

HHS Public Access

Adv Exp Med Biol. Author manuscript; available in PMC 2016 August 17.

Published in final edited form as:

Author manuscript

Adv Exp Med Biol. 2016; 854: 39-44. doi:10.1007/978-3-319-17121-0_6.

Therapeutic Approaches to Histone Reprogramming in Retinal Degeneration

Andre K. Berner and

740 S. Limestone St., Suite E-300, Lexington, KY 40536, USA

Mark E. Kleinman

Department of Ophthalmology and Visual Sciences, University of Kentucky, Lexington, KY 40536, USA

Andre K. Berner: andrekberner@gmail.com; Mark E. Kleinman: mark.kleinman@uky.edu

Abstract

Recent data have revealed epigenetic derangements and subsequent chromatin remodeling as a potent biologic switch for chronic inflammation and cell survival which are important therapeutic targets in the pathogenesis of several retinal degenerations. Histone deacetylases (HDACs) are a major component of this system and serve as a unique control of the chromatin remodeling process. With a multitude of targeted HDAC inhibitors now available, their use in both basic science and clinical studies has widened substantially. In the field of ocular biology, there are data to suggest that HDAC inhibition may suppress neovascularization and may be a possible treatment for retinitis pigmentosa and dry age-related macular degeneration (AMD). However, the effects of these inhibitors on cell survival and chemokine expression in the chorioretinal tissues remain very unclear. Here, we review the multifaceted biology of HDAC activity and pharmacologic inhibition while offering further insight into the importance of this epigenetic pathway in retinal degenerations. Our laboratory investigations aim to open translational avenues to advance dry AMD therapeutics while exploring the role of acetylation on inflammatory gene expression in the aging and degenerating retina.

Keywords

Retinal degeneration; Acetylome; Lysine deacetylases; Histone deacetylases; Valproic acid; Apoptosis; Inflammation; Aging Electronic supplementary material

6.1 Post-translational Acetylation Controls Gene Expression and Protein

Activity

Acetylation is a reversible post-translational modification that was first discovered in histones and occurs in a wide range of organisms. Histone proteins (H2A, H2B, H3 and H4) are integrated with 147 base-pairs of DNA in a complex called the nucleosome (Luger et al. 1997). Lysine acetylation and deacetylation of histones are carried out by two groups of

Correspondence to: Andre K. Berner, andrekberner@gmail.com.

Berner and Kleinman

enzymes: acetyl group addition by HATs (or lysine acetyltransferases KATs) and acetyl group removal by HDACs (or KDACs), respectively. A generalized epigenetic principle is that histone acetylation results in an open structure of the DNA enabling gene transcription whereas histone deacetylase activity tightens the nucleosome and compacts the chromatin making those sites inaccessible for transcription (de Ruijter et al. 2003). While this model has not applied to all systems, it is clear from current studies that there is a delicate balance of (de)acetyltransferase activity which may be dysregulated in aging diseases. Lysine acetylation is not limited to histones but are also present in innumerous other protein substrates giving acetylation a wider significance in developmental and disease states (Peserico and Simone 2011). Nuclear receptors (estrogen receptor, p300), proliferating factors (E2F/RB), hypoxia induced factors (HIF-1 α), transcription factors (NF κ B, p53, STAT3 and c-MYC) and other cellular proteins (α Tubulin, Ku70 and Hsp90) are all known non-histone targets of HATs and HDACs (Glozak et al. 2005).

