

## Invited commentary on David Fedson's article

Ian Clark, Lisa Alleva

Research School of Biology, Australian National University, Canberra, Australia  
E-mail: ian.clark@anu.edu.au

Please cite this paper as: Clark (2009) Invited commentary on David Fedson's article. *Influenza and Other Respiratory Viruses* 3(5), 199–201.

The recent H5N1 and H1N1 scares demonstrate that political, bureaucratic, and to a large extent scientific, thinking about how to ward off potentially large fatalities is currently restricted to stockpiling antivirals and generating new vaccines. Nevertheless, as David Fedson pointed out with undeniable logic in last month's issue of this journal (3: 129–142), an influenza research community that continues to confine its efforts to these approaches will fall far short, in any severe pandemic, in its brief to prevent worldwide high mortality.

Central to Fedson's answer to this challenge is for us to examine more closely the argument that a fatal outcome in influenza is largely a manifestation of excessive release of inflammatory cytokines, as embodied in the cytokine concept of disease. Given reasonable acceptance of this, he argues, we should then put more effort into testing the potential for treating influenza illness with cheap and readily available agents that are already, on other rationales, in therapeutic use for other purposes, and also known to suppress production of disease-inducing cytokines.

What is the cytokine concept of disease, and how wide is its relevance across the infectious diseases? The idea began nearly 30 years ago when a newly described endogenous anti-tumour agent<sup>1</sup> was used to rationalise the nature of malaria and systemic bacterial infections.<sup>2</sup> As reviewed,<sup>3</sup> when rTNF was later being tested as an anti-tumour agent in patients, the toxicity that prevented its widespread use so strikingly mimicked influenza that tumour researchers referred, in print, to it generating influenza-like side effects.<sup>4</sup> Symptoms, which included fatigue, fever, anorexia, chills, headache, pulmonary oedema, immunosuppression, myalgia, nausea, vomiting and diarrhoea<sup>5,6</sup> were worse with higher doses. Parenteral interleukin-2, by inducing TNF, also produces a very similar clinical picture.<sup>7</sup>

Together with influenza being the standard misdiagnosis of imported malaria in temperate countries, the experiences of these tumour researchers made it plausible that this disease model would explain the pathology of viral diseases, including influenza, as well as that of malaria and bacterial sepsis.<sup>8</sup> The concept of cytokine excess has also been adopted to rationalise the diseases caused by *Mycobacterium* spp.,<sup>9</sup>

*Salmonella typhi*,<sup>10</sup> *Leishmania* spp.,<sup>11</sup> *Toxoplasma gondii*,<sup>12</sup> *Coxiella brunetii*,<sup>13</sup> and *Listeria monocytogenes*.<sup>14</sup> It dominates the literature on the pathophysiological consequences of trauma, haemorrhagic shock, and burns because these, too, originate from cytokine excess.<sup>15,16</sup> Different triggers (gram-negative lipopolysaccharide, gram positive toxins, fungal or malarial toxins, or modulation of RIG-1 gene expression) and sites of production can be expected to generate different local patterns, so we must expect some clinical and pathological dissimilarities between systemic diseases that share this common fundamental origin.

TNF generation and circulating levels are increased in influenza,<sup>17</sup> particularly so for influenza caused by the more pathogenic strains. The evidence linking the excess cytokine concept with influenza disease will be well-known by most readers. In brief, influenza A virus stimulates the release of TNF from macrophages,<sup>18</sup> and the recent avian strain induces production of more TNF from human macrophages than do a range of less virulent strains of human influenza.<sup>19</sup> Likewise, this H5N1 influenza virus induces an inflammatory cytokine response in primary cultures of human alveolar and bronchial epithelial cells.<sup>20</sup> H5N1/97 upregulates TNF mRNA levels and TNF-related apoptosis-inducing ligand (TRAIL) in human monocyte-derived macrophages,<sup>21</sup> and higher levels of inflammatory cytokines and chemokines are associated with a fatal outcome.<sup>22</sup> Moreover, a reconstructed version of the strain of influenza virus responsible for massive human mortality in 1918–1919, but not non-virulent constructs or strains, induces a strong and prolonged pro-inflammatory cytokine response during the fatal infections it causes in mice<sup>23</sup> and macaque monkeys.<sup>24</sup> The literature's emphasis on TNF may be artificial, but it is now accepted as the progenitor of a cytokine superfamily, and is demonstrated to be a master regulator of the network of mediators it induces and interacts with.<sup>25</sup>

