Comparison of lymphocyte transformation and macrophage migration inhibition tests in the detection of beryllium hypersensitivity

WR WILLIAMS, W JONES WILLIAMS

From the Asthma Research Unit, Sully Hospital, Penarth and the Welsh National School of Medicine, Department of Pathology, Llandough Hospital, Penarth

SUMMARY In seven patients with chronic beryllium disease (Be) the Be lymphocyte transformation test was positive in 100%, independent of steroid therapy, and was reproducible. The Be macrophage migration inhibition test was only positive in four of seven patients (57%) not on steroids, and was not reproducible. In 72 potentially exposed healthy beryllium workers the lymphocyte transformation test was negative in all subjects. The macrophage test was positive in four of 78 and again the results were not reproducible. The workers with positive results showed no differences in age, type or duration of employment from those with negative results and showed no evidence of disease. In addition, the macrophage test was positive in two of 45 non-exposed control subjects.

We also confirmed the above advantages of the lymphocyte transformation technique by using tuberculin antigen (PPD). The PPD lymphocyte transformation test gave positive results in approximately 60% of healthy beryllium workers, but the PPD macrophage test was only positive in 7%.

We conclude that the Be lymphocyte transformation test is the most sensitive and reproducible and advocate its use in the diagnosis of disease and in monitoring the health of potentially exposed workers.

Delayed-type hypersensitivity reactions may be measured in vitro using lymphocyte cultures.¹ These tests are particularly relevant to beryllium disease as they avoid skin testing with an extremely potent sensitiser. Hypersensitivity is an important feature of chronic beryllium disease, and although not diagnostic, is of value in diagnosis.² Other applications of in vitro beryllium hypersensitivity tests include: (*a*) screening tests to detect the development of sensitivity in potentially exposed workers, who may be at increased risk of developing disease;

(b) as an indicator of industrial hygiene standard.

Two in vitro tests, macrophage migration inhibition and lymphocyte transformation, have been reported to be of value for detecting beryllium hypersensitivity (Table 1), although influenced to different degrees by steroid therapy. Previously, a survey of a beryllium metal factory using the Be macrophage migration inhibition test⁶ showed that 14% of workers were sensitised. However, the test was positive in only one of six patients.

In the present survey we have compared the

Table 1	Previous repo	rts of "in v	itro" beryllium
hypersens	sitivity tests		

Macrophage migration inhibition	Lymphocyte transformation		
Jones Williams et al ³	Hanifin <i>et al</i> ⁸		
2/7 patients positive (on steroids)	7 patients all positive		
Henderson et al ⁴	Deodhar ⁹		
3 patients positive	25/35 patients positive (on steroids)		
Marx and Burrell ⁵	Nishikawa et al ¹⁰		
6/7 patients positive (on steroids)	4 patients all positive (not on steroids)		
Price et al ⁸	Preuss ¹¹		
50 workers 14% positive	571 workers: 36% weak positives 		
1/6 patients positive (other patients on steroids)	37/47 patients positive—strong and persistent (on steroids)		

sensitivity and reproducibility of the two tests in a reinvestigation of the original factory, workers from a ceramic factory and patients with chronic beryllium disease. We also assessed the reliability of the two tests by the use of tuberculin antigen in the factory workers. In addition, the beryllium and tuberculin migration inhibition tests were performed on control subjects and patients with tuberculosis.

Material and methods

Details of the Be macrophage migration inhibition⁶ and Be lymphocyte transformation tests⁷ have previously been described.

Beryllium sulphate (BeSO₄) was used at $10^{-6} - 10^{-7}$ M and tuberculin (PPD; MAFF, Weybridge) at concentrations of 20 µg/ml (MIF test) and 7.5 µg/ml (LT test).

A MIF index

 $\frac{\text{(area of migration with antigen)}}{\text{(area of migration without antigen)}} \text{ of } < 0.8$

was taken as a positive value. In LT tests, a stimulation index (SI) =

$$\frac{(\text{cpm in cultures with antigen})}{(\text{cpm in cultures without antigen})} > 2.0$$

was taken as a positive test.

Various modifications to our standard MIF test, based on the technique of Rocklin¹² were made. A more pure lymphocyte population was obtained by preincubating Ficoll-Hypaque separated mononuclear cells for two hours in MEM with 5% human serum, in plastic Petri dishes. Most monocytes were removed by adherence.

Lymphocyte supernatants (lymphokines) were concentrated ($\times 3 - \times 39$) as follows. Lymphocytes were cultured for one to three days in flat-bottomed plastic tubes with or without PPD (5 μ g/ml). The medium was changed daily, pooled and control medium reconstituted with PPD prior to filtration (pore size <0.2 μ m) and dialysed. The contents of dialysis bags were evaporated to dryness with an air fan. These lymphokine preparations were reconstituted in MEM with 15% complement inactivated guinea pig serum and contained 15-105 μ g/ml PPD.

