



REVIEW PAPER

# Opportunities and challenges in the subsoil: pathways to deeper rooted crops

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## Abstract

**Greater exploitation of subsoil resources by annual crops would afford multiple benefits, including greater water and N acquisition in most agroecosystems, and greater sequestration of atmospheric C. Constraints to root growth in the subsoil include soil acidity (an edaphic stress complex consisting of toxic levels of Al, inadequate levels of P and Ca, and often toxic levels of Mn), soil compaction, hypoxia, and suboptimal temperature. Multiple root phenes under genetic control are associated with adaptation to these constraints, opening up the possibility of breeding annual crops with root traits improving subsoil exploration. Adaptation to Al toxicity, hypoxia, and P deficiency are intensively researched, adaptation to soil hardness and suboptimal temperature less so, and adaptations to Ca deficiency and Mn toxicity are poorly understood. The utility of specific phene states may vary among soil taxa and management scenarios, interactions which in general are poorly understood. These traits and issues merit research because of their potential value in developing more productive, sustainable, benign, and resilient agricultural systems.**

**Key words:** Calcium, carbon sequestration, crop breeding, nitrogen, phosphorus, root depth, soil impedance, subsoil, temperature water.

## Introduction

Sustaining a human population of 10 billion with a degrading soil resource base in a changing climate is a pre-eminent challenge of the 21st century. An important element of this effort will be the development of crops able to yield well with limited availability of water and nitrogen (N). Suboptimal availability of water and N are primary limitations to plant growth in terrestrial environments, including natural ecosystems and the low-input agroecosystems characteristic of developing nations. Food insecurity is already a massive problem in developing nations, and is projected to worsen in coming decades due to population growth, resource degradation, and climate change. In rich nations, intensive N fertilization meets crop requirements at the cost of substantial pollution of air and water resources (Cassman *et al.*, 2002; Ribaud *et al.*, 2011). Irrigation has limited potential to address expanding water needs in intensive agriculture

because of ongoing degradation and exhaustion of freshwater resources (Wallace and Gregory, 2002; Rosegrant *et al.*, 2009). Global climate change is projected to aggravate crop water limitation by increasing evaporative demand, accelerating soil degradation, and altering the distribution of precipitation in time and space (Wheeler and von Braun, 2013; IPCC, 2014). Crops with reduced requirements for water and N inputs would directly improve food production and therefore food security in developing nations, and would improve agricultural sustainability and productivity in rich nations (Lynch, 2007). Such crops would also benefit the global environment by converting more atmospheric CO<sub>2</sub> to plant biomass and eventually soil organic carbon (Lal, 2004).

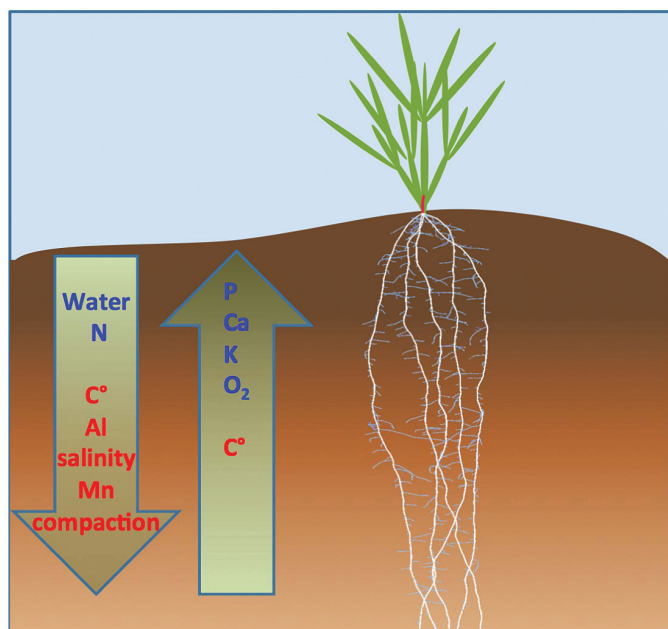
The development of crops with greater rooting depth addresses all three of these aims. Deeper rooting improves water and N capture in many agroecosystems, and increases

the stability of plant-derived carbon (C) in the soil. This is generally recognized and has recently been the subject of several reviews (Manschadi *et al.*, 2010; Kell, 2011; Wasson *et al.*, 2012; Lynch, 2013). The purpose of this article is to summarize these potential benefits very briefly, and then consider several of the constraints to achieving greater rooting depth (Fig. 1). These will be considered in the context of annual crop production, with an emphasis on opportunities for genetic improvement of crop rooting depth, as these are more generally applicable, but also some consideration of general issues regarding soil management. We do not attempt a comprehensive review of these topics but, where possible, cite recent reviews. Our goal is to contribute to the development of crop ideotypes as breeding targets for improved rooting depth.

