

RESEARCH ARTICLE

Open Access



Association of serum myokines and aerobic exercise training in patients with spinal cord injury: an observational study

Der-Sheng Han^{1,2,3†}, Ming-Yen Hsiao^{2†}, Tyng-Guey Wang², Ssu-Yuan Chen^{2*} and Wei-Shiung Yang^{4,5*}

Abstract

Background: Patients with spinal cord injury (SCI) have a higher prevalence of cardiovascular diseases compared to the healthy population. Aerobic exercise training is one of the recommended treatments. However, literature regarding the effect of aerobic training on patients with SCI is scarce. This study evaluated changes in parameters of exercise physiology and serum myokines immediately after exercise and after a training program among patients with SCI.

Methods: Male patients with SCI and age- and sex-matched healthy individuals were recruited. Cardio-pulmonary exercise testing (CPET) was used to determine oxygen uptake at peak exercise and anaerobic threshold in both groups. The patients with SCI attended aerobic exercise training for 36 sessions within 12–16 weeks. Basic data, hemodynamic and exercise physiology parameters, and serum myokine (myostatin, IGF-1, and follistatin) concentrations were measured pre- and post-exercise in both groups, and were repeated in patients with SCI post-training.

Results: Eleven patients with SCI underwent CPET and 5 completed the training. The 11 patients and 16 healthy adults had no differences in baseline serum myokine concentrations before CPET. Immediately after the CPET, the reference group had an $18 \pm 19\%$ increase in serum IGF-1, while the patients had no observable myokine changes. After aerobic exercise training, the 5 patients had a $48 \pm 18\%$ increase in serum myostatin compared to the pre-training level, although the body weight and exercise physiology parameters remained unchanged.

Conclusions: Acute exercise to exhaustion in CPET results in an immediate increase in serum IGF-1 in healthy individuals while aerobic exercise training results in increased serum myostatin in patients with SCI.

Keywords: Spinal cord injury, Aerobic capacity, Cardio-pulmonary exercise testing, Myostatin, Insulin-like growth factor-1

Background

Over the past decades, advances in medicine and health-care have greatly decreased risks of respiratory or urinary tract infection and renal failure in patients with spinal cord injury (SCI). However, mortality due to cardiovascular diseases continues to rise [1, 2]. There is a higher prevalence of cardiovascular diseases in patients with SCI compared to the healthy population [3, 4] partly because of decreased physical activity. Muscle weakness and atrophy, and diminished aerobic capacity are common in

patients with SCI [5], while autonomic dysfunction often results in altered heart rate (HR) and blood pressure (BP) response, further heightening cardiovascular risks [6, 7].

Aerobic exercise training is the standard management for preventing and treating cardiovascular diseases [8]. It is also one of the essential components of the recommended treatments for patients with SCI [9, 10]. However, the prescription of exercise training is challenging and since most patients with SCI lack adequate balance, motor control, and muscle strength of the lower extremities, arm ergometry is a frequently used exercise modality.

Recent evidence suggests that myostatin, a strong inhibitor of muscle hypertrophy counteracting IGF-1, is also related to aerobic metabolism of muscle [11–16]. IGF-1 is a growth controlling protein that affects the development of multiple systems, with the musculo-

* Correspondence: ssuyuan@ntu.edu.tw; wsyang@ntu.edu.tw

†Equal contributors

²Department of Physical Medicine and Rehabilitation, National Taiwan University Hospital, Taipei, Taiwan

⁴Department of Internal Medicine, National Taiwan University Hospital, No. 1, Chang-Teh St, Taipei, Taiwan

Full list of author information is available at the end of the article



skeletal as one of the most important [17–19]. In the hypothesis of the “accelerator-brake” model [11–14, 20], IGF-1 provides the initial muscle growth stimulus, and myostatin increases subsequently to prevent overgrowth of the muscle. Currently there are limited studies that explore the changes of serum myokines after chronic exercise training in patients with SCI. Only one small trial indicated no significant changes of serum myokines after activity-based training [21].

The present study aimed to evaluate the training effect of aerobic exercise using arm ergometry and changes in serum myokines immediately after exercise and after a training program in patients with SCI. It was hypothesized that exercise training might improve aerobic capacity and muscle strength, and result in elevated myokines.

