5 Caballero JD, Pastor MD, Vindel A *et al*. Emergence of *cfr*-mediated linezolid resistance in a methicillin-resistant *Staphylococcus aureus* epidemic clone isolated from patients with cystic fibrosis. *Antimicrob Agents Chemother* 2015; **60**: 1878–82.

6 Klitgaard RN, Ntokou E, Nørgaard K *et al*. Mutations in the bacterial ribosomal protein L3 and their association with antibiotic resistance. *Antimicrob Agents Chemother* 2015; **59**: 3518–28.

7 Griggs DJ, Marona H, Piddock LJ. Selection of moxifloxacin-resistant *Staphylococcus aureus* compared with five other fluoroquinolones. *J Antimicrob Chemother* 2003; **51**: 1403–7.

8 Brenciani A, Morroni G, Pollini S *et al.* Characterization of novel conjugative multiresistance plasmids carrying *cfr* from linezolid-resistant *Staphylococcus epidermidis* clinical isolates from Italy. *J Antimicrob Chemother* 2016; **71**: 307–13.

9 Campanile F, Bongiorno D, Perez M *et al*. Epidemiology of *Staphylococcus aureus* in Italy: First nationwide survey, 2012. *J Glob Antimicrob Res* 2015; **3**: 247–54.

10 Mendes RE, Deshpande LM, Smyth DS *et al*. Characterization of baseline methicillin-resistant *Staphylococcus aureus* isolates recovered from phase IV clinical trial for linezolid. *J Clin Microbiol* 2010; **48**: 568–74.

J Antimicrob Chemother 2016 doi:10.1093/jac/dkw132 Advance Access publication 26 April 2016

Evidence from a New York City hospital of rising incidence of genetically diverse carbapenem-resistant *Enterobacter cloacae* and dominance of ST171, 2007–14

Angela Gomez-Simmonds¹, Yue Hu¹, Sean B. Sullivan¹, Zheng Wang¹, Susan Whittier² and Anne-Catrin Uhlemann¹*

¹Department of Medicine, Division of Infectious Diseases, Columbia University Medical Center, New York, NY 10032, USA; ²Department of Pathology and Cell Biology, Clinical Microbiology Laboratory, Columbia University Medical Center, New York, NY 10032, USA

*Corresponding author. Tel: +1-212-342-4530; Fax: +1-212-305-5794; E-mail: au2110@cumc.columbia.edu

Sir,

The proliferation of carbapenem-resistant Enterobacteriaceae (CRE) over the past two decades signifies an escalating threat to modern healthcare. CRE are typically resistant to most antibiotic classes and produce infections associated with high mortality.¹ Carbapenem resistance mediated by *bla*_{KPC} has been described in all major Enterobacteriaceae, but can also result from alterations in porins combined with overexpression of endogenous AmpC enzymes.² However, clinical infections have been largely restricted to *Klebsiella pneumoniae*, especially epidemic *bla*_{KPC}-harbouring ST258.³

Recent studies report the local emergence of carbapenemresistant *Enterobacter cloacae* (CREC) internationally⁴ and in the USA,^{5–9} suggesting accelerating carbapenem resistance acquisition in this organism. The prevalence and population structure of CREC remain incompletely understood. Here we report an increase in CREC incidence at a tertiary care hospital in New York City over a 7 year period and characterize its molecular epidemiology.

We retrospectively reviewed the proportion of *E. cloacae* nonsusceptible to carbapenems from 2007 to 2014 at our hospital using antibiogram data. We then investigated the clinical and molecular epidemiology of CREC isolates since 2010 using MLST.¹⁰ Isolate susceptibilities were determined using Vitek 2 and interpreted using recent CLSI guidelines,¹¹ accounting for 2010 revisions to carbapenem MIC breakpoints. We PCR-amplified carbapenemase genes *bla*_{KPC}, *bla*_{VIM}, *bla*_{IMP}, *bla*_{NDM} and *bla*_{OXA-48}, *ampC* β -lactamase¹² and *ompC* and *ompF* porin genes.² PCR products were sequenced to differentiate subtypes or identify non-functional mutations. Our institutional review board approved this study.

