

Gray matter maturation and cognition in children with different *APOE* ϵ genotypes

Linda Chang, MD
Vanessa Douet, PhD
Cinnamon Bloss, PhD
Kristin Lee, MS
Alexandra Pritchett, BA
Terry L. Jernigan, PhD
Natacha Akshoomoff,
PhD
Sarah S. Murray, PhD
Jean Frazier, MD
David N. Kennedy, PhD
David G. Amaral, PhD
Jeffrey Gruen, MD
Walter E. Kaufmann, MD
B.J. Casey, PhD
Elizabeth Sowell, PhD
Thomas Ernst, PhD
For the Pediatric Imaging,
Neurocognition, and
Genetics (PING) Study
Consortium

Correspondence to
Dr. Chang:
lchang@hawaii.edu

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Supplemental data
at Neurology.org

ABSTRACT

Objective: The aims of the current study were to determine whether children with the 6 different *APOE* ϵ genotypes show differences in gray matter maturation, particularly for those with $\epsilon 4$ and $\epsilon 2$ alleles, which are associated with poorer outcomes in many neurologic disorders.

Methods: A total of 1,187 healthy children (aged 3–20 years, 52.1% boys, 47.9% girls) with acceptable data from the cross-sectional Pediatric Imaging Neurocognition and Genetics Study were evaluated for the effects of 6 *APOE* ϵ genotypes on macroscopic and microscopic cortical and subcortical gray matter structures (measured with 3-tesla MRI and FreeSurfer for automated morphometry) and on cognition (NIH Toolbox).

Results: Among *APOE* $\epsilon 4$ carriers, age-related changes in brain structures and cognition varied depending on genotype, with the smallest hippocampi in $\epsilon 2\epsilon 4$ children, the lowest hippocampal fractional anisotropy in younger $\epsilon 4\epsilon 4$ children, the largest medial orbitofrontal cortical areas in $\epsilon 3\epsilon 4$ children, and age-dependent thinning of the entorhinal cortex in $\epsilon 4\epsilon 4$ children. Younger $\epsilon 4\epsilon 4$ children had the lowest scores on executive function and working memory, while younger $\epsilon 2\epsilon 4$ children performed worse on attention tasks. Larger parietal gyri in the younger $\epsilon 2\epsilon 4$ children, and thinner temporal and cingulate isthmus cortices or smaller hippocampi in the younger $\epsilon 4\epsilon 4$ children, predicted poorer performance on attention or working memory.

Conclusions: Our findings validated and extended prior smaller studies that showed altered brain development in *APOE* $\epsilon 4$ -carrier children. The $\epsilon 4\epsilon 4$ and $\epsilon 2\epsilon 4$ genotypes may negatively influence brain development and brain aging at the extremes of age. Studying *APOE* ϵ polymorphisms in young children may provide the earliest indicators for individuals who might benefit from early interventions or preventive measures for future brain injuries and dementia. ***Neurology*® 2016;87:585–594**

GLOSSARY

AD = Alzheimer disease; **FA** = fractional anisotropy; **GAF** = genetic ancestry factor; **GAM** = general additive model; **PING** = Pediatric Imaging, Neurocognition, and Genetics; **ROI** = region of interest; **SES** = socioeconomic status; **WM** = working memory.

APOE $\epsilon 4$ is a well-known risk allele for Alzheimer disease (AD), especially late-onset AD, and may lead to poorer outcome in neurologic disorders.^{1–4} In addition, *APOE* $\epsilon 4$ may influence brain development.^{5–7} However, the *APOE* $\epsilon 4$ allele demonstrates antagonistic pleiotropy, with deleterious effects on cognition, brain morphometry, and activation primarily after 55 years of age, but no negative⁸ or even beneficial effects in adults younger than 50 years^{9,10} and children aged 6 to 15 years.^{11–13} Compared to non- $\epsilon 4$ carriers, healthy children carrying $\epsilon 4$ (8–20 years) tended to have thinner entorhinal cortex,⁵ while healthy infants carrying $\epsilon 4$ showed altered brain measures in regions affected by AD.^{6,7} Whether these structural differences influence cognitive performance in children with $\epsilon 4$ remains controversial.^{5,12–14}

From the Department of Medicine (L.C., V.D., K.L., A.P., T.E.), John A. Burns School of Medicine, University of Hawaii and The Queen's Medical Center, Honolulu; Department of Psychiatry, School of Medicine (C.B.), Departments of Psychiatry and Cognitive Science (T.L.J., N.A.), and Department of Pathology (S.S.M.), University of California San Diego, La Jolla; Department of Psychiatry (J.F., D.N.K.), University of Massachusetts Medical School, Boston; Department of Psychiatry and Behavioral Sciences (D.G.A.), University of California, Davis; Departments of Pediatrics and Investigative Medicine (J.G.), Yale Child Health Research Center, Yale University School of Medicine, New Haven, CT; Boston Children's Hospital (W.E.K.), Harvard Medical School, Boston, MA; Sackler Institute for Developmental Psychobiology (B.J.C.), Weil Cornell Medical College, New York, NY; Department of Pediatrics (E.S.), University of Southern California, Los Angeles; and Children's Hospital (E.S.), Los Angeles, CA.

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APOE $\epsilon 2$ also may affect brain variably. The $\epsilon 2$ carriers were less likely to develop clinical dementia^{15,16} and had greater cognitive reserve,¹⁷ since *APOE* $\epsilon 2$ may be neuroprotective.¹⁷ However, $\epsilon 2$ carriers also had more cortical amyloid plaques,¹⁸ with elevated risks of cerebral amyloid angiopathy¹⁷ and cortical infarctions.² Furthermore, children carrying $\epsilon 2$ performed worse on visuospatial tasks¹² and had thicker temporal cortices⁵ than non- $\epsilon 2$ carriers, similar to adults carrying $\epsilon 2$ with mild cognitive impairment or AD.¹⁹

Because of potentially opposite influences of *APOE* $\epsilon 4$ and *APOE* $\epsilon 2$, prior studies typically evaluated homozygous or heterozygous $\epsilon 4$ or $\epsilon 2$ individuals, but frequently excluded $\epsilon 2\epsilon 4$ participants.^{5,20,21} One study assessed all 6 *APOE* genotypes but only on IQ and academic achievements.¹³ Therefore, we evaluated group differences across all 6 *APOE* ϵ genotypes on gray matter morphometry and cognition in typically developing children. We hypothesized that children with different *APOE* ϵ genotypes, especially $\epsilon 4$ and $\epsilon 2$ carriers (including $\epsilon 2\epsilon 4$), would show differential gray matter measures and cognitive function across the age span of childhood.