Methods

Patient eligibility

Patients with SCI visiting the out-patient clinic of the Department of Physical Medicine and Rehabilitation of National Taiwan University Hospital were recruited. The inclusion criteria were age >20 years, male sex, and onset of injury more than 1 year. The diagnosis was based on the American Spinal Injury Association (ASIA) standards. Patients with known heart disease, atrial fibrillation/flutter, ventricular bigeminy, active inflammatory disease, malignancy, and cardiac pacemakers were excluded. Those who were unable to perform arm ergometry were further excluded.

For comparison of baseline serum concentration of myokines, sex- and age-matched healthy volunteers in the same facility were recruited as the reference group. We certify that all applicable institutional and governmental regulations concerning the ethical use of human volunteers were followed during the course of this research. The hospital's Research Ethic Committee approved the study protocol and all of the participants provided written informed consent.

Experimental protocol

Basic data, including age, height, and weight were obtained at the time of enrollment. Height was measured to the nearest 0.1 cm and weight to the nearest 0.1 kg on an electronic scale. The duration from injury to enrollment, neurologic level, and ASIA impairment scale (AIS) were recorded for the patients with SCI.

All of the participants underwent CPET. Their HR and blood BP were monitored throughout the test at rest and every 30 s. The peak responses of these parameters during CPET were also recorded. Serum myostatin, IGF-1, and follistatin concentrations were measured immediately before and after the CPET.

The patients with SCI were invited to undergo a 12- to 16-week exercise training by arm ergometry. The CPET

and measurements of hemodynamic parameters and cytokines were repeated after completion of the training.

All the blood samples were taken when the participants were resting, immediately before and after the CPET and after the exercise training.

Cardio-pulmonary exercise testing (CPET)

With the patient seated on the arm ergometer (Corival, Lode B.V., Zernikepark 16, Groningen, Netherlands) and after 2 min of baseline data collection, there was a 2 min warm-up arm cranking with no resistance before the load was increased by 10 W/min in a ramp protocol. Arm cranking was at 60–70 revolutions per min while the workload was gradually increased until volitional exhaustion or termination according to the guidelines of American College of Sports Medicine [22]. The CPET was conducted by a physiatrist and performed at least two hours after a meal.

During exercise, continuous 12-lead electrocardiogram and blood pressure (TANGO, SunTech Medical Instruments, NC, USA) were monitored. The expired air was analyzed by computerized breath-by-breath metabolic system (Quark b2 system, Cosmed s.r.l., Rome, Italy, or Cortex MetaMax 3B system, Leipzig, Germany). Exercise cardio-pulmonary parameters, including workload, minute ventilation (V_E), oxygen uptake (VO_2), carbon dioxide production (VCO_2), oxygen pulse, ventilator equivalent for oxygen (V_E/VO_2), and ventilatory equivalent for carbon dioxide (V_E/VCO_2), were processed.

Two independent observers with experience in CPET determined the ventilatory threshold. The anaerobic threshold (AT) was based on at least two of the following criteria: 1) V_E/VO_2 began to increase systematically without a corresponding increase in V_E/VCO_2 ; 2) the end tidal PO_2 began to increase without a decrease in the end tidal PCO_2 ; and 3) departure from linearity for minute ventilation. Discrepancy in the interpretation AT was solved via discussion. The peak oxygen uptake (peak VO_2) indicated the highest VO_2 value recorded using CPET and was used as the main outcome after the training period.

Exercise training by arm ergometry

The exercise training by arm ergometry for the patients with SCI was 30 min per session, with 3 sessions per week for 12 weeks. In case of a missed session, the training duration was prolonged to a maximum of 16 weeks to complete 36 training sessions. The intensity of training was guided by CPET results, targeting an intensity of VO_2 at AT throughout the training program. Using a non-invasive hemodynamic monitoring system (Polar Electro OY, Finland), HR was monitored throughout the exercise and the BP every 10 min.

Interval training was adopted, with 1 min of rest between exercises. The duration of exercise progressed as follows: one min for the 1st week, 2 min for the 2nd week, 3 min for

the 3rd and 4th weeks, 6 min for the 5th and 6th weeks, 12 min for the 7th and 8th weeks, 15 min for the 9th and 10th weeks, and 30 min for the 11th and 12th weeks.