## Deeper rooting affords several potential benefits

### *Rooting depth is related to water capture*

There is good evidence that in many drought environments, rooting depth is positively related to soil exploration and greater acquisition of water from deep soil strata. The utility of rooting depth for drought tolerance is well documented in the case of dryland wheat, in which genotypes with greater rooting depth are better able to exploit moisture stored from summer rainfall (Manschadi *et al.*, 2006; Wasson *et al.*, 2012). Evidence exists that deep rooting is also important in terminal and intermittent drought scenarios that are more common in rainfed agriculture. For example, rooting depth was positively associated with yield under terminal or intermittent drought in, for example, wheat (Lopes and Reynolds, 2010)



**Fig. 1.** Depiction of nutrient and physical constraints occurring in deeper soils, as discussed in the text. The root image is a barley root system as simulated in the structural-functional plant model *SimRoot*, courtesy of Johannes A. Postma.

and common bean (Sponchiado *et al.*, 1989). Steeper root growth angles increase rooting depth and drought tolerance in rice (Uga *et al.*, 2013) and common bean (Ho *et al.*, 2005). In maize, deeper rooting associated with genotypic variation for several root anatomical traits that reduce the metabolic cost of soil exploration substantially improves yield under terminal and intermittent drought (Zhu *et al.*, 2010; Jaramillo *et al.*, 2013; Chimungu *et al.*, 2014a, b; Lynch *et al.*, 2014). In agroecosystems with limited in-season precipitation, vigorous growth and rapid water extraction from the subsoil can reduce yield under drought by exhausting available soil water before the crop can mature (Manschadi *et al.*, 2006; Palta *et al.*, 2011). In some drought environments, regular precipitation that is insufficient to meet evapotranspirative demand following a prolonged dry season may result in greater availability of water in shallow soil strata, but such cases are relatively rare in the rainfed agriculture that sustains the majority of global crop production. In general, the ability to exploit water in deep soil strata is clearly advantageous, and has been posited as a central element of ideotypes for breeding more drought-tolerant crops (Wasson *et al.*, 2012; Lynch, 2013).

### *Rooting depth is related to N capture*

The predominant form of plant-available nitrogen in most agroecosystems is nitrate, which is highly soluble in water and therefore leaches to deeper soil horizons with precipitation. Rapid nitrate leaching below the root zone can be a significant source of N effluents in high-input agroecosystems (Wiesler and Horst, 1993; Raun, 1999; Cassman *et al.*, 2002; Chen *et al.*, 2010). Rapid development of root foraging in deep soil strata would increase the capture of nitrate in such environments (Wiesler and Horst, 1994; Dunbabin *et al.*, 2003), although surface soil strata can continue to be a source of N throughout the crop season (Wiesler and Horst, 1994), presumably as a result of mineralization, or fertility in excess of crop requirements. Mineralization of organic matter in the topsoil can also be a significant source of N in some systems, and is often the major source of N in low-input systems. In the case of low-input systems or systems in which N is provided in organic material (manure, crop residues, etc.), N availability may be greatest in shallow soil strata (Poudel *et al.*, 2001). In such cases, deeper rooting will not directly improve N capture, although, by improving water acquisition and thereby reducing a nearly universal limitation to crop growth, deep root phenotypes would have indirect benefits for plant growth, soil exploration, and N capture. Cereal root systems continually form nodal roots which pass through the topsoil as they descend to deeper soil strata, and so may not be as susceptible to the opportunity cost of exploring shallow versus deep soil strata, in contrast to dicotyledonous root architectures, which are dominated by lateral roots that form at distinct soil depths (Lynch, 2013). The hypothesis that deeper root phenotypes will enhance N acquisition in the majority of agricultural systems, despite the fact that N availability may be greater in surface soils in some situations, is consistent with available evidence from modelling and empirical studies (Pedersen *et al.*, 2009; Thorup-Kristensen *et al.*,

2009; Dai *et al.*, 2014; Dresbøll and Thorup-Kristensen, 2014). It is important to note that the effect of root traits on N capture will show strong interactions with environmental and management factors, especially as they affect N distribution and leaching rates (Dresbøll and Thorup-Kristensen, 2014). Accordingly, crop ideotypes for efficient N capture include deeper rooting itself (Mi *et al.*, 2010), and in the ‘steep, cheap, and deep’ (SCD) ideotype, architectural and anatomical traits that increase rooting depth (Lynch, 2013). Recent reports support specific elements of the SCD ideotype for improved N capture in maize, including reduced nodal root number; longer, fewer lateral roots, and increased root cortical aerenchyma (Postma and Lynch, 2011; Postma *et al.*, 2014; Saengwilai *et al.*, 2014a, b).

#### *Rooting depth is related to C sequestration*

A significant portion of plant photosynthate is deposited below-ground via roots, root symbionts, and root exudates, and by the senescence and mortality of plants and their organs (Lambers *et al.*, 2002; Farrar *et al.*, 2003; Jones *et al.*, 2004). Globally, soil C is estimated to be twice as large as the pool of atmospheric C (Kell, 2011, 2012), and the capacity of soils to retain C has not yet been saturated. Since the depth of C deposited in the soil by root activity is related to its residence time, deeper crop rooting, achieved by genetic selection or agronomic management, has been proposed as a viable option to sequester atmospheric CO<sub>2</sub> and partially mitigate global climate change (Kell, 2011, 2012).

### **Subsoil constraints to root growth**

In this review, we use the generic terms ‘topsoil’ and ‘subsoil’. By ‘topsoil’ we refer to the epipedon, the surface or A horizon, generally 15–30 cm in depth and highly influenced by plant growth and associated microbial activity, and therefore being more enriched in organic carbon, mineral nutrients, and generally characterized by more active nutrient transformations between organic and mineral forms. By ‘subsoil’ we refer to deeper soil strata, the E, B, and, in some cases, C horizons.