Between 2007 and 2014, we observed a hospital-wide increase in the proportion of carbapenem-non-susceptible *E. cloacae* from 1.6% to 8.7% of isolates (Figure 1a). This corresponded to a 5-fold absolute increase in incidence (Figure 1b), ranging from 4.6% to 19% in ICU inpatients, 1.3% to 10% in non-ICU inpatients and 0% to 2.6% in outpatients. During the same time period the incidence of carbapenem-resistant *K. pneumoniae, Escherichia coli* and *Proteus mirabilis* remained stable or decreased (Figure 1a).

We obtained clinical data for 56 consecutive patients with CREC, of whom 53 had initial isolates available for molecular typing. CREC largely affected patients requiring prolonged or recurrent hospitalizations and most had multiple comorbidities. The most common culture source was the respiratory tract (n=24), followed by blood (n=11) and urine (n=10). Most isolates (79%) had meropenem MICs of \geq 16 mg/L. While susceptibility to amikacin (95%) and polymyxin B (100%) was preserved, most isolates were resistant to gentamicin (73%), tobramycin (82%), levofloxacin (77%) and trimethoprim/sulfamethoxazole (89%). Both 30 day mortality (29%) and hospital mortality (46%) were high.

MLST revealed substantial clonal diversity, as CREC isolates belonged to 26 different STs. ST171 accounted for nearly half of all isolates (n=23, 43%), followed in frequency by ST78 (n=6) (Figure 1c). All other STs were singletons (n=24), including 11 novel STs. *bla*_{KPC-3} (n=30), *bla*_{KPC-2} (n=12) and *bla*_{KPC-4} (n=2) were the predominant putative mechanisms of carbapenem resistance, whereas *bla*_{NDM} occurred in one isolate (Figure 1d). ST171 and ST78 were found to harbour *bla*_{KPC-3}, while *bla*_{KPC-2}, *bla*_{KPC-4} and *bla*_{NDM} occurred in unique STs. All carbapenemase-negative isolates harboured *ampC* genes and four out of eight contained premature stop codons in the *ompC* porin (Table S1, available as Supplementary data at *JAC* Online). The mechanism of resistance remains undetermined in four isolates with lower meropenem MICs.

Our data support a highly concerning diversification of CRE beyond *K. pneumoniae*. While many CREC isolates belonged to diverse lineages, suggesting sporadic acquisition of bla_{KPC} -encoding plasmids, we also observed the emergence and rapid spread of two CREC clones, ST78 and ST171. ST171 was first detected at a Western Pennsylvania hospital⁷ and has spread in the Midwestern USA.⁹ ST78 and ST171 also accounted for the majority of 20 CREC isolates at a Boston hospital.⁸ Based on these findings and recent reports in the literature, we propose that ST171 is emerging as a dominant CREC clone in the USA.