ELISA of myostatin, IGF-1, and follistatin

Serum myostatin levels were measured using competitive immunoassay kits according to the manufacturer's protocol (Immunodiagnostik AG, Bensheim, Germany). The full-length myostatin peptide was measured with high specificity. The test sensitivity was 270 pg/ml, while the intra- and inter-assay variabilities were <10 and <15 %, respectively [23, 24].

Serum IGF-1 levels were measured using ELISA kits (Mediagnost, Reutlingen, Germany). Sensitivity was 0.09 ng/ml and the inter- and intra-assay variability were 6.8 and 6.7 %, respectively [25].

Serum follistatin was analyzed by sandwich ELISA (QuantikineH, R&D Systems, Minneapolis, MN, USA). Sensitivity was 29 pg/ml and intra- and inter-assay variability were 2.4, and 7.1 %, respectively [25].

Statistical analysis

The optical density values versus concentration for the standard curve were fitted to a four-parameter logistic regression model. The test of differences between the SCI and reference groups was performed non-parametrically using the Mann–Whitney test, while the Wilcoxon matched pairs test was used to analyze changes in the parameters before and after exercise, or before and after training. Statistical significance was set at $p < 0.05$. Descriptive statistics (mean, standard deviation and coefficient of variation [CV]) and the statistical tests were performed using the SPSS® 11.5 (SPSS Inc. Chicago, Illinois, USA).

Results

Basic data

There were 16 patients with SCI recruited. Five were excluded because of inability perform arm ergometry. Six patients dropped out during the exercise training program for personal reasons (Fig. 1), no adverse event was reported during the training. Table 1 lists the injury levels and the impairment scales of the patients with SCI. The mean duration from onset of injury to exercise test was 13.9 ± 5.0 years.

There were 16 healthy male adults in the reference group, and no differences existed in age and body weight and height when compared with the SCI group (Table 2). There were also no differences in resting serum myokine concentrations between groups. The reference group had significantly higher heart rate, work load and oxygen uptake at peak exercise (Table 2).

Cardio-pulmonary exercise testing (CPET)

Table 2 lists the baseline values and changes of concentrations of myokines after CPET. The reference group had significantly increased IGF-1 after CPET (18 ± 19 % above baseline, $p = 0.001$), while myostatin and follistatin remained unchanged ($p = 0.179$ and 0.918 , respectively). The serum myokine concentrations in the SCI group after CPET showed no differences when compared with the values before CPET ($p = 0.594$, 0.772 , 0.504 , for myostatin, IGF-1, and follistatin, respectively).

Exercise training by arm ergometry

Five patients with SCI completed the 12–16 weeks of exercise training. After training, serum myostatin concentration increased by 48 ± 18 %, which was significantly higher compared to the pre-training status (Table 3). IGF-1 and follistatin remained unchanged. The increase in peak VO_2 and peak V_E were not significant. The body weight and resting and peak hemodynamic parameters also remained the same. Two of the five patients did not reach AT VO_2 during CPET due to fatigue.