#### *Soil sodicity/salinity and boron*

In arid regions, subsoils can contain toxic levels of sodium (Na), boron (B), and salinity. Stress caused by saline and sodic soils has been intensively researched and recently reviewed, so will not be treated at length here (e.g. Munns and Tester, 2008). Substantial genetic differences in tolerance to these stresses exist among and within crop species, related generally to exclusion of toxic ions from the cytoplasm of active tissues, production of compatible solutes, and mechanisms to balance the competing osmotic needs for water acquisition and ion exclusion. Inhibition of calcium (Ca) transport and metabolism by Na is one aspect of salinity stress (Lynch and Läuchli, 1985) that may be ameliorated by reduced tissue Ca requirement (see below). B toxicity directly interferes with

root growth (Aquea *et al.*, 2012). Significant genotypic variation in B tolerance exists within species, and manipulation of B transporters has the potential to improve B tolerance further (Schnurbusch *et al.*, 2010).

#### *Soil acidity*

A large proportion of terrestrial vegetation is located in moist environments with weathered soils characterized by acid subsoils (Table 1). Acid subsoils (generally defined as having a pH <5) present several direct challenges to root growth, mainly aluminium (Al) toxicity and Ca deficiency, as well as several indirect challenges to plant health, mainly phosphorus (P) deficiency and manganese (Mn) toxicity. Crop adaptation to Al toxicity and P deficiency has been intensively researched, whereas crop adaptation to Ca deficiency and Mn toxicity has received less attention despite their global importance.

#### *Aluminium toxicity*

Al toxicity and tolerance are the subject of several recent reviews (Kochian *et al.*, 2005; Ryan *et al.*, 2011; Liu *et al.*, 2014). Below a pH of ~4.8, Al solubility increases and trivalent Al causes direct injury to root apices, reducing root growth, soil exploration, and hence water and nutrient acquisition. High Al<sup>3+</sup> activity in the soil solution also directly interferes with the uptake of Ca, magnesium (Mg), and potassium (K), and reduces P bioavailability. Al tolerance consists of both tolerance of tissue Al, presumably via mechanisms such as intracellular and tissue level compartmentation (Hiradate *et al.*, 1997; Feng Ma *et al.*, 1998), as well as exclusion of Al from the growing root tissues via the exudation of carboxylates, which complex and detoxify Al in the rhizosphere and apoplast. Al detoxification via carboxylate exudation, primarily as malate or citrate, accounts for substantial genotypic variation in Al tolerance in some crop species, and appears to have arisen through convergent evolution in multiple plant genera (Ryan and Delhaize, 2010), but does not account for genotypic variation in Al tolerance in other species (Kochian *et al.*, 2004). Al tolerance of root hairs is important since hairs are critical for P acquisition and in the formation of rhizosheaths which create favourable microenvironments for growing roots (Delhaize *et al.*, 2012).

Tolerance of trivalent Al is necessary for root exploration of acid subsoils. Inducible carboxylate exudation is an important mechanism of Al exclusion in several important crops, is well understood from genetic and physiological perspectives, and is already being deployed in crop breeding programmes in the tropics. However, additional research is needed to better understand mechanisms of Al tolerance in species for which carboxylates are not the main mechanism of tolerance, the genetic and physiological basis of Al detoxification or compartmentation in cells, tissues, and organs, and the role of rhizosphere communities in determining the fate of root exudates and in creating microenvironments with altered Al and nutrient bioavailability. This research should take into account the large spatiotemporal variation of root properties within and among root classes of mature plants, which might

**Table 1.** Principal characteristics and areas for the 12 orders of the USDA Soil Taxonomy system, with common subsoil constraints to root growth in representative suborders

Order	Description	Area (%) <sup>a</sup>	Common subsoil constraints
Alfisols	Soils from semi-arid to humid regions, typically developed under hardwood forest cover, with subsurface accumulation of clay	9.6	Hypoxia, hardness, temperature
Andisols	Developed from volcanic ash	0.7	Mn toxicity, low P
Aridisols	Desert soils without water for plants	12	Hardness, salinity/sodicity, low P
Entisols	Soils with minimal evolution, as in eroded or accumulation regions. No subsurface horizons	16.2	–
Gelisols	Permafrost (frozen soil) within 100 cm from the surface	8.6	Temperature, low P, Mn toxicity
Histosols	Organic rich, generally in cold latitudes	1.2	Hypoxia, low P, temperature, low Ca
Inceptisols	Soils with weakly developed subsurface horizons	9.8	–
Mollisols	Thick dark surface horizon	6.9	Hardness, hypoxia, temperature
Oxisols	Soils from tropical regions, highly weathered, deep, and uniform profiles	7.5	Acidity (low P, low Ca, K, Mg, Al, and Mn toxicity)
Spodosols	Bleached horizon over grey-brown (spodic) horizon	2.5	Hypoxia, acidity (low P, low Ca, Al, and Mn toxicity)
Ultisols	Weathered soils with low base saturation (low fertility) in subsurface	8.5	Acidity (low P, low Ca, K, Mg, Al, and Mn toxicity), hypoxia, hardness
Vertisols	Shrink and swell soils, that is soils that exhibit temporal variability in volume	2.4	Hypoxia, hardness

<sup>a</sup> Percentage of the total land area ( $1.3 \times 10^9$  km<sup>2</sup>); rows sum to 86%—the remainder (~14%) is from rock and ice-covered regions (Wilding, 2000).

be different from the primary or seminal roots of seedlings that have received the majority of research attention.