METHODS Participants. A total of 1,493 typically developing children aged 3 to 20 years were enrolled in the Pediatric Imaging, Neurocognition, and Genetics (PING) Study (<http://ping.chd.ucsd.edu>) at 10 US academic institutions from September 2010 to August 2012. The PING Study was designed to cross-sectionally investigate how genes influence brain maturation and cognitive measures across childhood. Children were enrolled from diverse ethnic groups and socioeconomic status (SES); detailed participant criteria were reported previously.²² Specifically, the 1,187 children in the current study were excluded for any confounding neurologic or psychiatric disorders, history of head trauma, mental retardation, preterm birth (<36 weeks), prenatal drug exposure (daily maternal illicit drug use >1 trimester), or any MRI contraindications (including pregnancy).

Standard protocol approvals, registrations, and patient consents. All participants provided written assents (older than 7 years) or consents (18 years or older), and parental consents (3–17 years), which along with the protocol were approved by each of the local institutional review boards for human subject studies.

Genotyping. Genomic DNA extracted from saliva was genotyped in 1,187 children for *APOE* ϵ (rs429358 and rs7412) using the iPLEX Gold assay at the Sequenom MassARRAY genotyping platform (Sequenom, San Diego, CA). Final genotypes were called using the MassARRAY Type, version 4.0. Replication and quality-control procedures were described previously.²² Genetic ancestry factor (GAF) was determined with the ADMIXTURE software (<https://www.genetics.ucla.edu/software/admixture/>) and the Illumina Human660W-Quad BeadChip for the 6 major

continental populations (African, Central Asian, East Asian, European, Native American, and Oceanic) in each child.

Magnetic resonance imaging. Image acquisition and processing were detailed previously.²² All scans were performed on 3-tesla scanners (9 Siemens, 2 General Electric, 2 Philips) using closely matched sequences. The protocol included 3-dimensional T1-weighted structural MRI (magnetization-prepared rapid-acquisition gradient echo, $1.0 \times 1.0 \times 1.2$ mm³, 8 minutes) and diffusion-tensor imaging (echo-planar imaging, 2.5-mm isotropic, $b = 1,000$ s/mm², 30 directions, 10 minutes) (<http://ping.chd.ucsd.edu>). Image processing included verification of protocol compliance and quality assurance. Automated morphometry using a modified FreeSurfer software was performed on the whole brain, and 7 subcortical and 20 cortical regions of interest (ROIs) from the standard FreeSurfer atlas were selected based on reported *APOE* ϵ effects in children⁵ and neonates,^{6,7} or patients with AD.⁴ Fractional anisotropy (FA) (corrected for B_0 inhomogeneities) in the same 7 subcortical ROIs was also assessed, since FA in these regions often increases during neurodevelopment.²³ Of 1,187 children, 1,080 met quality criteria for morphometry and 988 for FA.

NIH Toolbox. The NIH Toolbox Cognition Battery comprised 7 tests that assessed 8 cognitive domains.²⁴ Four domains found abnormal in patients with AD or $\epsilon 2$ or $\epsilon 4$ carriers^{25–27} were analyzed: (1) executive function–cognitive flexibility (Dimensional Change Card Sort Test); (2) visual attention (Flanker Inhibitory Control and Attention Test); (3) episodic memory (Picture Sequence Memory Test); and (4) working memory (WM) (List Sorting WM Test). For these domains, 1,060 children had acceptable data.

Statistical analyses. Statistical analyses were performed using the PING data portal (<https://ping-dataportal.ucsd.edu>).²² Genotype and genotype-by-age effects on morphometry and FA, and their relationships with cognition, were assessed with a general additive model (GAM) in R program (<http://www.r-project.org/>). GAM is a multiple linear regression model including smooth functions of variables that are data driven. Each model used age as a smooth independent variable and included a linear term for genotype and a smooth age-by-genotype interaction.²² GAMs used thin plate regression splines for the smoothing basis (using the $bs = "ts"$ specification), with basis dimensions $k = 4$ for main-effect smooth terms (e.g., age) and $k = 3$ for smooth interaction terms (e.g., age-by-sex).

All models covaried for SES (highest parental education and household income), sex, GAF, and scanner device. Subcortical volumes were adjusted for intracranial volume but not for average cortical thickness and area. For vertex-wise analyses, significance maps were thresholded at 5% using the false-discovery rate to correct for multiple comparisons. ROI-based analyses were corrected for multiple comparisons using the Holm-Bonferroni sequential method, which controls the family-wise error rate, using a stepwise procedure to adjust for significance levels instead of p values, and is uniformly more powerful than the Bonferroni correction.²⁸ Pairwise post hoc analyses were explored for contrasts with group differences on the GAM. Two children with $\epsilon 2\epsilon 2$ were not included in group analyses but are described separately.

RESULTS Participant characteristics. The 1,187 children were aged 12.1 ± 5.0 years; 569 were girls (table e-1 on the *Neurology*[®] Web site at Neurology.org). The 5 *APOE* ϵ allele groups were similar in sex proportion. The $\epsilon 2\epsilon 2$ children were the youngest and $\epsilon 2\epsilon 3$ children were the oldest; $\epsilon 3\epsilon 3$ was most common (61.78%), followed by $\epsilon 3\epsilon 4$ (21.8%), $\epsilon 2\epsilon 3$

(11.9%), $\epsilon 2\epsilon 4$ (2.6%), and $\epsilon 4\epsilon 4$ (1.75%), with $\epsilon 2\epsilon 2$ the rarest (0.17%). All genotype groups showed significant European GAF (60%–70%), except for $\epsilon 4\epsilon 4$ (35% European, but higher African ancestry than other groups). Parents/guardians of $\epsilon 4\epsilon 4$ children had the lowest household income, education, and occupation levels.