Histone deacetylases are a family of 18 known members, classified in four groups based on their homology to yeast proteins (Dokmanovic et al. 2007). Class I consists of HDAC1, 2, 3 and 8. HDAC1 and HDAC2 are ubiquitously expressed, strongly localized to nuclei and predominantly associated in megadalton complexes (Bantscheff et al. 2011; Di Marcotullio et al. 2011). Members of the Class II-family of HDACs are separated into Class IIa (HDAC4, 5, 7 and 9) which localize to both nuclear and cytosolic compartments and IIb (HDAC6 and 10) which are predominantly cytosolic. Class I/II HDACs are zinc dependent. Class III HDACs, also known as sirtuins (SIRTs), are evolutionarily unrelated to the other HDAC classes. SIRTs require NAD ⁺ as a co-factor making them highly sensitive to oxidative stress (Balaiya et al. 2012) and a hotly pursued potential therapeutic target for age-related and metabolic diseases (Imai and Guarente 2014). There is only one Class IV HDAC (HDAC11) which is also zinc dependent, localized to the nucleus and is heavily conserved in all living eukaryotes other than fungi (Gao et al. 2002).

6.2 Context and Tissue Dependent Effects of Histone Deacetylases

Expression levels of HATs and HDACs as well as targeted acetylation sites and proteins can dramatically change between tissues and within varying developmental, normal adult, and diseased states. There are several models of neuronal cell death in which HDAC inhibition (HDACi) exhibits a protective benefit whereas targeting identical pathways in cancer cells is pro-apoptotic. With the highly specialized and multicellular architecture of the eye, the dominant effects of HDAC inactivity remain very unclear and certainly become extremely difficult to interpret when comparing acute laboratory animal and cell culture models to chronic aging diseases. Still, we are gathering essential data that will allow us to develop a fundamental understanding of this critical and deeply conserved regulatory biologic system. Class I HDAC expression is considered ubiquitous though we have observed significant differences in immuno-localization patterns in the mouse retina with HDAC1/2. Class II HDACs are known to display highly specific tissue-dependent expression patters leading to variable sub-cellular localization and certainly tissue-specific biological effects of enzyme inactivity and subsequent imbalances in the acetylome. Many diseases have been associated with altered global acetylation patterns including cancer, cardiovascular disease and inflammatory diseases. Hyper-acetylation via HDA-Ci is known to be cytoprotective in

Berner and Kleinman

models of neuronal ischemic injury (Kim and Chuang 2014; Murphy et al. 2014), Huntington's disease (Ferrante et al. 2003), and stroke (Liu et al. 2012). Yet, HDACi is nearly uniformly cytotoxic in cancer models (McConkey et al. 2012). HDACi also has opposing effects on critical immune system mediators. Toll-like receptors (TLRs) are potent cell-signaling gateways to innate immune pathways and downstream inflammatory responses. Treatment of cultured human macrophages with HDACi leads to caspasedependent apoptosis and release of pro-inflammatory cytokines; however, this effect is reversed by pre-treatment with TLR agonists including LPS and poly I:C (Tsolmongyn et al. 2013). Data revealing the protean biology of the acetylome must be seriously addressed and rigorously studied in the laboratory prior to pharmacologically approaching HDAC/HAT manipulation for the treatment of human diseases.

A potent and well-characterized Class I/II HDACi is suberoylanilide hydroxamic acid (SAHA also known as vorinostat) which is currently in advanced phase clinical phase trials for multiple myeloma and several solid tumors. Similar to previous HDACi results, the pharmacologic effects of SAHA are highly dependent on cell-type and state coupled with a limited therapeutic window. While low concentrations of SAHA may be significantly cytoprotective, higher concentrations are pro-apoptotic in many immune cell types (Li et al. 2008). Nearly identical data exists for valproic acid (VPA), another small molecule Class I/II HDACi which is most widely used for seizure prophylaxis. VPA has been shown to reduce brain damage in an animal model of transient cerebral ischemia (Ren et al. 2004), provide acute neuro-protection in ischemic retinal injury (Alsarraf et al. 2014), and stimulate axonal regrowth after optic nerve crush (Biermann et al. 2010). These data have rapidly opened translational avenues of pharmacologic induced chromatin remodeling as a novel target for the epigenetic regulation of critical cell death and survival pathways in aging and neurodegenerative diseases.