#### Low calcium availability

Low Ca availability is another primary challenge to root growth in acid subsoils. It has long been recognized that Ca plays an integral role in Al toxicity (e.g. Foy *et al.*, 1969). Trivalent Al interferes with Ca transport and metabolism (Rengel and Zhang, 2003), and reduces the activity of Ca at the surface of the plasma membrane, an effect which is associated with Al toxicity and tolerance (Kobayashi *et al.*, 2013). Indeed, supplemental Ca ameliorates Al toxicity in moderately acid soils. This is especially important in acid subsoils, since Ca availability declines with depth in such soils, due to decreasing Ca content and increasing saturation of soil exchange sites with soluble Al (Table 1). Although other nutrients including K and Mg may also be limited in acid subsoils, low Ca availability is particularly problematic for root growth since Ca is poorly mobile in the phloem. Demands of root apices for Ca, primarily for cell wall biosynthesis and stability, must therefore be satisfied by Ca supplied directly by soil adjacent to the growing root, with only small amounts provided by the plant itself (Marschner, 1995). Tissue requirements for Ca are primarily in the apoplast, and vary among and within plant species, a large difference being evident between grasses and dicots, since pectins are more abundant in the cell walls of dicots, which consequently have a much greater Ca requirement (Marschner, 1995). Tissue Ca requirement also varies among genotypes of the same species

(Spehar and Galwey, 1997), which may be due to differences in cell wall composition, since genotypes having less pectin or greater pectin methylation might have a reduced Ca requirement (Blamey *et al.*, 2014). It is possible that genetic variation for anatomical traits that influence the amount of cell wall material per unit volume, such as cell size, also affect tissue Ca requirement. Selecting plants with reduced internal Ca requirement may be more fruitful than selecting plants with greater Ca acquisition, since Ca is primarily drawn to roots via transpiration-driven mass flow (Barber, 1995), and increasing transpiration would have significant trade-offs for water economy in most agroecosystems.

Crop genotypes with reduced Ca requirement would be better able to explore acid subsoils despite reduced Ca availability and Al toxicity. Other than the possibility of selecting for reduced pectin content or greater pectin methylation in dicot species, it is not clear how this might be achieved, since this research topic is neglected. Given the importance of soil acidity for crop production, and the importance of Ca for root growth in acid soil, further research is warranted.

#### Low phosphorus availability

Low P availability is an inherent challenge to root growth in acid subsoil. Unlike Ca, P moves readily in the phloem and can be provided to growing root tips by the plant. However, bioavailable P is concentrated in the topsoil because of the low mobility of P in soil and the accumulated effects of biomass deposition and greater microbiological activity in the topsoil (Lynch and Brown, 2001). Plant adaptations to low P

availability consist of mechanisms to increase soil foraging, especially in the topsoil where P availability is greatest, mechanisms to increase the availability of P in the rhizosphere, and mechanisms to reduce internal P requirements (Vance *et al.*, 2003; Lynch, 2011; Richardson *et al.*, 2011).

Genotypic variation for plant P requirements through such mechanisms as prolonged phenology (Nord and Lynch, 2008, 2009; Nord *et al.*, 2011), low tissue P requirement (Lambers *et al.*, 2011), or reduced seed P content (Rose *et al.*, 2010) are potential, largely unexploited breeding targets. One reason for this is that they may entail significant trade-offs. For example, extended phenology may incur fitness penalties when the length of the growing season is limited by temperature or water, and reducing seed P content may have fitness impacts on young seedlings growing in low P soil. Species adapted to extremely low P ecosystems with low tissue P content also have slow growth, a trait that is incompatible with annual crops. *A priori*, it is reasonable to expect that the evolution of land plants would have already optimized the cellular and tissue level utilization of P, which is a nearly universal limitation in terrestrial environments (Vance *et al.*, 2003; Lynch, 2007).

The prospect of improving P acquisition by genetic modification of the production of P-solubilizing compounds has been intensively researched, but despite promising results in synthetic media, results in actual soil have been disappointing (Richardson *et al.*, 2011). For example, a recent comparison of near-isogenic wheat lines found no benefit of citrate efflux for P uptake (Ryan *et al.*, 2014). Transgenic *Trifolium subterraneum* lines expressing a fungal phytase had greater P uptake in only one of five soils tested (George *et al.*, 2005b). Such results highlight the need to confirm results from studies conducted in artificial media with studies in living soil, where complex interactions with soil chemistry and in particular with the microbial communities in the rhizosphere occur. Microorganisms can immobilize and metabolize plant exudates (Jones *et al.*, 2004), and have their own P-solubilizing mechanisms, which can dramatically change fitness outcomes resulting from plant phenotypes (George *et al.*, 2005a, b, 2007).