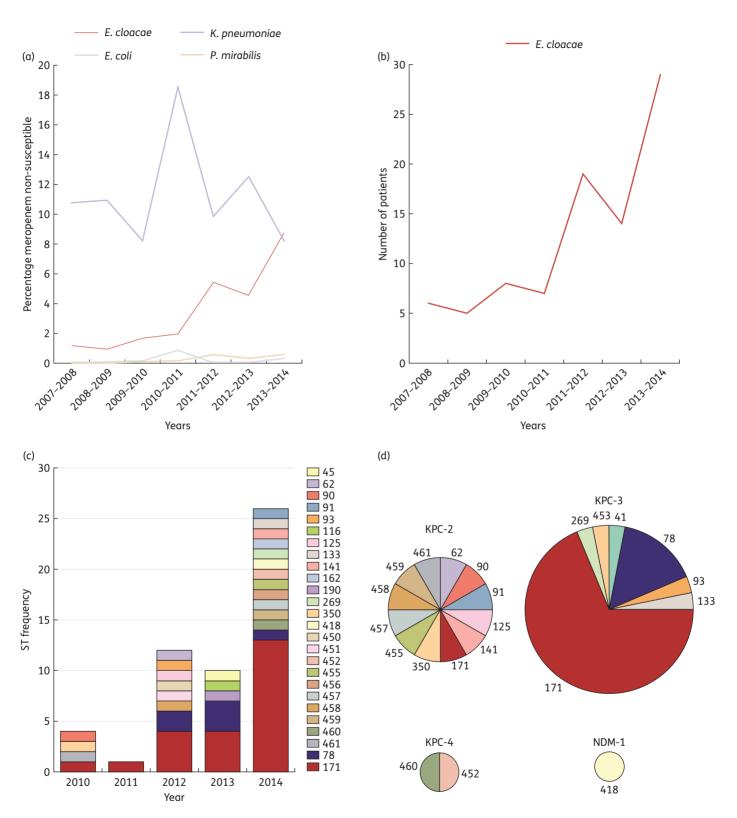


Figure 1. Emergence of CREC and corresponding multilocus STs. (a) Percentage of single-patient isolates of various Enterobacteriaceae non-susceptible to meropenem between 2007 and 2014 based on antibiogram data. (b) Absolute number of patients infected with meropenem-non-susceptible *E. cloacae* isolates per year. (c) STs of CREC isolates between 2007 and 2014. (d) Carbapenemase gene carriage amongst CREC isolates according to ST. This figure appears in colour in the online version of *JAC* and in black and white in the print version of *JAC*.

While international CREC isolates primarily harboured metallo- β -lactamases and OXA-48,⁴ our data support a growing role for $bla_{\rm KPC}$ -mediated CREC in the USA. At least three different $bla_{\rm KPC}$ -encoded carbapenemases have been described in ST171 alone.⁸ This points to the efficiency of *E. cloacae* in acquiring a variety of resistance plasmids, potentiating the emergence of new resistant strains. Moreover, *E. cloacae* ST171 and to a lesser extent ST78 appear to be able to acquire and maintain $bla_{\rm KPC}$ -harbouring plasmids and persist as hospital-associated CRE strains.

This was a single-centre, retrospective study, and thus our findings may have limited generalizability. Notably, the increased incidence was unlikely to result from changes in CLSI breakpoints as only revised guidelines were applied, or from a system-wide breakdown of infection control as it was restricted to *E. cloacae*.

The alarming increase in CREC infections was driven by diverse clones, suggesting frequent acquisition of $bla_{\rm KPC}$ -encoded resistance as well as an increase in ST78 and ST171. The emergence of $bla_{\rm KPC}$ -harbouring ST171 across the USA suggests its potential as an epidemic CREC clone. Future molecular surveillance studies are urgently needed to further assess the spread of CREC including ST171.

Funding

This work was supported in part by the National Institute of Allergy and Infectious Diseases, National Institutes of Health (1R01AI116939 and 1R01AI116939-S01 to A.-C. U. and 5T32AI100852 to A. G.-S.) and the Columbia University Irving scholarship (A.-C. U.).

Transparency declarations

None to declare.

Supplementary data

Table S1 is available as Supplementary data at JAC Online (http://jac. oxfordjournals.org/).

References

1 CDC. Antibiotic Resistance Threats in the United States, 2013. http://www.cdc.gov/drugresistance/threat-report-2013/.

2 Doumith M, Ellington MJ, Livermore DM *et al.* Molecular mechanisms disrupting porin expression in ertapenem-resistant *Klebsiella* and *Enterobacter* spp. clinical isolates from the UK. *J Antimicrob Chemother* 2009; **63**: 659–67.

3 Nordmann P, Naas T, Poirel L. Global spread of carbapenemaseproducing Enterobacteriaceae. *Emerg Infect Dis* 2011; **17**: 1791–8.