Thus Fedson's proposal – that any agent known to reduce inflammatory cytokine production and to ameliorate any one of the diseases or conditions mentioned earlier warrants testing in the others – is logical and compelling. Examples are two peroxisome proliferator-activated receptor (PPAR) agonists: gemfibrozil, a fibrate (PPAR- $\alpha$  agonist), reported

by our group to reduce mortality in mouse infection with a H2N2 strain of influenza A<sup>26</sup> and rosiglitazone, a glitazone (PPAR- $\gamma$  agonist) that does the same in a mouse model for malaria.<sup>27</sup> Both classes of agents reduce inflammatory cytokine production, largely through antagonising the signal transducer, nuclear factor-kappaB (NF $\kappa$ B).<sup>28</sup>

The immunosuppression that can accompany influenza<sup>29</sup> is in fact characteristic of systemic inflammatory disease in general. It is much studied, in terms of cytokine imbalance, in conditions such as sepsis,<sup>30</sup> malaria,<sup>31</sup> trypanosomiasis,<sup>32</sup> and trauma.<sup>33</sup> In malaria, for example, its mechanism has been shown to depend on nitric oxide,<sup>34</sup> a downstream mediator of TNF, which inhibits the function of dendritic cells in malaria, thus limiting antigen presentation.<sup>35</sup> Thus, apart from its clinical relevance as the cause of the secondary bacterial pneumonia often seen in severe influenza, as Fedson discusses, this immunosuppression is a reliable, though indirect, indicator that influenza belongs to the family of conditions caused by excess cytokine production.

Likewise, the rarity of pulmonary oedema in childhood compared with adult influenza,<sup>36</sup> malaria,<sup>37</sup> and trauma,<sup>38</sup> also casts influenza into the same cytokine-mediated mould as malaria and sepsis. Already observed differences in the anti-inflammatory versus pro-inflammatory cytokine ratios between paediatric and adult macrophages stimulated with LPS<sup>39</sup> can plausibly be attributed to differences in PPAR function, as its activation is prolonged in young mice compared with older mice.<sup>40</sup> These insights can only be appreciated if influenza scientists seek, as Fedson urges, the expertise of researchers outside of their immediate discipline.

Others have suggested that any treatment directed against the inflammatory cytokines that cause illness will also inhibit the protective innate response against the virus. This is a valid possibility, as TNF has been reported to exert an *in vitro* effect against influenza virus in human epithelial cells.<sup>41</sup> But what happens *in vivo* is what matters, and others have found that anti-TNF antibody improves experimental influenza disease without influencing virus clearance.<sup>17,42</sup> With close to a million patients having received long-term TNF-neutralising drugs for rheumatoid arthritis or Crohn's disease by 2004,<sup>43</sup> and a call being made in that year for alertness to the possibility of hepatitis or HIV exacerbation,<sup>44</sup> so far as we are aware there are as yet no reports of enhancement of viral disease, including influenza. Likewise, we are not aware of any reports of viral enhancement in patients taking fibrates, glitazones or statins for other reasons. While not as powerful as anti-TNF therapy, these less specific agents are advantageous when the target cytokines are not yet fully defined, as they inhibit a range of them.

Certainly, the simplicity and logic embodied in Fedson's approach to the practical treatment of severe influenza

should capture the attention and imagination of researchers interested in intractable infectious disease.