Several architectural, morphological, and anatomical traits improve P acquisition by increasing topsoil foraging. Since these have been recently reviewed (Lynch, 2011; Richardson *et al.*, 2011), we focus here on the specific case of subsoil exploration. Root architectural traits that improve P acquisition by increasing topsoil foraging may incur trade-offs for subsoil resources such as water (Ho *et al.*, 2005). The metabolic costs of soil exploration by plant roots and their symbionts are large (Lambers *et al.*, 2002; Lynch and Ho, 2005), so the need to explore both deep and shallow soil domains without sacrificing yield is a challenge. One set of traits that may be useful in this context are those that reduce or optimize the metabolic costs of soil exploration. The formation of root cortical aerenchyma (RCA), which converts living cortical tissue to air space through programmed cell death, improves P acquisition by reducing the metabolic cost of soil exploration (Fan *et al.*, 2003; Postma and Lynch, 2010, 2011; Lynch, 2014). Other anatomical traits that reduce the cortical burden of root tissue merit research in this context, including root etiolation, cortical senescence, variation in cortical

cell file number, and cortical cell size. Root hairs are metabolically inexpensive and critically important for P acquisition by expanding the effective depletion zone around the root axis (Gahoonia *et al.*, 1997; Bates and Lynch, 2000a, b; Ma *et al.*, 2001; Zhu *et al.*, 2009). Genotypic variation in root hair length and density is therefore an excellent avenue for breeding more P-efficient crops (Richardson *et al.*, 2011). Arbuscular mycorrhizal (AM) symbioses are also important for P acquisition in most crops, but in the context of subsoil exploration it is noteworthy that AM fungal inoculum and colonization decline substantially with soil depth (Higo *et al.*, 2013; Shukla *et al.*, 2013). It is also not clear how much progress may be made by breeding for more effective AM symbioses, as AM plants are typically highly effective already, and associate with a variety of fungal symbionts already present in agricultural soils. Several root architectural traits may be useful in balancing the needs for topsoil and subsoil foraging. Lateral root branching density can be optimized to balance the needs for intensive soil foraging for P and more extensive, deeper soil foraging for mobile resources such as nitrate (Postma *et al.*, 2014). Dimorphic root architectures exist, possessing axial roots with a greater range of growth angles, or combinations of traits for topsoil foraging such as hypocotyl-borne roots with traits for subsoil foraging such as steep axial growth angles (Miller *et al.*, 2003; Miguel *et al.*, 2013). The utility of such dimorphic root architectures merits validation.

#### *Manganese toxicity*

Mn toxicity is an important but widely ignored constraint to plant growth in weathered soils. This is partially because of the complexity of the processes regulating Mn bioavailability in the rhizosphere as well as Mn toxicity in leaves, both of which have substantial interactions with other environmental variables. Plant-available Mn is divalent, created by reduction of Mn oxides in the soil (Sparrow and Uren, 2014). Mn bioavailability is therefore sensitive to soil redox potential (Eh/pe) as well as pH and soil Mn content. Mn toxicity often occurs at pH 5.3 and below (in contrast to pH 4.8 and below for Al toxicity), or even at a pH as high as 7 in hypoxic conditions (Schlichting and Sparrow, 1998). Mn bioavailability is also affected by the rhizosphere pH, which in turn is affected by the balance of anion and cation uptake by the root, usually governed by the ratio of nitrate and ammonium uptake (Elamin and Wilcox, 1986). Mn uptake is also greatly affected by competition with base cations such as Mg<sup>2+</sup> and Ca<sup>2+</sup>, and the other transition metals, especially Fe (Marschner, 2002). The importance of redox status and therefore localized or temporary hypoxia is an important factor in the subsoil, which generally has low oxygen status and may be compacted (see below). Mn toxicity in the plant may be related to photo-oxidative stress in photosynthetic tissue (Gonzalez *et al.*, 1998; St.Clair and Lynch, 2005). Substantial genotypic variation in Mn tolerance may be related to antioxidant mechanisms and subcellular compartmentation (Gonzalez *et al.*, 1998; Gonzalez and Lynch, 1999).

Crop tolerance to Mn toxicity in the subsoil may be improved by RCA formation, which would increase the

oxygen status of the rhizosphere, as shown in flooded soils (Jackson and Armstrong, 1999), as well as mechanisms to tolerate Mn in leaf tissue, through tissue and cellular level compartmentation as well as greater antioxidant capacity.

#### *Deep rooting is limited by soil physical properties*

Deep rooting is generally limited by abiotic factors such as temperature, aeration, and high mechanical soil impedance. For example, in a wet compacted soil the elongation of roots may be limited due to mechanical soil impedance and hypoxia, while in a compacted dry soil, root elongation may be limited by water deficit and soil strength as root elongation is significantly limited under the following conditions: penetrometer resistances >2 MPa, air-filled volume of <10%, and a matric potential greater than -1.5 MPa (Bengough *et al.*, 2011).