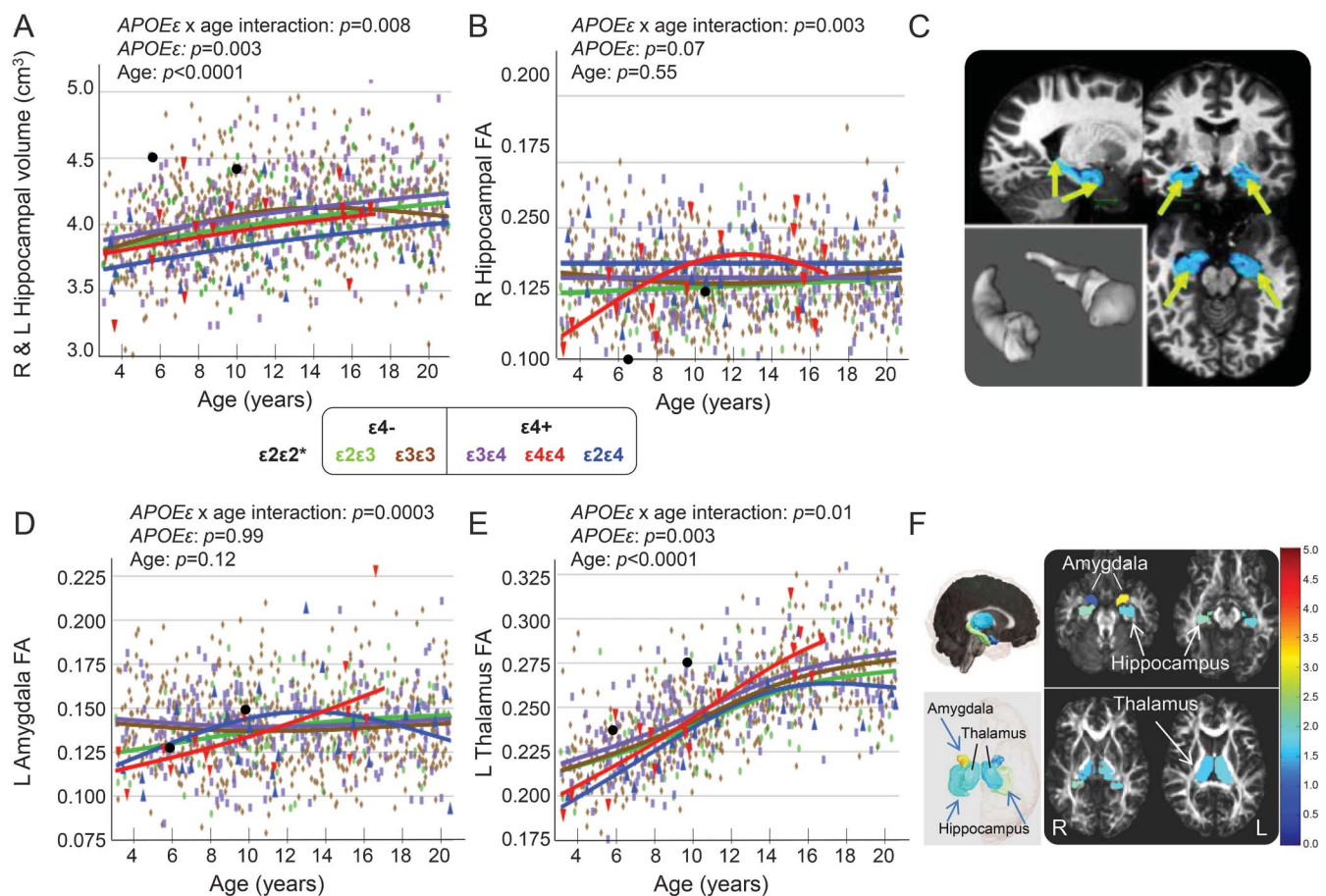
Subcortical volume differences across genotypes. The hippocampi differed across genotype groups (figure 1, A and B; table e-2). Independent of age, $\epsilon 3\epsilon 4$ children had the largest, while $\epsilon 2\epsilon 4$ children had the smallest hippocampi across groups (figure 1B). Hippocampal volumes increased linearly with age but differed by *APOE* ϵ genotype, with an inverted U shape in $\epsilon 3\epsilon 3$ children (peaking at 13.2 years). The 2 children with $\epsilon 2\epsilon 2$ had relatively large hippocampi.

Genotype-by-age on subcortical FA. Age-dependent FA changes were evaluated in all subcortical structures

(table e-2; figure 1, B–F). In the right hippocampus, FAs in $\epsilon 4\epsilon 4$ children were lower at younger ages (younger than 7 years) but normalized thereafter, with no changes with age (figure 1B). As a group, $\epsilon 2\epsilon 4$ children had the highest FA, while the younger $\epsilon 2\epsilon 2$ child showed the lowest hippocampal FA. Conversely, in the left amygdala, younger children (<7 years) with $\epsilon 4\epsilon 4$, $\epsilon 2\epsilon 4$, and $\epsilon 2\epsilon 3$ had lower FA than $\epsilon 3\epsilon 3$ and $\epsilon 3\epsilon 4$ groups, whose FA was constant with age (figure 1D). The left thalamus FA increased with age in all children (figure 1E), but children younger than 7 years with $\epsilon 2\epsilon 4$ and $\epsilon 4\epsilon 4$ had lower FA, while children older than 12 years with $\epsilon 4\epsilon 4$ showed higher FA (figure 1E).

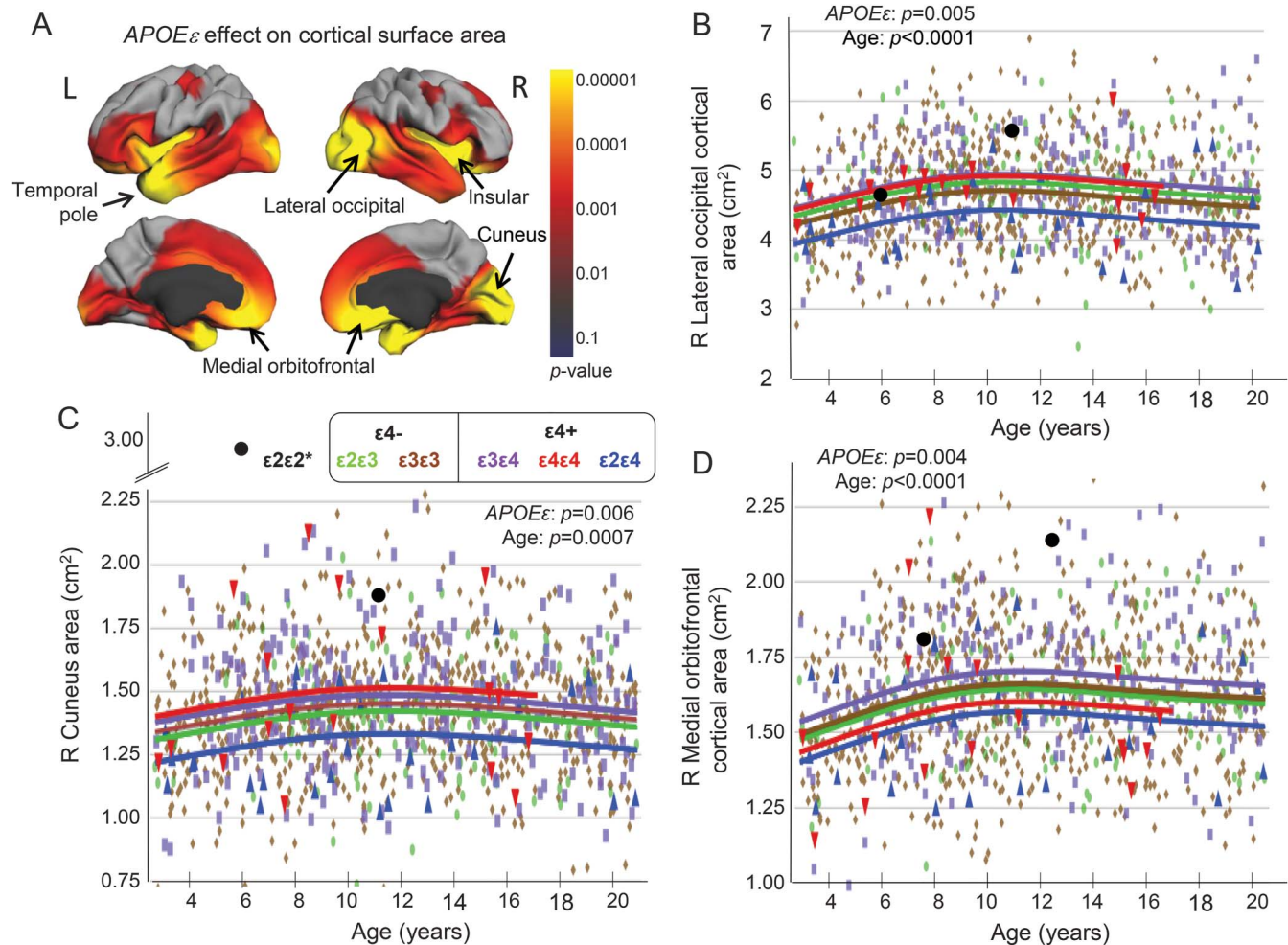
***APOE* ϵ genotype on cortical measures.** Several cortical areas differed by *APOE* ϵ genotype (figure 2A), with parallel age-related trajectories (validated in the ROI model) (figure 2, A–D; table e-3). For these regions,