## References

- 1 Carswell EA, Old LJ, Kassel RL, Green S, Fiore N, Williamson B. An endotoxin-induced serum factor that causes necrosis of tumors. *Proc Natl Acad Sci USA* 1975; 72:3666–3670.
- 2 Clark IA, Virelizier J-L, Carswell EA, Wood PR. Possible importance of macrophage-derived mediators in acute malaria. *Infect Immun* 1981; 32:1058–1066.
- 3 Clark IA. How TNF was recognized to be a key mechanism of disease. *Cytokine Growth FR* 2007; 18:335–343.
- 4 Rauthe G, Siermanns J. Recombinant tumour necrosis factor in the local therapy of malignant pleural effusion. *Eur J Cancer* 1997; 33:226–231.
- 5 Steinmetz T, Schaadt M, Gahl R, Schenk V, Diehl V, Pfreundschuh M. Phase 1 study of 24-hour continuous intravenous infusion of recombinant human tumor necrosis factor. *J Biol Resp Mod* 1988; 7:417–423.
- 6 Jakubowski AA, Casper ES, Gabrilove JL, Templeton M-A, Sherwin SA, Oettgen HF. Phase 1 trial of intramuscularly administered tumor necrosis factor in patients with advanced cancer. *J Clin Oncol* 1989; 7:298–303.
- 7 Mier JW, Vachino G, Van der Meer JWM *et al.* Induction of circulating tumor necrosis factor (TNF- $\alpha$ ) as the mechanism for the febrile response to interleukin-2 (IL-2) in cancer patients. *J Clin Immunol* 1988; 8:426–436.
- 8 Clark IA, Cowden WB. Is TNF a key to acute infectious illness? *Today's Life Science* 1989; 1:26–29.
- 9 Rook GAW, Taverne J, Leveton C, Steele J. The role of gamma-interferon, vitamin D<sub>3</sub> metabolites and tumour necrosis factor in the pathogenesis of tuberculosis. *Immunology* 1987; 62:229–234.
- 10 Bhutta ZA, Mansoorali N, Hussain R. Plasma cytokines in paediatric typhoidal salmonellosis: correlation with clinical course and outcome. *J Infect* 1997; 35:253–256.
- 11 Raziuddin S, Abdalla RE, el Awad EH, al Janadi M. Immunoregulatory and proinflammatory cytokine production in visceral and cutaneous leishmaniasis. *J Infect Dis* 1994; 170:1037–1040.
- 12 Arsenijevic D, Girardier L, Seydoux J, Chang HR, Dulloo AG. Altered energy balance and cytokine gene expression in a murine model of chronic infection with *Toxoplasma gondii*. *Am J Physiol* 1997; 272:E908–E917.
- 13 Mege JL, Maurin M, Capo C, Raoult D. *Coxiella burnetii* – the Query fever bacterium – a model of immune subversion by a strictly intracellular microorganism. *FEMS Microbiol Rev* 1997; 19:209–217.
- 14 Nakane A, Yamada K, Hasegawa S *et al.* Endogenous cytokines during a lethal infection with *Listeria monocytogenes* in mice. *FEMS Microbiol Lett* 1999; 175:133–142.
- 15 Roumen RMH, Hendriks T, Van der Venjongekrijg J *et al.* Cytokine patterns in patients after major vascular surgery, hemorrhagic shock, and severe blunt trauma - relation with subsequent adult respiratory distress syndrome and multiple organ failure. *Ann Surg* 1993; 218:769–776.
- 16 Giroir BP, Horton JW, White DJ, McIntyre KL, Lin CQ. Inhibition of tumor necrosis factor prevents myocardial dysfunction during burn shock. *Am J Physiol* 1994; 267:H118–H124.
- 17 Peper RL, Vancampen H. Tumor necrosis factor as a mediator of inflammation in influenza A viral pneumonia. *Microb Pathog* 1995; 19:175–183.
- 18 Hinder F, Schmidt A, Gong JH *et al.* Influenza A virus infects macrophages and stimulates release of tumor necrosis factor-alpha. *Pathobiology* 1991; 59:227–231.

