Technical method

A selective enrichment broth for the isolation of *Clostridium difficile*

SHEILA O'FARRELL, M WILKS, JQ NASH, SOAD TABAQCHALI Department of Medical Microbiology, St Bartholomew's Hospital, London EC1

The cycloserine, cefoxitin, fructose agar medium (CCF agar) devised by George *et al*¹ and the modification of it where egg yolk is replaced by horse blood (Oxoid CCF agar), have been widely and successfully used for the isolation of Clostridium difficile from faecal specimens.¹⁻⁴ For epidemiological and environmental work where only small numbers of organisms may be present, it is likely that a more sensitive method of culture is required. Hafiz et al used a broth containing p-cresol in studying vaginal carriage of C difficile⁵ whilst Wilson et al⁶ have reported that the incorporation of sodium taurocholate in CCF agar (in place of egg yolk) enhances the recovery of spore forms of C difficile from solid media. We describe the use of cycloserine cefoxitin fructose broth containing 0.1% sodium taurocholate (CCFT broth) for the isolation of Cdifficile from vaginal and faecal specimens.

Material and methods

MEDIA

Solid media

1 Reinforced clostridial agar (RCA, Oxoid) supplemented with 0.2% p-cresol (BDH)—group II patients only.

2 Cycloserine cefoxitin fructose blood agar (CCF agar Oxoid).

Enrichment broth

The broth was made up in our laboratory to the same formula as the commercial CCF medium (Oxoid), but omitting the agar and including 1 g/l of sodium taurocholate.

PATIENTS

Five groups of subjects were studied independently (Table). Groups I-III were unselected consecutive

Accepted for publication 23 August 1983

women attending the Department of Genital Medicine (DGM). Groups IVa and IVb were mothers attending the maternity unit. Predelivery high vaginal swabs were taken by the midwife at onset of labour. Post delivery vaginal swabs were taken just prior to discharge. Group V were the babies delivered to mothers in group IV.

Culture method

Swabs were inoculated onto solid media and then discarded. When enrichment cultures were performed (see Table), a duplicate swab was broken off into the CCFT broth.

Cultures were incubated in an anaerobic cabinet $(10\% \text{ CO}_2)$ for 7 days. Plates were examined for typical *C difficile* colonies after 48 h and seven days. Suspect colonies were subcultured to blood agar for "purity" prior to further tests.

ENRICHMENT

CCFT broths were subcultured to blood agar after 48 h and seven days. Suspect colonies on blood agar after 48 h were streaked out for purity and treated as for the direct plate subcultures. Isolates were identified as *C difficile* by their typical colonial and Gram stain morphology, distinctive odour and pattern of volatile fatty acid production as detected by gas liquid chromatography.⁷

Results and discussion

High vaginal swabs

The results are summarised in the Table. Only on C*difficile* isolation was made from 132 high vaginal swabs examined using CCF agar in groups I and II. No positive isolates were obtained using the RCA medium containing p-cresol (group II). However the use of CCFT broth greatly increased the isolation rates from vaginal swabs. In groups III and IVa the isolation rates from broth were 11% and 18% respectively compared with 1.2% and 0% from CCF agar alone. These differences in isolation rates are significant (p < 0.01). In group IVb (post-delivery mothers) CCFT broth was again superior to CCF agar but the total numbers are small and not statistically significant. However, when the overall isolation rates from vaginal specimens are examined for groups II, III and IV combined, the rate of isolation from CCFT 22/177 (12%) is significantly greater than that from CCF agar 3/177 (p < 0.001).

