciferol) replacement therapy on blood pressure in type 2 diabetes patients; a randomized, double-blind, placebo controlled clinical trial.

- *J. Nephropathol.* **3**, 29–33. (doi:10.12860/jnp. 2014.07)
- 83. Romero JC, Reckelhoff JF. 1999 Role of angiotensin and oxidative stress in essential hypertension. *Hypertension* **34**, 943 – 949. (doi:10.1161/01.HYP. 34.4.943)
- Ortiz MC, Manriquez MC, Romero JC, Juncos JA.
 2001 Antioxidants block angiotensin II-induced increases in blood pressure and endothelin.
 Hypertension 38, 655 659. (doi:10.1161/01.HYP.
 38.3.655)
- Berridge MJ. 2012 Calcium signalling remodelling and disease. *Biochem. Soc. Trans.* 40, 297 – 309. (doi:10.1042/BST20110766)
- 86. Berridge MJ. 2006 Remodelling Ca²⁺ signalling systems and cardiac hypertrophy. *Biochem. Soc. Trans.* **34**, 228–231. (doi:10.1042/BST0340228)
- Lipp P, Laine M, Tovey SC, Burrell KM, Berridge MJ, Li W, Bootman MD. 2000 Functional InsP₃ receptors that may modulate excitation-contraction coupling in the heart. *Curr. Biol.* 10, 939–942. (doi:10.1016/ S0960-9822(00)00624-2)
- 88. Wu X *et al.* 2006 Local InsP₃-dependent perinuclear Ca²⁺ signaling in cardiac myocyte excitation transcription coupling. *J. Clin. Invest.* **116**, 675 682. (doi:10.1172/JCI27374)
- 89. Molkentin JD. 2006 Dichotomy of Ca²⁺ in the heart: contraction versus intracellular signaling. *J. Clin. Invest.* **116**, 623–626. (doi:10.1172/JCl27824)
- Luo D et al. 2008 Nuclear Ca²⁺ sparks and waves mediated by inositol 1,4,5-trisphosphate receptors in neonatal rat cardiomyocytes. *Cell Calcium* 43, 165 174. (doi:10.1016/j.ceca. 2007.04.017)
- Harzheim D, Movassagh M, Foo RS, Ritter O, Tashfeen A, Conway SJ, Bootman MD, Roderick HL. 2009 Increased InsP₃Rs in the junctional sarcoplasmic reticulum augment Ca²⁺ transients and arrhythmias associated with cardiac hypertrophy. *Proc. Natl Acad. Sci. USA* **106**, 11 406 – 11 411. (doi:10.1073/pnas.0905485106)
- Higazi DR et al. 2009 Endothelin-1-stimulated InsP₃-induced Ca²⁺ release is a nexus for hypertrophic signalling in cardiac myocytes. Mol. Cell 33, 472–482. (doi:10.1016/j.molcel.2009. 02.005)
- 93. Nakayama H *et al.* 2010 The IP₃ receptor regulates cardiac hypertrophy in response to select stimuli. *Circ. Res.* **107**, 659–666. (doi:10.1161/CIRCRESAHA. 110.220038)
- 94. Sankar N, deTombe PP, Mignery GA. 2014
 Calcineurin-NFATc regulates type 2 inositol 1,4,5trisphosphate receptor (InsP3R2) expression during
 cardiac remodeling. *J. Biol. Chem.* **289**, 188–198.
 (doi:10.1074/jbc.M113.495242)
- 95. Drawnel FM *et al.* 2012 Mutual antagonism between IP3RII and miRNA-133a regulates calcium signals and cardiac hypertrophy. *J. Cell Biol.* **199**, 783 798. (doi:10.1083/jcb.201111095)
- 96. Choudhury S, Bae S, Ke QJY, Lee JY, Singh SS, St-Arnaud R, del Monte F, Kang PM. 2014 Abnormal calcium handling and exaggerated cardiac dysfunction in mice with defective Vitamin D

- signaling. *PLoS ONE* **9**, e108382. (doi:10.1371/journal.pone.0108382)
- 97. Sag CM, Santos CX, Shah AM. 2014 Redox regulation of cardiac hypertrophy. *J. Mol. Cell Cardiol.* **73**, 103–111. (doi:10.1016/j.yjmcc.2014. 02.002)
- 98. Köhler AC, Sag CM, Maier LS. 2014 Reactive oxygen species and excitation-contraction coupling in the context of cardiac pathology. *J. Mol. Cell Cardiol.* **73**, 92 102. (doi:10.1016/j.yjmcc.2014.03.001)
- Bendall JK, Cave AC, Heymes C, Gall N, Shah AM.