4 Girlich D, Poirel L, Nordmann P. Clonal distribution of multidrug-resistant *Enterobacter cloacae. Diagn Microbiol Infect Dis* 2015; **81**: 264–8.

5 Marchaim D, Chopra T, Perez F *et al*. Outcomes and genetic relatedness of carbapenem-resistant Enterobacteriaceae at Detroit Medical Center. *Infect Control Hosp Epidemiol* 2011; **32**: 861–71.

6 Kiedrowski LM, Guerrero DM, Perez F *et al.* Carbapenem-resistant *Enterobacter cloacae* isolates producing KPC-3, North Dakota, USA. *Emerg Infect Dis* 2014; **20**: 1583–5.

7 Ahn C, Syed A, Hu F et al. Microbiological features of KPC-producing *Enterobacter* isolates identified in a U.S. hospital system. *Diagn Microbiol Infect Dis* 2014; **80**: 154–8.

8 Pecora ND, Li N, Allard M *et al*. Genomically informed surveillance for carbapenem-resistant Enterobacteriaceae in a health care system. *MBio* 2015; **6**: e01030.

9 Hargreaves ML, Shaw KM, Dobbins G *et al.* Clonal dissemination of *Enterobacter cloacae* harboring bla_{KPC-3} in the upper midwestern United States. *Antimicrob Agents Chemother* 2015; **59**: 7723–34.

10 Miyoshi-Akiyama T, Hayakawa K, Ohmagari N *et al.* Multilocus sequence typing (MLST) for characterization of *Enterobacter cloacae*. *PLoS One* 2013; **8**: e66358.

11 Clinical and Laboratory Standards Institute. *Performance Standards for Antimicrobial Susceptibility Testing: Twenty-fifth Informational Supplement M100-S25.* CLSI, Wayne, PA, 2015.

12 Voets GM, Fluit AC, Scharringa J *et al.* A set of multiplex PCRs for genotypic detection of extended-spectrum β -lactamases, carbapenemases, plasmid-mediated AmpC β -lactamases and OXA β -lactamases. *Int J Antimicrob Agents* 2011; **37**: 356–9.

J Antimicrob Chemother 2016 doi:10.1093/jac/dkw137 Advance Access publication 26 April 2016

Characterization of an outbreak due to CTX-M-15-producing *Klebsiella pneumoniae* lacking the *bla*_{OXA-48} gene belonging to clone ST405 in a neonatal unit in southern Spain

Jesús Machuca¹, Lorena López-Cerero^{1*}, Felipe Fernández-Cuenca¹, Irene Gracia-Ahufinger², Guillermo Ruiz-Carrascoso³, Fernando Rodríguez-López² and Álvaro Pascual^{1,4}

¹Unidad intercentros de Enfermedades Infecciosas, Microbiología y Medicina Preventiva, Hospital Universitario Virgen Macarena, Seville, Spain; ²Unidad de Microbiología Clínica, Hospital Universitario Reina Sofía-IMIBIC-Universidad de Cordoba, Cordoba, Spain; ³Unidad de Microbiología Clínica, Hospital Universitario La Paz-IdiPaz, Madrid, Spain; ⁴Department of Microbiology, University of Seville, Seville, Spain

*Corresponding author. Tel: +34-954-55-28-63; Fax: +34-954-37-74-13; E-mail: llopez@us.es

Sir,

The dissemination of high-risk clones presents a major challenge for the worldwide health system because of their ability to overcome control measures and cross institutional and national borders and so provide a stable platform for the spread of resistance determinants. The traceability of such successful clones is important for the design of control strategies.¹ The dissemination of different high-risk MDR *Klebsiella pneumoniae* clones belonging to clonal complex 258/340 (ST11, ST258, ST340, ST437 and ST512) has been a key factor in the propagation of carbapenemase genes, especially *bla*_{KPC}.² The role played by other clones is less well known. In 2012, a nosocomial large-hospital outbreak