6.3 Differential Effects of Histone Deacetylase in the Retina and Retinal Pigment Epithelium

Investigations of the *rd1* mouse demonstrated significant protection form loss of photoreceptors after broad inhibition of Class I/II HDACs with trichostatin A (TSA) (Sancho-Pelluz et al. 2010). A single report was then published suggesting the therapeutic efficacy of VPA in the treatment of retinitis pigmentosa (Clemson et al. 2011). Similar benefits were reported for VPA in *rd1* mice (Mitton et al. 2012); however, the same treatment had the contrary effect in *rd10* mice (Guzman et al. 2014). Additional studies were performed even though the original data had been hotly contested with multiple letters in the literature with severe limitations to the study design and reports of inefficacy and even loss of vision associated with the use of VPA for retinal degeneration (van Schooneveld et al. 2011; Sisk 2012). Recently, a long-term follow-up study confirmed visual decline and adverse side-effects associated with VPA therapy in patients with retinitis pigmentosa (Bhalla et al. 2013).

Four independent groups have presented varying data regarding VPA and retinal degeneration. Reports included positive (Iraha et al. 2014), variable (Guzman et al. 2014;

Lai et al. 2014) or outright negative findings (Berner et al. 2014; Kumar et al. 2014). In a transgenic Xenopus model expressing various human rhodopsin mutations, only retinal degeneration secondary to the P23H mutation was favorably treated with VPA (Lai et al. 2014). Despite VPA's described neuro-protective and anti-inflammatory properties, in just these few studies, significant retinotoxicity was encountered in numerous animal and cellculture models. We have demonstrated that VPA up-regulates caspase-3 activation and cell death in primary human RPE isolates, a finding which has been confirmed in other studies (Suuronen et al. 2007; Kumar et al. 2014). VPA treatment exhibits a significant proinflammatory response *in vitro* and *in vivo* with an array of cytokines, cytokine receptors, mediating enzymes and transcription factors (Kleinman et al. 2013; Kleinman et al. 2014). This pro-inflammatory signature is in accordance to the known immune response in AMD (Suuronen et al. 2007; Shakespear et al. 2011; Miao et al. 2012; Whitcup et al. 2013). Further investigations into this powerful epigenetic regulatory system will continue to yield important features of HDAC involvement in the pathogenesis and treatment of retinal degenerations; however, at this time we urge caution using VPA as a treatment option for these diseases given the variable treatment effect dependent on tissue-type and cellular target (Fig. 6.1).

Acknowledgments

M.E.K. was supported by NEI/NIH, Career Development Awards from the Foundation Fighting Blindness and Research to Prevent Blindness, and the American Federation for Aging Research.