Figure 1 *APOE* ϵ genotypic variations on age-dependent changes of subcortical volumes and FA



(A) Right and left hippocampal volumes are shown as averaged values (since they were not significantly different) across the 6 *APOE* ϵ genotypes. Note the relatively larger volumes in the 2 individuals homozygous for $\epsilon 2$ (black) and the smallest volumes across the age range and genotype groups in those with $\epsilon 2\epsilon 4$ (blue, $\epsilon 2\epsilon 4 < \epsilon 3\epsilon 3$, $p = 0.007$). (B) Right hippocampal FA is relatively stable across the age range except for the children with $\epsilon 4\epsilon 4$ (red), especially those at younger ages (similar age-related curve is seen in the left hippocampus; table e-2). The younger child with $\epsilon 2\epsilon 2$ also showed the lowest FA in the hippocampus (black). (C) Segmented hippocampi shown in 3 orientations (arrows). (D) The younger children (<10 years) with $\epsilon 2$ allele ($\epsilon 2\epsilon 2$, $\epsilon 2\epsilon 3$, and $\epsilon 2\epsilon 4$) and $\epsilon 4\epsilon 4$ showed relatively lower FA in left amygdala. (E) Younger children with $\epsilon 2\epsilon 4$ and $\epsilon 4\epsilon 4$ also showed relatively lower FA in left thalamus (post hoc test: $\epsilon 2\epsilon 4 < \epsilon 3\epsilon 3$, $p = 0.01$). (F) Log p value maps of the brain regions showing age-by-genotype interactions (see also table e-2). *Data for the $\epsilon 2\epsilon 2$ children are not included in the group analyses. FA = fractional anisotropy.

Figure 2 APOE ϵ genotypic variations on cortical areas



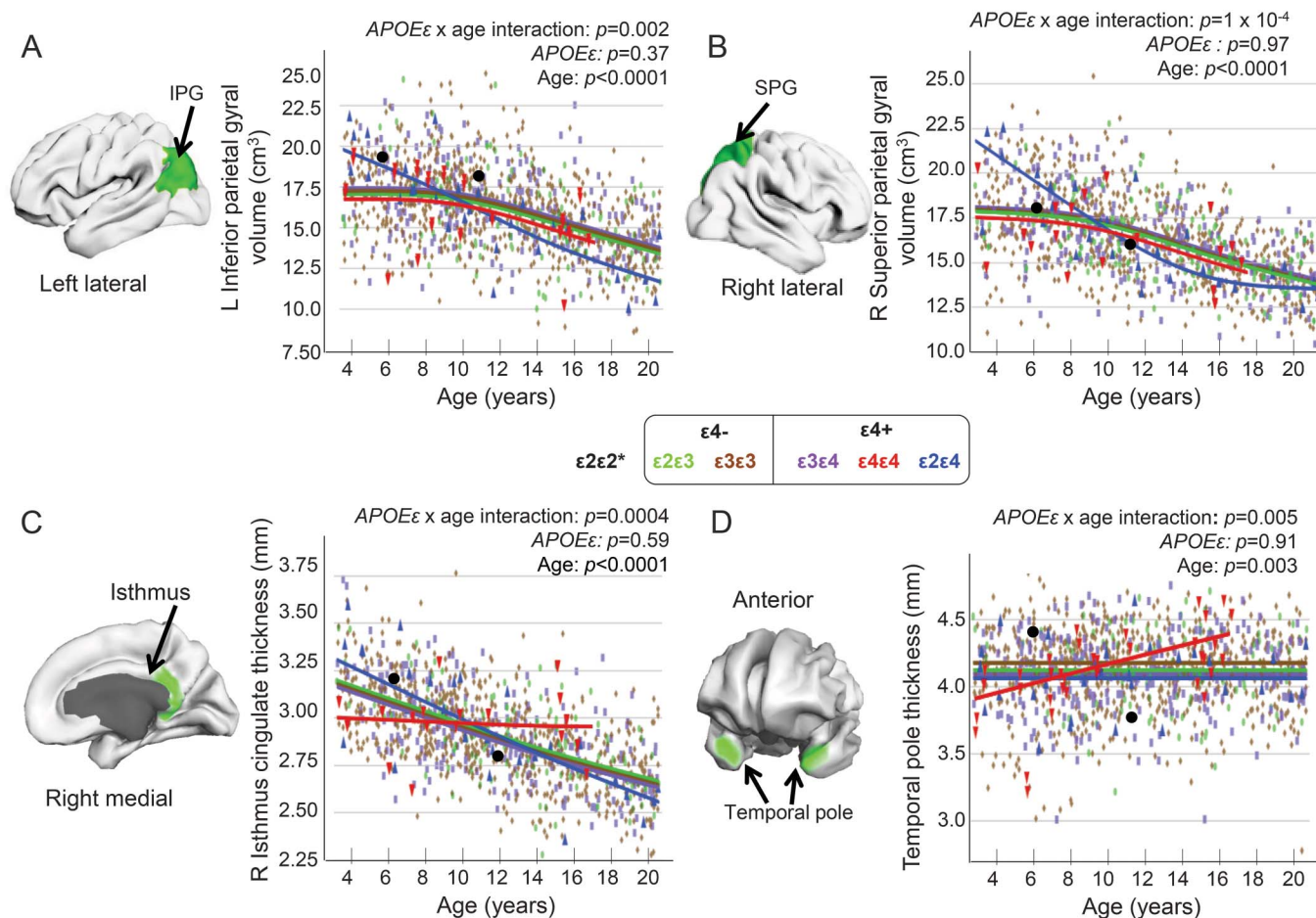
(A) The p value maps using the vertex model reveal significant APOE ϵ effects (red and yellow) on cortical surfaces of the insular cortex, the temporal poles, as well as the 3 right hemisphere regions (lateral occipital, medial orbitofrontal, and cuneus) shown to be different across genotypes on the region-of-interest model (see B–D). The $\epsilon 2\epsilon 4$ group showed the smallest cortical areas across the age range in the right lateral occipital cortex (B), right cuneus (C), and right medial orbitofrontal cortex (D). Post hoc analysis shows that, relative to the reference $\epsilon 3\epsilon 3$ group (brown), the $\epsilon 2\epsilon 4$ group (blue) had significantly smaller right medial orbitofrontal cortical areas ($p = 0.006$) and smaller right cuneus areas ($p = 0.002$), while the $\epsilon 3\epsilon 4$ group (purple) had larger lateral occipital ($p = 0.016$) and medial orbitofrontal cortical areas ($p = 0.006$). The $\epsilon 4\epsilon 4$ group (red) also had the largest areas among all groups in the right cuneus ($\epsilon 4\epsilon 4 > \epsilon 2\epsilon 4$, $p = 0.07$) and right lateral occipital area ($\epsilon 4\epsilon 4 > \epsilon 2\epsilon 4$, $p = 0.04$). The children with $\epsilon 2\epsilon 2$ (black) had relatively large cortical areas in the cuneus and right medial orbitofrontal regions, largest in the younger $\epsilon 2\epsilon 2$ child. See also table e-3 for additional results. *Data for the $\epsilon 2\epsilon 2$ children are not included in the group analyses.