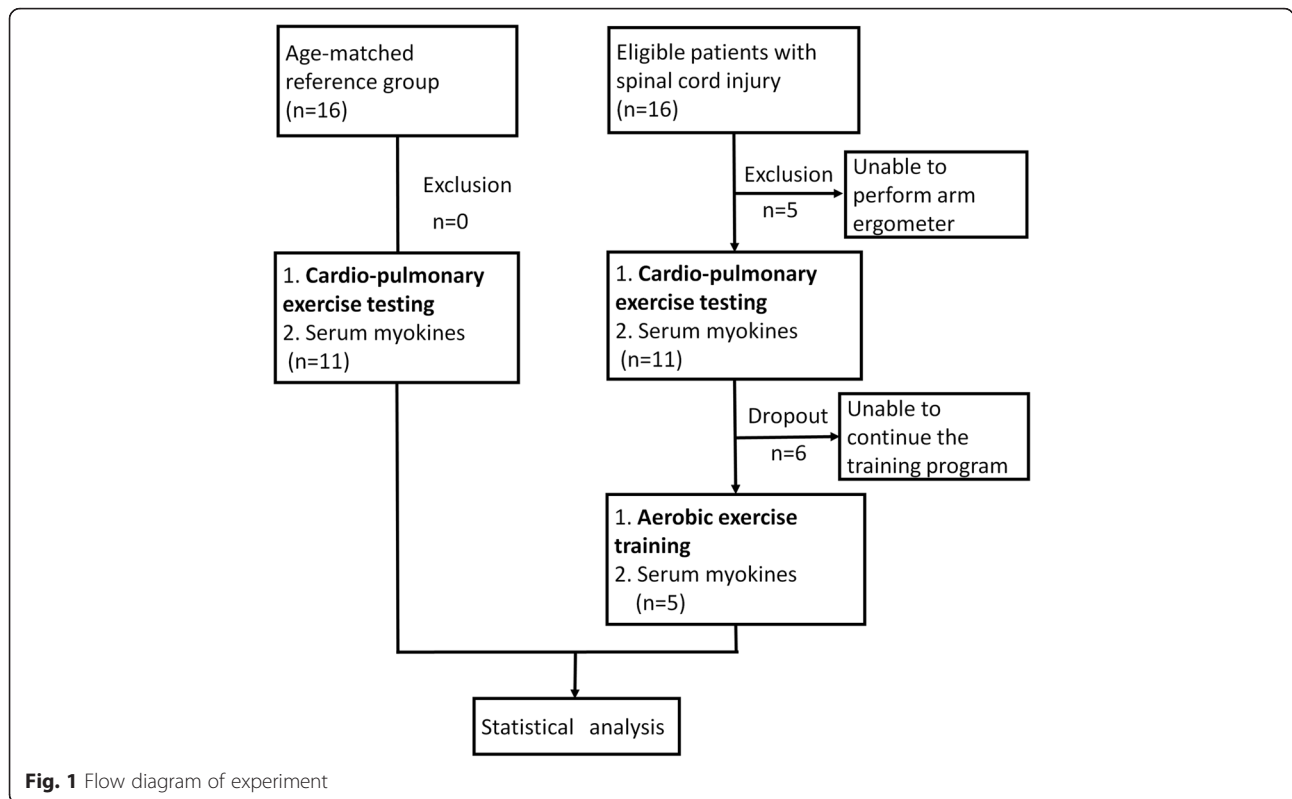
Discussion

The present study demonstrates that aerobic exercise training by arm ergometry results in increased serum myostatin level in patients with chronic SCI, even though the aerobic capacity and hemodynamic parameters do not change significantly. Acute exercise to exhaustion in the CPET leads to an immediate increase in serum IGF-1 in the reference group.

Myostatin has long been considered a cytokine that negatively regulates muscle growth. Recent evidence also point to its role in regulating the energy system of muscles. In the proposed “accelerator-brake” model, myostatin and IGF-1 act as counter-regulatory molecules for muscle hypertrophy [11–14, 20]. Thus, myostatin expression theoretically increases to limit the growth signals of muscle tissue such as exercise or growth factors.

On the other hand, animal studies suggest that myostatin is also related to aerobic capacity. *In vitro* study have shown that myostatin deficiency result in muscle fiber type conversion towards anaerobic metabolism [15, 16]. *In vivo*, there is also decreased exercise tolerance and oxygen consumption in myostatin-deficient mice [26]. Literature regarding responses of serum myostatin after exercise training in humans are limited and mainly focused on resistance training [27, 28]. Relevant data is limited for patients with SCI.

To date, this is the first study to show that aerobic exercise training by arm ergometry in patients with chronic SCI leads to elevated serum myostatin concentration. In the hypothesis of the “accelerator-brake” model [11–14, 20], IGF-1 increases first after exercise as muscle growth stimulus, followed by myostatin, which



increases correspondingly to provide negative feedback signal. Our study failed to show increased IGF-1 after exercise training which may be due to inadequate exercise intensity, although further evidence is required to test the hypothesis. On the other hand, the increased myostatin may indicate an initial change of aerobic metabolism of muscle after aerobic training, although there are no detectable peak VO₂ changes in CPET.