 2002 Pivotal role of a gp91(phox)-containing NADPH oxidase in angiotensin II-induced cardiac hypertrophy in mice. *Circulation*. **105**, 293–296. (doi:10.1161/hc0302.103712)
- 100. Li JM, Gall NP, Grieve DJ, Chen M, Shah AM. 2002 Activation of NADPH oxidase during progression of cardiac hypertrophy to failure. *Hypertension* 40, 477 – 484. (doi:10.1161/01.HYP. 0000032031.30374.32)
- Burgoyne JR, Mongue-Din H, Eaton P, Shah AJ.
 Redox signalling in cardiac physiology and pathology. *Circ. Res.* 111, 1091 – 1106. (doi:10. 1161/CIRCRESAHA.111.255216)
- 102. Mancuso P, Rahman A, Hershey SD, Dandu L, Nibbelink KA, Simpson RU. 2008 1,25-dihydroxyvitamin-D₃treatment reduces cardiac hypertrophy and left ventricular diameter in spontaneously hypertensive heart failure-prone (cp/+) rats independent of changes in serum leptin. *J. Cardiovasc. Pharm.* **51**, 559−564. (doi:10.1097/FJC.0b013e3181761906)
- 103. Gotsman I, Shauer A, Zwas DR, Hellman Y, Keren A, Lotan C, Admon D. 2012 Vitamin D deficiency is a predictor of reduced survival in patients with heart failure; vitamin D supplementation improves outcome. Eur. J. Heart Fail. 14, 357—366. (doi:10. 1093/eurjhf/hfr175)
- 104. Khachaturian ZS. 1989 Calcium, membranes, aging, and Alzheimer's disease. Introduction and overview. Ann. N Y Acad. Sci. **568**, 1–4. (doi:10.1111/j.1749-6632.1989.tb12485.x)
- LaFerla FM. 2002 Calcium dyshomeostasis and intracellular signaling in Alzheimer's disease. *Nat. Rev. Neurosci.* 3, 862–872. (doi:10.1038/nrn960)
- 106. Stutzmann GE. 2007 The pathogenesis of Alzheimer's disease is it a lifelong 'calciumopathy'. Neuroscientist 13, 546–559. (doi:10.1177/1073858407299730)
- 107. Bezprozvanny I, Mattson MP. 2008 Neuronal calcium mishandling and the pathogenesis of Alzheimer's disease. *Trends Neurosci.* **31**, 454–463. (doi:10.1016/j.tins.2008.06.005)
- Bojarski L, Herms J, Kuznicki J. 2008 Calcium dysregulation in Alzheimer's disease.
 Neurochem. Int. 52, 621 – 633. (doi:10.1016/j. neuint.2007.10.002)
- 109. Stutzmann GE, Mattson MP. 2011 Endoplasmic reticulum Ca²⁺ handling in cells in health and disease. *Pharmacol. Rev.* **63**, 700–727. (doi:10. 1124/pr.110.003814)
- 110. Kuchibhotla KV, Goldman ST, Lattarulo CR, Wu H-Y, Hyman BT, Bacskai BJA. 2008 Abeta plaques lead to

- aberrant regulation of calcium homeostasis in vivo resulting in structural and functional disruption of neuronal networks. *Neuron* **59**, 214–225. (doi:10. 1016/j.neuron.2008.06.008)
- 111. Lopez JR, Lyckman A, Oddo S, Laferla FM, Querfurth HW, Shtifman A. 2008 Increased intraneuronal resting [Ca²⁺] in adult Alzheimer's disease mice. J. Neurochem. **105**, 262–271. (doi:10.1111/j.1471-4159.2007.05135.x)
- 112. Berridge MJ. 2012 Dysregulation of neural calcium signalling in Alzheimer disease, bipolar disorder and schizophrenia. *Prion* **6**, 1–12. (doi:10. 4161/pri.21767)
- Berridge MJ. 2014 Calcium regulation of neural rhythms, memory and Alzheimer's disease.