$\epsilon 2\epsilon 4$ children showed the smallest areas, while $\epsilon 4\epsilon 4$ or $\epsilon 3\epsilon 4$ children had the largest areas (figure 2, B–D). The 2 children with $\epsilon 2\epsilon 2$ had exceptionally large areas. The selected cortical volumes decreased with age for all genotypes. Compared with other groups, $\epsilon 2\epsilon 4$ children had larger left inferior parietal gyrus and right superior parietal gyrus at younger age (<10 years) but smaller volumes during adolescence (figure 3, A and B).

For cortical thickness, the right isthmus cingulate showed age-dependent thinning, except for $\epsilon 4\epsilon 4$ children who showed no change with age (figure 3C). Conversely, temporal pole thickness was constant with age, except for $\epsilon 4\epsilon 4$ children who showed age-related increases, with thinner cortices in younger and thicker

cortices in older individuals (figure 3D). Furthermore, since prior reports compared $\epsilon 4$ carriers ($\epsilon 4\epsilon 4$, $\epsilon 4\epsilon 3$) with non- $\epsilon 4$ carriers ($\epsilon 3\epsilon 3$, $\epsilon 2\epsilon 3$),^{5,29} we performed vertex-based, whole-brain analyses using the same grouping. Compared to non- $\epsilon 4$ carriers, $\epsilon 4$ carriers had nonsignificantly thicker left entorhinal cortex (figure 4A, yellow region), verified on ROI analyses (figure 4B), and nonsignificantly thinner dorsal postcentral and lateral temporal cortices (figure 4A, lighter blue). However, cortical thickness showed a trend for group differences on age-dependent measures in parahippocampal regions and the left postcentral and entorhinal cortices (figure 4C, yellow regions). ROI analyses verified group differences in age-dependent thinning in the left entorhinal cortex. Specifically,

Figure 3 APOE ϵ -by-age interactions or APOE ϵ effects on cortical volumes and thickness



Brain regions with APOE ϵ -by-age interactions are shown (green). (A and B) The $\epsilon 2\epsilon 4$ group showed the largest average volumes in inferior and superior parietal cortices at younger age (<10 years), but the smallest average volumes during adolescence. (C) The isthmus of the cingulate showed age-dependent thinning in all children except for those with $\epsilon 4\epsilon 4$. (D) In contrast, all children showed relatively stable temporal pole thickness (averaged left and right) across the age range, except for the children homozygous for $\epsilon 4$. All models for regions of interest were generated from the general additive model with thickness or volume of the region of interest as dependent variable, covarying for sex, scanner device, socioeconomic status, and genetic ancestry factor. *Data for the $\epsilon 2\epsilon 2$ children are not included in the group analyses. IPG = inferior parietal gyrus; SPG = superior parietal gyrus.

$\epsilon 4\epsilon 4$ children showed the steepest slope ($r = -0.66$, $p = 0.02$), with thicker cortices in younger children but thinner cortices in older children (>11 years; figure 4D), as reported for adolescents.⁵

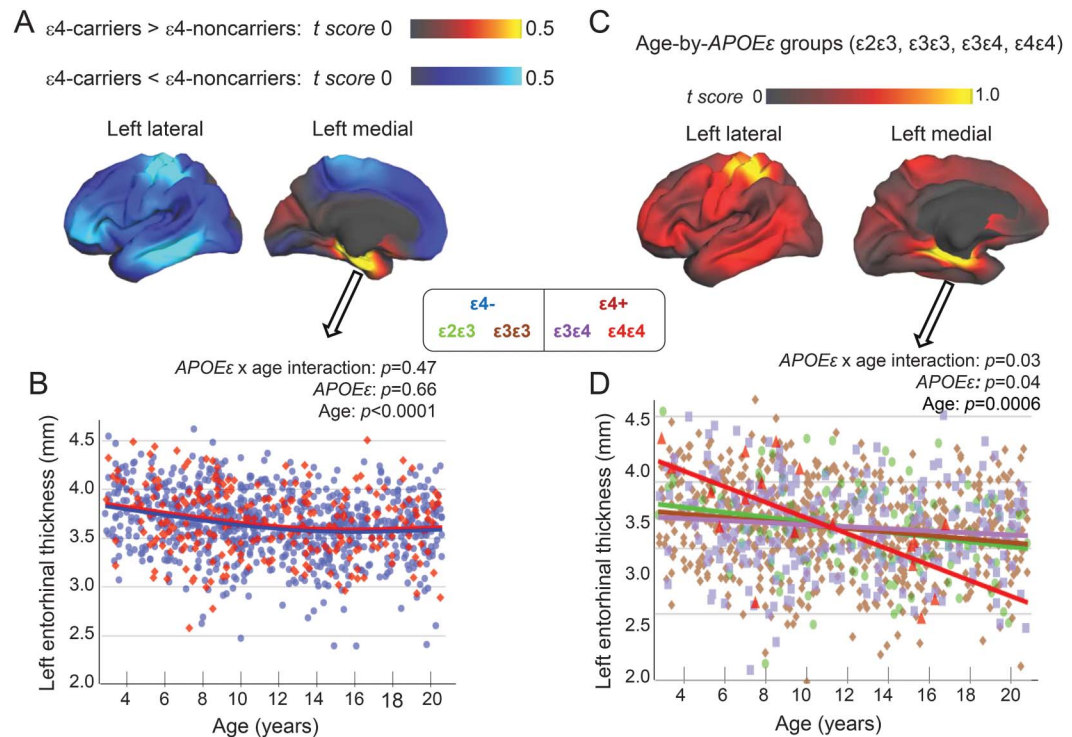
APOE ϵ genotypes and age on cognitive performance. Age-by-genotype interactions were found on executive function, attention, and WM (figure 5, A–C) but not episodic memory. Compared to other genotype groups, younger $\epsilon 4\epsilon 4$ children had lower scores on executive function and WM, but similar or better performance after age 8 years (figure 5, A and B). On the attention task, younger $\epsilon 2\epsilon 4$ children performed worse than other genotype groups, but their scores normalized after age 10 years (figure 5C).