SUBJECTS

Beryllium antigen

Beryllium patients The two tests were performed on seven patients, fulfilling most of the criteria for diagnosis.² All were men working with beryllium

metal, alloys, or both. The predominant symptoms included, to varying degrees, lethargy, dyspnoea, cough and weight loss. All showed radiological evidence of pulmonary fibrosis. Kveim tests were made to assist in excluding sarcoidosis. Positive biopsies showed epithelioid cell granulomas (Table 2).

Beryllium workers The in vitro tests were compared in 72 factory workers. Factory No 1: 35 workers, all engaged in machining pure beryllium, including 30 previously tested.⁶ Factory No 2: a total of 37 workers involved in ceramic manufacture using beryllium oxide.

Controls The macrophage test was further assessed in control subjects. These included 35 hospitalised male patients, who were tested before or 10 days after undergoing surgery (for example, piles, varicose veins, hernias, etc). Ten were patients with tuberculosis. They were all within the working population age group, and none had any known exposure to beryllium.

Tuberculin antigen

The macrophage test was also assessed in the same factory population, controls and tuberculosis patients. The lymphokine concentration modification of the macrophage test was carried out on 12 additional patients with tuberculosis and seven tuberculin-positive normal subjects.

Results

BERYLLIUM ANTIGEN

Chronic beryllium patients (Table 3)

The macrophage test was positive in four of seven (57%) and the transformation test was positive in all seven (100%). The transformation results were unaffected by steroid treatment, but in two of the six patients on steroids the migration test was negative.

Of three patients previously reported as macrophage migration-positive^{3 6} only one was positive on this occasion.

Table 2Details of chronic beryllium patients

Patient	Age (yr)	Length of employment (yr)	Kveim test	Mantoux test	Be patch test	Be urine/tissues	Biopsy
1	47	10	-	ND	+	ND	+
;	34	13	-	ND	+	-/-	-
3	59	20	-	-	-	-/ND	ND
4	37	7	ND	ND	ND	-/ND	ND
5	48	4	_	-	+	-/+	+
6	43	18	_	-	+	-/ND	+
7	54	3	_	ND	-	ND	+

Table 3	Comparison of Be MIF and Be LT tests on
patients	with chronic bervllium disease

Patients	Be MIF ratio	Be LT (20 % serum)		
		CPM with Be	SI .	
1*	0.78+	5496	2.9+	
2*	0.78+	11070	3.7+	
3*	0.72+	7850	10.0+	
4*	0.65+	5700	7·6 +	
5*	0.86-	36557	17.4+	
6*	0.98	14620	9.3+	
7	0.93 -	13871	22.3+	
Positive rate	4/7		7/7	

*on steroids; + = positive; - = negative; CPM = counts per minute; SI = stimulation index.

Healthy beryllium workers (Table 4)

The macrophage test was initially positive in 4 of 72 (5%) tested. Two of the four were available for retesting and one gave a negative result.

We were able to compare the results of macrophage tests in four workers previously reported as positive, Be MIF ratio $<0.87.^{6}$ Three were initially positive but on repeat testing all were negative. Four workers not examined in the 1977 survey had MIF ratios <0.87, but only two were positive at <0.8(included in Table 4).

In all 72 workers, the lymphocyte transformation results were negative.

Control population

The macrophage test was positive in two of 45 (4%) subjects with no known beryllium exposure.

TUBERCULIN ANTIGEN

The PPD lymphocyte transformation test (Table 4) was positive in 58% of the healthy factory population and in all seven patients with chronic beryllium disease. The PPD macrophage test, however, showed a very low positive rate (7%) in the factory population.

In addition, the tuberculin macrophage test was only positive in one of six patients with tuberculosis and in only two of 45 of the control group.

The modified tuberculin macrophage migration inhibition test in patients with tuberculosis was positive in only two (0.4 and 0.74) of 12 using three-

day culture supernatants and in only one (0.79) of four using one-day cultures, and were independent of lymphokine and PPD concentrations. In tuberculin skin-positive subjects there were no positive results (0 of 6) using one-day culture supernatants, and only one (0.78) of five was positive with three-day culture supernatants. This test was again therefore unsatisfactory.

Discussion

Our comparison of two in vitro beryllium hypersensitivity tests have demonstrated the advantages of the lymphocyte transformation test. The beryllium transformation test gave a positive result in all seven patients with chronic beryllium disease, and was independent of steroid therapy. In contrast, the Be macrophage test was not reproducible and in three patients, two on steroids, gave negative results.

In healthy potentially exposed workers, from two factories, the Be transformation test was negative in all 72 tested. The Be macrophage test was positive in four of 78 (3%) and of two retested, one was negative. These positive workers were indistinguishable from those with negative results, as regards details of potential exposure and length of employment, and showed no evidence of beryllium or other disease.