- 19 Cheung CY, Poon LLM, Lau AS *et al.* Induction of proinflammatory cytokines in human macrophages by influenza A (H5N1) viruses: a mechanism for the unusual severity of human disease. *Lancet* 2002; 360:1831–1837.
- 20 Chan MCW, Cheung CY, Chui WH *et al.* Proinflammatory cytokine responses induced by influenza A (H5N1) viruses in primary human alveolar and bronchial epithelial cells. *Resp Res* 2005; 6:135.
- 21 Zhou JF, Law HKW, Cheung CY, Ng IHY, Peiris JSM, Lau YL. Functional tumor necrosis factor-related apoptosis-inducing ligand production by avian influenza virus-infected macrophages. *J Infect Dis* 2006; 193:945–953.
- 22 de Jong MD, Simmons CP, Thanh TT *et al.* Fatal outcome of human influenza A (H5N1) is associated with high viral load and hypercytokinemia. *Nat Med* 2006; 12:1203–1207.
- 23 Kash JC, Tumpey TM, Proll SC *et al.* Genomic analysis of increased host immune and cell death responses induced by 1918 influenza virus. *Nature* 2006; 443:578–581.
- 24 Kobasa D, Jones SM, Shinya K *et al.* Aberrant innate immune response in lethal infection of macaques with the 1918 influenza virus. *Nature* 2007; 445:319–323.
- 25 Charles P, Elliott MJ, Davis D *et al.* Regulation of cytokines, cytokine inhibitors, and acute-phase proteins following anti-TNF-alpha therapy in rheumatoid arthritis. *J Immunol* 1999; 163:1521–1528.
- 26 Budd A, Alleva L, Alsharifi M *et al.* Increased survival after gemfibrozil treatment of severe mouse influenza. *Antimicrob Agents Chemother* 2007; 51:2965–2968.
- 27 Serghides L, Patel SN, Ayi K *et al.* Rosiglitazone modulates the innate immune response to *Plasmodium falciparum* infection and improves outcome in experimental cerebral malaria. *J Infect Dis* 2009; 199:1536–1545.
- 28 Delerive P, Fruchart JC, Staels B. Peroxisome proliferator-activated receptors in inflammation control. *J Endocrinol* 2001; 169:453–459.
- 29 Kantzler GB, Lauterier SF, Cusumano CL, Lee JD, Ganguly R, Waldman RH. Immunosuppression during influenza virus infection. *Infect Immun* 1974; 10:996–1002.
- 30 Weighardt H, Heidecke CD, Emmanuilidis K *et al.* Sepsis after major visceral surgery is associated with sustained and interferon-gamma-resistant defects of monocyte cytokine production. *Surgery* 2000; 127:309–315.
- 31 Voller A, Gall D, Manawadu BR. Depression of the antibody response to tetanus toxoid in mice infected with malaria parasites. *Z Tropenmed Parasitol* 1972; 23:152–155.
- 32 Mabbott NA, Coulson PS, Smythies LE, Wilson RA, Sternberg JM. African trypanosome infections in mice that lack the interferon-gamma receptor gene: nitric oxide-dependent and -independent suppression of T-cell proliferative responses and the development of anaemia. *Immunology* 1998; 94:476–480.
- 33 Zellweger R, Ayala A, Demaso CM, Chaudry IH. Trauma-hemorrhage causes prolonged depression in cellular immunity. *Shock* 1995; 4:149–153.
- 34 Rockett KA, Awburn MM, Rockett EJ, Cowden WB, Clark IA. Possible role of nitric oxide in malarial immunosuppression. *Para Immunol* 1994; 16:243–249.
- 35 Wykes MN, Liu XQ, Jiang S, Hirunpetcharat C, Good MF. Systemic tumor necrosis factor generated during lethal *Plasmodium* infections impairs dendritic cell function. *J Immunol* 2007; 179:3982–3987.
- 36 Brundage JF, Shanks GD. Deaths from bacterial pneumonia during 1918–19 influenza pandemic. *Emerg Infect Dis* 2008; 14:1193–1199.
- 37 Dondorp AM, Lee SJ, Faiz MA *et al.* The relationship between age and the manifestations of and mortality associated with severe malaria. *Clin Infect Dis* 2008; 47:151–157.
- 38 Calkins CM, Bensard DD, Moore EE *et al.* The injured child is resistant to multiple organ failure: a different inflammatory response? *J Trauma* 2002; 53:1058–1063.
- 39 Barsness KA, Bensard DD, Partrick DA, Calkins CM, Hendrickson RJ, McIntyre RC. Endotoxin induces an exaggerated interleukin-10 response in peritoneal macrophages of children compared with adults. *J Ped Surg* 2004; 39:912–915.
- 40 Shin T, Kuboki S, Huber N *et al.* Activation of peroxisome proliferator-activated receptor-gamma during hepatic ischemia is age-dependent. *J Surg Res* 2008; 147:200–205.
- 41 Seo SH, Webster RG. Tumor necrosis factor alpha exerts powerful anti-influenza virus effects in lung epithelial cells. *J Virol* 2002; 76:1071–1076.
- 42 Hussell T, Pennycook A, Openshaw PJM. Inhibition of tumor necrosis factor reduces the severity of virus-specific lung immunopathology. *Eur J Immunol* 2001; 31:2566–2573.
- 43 Vilcek J, Feldmann M. Historical review: cytokines as therapeutics and targets of therapeutics. *Trends Pharmacol Sci* 2004; 25:201–209.
- 44 Calabrese LH, Zein N, Vassilopoulos D. Safety of antitumor necrosis factor (anti-TNF) therapy in patients with chronic viral infections: hepatitis C, hepatitis B, and HIV infection. *Ann Rheum Dis* 2004; 63:18–24.