Relationships between brain morphometry and cognition. After adjustments for sex, SES, and GAF, brain regions with age-related genotype variations also showed differential correlations with attention or WM. In the

right superior parietal gyrus, $\epsilon 2\epsilon 4$ children differed from other groups; those with larger superior parietal gyral volumes had the lowest attention ($r = -0.58$, $p = 0.001$; figure 5D) or WM scores ($r = -0.59$, $p = 0.0005$; figure 5E). Similarly, across genotypes, $\epsilon 2\epsilon 4$ children with thicker cortices had the poorest attention (right isthmus cingulate: $r = -0.62$, $p = 0.0002$, figure 5F; temporal pole: $r = -0.43$, $p = 0.01$, figure 5G), while $\epsilon 4\epsilon 4$ children with thinner temporal poles had poorer attention ($r = +0.5$, $p = 0.02$; figure 5G). All children with smaller hippocampal volumes had poorer WM performance, especially $\epsilon 4\epsilon 4$ children ($r = +0.48$, $p = 0.03$; figure 5H).

DISCUSSION The APOE ϵ gene is polymorphic with 3 alleles, with $\epsilon 3$ being the most common (approximately 78%), followed by $\epsilon 4$ (14%) and $\epsilon 2$ (8%).¹⁵ Evaluating all 6 APOE ϵ genotypes in a large cohort of typically developing children clarified the

Figure 4 Cortical thickness differences between $\epsilon 4$ carriers and non- $\epsilon 4$ carriers



These analyses were performed using the same grouping as prior studies.⁵ (A) On the vertex-based analyses (covaried for age, sex, scanner device, socioeconomic status, and genetic ancestry factor), compared to $\epsilon 4$ noncarriers (668 $\epsilon 3\epsilon 3+$ and 124 $\epsilon 2\epsilon 3$), $\epsilon 4$ carriers (235 $\epsilon 3\epsilon 4+$ and 21 $\epsilon 4\epsilon 4$) showed nonsignificantly thicker right precuneus (data not shown) and left entorhinal cortices (red-yellow regions), and nonsignificantly thinner left postcentral (parietal) and left lateral temporal cortices (blue areas). (B) Region-of-interest analysis of the left entorhinal region verified the nonsignificantly thicker cortex in $\epsilon 4$ carriers (red dots) than non- $\epsilon 4$ carriers (blue dots). (C) The 4 genotype groups showed a trend for group differences ($p = 0.13$) on age-dependent changes in cortical thickness in the left entorhinal and parahippocampal regions as well as the left postcentral cortex (yellow regions). (D) Region-of-interest analyses verified the age-dependent group differences; however, the $\epsilon 4\epsilon 4$ children (red dots) showed steepest age-dependent thinning in this brain region, with thicker cortices in the younger children but thinner cortices in the older children (>11 years).

associations of heterozygous vs homozygous $\epsilon 2$ and $\epsilon 4$ on brain development. The major findings are as follows: (1) compared to other genotype groups, $\epsilon 4\epsilon 4$, $\epsilon 2\epsilon 2$, and $\epsilon 2\epsilon 4$ children had altered age-related slopes in brain regions often affected in AD³⁰; (2) smaller hippocampal volumes in younger $\epsilon 2\epsilon 4$ children and lower hippocampal FA in younger $\epsilon 4\epsilon 4$ children mirror the smaller volumes and steeper age-dependent atrophy of the hippocampi in elderly $\epsilon 4$ and/or $\epsilon 2$ carriers³¹; and (3) the younger $\epsilon 2\epsilon 4$ and $\epsilon 4\epsilon 4$ children with altered age-related changes in brain measures also showed poorer performance on attention or WM tasks.

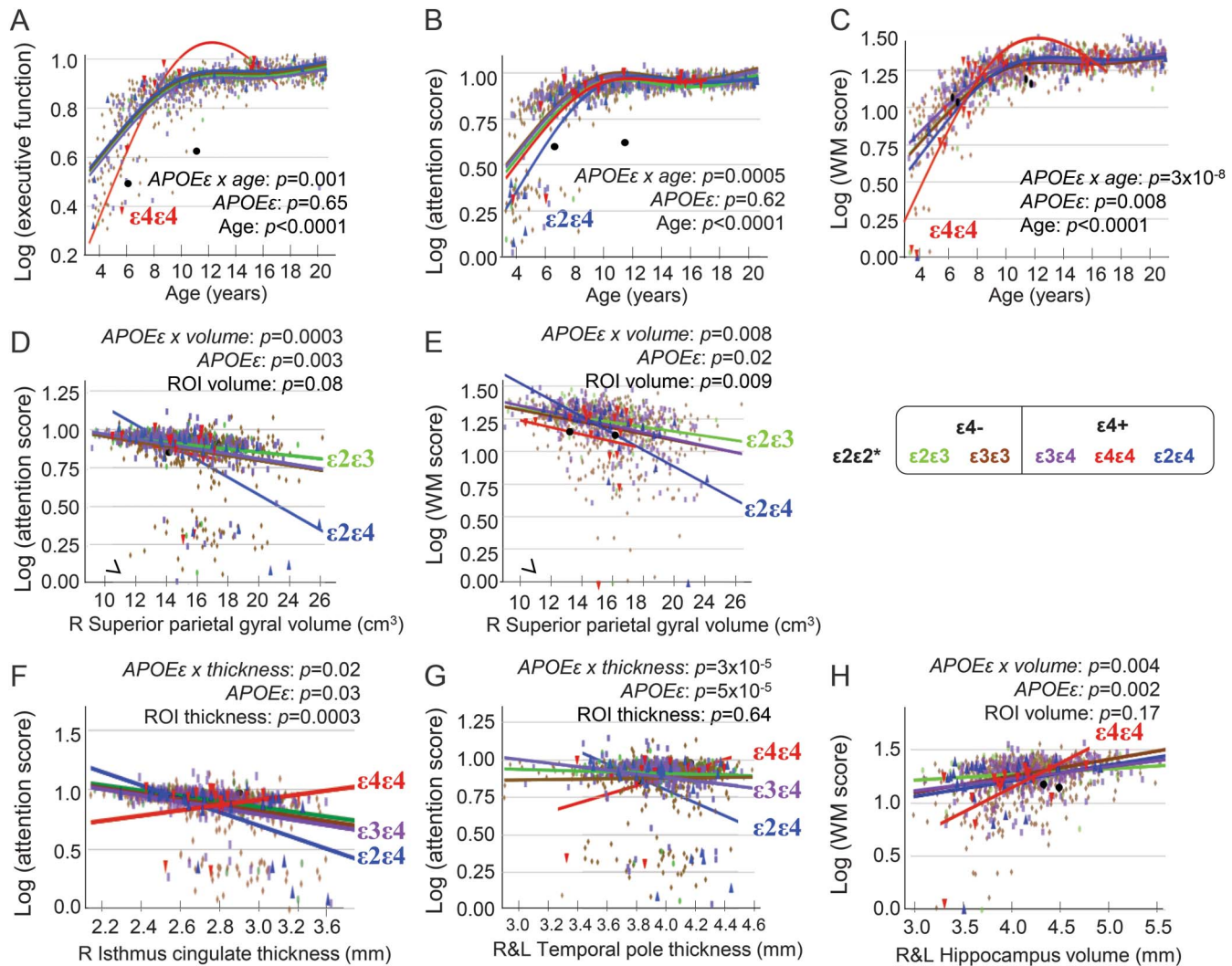
Children with the most common $\epsilon 3\epsilon 3$ genotype served as our reference group. These children showed typical age-related increases in hippocampal volumes³² and the medial orbitofrontal and occipital cortical areas until early adolescence.³³ They also showed the typical age-dependent increase in thalamic FA,²³ reflecting ongoing myelination, but not age-related increases in hippocampal and amygdala FA, as reported in healthy children without genotype groupings.²³ Age-dependent