Although the two factories were using beryllium in different manufacturing processes, the results of both Be tests and the PPD macrophage test were very similar. Positive PPD transformation tests were, however, fewer in the ceramic factory (50%) than in the machining factory (65%). In the control healthy subjects, the Be macrophage test was positive in two subjects (4%), neither of which had any known beryllium exposure.

The reliability of the tests was also assessed using tuberculin. The PPD transformation test was positive in 58% of the healthy factory workers. This incidence compares with the 60% positive Mantoux reaction found in the factory 1 population.⁶ The PPD macrophage test gave an unacceptably low positive rate in the factory population, tuberculosis patients and Mantoux-positive subjects. Attempts to improve the technique by concentrating lymphokine supernatants were ineffective.

 Table 4 Comparison of the macrophage migration inhibition, MIF, and lymphocyte transformation (LT) tests (Be and PPD) in healthy beryllium workers

Factory	Positivity	Be		PPD	
		MIF	LT	MIF	LT
1	+	2	0	2	26
	_	33	35	26	14
2	+	2	0	3	17
	-	35	37	36	17
Positive rate:		4/72 (5%)	0/72	5/67 (7%)	43/74 (58%)

Our results contrast with those of Rocklin¹² who found the MIF assay of human lymphocyte supernatants, in most instances, a reliable test for assessing tuberculin hypersensitivity. Even so, the sensitivity of the Rocklin technique does not match that of the direct macrophage migration inhibition test for detecting hypersensitivity in experimental animals.¹³ Loss of sensitivity may be associated with dialysis of lymphokine preparations which tend to disrupt MIF molecules.¹⁴

We conclude that the lymphocyte transformation test is superior to the macrophage migration inhibition assay as a means of detection of hypersensitivity using both beryllium and tuberculin antigen. Also, the transformation tests is technically easier and does not require laboratory animals. We therefore advocate the use of the beryllium lymphocyte transformation test in the diagnosis of beryllium disease and in monitoring the health of potentially exposed workers.

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References

- ¹ Bice DE, Lopez M, Rothschild H, Salvaggio J. Comparison of Candida-delayed hypersensitivity skin test size with lymphocyte transformation, migration inhibitory factor and antibody titre. *Int Arch Allergy Appl Immunol* 1974;47:54-62.
- ² Jones Williams W. Beryllium disease—pathology and diagnosis. J Soc Occup Med 1977;27:93-6.
- ³ Jones Williams W, Grey J, Pioli EM. Diagnosis of chronic

beryllium disease. Br Med J 1972;iv:175.

- ⁴ Henderson WR, Fukuyama K, Epstein WL, Spitler LE. "In vitro" demonstration of delayed hypersensitivity in patients with berylliosis. J Invest Dermatol 1972;58:5-7.
- ⁵ Marx JJ, Burrell R. Delayed hypersensitivity to beryllium compounds. J Immunol 1973;111:590-8.
- ⁶ Price CD, Jones Williams W, Pugh A, Joynson DH. Role of "in vitro" tests of hypersensitivity in beryllium workers. J Clin Pathol 1977;30:24-8.
- ⁷ Williams WR Jones Williams W. Development of beryllium lymphocyte transformation test in chronic beryllium disease. Int Arch Allergy Appl Immunol 1982;67:175-80.
- 8 Hanifin JM, Epstein WL, Cline MJ. "In vitro" studies of granulomatous hypersensitivity to beryllium. J Invest Dermatol 1970;55:284-8.
- ⁹ Deodhar SD, Barna B, van Ordstrand HS. A study of the immunological aspects of chronic berylliosis. *Chest* 1973;63:309-13.
- ¹⁰ Nishikawa S, Hirata T, Kitaichi M, Izumi I. Three year prospective study of Mantoux reactions in factory workers exposed to beryllium oxide. In: Jones Williams W, Davies BH, eds. Proceedings of the 8th International Conference on sarcoidosis and other granulomatous diseases. Cardiff: Alpha & Omega Press, 1980:722-7.
- ¹¹ Preuss OP, Deodhar JD, van Ordstrand HS. Lymphocyte transformation in beryllium workers. In: Jones Williams W, Davies BH, eds. Proceedings of the 8th International Conference on sarcoidosis and other granulomatous diseases. Cardiff: Alpha & Omega Press, 1980:711-4.
- ¹² Rocklin RE, Meyers OL, David JR. An "in vitro" assay for cellular hypersensitivity in man. J Immunol 1970;104: 95-102.
- ¹³ David JR, Al-Askari S, Lawrence HS, Thomas L. Delayed hypersensitivity "in vitro". J Immunol 1964;93:264-73.
- ¹⁴ Possanza G, Cohen MC, Yoshida T, Cohen S. Human macrophage migration inhibition factor: evidence for subunit structure. Science 1979;205:300-1.

Requests for reprints to: Dr W Jones Williams, Department of Pathology, Llandough Hospital, Penarth, Glam CF6 1XX, Wales.