Technical methods

Isolation rates of C difficile in five groups of subjects

| Group | No of subjects | Description of subjects | Specimen | Media | Isolation rate |
|--------|----------------|-------------------------------------|--|------------------------------------|------------------------------------|
| I | 62 | Consecutive women attending DGM* | Single HVS | CCF agar | 0/82 |
| II | 90 | Consecutive women attending DGM* | Single HVS | CCF agar RCA + 0.1% p-cresol | 1/50 0/50 |
| III | 82 | Consecutive women attending DGM* | Duplicate HVS | CCF agar CCF broth | 1/82 (1·2%) p < 0·01 9/82 (11%) |
| IV (a) | 90 | Mothers predelivery | Duplicate HVS | CCG agar CCF broth | 0/50 (0%) p < 0.0027 9/50 (18%) |
| IV (b) | 45 | Mothers | Duplicate HVS | CCF agar | 2/45 (4·4%) 4/45 (8·8%) |
| v | 50 | Neonates 2nd-5th day | Swab from rectum or soiled nappy | CCF agar CCF broth | 29/50 (58%) 28/50 (56%) |

*Department of Genital Medicine.

p was derived from standard error (SE) using SE of differences in proportion = $\int \frac{p(100 - p)}{p(100 - p)} +$

HVS = high vaginal swab.

NEONATAL STOOL SPECIMENS (GROUP V) In contrast to the results obtained with vaginal swabs, the isolation rate obtained using CCFT broth was not significantly different to that obtained using CCF agar. This may be explained by the relatively high counts of *C difficile* organisms present in many faecal specimens and by the occasional failure of CCFT broth to yield *C difficile* when overgrowth of

"coliform" organisms had taken place in the broth. In conclusion we suggest that a liquid culture medium, such as that described here, is a useful addition to conventional agar culture when only small numbers of C difficile organisms are likely to be present.

References

¹ George, LW, Sutter, LV, Citron, D, Finegold, SM. Selective and differential medium for isolation of *C difficile. J Clin Microbiol*

1979;9:214-9.

² Enevold Falsen E, Bertil Kaijger B, Lars Nehls L, Börje Nygren B, Svedhem A. *Clostridium difficile* in relation to enteric bacterial pathogens. J Clin Microbiol 1980;12:297-300.

p (100 – p)

- ³ Holst E, Helin I, Mardh P-A. Recovery of *Clostridium difficile* from children. *Scand J Infect Dis* 1981;13:41-5.
- ⁴ Nash JQ, Chattopadhyay B, Honeycombe J, Tabaqchali S. Clostridium difficile and cytotoxin in routine faecal specimens. J Clin Pathol 1982;35:561-5.
- ⁵ Hafiz S, McEntegart MG, Morton RS, Waitkim SA. C difficile in the urogenital tract of males and females. Lancet 1975;i:420-1.
- ⁶ Wilson KH, Kennedy MJ, Fekety FR. Use of sodium taurocholate to enhance spore recovery in a medium for *Clostridium difficile. J Clin Microbiol* 1982;15:443-6.
- ⁷ Holdemann LV, Cato EP, Moore WEC. Anaerobic laboratory manual 4th ed. Blacksburg, Virginia: Virginia Polytechnic Institute, 1977.

Requests for reprints to: Dr JQ Nash, Department of Medical Microbiology, St. Bartholomew's Hospital, West Smithfield, London EC1A 7BE, England.

Rapid demonstration of nucleic acids using "oxidised" gallocyanin and chromic potassium sulphate: methods and applications

OAN HUSAIN, KC WATTS Department of Cytopathology, Charing Cross Hospital, London W6

Studies of metallic salt lakes of the oxazine dyes gallamin blue, celestine blue and gallocyanin led to the introduction of techniques for selective staining of Nissl and nuclear substance in nerve cells¹ and specific nuclear stains²⁻⁴ and to the theory of gal-

Accepted for publication 21 September 1983

locyanin chromalum staining and its application for quantitative estimation of basophilia.⁵

The gallocyanin chromalum technique of Einarson¹ was reviewed by us for use with the Quantimet image analysing computer⁶ and has now been adopted as a nuclear stain for the automated interactive cervical cancer screening system CERVIFIP where detection by integrated optical density is utilised, but its use in routine service conditions is limited by the fact that the staining technique requires incubation at 42°C for 16 h.⁷

We have therefore conducted experiments to reduce the staining time required to a minimum but still retain its degree of stoichiometry and stain density. Our results have also shown that the rapid staining technique is useful for staining cells other than those exfoliated from the cervix, namely