References

- Alsarraf O, Fan J, Dahrouj M, et al. Acetylation: A lysine modification with neuroprotective effects in ischemic retinal degeneration. Exp Eye Res. 2014; 127C:124–131. [PubMed: 25064603]
- Balaiya S, Khetpal V, Chalam KV. Hypoxia initiates sirtuin1-mediated vascular endothelial growth factor activation in choroidal endothelial cells through hypoxia inducible factor-2alpha. Mol Vis. 2012; 18:114–120. [PubMed: 22275802]
- Bantscheff M, Hopf C, Savitski MM, et al. Chemoproteomics profiling of HDAC inhibitors reveals selective targeting of HDAC complexes. Nat Biotechnol. 2011; 29:255–265. [PubMed: 21258344]
- Berner A, Mohan K, Lou D-Y, et al. RPE Cytotoxicity and Caspase Activation after Treatment with Valproic Acid. ARVO Meeting Abstracts. 2014; 55:5991.
- Bhalla S, Joshi D, Bhullar S, et al. Long-term follow-up for efficacy and safety of treatment of retinitis pigmentosa with valproic acid. Br J Ophthalmol. 2013; 97:895–899. [PubMed: 23603755]
- Biermann J, Grieshaber P, Goebel U, et al. Valproic acid-mediated neuroprotection and regeneration in injured retinal ganglion cells. Investigative ophthalmology & visual science. 2010; 51:526–534. [PubMed: 19628741]
- Clemson CM, Tzekov R, Krebs M, et al. Therapeutic potential of valproic acid for retinitis pigmentosa. Br J Ophthalmol. 2011; 95:89–93. [PubMed: 20647559]
- de Ruijter AJ, van Gennip AH, Caron HN, et al. Histone deacetylases (HDACs): characterization of the classical HDAC family. Biochem J. 2003; 370:737–749. [PubMed: 12429021]
- Di Marcotullio L, Canettieri G, Infante P, et al. Protected from the inside: endogenous histone deacetylase inhibitors and the road to cancer. Biochim Biophys Acta. 2011; 1815:241–252. [PubMed: 21277938]
- Dokmanovic M, Clarke C, Marks PA. Histone deacetylase inhibitors: overview and perspectives. Mol Cancer Res. 2007; 5:981–989. [PubMed: 17951399]
- Ferrante RJ, Kubilus JK, Lee J, et al. Histone deacetylase inhibition by sodium butyrate chemotherapy ameliorates the neurodegenerative phenotype in Huntington's disease mice. The Journal of

neuroscience: the official journal of the Society for Neuroscience. 2003; 23:9418–9427. [PubMed: 14561870]

- Gao L, Cueto MA, Asselbergs F, et al. Cloning and functional characterization of HDAC11, a novel member of the human histone deacetylase family. The Journal of biological chemistry. 2002; 277:25748–25755. [PubMed: 11948178]
- Glozak MA, Sengupta N, Zhang X, et al. Acetylation and deacetylation of non-histone proteins. Gene. 2005; 363:15–23. [PubMed: 16289629]
- Guzman E, Despande M, Byrd DW, et al. Systemic Valproic Acid can Accelerate Photoreceptor Loss in rd10 mice. Invest Ophthalmol Vis Sci. 2014; 55:1281.
- Imai SI, Guarente L. NAD and sirtuins in aging and disease. Trends in cell biology. 2014; 24:464–471. [PubMed: 24786309]
- Iraha S, Hirami Y, Oota S, et al. The efficacy of valproic acid for retinitis pigmentosa patients. ARVO Meeting Abstracts. 2014; 55:1390.
- Kim HJ, Chuang DM. HDAC inhibitors mitigate ischemia-induced oligodendrocyte damage: potential roles of oligodendrogenesis, VEGF, and anti-inflammation. Am J Transl Res. 2014; 6:206–223. [PubMed: 24936215]
- Kleinman M, Berner A, Lou D, et al. Epigenetic Regulation of Eotaxin Expression in Human Retinal Pigment Epithelium. Invest Ophthalmol Vis Sci. 2013; 54:344.
- Kleinman ME, Berner A, Mohan K, et al. Histone Deacetylase Expression and Inhibition in Age Related Macular Degeneration. ARVO Meeting Abstracts. 2014; 55:3457.
- Kumar A, Kothary PC, Rossi B, et al. Valproic Acid Induced Inhibition of Fibroblast Growth Factor 2 Synthesis in Human Retinal Pigment Epithelial Cells. Invest Ophthalmol Vis Sci. 2014; 55:364.
- Lai RYJ, Zong Z, Tam BM, et al. Opposing effects of valproic acid treatment in four animal models of retinitis pigmentosa. ARVO Meeting Abstracts. 2014; 55:4370.
- Li N, Zhao D, Kirschbaum M, et al. HDAC inhibitor reduces cytokine storm and facilitates induction of chimerism that reverses lupus in anti-CD3 conditioning regimen. Proc Natl Acad Sci U S A. 2008; 105:4796–4801. [PubMed: 18347343]
- Liu XS, Chopp M, Kassis H, et al. Valproic acid increases white matter repair and neurogenesis after stroke. Neuroscience. 2012; 220:313–321. [PubMed: 22704966]
- Luger K, Mader AW, Richmond RK, et al. Crystal structure of the nucleosome core particle at 2.8 A resolution. Nature. 1997; 389:251–260. [PubMed: 9305837]
- McConkey DJ, White M, Yan W. HDAC inhibitor modulation of proteotoxicity as a therapeutic approach in cancer. Adv Cancer Res. 2012; 116:131–163. [PubMed: 23088870]
- Miao H, Tao Y, Li XX. Inflammatory cytokines in aqueous humor of patients with choroidal neovascularization. Mol Vis. 2012; 18:574–580. [PubMed: 22419849]
- Mitton KP, Guzman EE, Byrd D, et al. Rescue Of Photoreceptor Degeneration In Rd1 Mice By Systemic Treatment With Valproic Acid. Invest Ophthalmol Vis Sci. 2012; 53:5585. [PubMed: 22836761]
- Murphy SP, Lee RJ, McClean ME, et al. MS-275, a class I histone deacetylase inhibitor, protects the p53-deficient mouse against ischemic injury. J Neurochem. 2014; 129:509–515. [PubMed: 24147654]
- Peserico A, Simone C. Physical and functional HAT/HDAC interplay regulates protein acetylation balance. J Biomed Biotechnol. 2011; 2011:371832. [PubMed: 21151613]
- Ren M, Leng Y, Jeong M, et al. Valproic acid reduces brain damage induced by transient focal cerebral ischemia in rats: potential roles of histone deacetylase inhibition and heat shock protein induction. J Neurochem. 2004; 89:1358–1367. [PubMed: 15189338]
- Sancho-Pelluz J, Alavi MV, Sahaboglu A, et al. Excessive HDAC activation is critical for neurodegeneration in the rd1 mouse. Cell Death Dis. 2010; 1:e24. [PubMed: 21364632]
- Shakespear MR, Halili MA, Irvine KM, et al. Histone deacetylases as regulators of inflammation and immunity. Trends Immunol. 2011; 32:335–343. [PubMed: 21570914]
- Sisk RA. Valproic acid treatment may be harmful in non-dominant forms of retinitis pigmentosa. Br J Ophthalmol. 2012; 96:1154–1155. [PubMed: 22581401]