The present study shows an immediate increase in serum IGF-1 level after acute exercise to exhaustion in

Table 1 Injury level and severity of SCI among the study patients

| Patient | Bone level | Motor level | Sensory level | Neurologic level | AIS grade | Dropout |
|---------|------------|-------------|---------------|------------------|-----------|---------|
| 1 | C3 | NA | C4 | C4 | D | - |
| 2 | C5 | C5 | C5 | C5 | A | - |
| 3 | NA | T6 | T6 | T6 | A | - |
| 4 | L1-2 | L1 | L1 | L1 | A | - |
| 5 | C5 | C6 | C5 | C5 | B | - |
| 6 | C4-5 | C4 | C4 | C4 | A | + |
| 7 | L4 | L3 | L2 | L2 | D | + |
| 8 | L1 | L1 | L1 | L1 | A | + |
| 9 | C4-5 | C5 | C4 | C4 | A | + |
| 10 | C5-6 | C6 | C6 | C6 | A | + |
| 11 | NA | T1 | C5 | C5 | C | + |

Abbreviations: AIS the American spinal injury association impairment scale; NA not applicable

CPET, but only in healthy participants and not in patients with SCI. IGF-1 is an age-related serum protein with growth controlling ability [17]. It affects the development of multiple systems, with the musculo-skeletal as one of the most important [18, 19]. Previous trials report an immediate increase in serum IGF-1 after bouts of exercise in healthy adults [29–31]. It was hypothesized that IGF-1 is released from skeletal muscle independent of growth hormone stimulation. The mean percentage of increase ranges from 13 % to 33 % in different studies [29–31], while the results here show an average of 18 % increase. Of note, the increase in IGF-1 is also influenced by exercise intensity [31]. All of the trials used an intensity of exercise training at or above AT VO₂. Part of the reason for the unresponsiveness of serum IGF-1 level after exercise in patients with SCI in the present study may be inadequate exercise intensity during CPET due to early fatigue.

There are few related studies available and a recent trial indicates no change in myostatin and IGF-I, as well as in body weight, among patients with SCI after chronic exercise training [21]. In contrast to the aerobic training by arm ergometry in this study, the cited trial used activity-based training that mainly consisted of resistance and locomotor training focused on the lower extremities.

The study had several limitations. Since this study did not measure the change in skeletal muscle mass or

Table 2 Basic data and change of serum myokines after exercise testing in reference and SCI (pre-training) groups

| | | Reference (n = 16) | SCI (n = 11) | p value ^b |
|--------------------|-----------------------------|--------------------------|-------------------|----------------------|
| Basic data | Age (year) | 44.7 ± 6.4 | 39.9 ± 7.1 | 0.093 |
| | Height (cm) | 171.8 ± 4.6 | 169.7 ± 5.7 | 0.298 |
| | Weight (kg) | 71.9 ± 9.9 | 72.0 ± 11.5 | 0.786, |
| | BMI (kg/m ²) | 24.3 ± 2.4 | 25.0 ± 3.7 | 0.693 |
| Resting | myostatin (ng/ml) | 6.1 ± 1.6 | 6.1 ± 1.8 | 0.980 |
| | IGF-1 (ng/ml) | 223.4 ± 62.2 | 216.4 ± 49.5 | 0.805 |
| | follistatin (ng/ml) | 1284.4 ± 809.0 | 1100.9 ± 354.7 | 0.587 |
| Change of myokines | Δmyostatin (ng/ml) | 0.4 ± 1.6 | 0.3 ± 1.1 | 0.348 |
| | ΔIGF-1 (ng/ml) | 40.6 ± 43.0 ^a | 2.1 ± 48.3 | 0.030 |
| | Δfollistatin (ng/ml) | 59.4 ± 349.7 | 55.5 ± 498.4 | 0.657 |
| Peak exercise | HR (beat/min) | 138.8 ± 13.8 | 117.4 ± 23.8 | 0.031 |
| | VO ₂ (ml/min) | 1454.5 ± 263.9 | 877.5 ± 426.2 | 0.012 |
| | VO ₂ (ml/min/kg) | 20.5 ± 4.0 | 12.3 ± 5.5 | 0.026 |
| | V _E (ml/min) | 63912.5 ± 15181.1 | 32845.5 ± 15628.5 | 0.032 |
| | Work load (watt) | 38.0 ± 31.0 | 80.0 ± 19.0 | 0.009 |