#### *Soil compaction*

Soil compaction limits root elongation in deep soils (Materchera *et al.*, 1991; Bengough *et al.*, 2006). Soil drying also increases soil impedance in many soils, which reduces root growth (e.g. Bengough *et al.*, 1997, 2006; Hinsinger *et al.*, 2009; Whitmore and Whalley, 2009; Bingham *et al.*, 2010). Large impedance is associated with strong soils of high soil bulk density or low water content. Bengough *et al.* (2006) reported that roots prefer to grow in looser soils than in denser soils but that this response depended on genotype and the spatial arrangement of the soil with respect to the main root axes. Impeded soils are known to restrict plant root growth, reduce water and nutrient uptake, and thereby restrict plant development (Ishaq *et al.*, 2001). The extent of mechanical impedance of the soil depends on the physical properties of the soil including its bulk density, matric potential and soil structure, and volumetric water content (reviewed by Bengough *et al.*, 2011; Valentine *et al.*, 2012). A survey of a range of field soils suggested that root elongation was generally limited by mechanical impedance, even in relatively wet field soils (Bengough *et al.*, 2011).

Previous research has focused on the biophysical relationships between cell wall extension and water potential (Watt *et al.*, 2005; Bengough *et al.*, 2006; Hinsinger *et al.*, 2009). A strategy for annual plant roots to avoid soil hardening due to receding water tables, and, subsequently, increasing soil impedance, would be to adapt elongation rates accordingly, allowing the root tip to remain in moist and favourable soil conditions. This may be achieved through changes in the relative length of the elongation zone since the elongation rate and length of the elongation zone are correlated (e.g. Sharp *et al.*, 2004; Bengough *et al.*, 2006). The adaptations could be achieved through changes in the cell wall properties of the expanding cells in the elongation zone. Increased impedance reduces the rate of root elongation and increases radial thickening (Bengough and Mullins, 1990; Clark *et al.*, 2003). Similarly, increased soil strength decreases the length of the elongation zone of the root, and can as much as double the root diameter (Bengough and Mullins, 1990; Watt *et al.*, 2005; Hinsinger *et al.*, 2009; Bengough *et al.*, 2011). RCA

formation reduces the radial transport of water and nutrients (Fan *et al.*, 2007; Hu *et al.*, 2014), but the location of RCA formation in older root segments might not have a large net impact on plant resource acquisition. Therefore, we suggest that RCA formation might be beneficial, preventing lateral water and nutrient loss along the root in soils with a low water potential, functioning as a barrier to water loss in drier topsoils. It may also be advantageous to avoid desiccation of the root tip and surrounding soil by limiting water transport from root apices to mature xylem through such mechanisms as delayed xylem maturation, increased suberization of the endodermis, and hydraulic isolation of rhizosheaths (Lynch, 2014). Architectural traits that position roots in deeper soil domains may also aid in the avoidance of soil hardening in drying surface strata (Lynch, 2013).

Genetic diversity has potential for trait discovery to overcome the constraints of soil compaction. For example, wheat (e.g. Botwright Acuña and Wade, 2012) and lupin (e.g. Chen *et al.*, 2014) genotypes differ in their response to mechanical soil impedance. In general, root systems responded to mechanical impedance as roots are repelled, reduce their elongation rate, and thicken radially on encountering the physical barrier (Materchera *et al.*, 1992). Radial thickening makes roots more resistant to buckling in hard soils (Materchera *et al.*, 1992; Bengough *et al.*, 1997; Clark *et al.*, 2008; Whalley *et al.*, 2008), which increases soil penetration. Bengough *et al.* (2011) and Haling *et al.* (2013) hypothesized a further root trait—root anchorage—to facilitate root penetration into harder soils. The anchorage of roots might be attained by friction of soil particles and maturing tissues behind the elongation zone by producing root hairs and lateral roots. The anchoring of the root apex through root hair growth may support the turgor pressure to force the root tip further into the soil. Another root trait may be the ability to change trajectory as a reaction to soil compaction, using the soil matrix at the bend of the root as an anchor point which reflects the reaction force entering the compacted soil layer. Future research will have to address soil environments with multiple stresses such as limited water availability in compacted soils. A strategy for soil exploration for water in compacted soils would be to exploit paths of less resistance using cracks and biopores, and sensing water availability within soils. Studies showed that roots were found more frequently in artificially created pores in compacted soils than expected by chance alone (Stirzaker *et al.*, 1996). Recently, Bao *et al.* (2014) indicated that root systems are able to respond to local hydraulic changes of the soil environment, which they called ‘hydropatterning’. This recent study reported that plants can distinguish between wet and drier soil domains, developing root hairs and lateral roots in the wet environments. Preferential root growth in wetter soil domains would result in reshaping of root architecture over time. In the common case in which the topsoil dries before the subsoil, this would result in deeper root architecture, which would be favourable for water acquisition. If genotypic differences in sensing such localized resources exist, this may be an opportunity to increase resource acquisition efficiency of crop roots in deeper soil profiles.