decreases in parietal cortical volume and thinning of the isthmus likely reflect pruning of neuronal synapses and cell shrinkage.³³ Furthermore, our $\epsilon 3\epsilon 3$ children with thinner isthmus had similarly better performance in WM and attention.³⁴

The children with $APOE$ $\epsilon 2\epsilon 3$ genotype had comparable brain morphometry and cognitive performance as $\epsilon 3\epsilon 3$ children. However, younger (<7 years) $\epsilon 2\epsilon 3$ children, like those with $\epsilon 2\epsilon 4$ or $\epsilon 4\epsilon 4$, had relatively lower FA in left amygdala, suggesting lesser microstructural integrity, such as lower cellular density or lesser myelination. Since the amygdala is involved in emotional processing, these children may have greater vulnerability to emotional problems. Lower amygdala FA was also found in infants whose mothers had greater prenatal depressive symptoms³⁵; hence, future studies should include maternal depressive symptoms as a covariate to determine whether these symptoms might account for the lower amygdala FA in younger $\epsilon 2\epsilon 3$ children.

Relative to $\epsilon 3\epsilon 3$ children, those with one $\epsilon 4$ allele, specifically $APOE$ $\epsilon 3\epsilon 4$, had larger hippocampi, occipital and frontal cortical areas, and thicker

Figure 5 Cognitive performance in relation to age, brain volumes, or cortical thickness



APOE ϵ genotype-by-age interactions on executive function (A), attention (B), and WM (C). Children with larger right superior parietal gyral volumes had poorer attention scores (D) or WM scores (E) across all genotype groups, especially those with *APOE* $\epsilon2\epsilon4$. Similarly, those with thicker cortices in the isthmus cingulate cortical thickness (F) and temporal poles (G) had poorer attention and WM scores, especially children with *APOE* $\epsilon2\epsilon4$. In contrast, children with *APOE* $\epsilon4\epsilon4$ with thicker cortices had better attention and WM scores (F and G). All children with larger hippocampal volumes had higher WM scores (H), especially children homozygous for *APOE* $\epsilon4$ (H). *Data for the $\epsilon2\epsilon2$ children (black dots) are not included in the group analyses. ROI = region of interest; WM = working memory.

temporal poles, but similar cognitive functioning. These larger brain measures are consistent with an antagonistic pleiotropic effect of $\epsilon4$, similar to findings in middle-aged (51–59 years) heterozygous $\epsilon4$ carriers (primarily $\epsilon3\epsilon4$).^{3,10} Normal or thicker cortices in our $\epsilon3\epsilon4$ children contrast with prior findings of thinner left entorhinal and orbitofrontal cortices (92% $\epsilon3\epsilon4$ compared to $\epsilon3\epsilon3$ children).⁵ However, larger cortical areas in our $\epsilon3\epsilon4$ children resemble larger parietal volumes in $\epsilon3\epsilon4$ infants (ages 8.5–14 months)⁷ relative to $\epsilon3\epsilon3$ participants. Moreover, consistent with our results, prior studies found similar IQ or cognitive performance between $\epsilon3\epsilon4$ and $\epsilon3\epsilon3$ children.^{5,13}

Despite the relatively normal hippocampal volumes, the younger $\epsilon4\epsilon4$ participants had the lowest

FA in hippocampus, amygdala, and thalamus, suggesting slower development initially with lesser myelination or lower cellularity in these regions. These children also did not show typical age-dependent cortical thinning in posterior cingulate (isthmus) cortex, which suggests aberrant brain maturation, possibly due to reduced synaptic pruning. Such possible aberrant brain maturation with lesser cortical thinning was found in children with prenatal alcohol exposure and fetal alcohol syndrome,³⁶ although such children were excluded in the current study. Furthermore, $\epsilon4\epsilon4$ children showed age-dependent thickening of the temporal pole, which along with the isthmus, is often affected in AD^{30,37} and in cognitively healthy $\epsilon4$ carriers,³⁰ who showed age-dependent increases in amyloid deposition.¹⁸

Younger $\epsilon 4\epsilon 4$ children showed the poorest executive function and WM among groups, even after SES adjustment, similar to school-age $\epsilon 4$ carriers with AD family history.¹⁴ The normal cognition of our older $\epsilon 4\epsilon 4$ children is consistent with prior findings in older $\epsilon 4\epsilon 4$ children^{5,13} and in young and middle-aged adults.¹⁰ The $\epsilon 4\epsilon 4$ children with smaller hippocampal volumes and thinner temporal pole had poorer WM and attention; these findings resemble the thinnest frontal cortices²⁹ in middle-aged $\epsilon 4\epsilon 4$ individuals without dementia, who showed the most rapid decline on mental arithmetic tasks requiring WM.²⁶ Hence, $\epsilon 4$ homozygosity might slow maturation of the hippocampus and cortical thickness, which in turn might negatively affect WM and attention. Mirroring these findings, older individuals with $\epsilon 4$ homozygosity had the highest prevalence for AD (50%–91%)^{4,15} and the greatest hippocampal and temporal lobe atrophy³⁰ among genotypes.