 J. Physiol. 592, 281 – 293. (doi:10.1113/jphysiol. 2013.257527)
- 114. Jensen LE, Bultynck G, Luyten T, Amijee H, Bootman MD, Roderick HL. 2013 Alzheimer's disease-associated peptide Aβ42 mobilizes ER Ca²⁺ via InsP₃R-dependent and -independent mechanisms. *Front. Mol. Neurosci* 6, 36. (doi:10.3389/fnmol. 2013.00036)
- 115. Brawek B, Garaschuk O. 2014 Network-wide dysregulation of calcium homeostasis in Alzheimer's disease. *Cell Tissue Res.* **357**, 427 438. (doi:10. 1007/s00441-014-1798-8)
- 116. Demoro A, Parker I. 2013 Cytotoxicity of intracellular Aβ42 amyloid oligomers Involves Ca²⁺ release from the endoplasmic reticulum by stimulated production of inositol trisphosphate. *J. Neurosci.* 33, 3824–3833. (doi:10.1523/JNEUROSCI.4367-12.2013)
- 117. Um JW et al. 2013 Metabotropic glutamate receptor 5 is a coreceptor for Alzheimer abeta oligomer bound to cellular prion protein. *Neuron* 79, 887–902. (doi:10.1016/j.neuron.2013.06.036)
- 118. Armato U, Bonafini C, Chakravarthy B, Pacchiana R, Chiarini A, Whitfield JF, Dal Prà I. 2012 The calciumsensing receptor: a novel Alzheimer's disease crucial target? J. Neurol. Sci. 322, 137 – 140. (doi:10.1016/ j.jns.2012.07.031)
- Popovics P, Stewart AL. 2012 Phospholipase C-η activity may contribute to Alzheimer's disease-associated calciumopathy. *J. Alzheimer's Dis.* 30, 737 744. (doi:10.3233/JAD-2012-120241)
- 120. Chen X, Lin R, Chang L, Xu S, Wei X, Zhang J, Wang C, Anwyl C, Wang Q. 2013 Enhancement of long-term depression by soluble amyloid β protein in rat hippocampus is mediated by metabotropic glutamate receptor and involves activation of p38MAPK, STEP and Caspase 3. *Neuroscience* 253, 435 443. (doi:10.1016/j.neuroscience.2013. 08.054)
- 121. Cheung KH, Mei L, Mak DO, Hayashi I, Iwatsubo T, Kang DE, Foskett JK. 2010 Gain-of-function enhancement of IP₃receptor modal gating by familial Alzheimer's disease-linked presenilin mutants in human cells and mouse neurons. *Sci. Signal.* **3**, ra22. (doi:10.1126/scisignal.2000818)
- 122. Shilling D, Müller M, Takano H, Mak DO, Abel T, Coulter DA, Foskett JK. 2014 Suppression of InsP₃ receptor-mediated Ca²⁺ signaling alleviates mutant presenilin-linked familial Alzheimer's disease

- pathogenesis. J Neurosci. 34, 6910-6923. (doi:10. 1523/JNEUROSCI.5441-13.2014)
- 123. Tuohimaa P, Keisala T, Minasyan A, Cachat J, Kalueff A. 2009 Vitamin D nervous system and aging. Psychoneuroendocrinology 34(Suppl. 1), S278-S286. (doi:10.1016/j.psyneuen.2009.07.003)
- 124. Annweiler C, Schott AM, Allali G, Bridenbaugh S, Kressig RW, Allain P, Herrmann FR, Beauchet O. 2010 Association of vitamin D deficiency with cognitive impairment in older women: crosssectional study. Neurology 74, 27-32. (doi:10. 1212/WNL.0b013e3181beecd3)
- 125. Annweiler C, Fantino B, Le Gall D, Schott AM, Berrut G, Beauchet O. 2011 Severe vitamin D deficiency is associated with advanced-stage dementia in geriatric inpatients. J. Am. Geriatr. Soc. 59, 169 – 171. (doi:10.1111/j.1532-5415.2010.03166.x)
- 126. Wang L et al. 2012 Vitamin D receptor and Alzheimer's disease: a genetic and functional study. Neurobiol. Aging 33, 1844e1-1844e9. (doi:10. 1016/j.neurobiolaging.2011.12.038)
- 127. DeLuca GC, Kimball SM, Kolasinski J, Ramagopalan SV, Ebers GC. 2013 Review: the role of vitamin D in nervous system health and disease. Neuropath. Appl. Neurobiol. 39, 458-484. (doi:10.1111/nan. 12020)
- 128. Gezen-Ak AD, Yilmazer S, Dursun E. 2014 Why vitamin D in Alzheimer's disease? The hypothesis. J. Alzheimer's Dis. 40, 257 – 269. (doi:10.3233/JAD-
- 129. Annweiler C, Rolland Y, Schott AM, Blain H, Vellas B, Herrmann FR, Beauchet O. 2012 Higher vitamin D dietary intake is associated with lower risk of Alzheimer's disease: a 7-year follow-up. J. Gerontol. A Biol. Sci. Med. Sci. 67, 1205 – 1211. (doi:10.1093/ gerona/gls107)