## Hypoxia

Hypoxia or anoxia affects roots in soils if the air-filled pore space is limited (<10 %) due to the soil structure or the water saturation of the soil. Survival of roots in anaerobic soil conditions requires avoidance mechanisms (reviewed by Vartapetian and Jackson, 1997). This is achieved in some species by changing the growth trajectory of the roots, increasing root growth in oxygenated soil profiles or, in flooded environments, the formation of aerial roots. Internally interconnected RCA will also overcome the restriction of low oxygen levels in the soil. In well-adapted species, the aerenchyma can extend from the leaf stomata to the root tip (Vartapetian and Jackson, 1997). Plants are known to form RCA under waterlogged conditions (Jackson and Armstrong, 1999), which may occur during the rainy season or flooding as an agricultural practice in tropical regions. Oxygen transport in RCA may be facilitated by concentration gradients or through temperature-driven pressure gradients. Many wetland plants, including rice, develop a barrier to radial oxygen loss through suberization of the hypodermis (Colmer, 2003; Garthwaite *et al.*, 2008). Temperate cereals may follow another strategy by removing the cortex through cortical senescence, subsequently reducing the requirement for oxygen. Other anatomical traits may also reduce the cortical burden (Jaramillo *et al.*, 2013) in hypoxia and therefore conserve oxygen for the growing tip in deeper soils, including cell file numbers or cortical cell size (Chimungu *et al.*, 2014; Lynch *et al.*, 2014), which may facilitate root elongation under low oxygen conditions. However, these hypotheses need to be tested.

## Temperature

Vertical temperature gradients, often of substantial magnitude, are common in natural soils (Table 1). Temperature effects on root growth have been studied for many years. Bauer and Bradow (1996) screened cotton genotypes that were impacted less by cold temperatures, finding that root length was genotype dependent under low temperature. Maize root growth and development decreased linearly with decreasing temperatures in a nutrient solution experiment using a temperature range from 22 °C to 13 °C (Nagel *et al.*, 2009). A similar effect of temperature on root elongation and lateral root growth in other plant species such as canola, sunflower, barley, and cotton were reported in several studies (Cumbus and Nye, 1982; Abbas Al-ani and Hay, 1983; Macduff *et al.*, 1986; Nagel *et al.*, 2009). Low temperatures may inhibit deep rooting of plants, restricting apical root elongation as low temperature decreases metabolic activity and therefore sink strength of roots, suggesting feedback mechanisms on phloem loading and photosynthesis (Minchin, 2002). The effects of vertical temperature gradients on assimilate transport depend on the fraction of roots exposed to lower temperatures (Sowinski *et al.*, 1998). Plant breeding for cold tolerance may increase deep rooting of crops, allowing root elongation in deeper and, subsequently, colder environments. For example, a chilling-sensitive maize variety had decreased

root length and root surface area when compared with a chilling-tolerant variety in low temperature treatments (Richner *et al.*, 1996). Hund *et al.* (2007) investigated the effects on root morphology of low temperature in 21 modern inbred maize lines under low temperature and found great variation of root morphology among the genotypes, concluding that variation of root traits accounted for early vigour under low temperatures and could be a target for future plant breeding. Furthermore, high temperature may also inhibit root growth as roots and the soil respire rapidly in warm temperatures, which may lead to anaerobic conditions in the root zone within a short period, and, subsequently, may decrease root growth into the deeper soil profile. The formation of RCA would help to provide oxygen under those growth conditions, facilitating deeper root growth along the vertical temperature gradient.

## Agronomic practices facilitating deep rooting

Production system, timing, and frequency of cultivation affect soils; the impact depends on soil type, topography, and climate (Stoate *et al.*, 2001). For example, increases in field size and increased use of heavy machinery have contributed to greater levels of soil erosion (Evans, 1996). Additionally, the use of heavier farm machinery may increase soil impedance in arable production systems (reviewed by Batey, 2009). Soil compaction is a particular problem on soils with low organic matter where earthworm abundance and activity are low (Makeschin, 1997). Mechanical soil impedance is a major limitation to agricultural production systems, unless a network of channels or biopores exists for roots to exploit (White and Kirkegaard, 2010), as roots are able to grow in continuous biopores or channels and therefore explore compacted soils for nutrients and water in deeper soils. For example, wheat root growth in hard subsoils is almost confined entirely to biopores in very hard Australian soils (White and Kirkegaard, 2010). Some no-tillage farms use rotations to encourage the formation of biopores/channels (Passioura, 2002). Alternatively, deep-ripping during cultivation (Delroy and Bowden, 1986; reviewed by Batey, 2009) or the application of gypsum to alter soil structure and aggregate stability, could be used to facilitate deeper root growth in compacted soils. Certain plant species could be used as pioneer species creating such biopores/channels. Materechera *et al.* (1992) found that dicotyledonous plants had larger diameters and penetrated harder soil layers better than monocotyledonous species. Therefore, dicots such as alfalfa and lupin may be used as pioneer species to avoid unsuitable topsoil conditions and to exploit nutrients and water in deeper subsoils, while creating root channels that can be exploited by subsequent cereal crops. For example, *Lupinus albus* can penetrate subsoil to depths of >100 cm, avoiding unfavourable topsoils (Pennisi, 2008). However, White and Kirkegaard (2010) found that >85% of roots were clustered together using those channels and biopores, which may limit the exploration of those soils as roots are unable to explore the soils for nutrients and water except within the biopores and channels.

Frequent cultivation decreases soil organic matter content and, subsequently, impacts soil structure and composition (Stoate *et al.*, 2001). The use of fertilizers (organic or inorganic), herbicides, and pesticides also influences soil structure directly or indirectly through impacts on the soil fauna (e.g. Seghers *et al.*, 2005; reviewed by Bünemann *et al.*, 2006). For example, earthworms may improve soil structure in soils that are not compacted and maintain high organic matter, improving aeration, crop root growth, and drainage (Marinissen and Bok, 1988; Makeschin, 1997). Other agricultural practices such as minimum tillage can increase soil organic matter (Logan *et al.*, 1991; Fox, 2000; Bescansa *et al.*, 2006; McCarthy *et al.*, 2008), which would also facilitate deeper rooting and deeper C sequestration through greater water availability in the soil, as organic matter improves soil moisture retention (Benckiser, 1997).