Abbreviations: SCI spinal cord injury, BMI body mass index, IGF-1 insulin-like growth factor-1, HR heart rate, VO₂ oxygen uptake, V_E minute ventilation

^ap < 0.05 by comparison between resting and peak exercise status by Wilcoxon matched pairs test

^bComparison between reference and SCI groups by Mann–Whitney test

Table 3 The comparison in the SCI groups before and after 12–16 weeks of training

| | | SCI (n = 5) | | P value ^a |
|---------------|-----------------------------|-------------------|-------------------|----------------------|
| | | Pre-training | Post-training | |
| Basic data | Age (year) | 39.6 ± 7.4 | - | - |
| | Height (cm) | 167.6 ± 6.0 | 167.6 ± 6.0 | - |
| | Weight (kg) | 65.6 ± 6.6 | 67.9 ± 7.3 | 0.180 |
| | BMI (kg/m ²) | 23.5 ± 3.4 | 24.3 ± 3.8 | 0.180 |
| Resting | HR (beat/min) | 72.5 ± 13.8 | 71.8 ± 13.8 | 0.992 |
| | SBP (mmHg) | 109.5 ± 24.8 | 110.6 ± 32.1 | 0.128 |
| | DBP (mmHg) | 65.9 ± 11.6 | 67.4 ± 15.0 | 0.799 |
| Peak exercise | HR (beat/min) | 117.4 ± 23.8 | 120.4 ± 19.7 | 0.866 |
| | SBP (mmHg) | 136.8 ± 28.0 | 145.6 ± 34.1 | 0.446 |
| | DBP (mmHg) | 77.8 ± 14.9 | 78.6 ± 18.5 | 0.204 |
| | VO ₂ (ml/min) | 877.5 ± 426.2 | 902.4 ± 360.7 | 0.553 |
| | VO ₂ (ml/min/kg) | 12.3 ± 5.5 | 13.9 ± 5.5 | 0.866 |
| | V _E (ml/min) | 32845.5 ± 15628.5 | 33580.0 ± 13222.2 | 0.398 |
| | Work load (watt) | 39.6 ± 27.9 | 59.6 ± 32.1 | 0.312 |
| Myokines | myostatin (ng/ml) | 6.1 ± 2.0 | 9.0 ± 1.5 | 0.018 |
| | IGF-1 (ng/ml) | 219.7 ± 51.7 | 204.4 ± 59.6 | 0.176 |
| | follistatin (ng/ml) | 1042.7 ± 351.1 | 909.8 ± 409.0 | 0.398 |

Abbreviations: SCI spinal cord injury, BMI body mass index, HR heart rate, SBP systolic blood pressure, DBP diastolic blood pressure, VO₂ oxygen uptake, V_E minute ventilation; IGF-1 insulin-like growth factor-1

^aby comparison between pre- and post-training groups using the Wilcoxon matched pairs test

strength after exercise training, whether increased myostatin reflects a signal of muscle growth or a change in aerobic metabolism remains unanswered. Some of the patients with SCI in this study did not attain AT VO_2 during CPET due to early limb fatigue, and the true aerobic capacity may be underestimated. Similarly, the intensity of training (targeting an intensity of VO_2 at AT) may also be inadequate for most of the patients because of early limb fatigue. Furthermore, the reference group did not receive exercise training and as such, there is no comparison of training effect for patients with SCI. The limited case number due to difficulty in enrollment further lessens the discriminative power for the effect of exercise training. Moreover, there is a substantial heterogeneity in the SCI group, with varying severities and levels of injury, and large inter-individual differences in serum myokines. Further subgroup analysis is hampered by the small case number. Nevertheless, the results still show a marked increase in serum myostatin level after aerobic exercise training in chronic patients with SCI, although further studies are warranted to clarify the interaction between serum myostatin and changes in aerobic metabolism or muscle growth after training.

Conclusions

This study evaluates the effects of aerobic exercise training by arm ergometry on serum concentrations of myokines in patients with chronic SCI. Acute exercise to exhaustion in CPET results in an immediate increase in serum IGF-1 in healthy individuals while aerobic exercise training results in increased serum myostatin in patients with SCI. The interaction between serum myostatin and changes in aerobic metabolism or muscle growth after training requires elucidation by further large-scale prospective studies.