- Suuronen T, Nuutinen T, Ryhanen T, et al. Epigenetic regulation of clusterin/apolipoprotein J expression in retinal pigment epithelial cells. Biochem Biophys Res Commun. 2007; 357:397–401. [PubMed: 17420006]
- Tsolmongyn B, Koide N, Odkhuu E, et al. Lipopolysaccharide prevents valproic acid-induced apoptosis via activation of nuclear factor-kappaB and inhibition of p53 activation. Cellular immunology. 2013; 282:100–105. [PubMed: 23770718]
- van Schooneveld MJ, van den Born LI, van Genderen M, et al. The conclusions of Clemson et al concerning valproic acid are premature. Br J Ophthalmol. 2011; 95:153–154. author reply. [PubMed: 20971790]
- Whitcup SM, Sodhi A, Atkinson JP, et al. The role of the immune response in age-related macular degeneration. Int J Inflam. 2013; 2013:348092. [PubMed: 23762772]

Berner and Kleinman

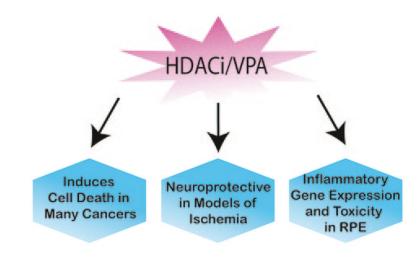


Fig. 6.1.

While HDAC inhibition is cytoprotective in many models of neuronal cell death, it is also capable of inducing significant cytotoxicity in various cancers and up-regulating inflammatory gene expression and cell death in the RPE