Agronomic practices have substantial effects on soil properties, crop growth, and subsoil exploration by crop roots. The development of crop genotypes with root traits improving subsoil exploration will have to consider not only the target soil environment but also the target management environment. Such ‘G×E×M’ interactions are in general poorly understood. The complexity and scope of possible scenarios, including future climate scenarios, call for the use of multi-scale mechanistic modelling approaches to inform and guide empirical research.

## Updated ideotype for deep rooting

The ‘steep, cheap, and deep’ ideotype consists of architectural, anatomical, and physiological traits that could improve the capture of soil water and N by maize root systems by improving subsoil exploration (Lynch, 2013). In this review we propose several additional elements of this ideotype as adaptations to subsoil constraints to root growth, as summarized in Table 2.

In acid subsoils, Al tolerance is critically important, and low Ca requirement, P efficiency, and Mn tolerance would be useful. Soil impedance is a pervasive subsoil constraint to root growth. Traits improving root penetration in hard subsoil include ‘hydropatterning’ and the ability to find cracks and biopores which provide optimal deep soil exploration for nutrients and water. Root hairs and lateral root formation provide anchorage and reduce buckling of roots in compacted soils. Tolerance to low soil temperature is important for subsoil exploration in temperate soils. The formation of RCA and reduction of cortical cell file number and cortical cell size would facilitate growth by reducing metabolic burden and oxygen requirements, allowing deeper penetration of roots, and, potentially, greater tolerance of hypoxia or low temperature. Additionally, RCA could increase oxygen levels, thereby reducing Mn toxicity in low pH soils. In fact, any traits reducing the respiratory requirements of root

**Table 2.** Deep soil constraints, root traits/phenes, and example references

Soil constraint	Root trait(s)/phene(s)	Reference
Al toxicity	Carboxylate exudation	Kochian <i>et al.</i> (2004)
	Al tolerance of root hairs	Delhaize <i>et al.</i> (2012)
	Rhizosheaths	Delhaize <i>et al.</i> (2012)
	Tissue tolerance	Hiradate <i>et al.</i> (1997); Feng Ma <i>et al.</i> (1998)
Low Ca availability	Reduced tissue Ca requirement	Spehar and Galwey (1997)
	Larger cell size	Hypothetical
Low P availability	Lower pectin content or greater pectin methylation	Blamey <i>et al.</i> (2014)
	RCA	Fan <i>et al.</i> (2003)
	Cortical senescence, reduction of cortical cell file number, and cortical cell size	Hypothetical
	Root architecture	Richardson <i>et al.</i> (2011)
	Root hairs	Richardson <i>et al.</i> (2011)
Mn toxicity	Mycorrhizal symbiosis	Lambers <i>et al.</i> (2002); Lynch and Ho (2005)
	Root exudates	Jones <i>et al.</i> (2004); George <i>et al.</i> (2005b)
	RCA	Hypothetical
	Antioxidant mechanisms	Gonzalez <i>et al.</i> (1998)
Soil compaction	Subcellular compartmentation	Gonzalez and Lynch (1999)
	Adaptation of the elongation zone	Hypothetical
	Reduction of water transport from the root tip	Hypothetical
	Radial thickening	Materechera <i>et al.</i> (1992); Bengough <i>et al.</i> (1997); Clark <i>et al.</i> (2008); Whalley <i>et al.</i> (2008)
	Root anchorage (root hairs and lateral roots)	Hypothetical
Hypoxia	‘Hydropatterning’	hypothetical
	RCA	Jackson and Armstrong (1999); Colmer (2003); Garthwaite <i>et al.</i> (2008); Henry <i>et al.</i> (2012)
Temperature	Suberization	Hypothetical
	Breeding for cold tolerance	Hund <i>et al.</i> (2007)
	RCA	Hypothetical



tissue, including anatomical traits such as RCA, cortical cell file number, cortical senescence, and cortical cell size, should afford multiple benefits for subsoil exploration. Reduced respiratory requirements would result in greater soil exploration and therefore better acquisition of P and Ca, greater tolerance to hypoxia, greater tolerance to the increased respiratory demand of high soil temperature, greater tolerance of the challenge to maintain respiration under low soil temperature, and potentially greater tolerance of Mn toxicity by diminishing oxygen consumption in the rhizosphere, thereby maintaining Mn in a more oxidized state. Traits that slow desiccation of soil surrounding root apices, such as delayed xylem maturation and suberization of the endodermis, may reduce resistance to root penetration. The utility of any of these traits will of course depend on the nature of the soil environment and crop management. We propose that many of these traits will exhibit synergism; that is, their value in combination will exceed their additive value. For example, alleviation of Al toxicity will improve plant root growth and therefore plant acquisition of Ca and P, other limiting resources. These traits and issues merit research because of their potential value in developing more productive, sustainable, benign, and resilient agricultural systems.

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