Abbreviations

SCI, spinal cord injury; CPET, Cardio-pulmonary exercise testing; VO_2 , oxygen uptake; AT, anaerobic threshold; VCO_2 , carbon dioxide production; IGF-1, insulin-like growth factor-1; \dot{V}_E , minute ventilation; HR, heart rate; BP, blood pressure; ASIA, American Spinal Injury Association

Acknowledgements

The authors thank all study investigators, participants, and care partners who assisted with this trial.

Funding

This research was supported by grants from the National Science Council of Taiwan (NSC 100-2314-B-002-014 and NSC 99-2314-B-002-038-MY3).

Availability of data and materials

Data that have been used to draw conclusions within this manuscript are reported in full. The study was retrospectively registered at Clinicaltrials.gov. (Trial registration protocol ID: 201007031R; ClinicalTrials.gov Identifier: NCT02850133; date: Aug 1st, 2010 to Jul 31st, 2013; available at <https://clinicaltrials.gov/show/NCT02850133>).

Authors' contributions

D-SH: conception and design of the study, acquisition of data, interpretation of data; revision of the manuscript. M-YH: acquisition and analysis of data, interpretation of data; drafting and revision of the manuscript. T-GW:

interpretation of data; drafting and revision of the manuscript. S-YC: design of the study, interpretation of data, revision of the manuscript. W-SY: design of the study, interpretation of data, revision of the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval and consent to participate

National Taiwan University Hospital Research Ethics Committee reviewed and approved the protocol. Written informed consent was obtained from all participants or from legally authorized representatives.

Author details

¹Department of Physical Medicine and Rehabilitation, National Taiwan University Hospital Beihu Branch, Taipei, Taiwan. ²Department of Physical Medicine and Rehabilitation, National Taiwan University Hospital, Taipei, Taiwan. ³Community and Geriatric Medicine Research Center, National Taiwan University Hospital Beihu Branch, Taipei, Taiwan. ⁴Department of Internal Medicine, National Taiwan University Hospital, No. 1, Chang-Teh St, Taipei, Taiwan. ⁵Research Center for Developmental Biology and Regenerative Medicine, National Taiwan University, Taipei, Taiwan.

Received: 16 December 2015 Accepted: 2 August 2016

Published online: 17 August 2016

References

- Garshick E, Kelley A, Cohen SA, et al. A prospective assessment of mortality in chronic spinal cord injury. *Spinal Cord*. 2005;43(7):408–16.
- DeVivo MJ, Krause JS, Lammertse DP. Recent trends in mortality and causes of death among persons with spinal cord injury. *Arch Phys Med Rehabil*. 1999;80(11):1411–9.
- Krum H, Howes LG, Brown DJ, et al. Risk factors for cardiovascular disease in chronic spinal cord injury patients. *Paraplegia*. 1992;30(6):381–8.
- Myers J, Lee M, Kiratli J. Cardiovascular disease in spinal cord injury: an overview of prevalence, risk, evaluation, and management. *Am J Physical Med Rehabil/Assoc Acad Physiatrists*. 2007;86(2):142–52.
- Phillips WT, Kiratli BJ, Sarkarati M, et al. Effect of spinal cord injury on the heart and cardiovascular fitness. *Curr Probl Cardiol*. 1998;23(11):641–716.
- Karlsson AK. Autonomic dysfunction in spinal cord injury: clinical presentation of symptoms and signs. *Prog Brain Res*. 2006;152:1–8.
- Myers JN, Hsu L, Hadley D, Lee MY, Kiratli BJ. Post-exercise heart rate recovery in individuals with spinal cord injury. *Spinal Cord*. 2010;48(8):639–44.
- Leon AS, Franklin BA, Costa F, et al. Cardiac rehabilitation and secondary prevention of coronary heart disease: an American Heart Association scientific statement from the Council on Clinical Cardiology (Subcommittee on Exercise, Cardiac Rehabilitation, and Prevention) and the Council on Nutrition, Physical Activity, and Metabolism (Subcommittee on Physical Activity), in collaboration with the American association of Cardiovascular and Pulmonary Rehabilitation. *Circulation*. 2005;111(3):369–76.
- Cragg JJ, Stone JA, Krassioukov AV. Management of cardiovascular disease risk factors in individuals with chronic spinal cord injury: an evidence-based review. *J Neurotrauma*. 2012;29(11):1999–2012.
- Jacobs PL, Nash MS. Exercise recommendations for individuals with spinal cord injury. *Sports Med*. 2004;34(11):727–51.
- Shyu KG, Ko WH, Yang WS, Wang BW, Kuan P. Insulin-like growth factor-1 mediates stretch-induced upregulation of myostatin expression in neonatal rat cardiomyocytes. *Cardiovasc Res*. 2005;68(3):405–14.
- Gaussin V, Depre C. Myostatin, the cardiac chalone of insulin-like growth factor-1. *Cardiovasc Res*. 2005;68(3):347–9.
- Mak RH, Rotwein P. Myostatin and insulin-like growth factors in uremic sarcopenia: the yin and yang in muscle mass regulation. *Kidney Int*. 2006;70(3):410–2.
- Han DS, Chen YM, Lin SY, et al. Serum myostatin levels and grip strength in normal subjects and patients on maintenance hemodialysis. *Clin Endocrinol (Oxf)*. 2011;75(6):857–63.