- 130. Annweiler C, Llewellyn DJ, Beauchet O. 2013 Low serum vitamin D concentrations in Alzheimer's disease: a systematic review and meta-analysis. J. Alzheimers Dis. 33, 659-674. (doi:10.3233/JAD-2012-121432)

- 131. Lehmann DJ, Refsum H, Warden DR, Medway C, Wilcock GK, Smith AD. 2011 The vitamin D receptor gene is associated with Alzheimer's disease. *Neurosci. Lett.* **504**, 79-82. (doi:10.1016/j.neulet. 2011.08.057)
- 132. Gezen-Ak DE et al. 2007 Association between vitamin D receptor gene polymorphism and Alzheimer's disease. Tohoku J. Exp. Med. 212, 275 – 282. (doi:10.1620/tjem.212.275)
- 133. Durk MR, Han K, Chow EC, Ahrens R, Henderson JT, Fraser PE, Pang KS. 2014 1α ,25-Dihydroxyvitamin D3 reduces cerebral amyloid- β accumulation and improves cognition in mouse models of Alzheimer's disease. J Neurosci. 34, 7091-7101. (doi:10.1523/ JNEUROSCI.2711-13.2014)
- 134. Sutherland MK, Somerville MJ, Yoong LKK, Bergeron C, Haussler MR, Mclachlan DRC. 1992 Reduction of vitamin D hormone receptor mRNA levels in Alzheimer as compared to Huntington hippocampus: correlation with calbindin-28 k mRNA levels. Mol. Brain Res. 13, 239-250. (doi:10.1016/ 0169-328X(92)90032-7)
- 135. Palop JJ, Jones B, Kekonius L, Chin J, Yu G-Q, Raber J, Masliah E, Mucke L. 2003 Neuronal depletion of calcium-dependent proteins in the dentate gyrus is tightly linked to Alzheimer's disease-related cognitive deficits. Proc. Natl Acad. Sci. USA 100, 9572-9577. (doi:10.1073/pnas. 1133381100)
- 136. Niture SK, Jaiswal AK. 2012 Nrf2 protein upregulates antiapoptotic protein Bcl-2 and prevents cellular apoptosis. J. Biol. Chem. 287, 9873-9886. (doi:10.1074/jbc.M111.312694)
- 137. Rohn TT, Vyas V, Hernandez-Estrada T, Nichol KE, Christie L-A, Head E. 2008 Lack of pathology in a triple transgenic mouse model of Alzheimer's disease after overexpression of the anti-apoptotic protein Bcl-2. J. Neurosci. 28, 3051-3059. (doi:10. 1523/JNEUROSCI.5620-07.2008)
- 138. Kanninen K et al. 2009 Intrahippocampal injection of a lentiviral vector expressing Nrf2 improves

- spatial learning in a mouse model of Alzheimer's disease. Proc. Natl Acad. Sci. USA 106, 16 505-16 510. (doi:10.1073/pnas.0908397106)
- 139. Semba RD, Moghekar AR, Hu J, Sun K, Turner R, Ferrucci L, O'Brien R. 2014 Klotho in the cerebrospinal fluid of adults with and without Alzheimer's disease. Neurosci. Lett. **558**, 37 – 40. (doi:10.1016/i.neulet.2013.10.058)
- 140. Kuang K et al. 2014 Klotho upregulation contributes to the neuroprotection of liqustilide in an Alzheimer's disease mouse model. Neurobiol. Aging **35**, 169 – 178. (doi:10.1016/j.neurobiolaging.2013. 07.019)
- 141. Querfurth HK, Selkoe DJ. 1994 Calcium ionophore increases amyloid beta peptide production by cultured cells. *Biochemistry* **33**, 4550 – 4561. (doi:10.1021/bi00181a016)
- 142. Itkin A, Dupres V, Dufrêne YF, Bechinger B, Ruysschaert JM, Raussens V. 2011 Calcium ions promote formation of amyloid β -peptide (1-40) oligomers causally implicated in neuronal toxicity of Alzheimer's disease. PLoS ONE 6, e18250. (doi:10. 1371/journal.pone.0018250)
- 143. Pierrot N, Santos SF, Feyt C, Morel M, Brion JP, Octave JN. 2006 Calcium-mediated transient phosphorylation of tau and amyloid precursor protein followed by intraneuronal amyloid-beta accumulation. J. Biol. Chem. 281, 39 907 – 39 914. (doi:10.1074/jbc.M606015200)
- 144. Green KN, LaFerla, FM. 2008 Linking calcium to AB and Alzheimer's disease. Neuron **59**, 190 – 194. (doi:10.1016/j.neuron.2008.07.013)
- 145. Del Prete D, Checler F, Chami M. 2014 Ryanodine receptors: physiological function and deregulation in Alzheimer disease. *Mol. Neurodegener.* **9**, 21–36. (doi:10.1186/1750-1326-9-21)
- 146. DeCaluwé J, Dupont G. 2013 The progression towards Alzheimer's disease described as a bistable switch arising from the positive loop between amyloids and Ca^{2+} . J. Theor. Biol. **331**, 12–18. (doi:10.1016/j.jtbi.2013.04.015)