Unlike $\epsilon 3\epsilon 4$ children with the largest hippocampi, $\epsilon 2\epsilon 4$ children had the smallest hippocampi and orbitofrontal and occipital surface areas among groups and across ages. Therefore, $\epsilon 4$ allele effects differ greatly when combined with $\epsilon 2$ vs $\epsilon 3$. Although the younger $\epsilon 2\epsilon 4$ children had larger parietal cortices and thicker posterior cingulate cortices, they had poorer attention and WM. These findings also mirror those in the 3 $\epsilon 2\epsilon 4$ oldest old (>90 years) among 89 participants, since all 3 met criteria for dementia but not neuropathology for AD.³⁸ While $\epsilon 2$ carriers showed reduced cognitive decline, $\epsilon 2\epsilon 4$ is a risk genotype for AD across ethnicity.¹⁵ Unfortunately, $\epsilon 2\epsilon 4$ participants are often excluded from studies because of potentially opposite effects of $\epsilon 4$ and $\epsilon 2$.^{11,12,21,38}

In the $\epsilon 2\epsilon 2$ children, the relatively large hippocampi are similar to the case reports of $\epsilon 2\epsilon 2$ adults,³¹ and the lower hippocampal FA in the younger child suggests less coherent fibers in the large hippocampi. They also had relatively higher thalamic FA, similar to findings in $\epsilon 2$ heterozygous adults.³⁹ Our $\epsilon 2\epsilon 2$ children also had poorer attention and executive function, while $\epsilon 2\epsilon 2$ children in a prior study showed above-average IQ.¹³ Furthermore, a 92-year-old $\epsilon 2\epsilon 2$ woman showed no cognitive deficits until her stroke, despite the postmortem finding of prominent AD neuropathology.¹⁶ The relative preservation of cognition in aging $\epsilon 2\epsilon 2$ individuals may be due to ApoE $\epsilon 2$'s antioxidant, anti-inflammatory, and antiproteolytic effects.¹⁷ However, these processes may not be relevant in the developing brain.

For comparison with the literature, we also evaluated all $\epsilon 4$ carriers. Hippocampal volumes in $\epsilon 4$ carriers ranged from smaller ($\epsilon 2\epsilon 4$) to no difference ($\epsilon 4\epsilon 4$) or slightly larger ($\epsilon 3\epsilon 4$) relative to $\epsilon 3\epsilon 3$ children. Similarly, cortical surface areas were smallest in $\epsilon 2\epsilon 4$ but largest in $\epsilon 3\epsilon 4$ or $\epsilon 4\epsilon 4$. Hence,

combining all $\epsilon 4$ carriers might have attenuated or abolished group differences compared to non- $\epsilon 4$ carriers,^{5–7,12,14,40} and the larger parietal volumes in the youngest $\epsilon 2\epsilon 4$ children or the thinnest isthmus gyrus and temporal poles in the youngest $\epsilon 4\epsilon 4$ children might have been missed. In fact, our $\epsilon 4$ carriers collectively had nonsignificantly thicker temporal lobes, contrasting with thinner entorhinal cortices in a prior study of healthy $\epsilon 4$ children.⁵ However, further analysis showed that $\epsilon 4\epsilon 4$ children had steeper age-dependent thinning in this region, leading to thinner entorhinal cortices in adolescents, similar to prior findings.⁵ Moreover, our youngest $\epsilon 4$ carriers performed poorer on WM than the non- $\epsilon 4$ carriers, which resemble WM deficits in older healthy $\epsilon 4$ carriers.²⁵

The age-dependent brain measures of $\epsilon 4\epsilon 4$ and $\epsilon 2\epsilon 4$ children often deviated from those in the other genotype groups. The youngest children with one of these genotypes had less mature brain structures and poorer cognitive function, but tended to normalize or exceed the other genotype groups during late adolescence. Prior studies of $\epsilon 4\epsilon 4$ and $\epsilon 2\epsilon 4$ young adults showed larger specific brain structures and better cognitive performance, whereas older adults showed poorer cognitive performance or less efficient neural networks relative to other genotype groups.^{9,10} Incidentally, $\epsilon 2\epsilon 4$ participants have a low odds ratio of AD until age 50 years, but the highest odds at age 70 years,¹⁵ while $\epsilon 4$ homozygosity leads to earliest AD onset (approximately 68 years).⁴

This study has several limitations. Despite this relatively large sample, the age-related brain measures may be biased by the cohort effect, driven by participants with these rare genotypes at the extremes of age range, in this cross-sectional study. Longitudinal follow-ups are needed to confirm the true developmental trajectories in the less prevalent $\epsilon 4$ or $\epsilon 2$ carriers. In addition, some younger children (<5 years) could not perform all NIH Toolbox tasks; future studies with more young children using age-appropriate assessments are needed. Lastly, although we covaried for GAF, ethnicity may influence the effects of $\epsilon 4$ and $\epsilon 2$ on brain and cognitive measures.^{4,15} However, repeating all the analyses only in children with >50% European ancestry yielded similar results (table e-4, figures e-1 to e-3). Future studies should include a larger sample of children with other ancestry and evaluate them separately.

This large sample of children validated and extended prior smaller studies that showed altered brain development in $\epsilon 4$ carriers. The $\epsilon 4\epsilon 4$ and $\epsilon 2\epsilon 4$ carriers appear to show the strongest antagonistic pleiotropic effects, with negative influences on brain structures and cognition at younger age, mirroring those in elderly participants and patients with

AD. Future studies of *APOE* ϵ should evaluate each genotype separately, since brain development, and possibly brain aging and recovery, may vary substantially across specific $\epsilon 4$ or $\epsilon 2$ genotypes. Finally, studying *APOE* ϵ polymorphism in young children may provide early indications of risk of future brain injuries and dementia. Given the urgent need to determine how early patients with AD should receive interventions or preventive treatments, a thorough understanding of how AD risk genes, such as *APOE* $\epsilon 4$, might independently or interactively influence the brain across the ages, is needed.

AUTHOR CONTRIBUTIONS

Linda Chang, Vanessa Douet, and Thomas Ernst developed the concept, designed the study, performed the statistical analyses, interpreted the data, drafted and revised the manuscript. Linda Chang, Thomas Ernst, Alexandra Pritchett, Kristin Lee, Terry Jernigan, Natacha Akshoomoff, Sarah Murray, Cinnamon Bloss, David Kennedy, Jean Frazier, David Amaral, Jeffrey Gruen, Walter Kaufmann, B.J. Casey, and Elizabeth Sowell were all involved in the data acquisition. Sarah Murray was responsible for all genotyping and genetic imputations. The USCD team was responsible for all image processing. All authors were involved in the critical revision and approval of the manuscript.

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DISCLOSURE

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