15. Hennebry A, Berry C, Siriatt V, et al. Myostatin regulates fiber-type composition of skeletal muscle by regulating MEF2 and MyoD gene expression. *American journal of physiology. Cell Physiol.* 2009;296(3):C525–34.
16. Baligand C, Gilson H, Menard JC, et al. Functional assessment of skeletal muscle in intact mice lacking myostatin by concurrent NMR imaging and spectroscopy. *Gene Ther.* 2010;17(3):328–37.
17. Grimberg A, Cohen P. Role of insulin-like growth factors and their binding proteins in growth control and carcinogenesis. *J Cell Physiol.* 2000;183(1):1–9.
18. Daughaday WH, Rotwein P. Insulin-like growth factors I and II. Peptide, messenger ribonucleic acid and gene structures, serum, and tissue concentrations. *Endocr Rev.* 1989;10:68–91.
19. Schiaffino S, Mammucari C. Regulation of skeletal muscle growth by the IGF1-Akt/PKB pathway: insights from genetic models. *Skelet Muscle.* 2011;1(1):4.
20. Chien YH, Han DS, Hwu WL, Thurberg BL, Yang WS. Myostatin and insulin-like growth factor I: potential therapeutic biomarkers for pompe disease. *PLoS One.* 2013;8(8):e71900.
21. Astorino TA, Harness ET, Witzke KA. Chronic activity-based therapy does not improve body composition, insulin-like growth factor-I, adiponectin, or myostatin in persons with spinal cord injury. *J Spinal Cord Med.* 2015;38(5):615–25.
22. Balady GJ, Arena R, Sietsema K, et al. Clinician's Guide to cardiopulmonary exercise testing in adults: a scientific statement from the American Heart Association. *Circulation.* 2010;122(2):191–225.
23. Wintgens KF, Dschietzig T, Stoeva S, Paulsson M, Armbruster FP. Plasma myostatin measured by a competitive ELISA using a highly specific antiserum. *Clin Chim Acta.* 2012;413:1288–94.
24. Jones G, Monika W, Kreissig SB, et al. Extension of the four-parameter logistic model for ELISA to multianalyte analysis. *J Immunol Methods.* 1994;177:1–7.
25. Haidet AM, Rizo L, Handy C, et al. Long-term enhancement of skeletal muscle mass and strength by single gene administration of myostatin inhibitors. *Proc Natl Acad Sci U S A.* 2008;105(11):4318–22.
26. Mouisel E, Relizani K, Mille-Hamad L, et al. Myostatin is a key mediator between energy metabolism and endurance capacity of skeletal muscle. *Am J Physiol Regulatory Int Comparative Physiol.* 2014;307(4):R444–54.
27. Paoli A, Pacelli QF, Neri M, et al. Protein Supplementation Increases Postexercise Plasma Myostatin Concentration After 8 Weeks of Resistance Training in Young Physically Active Subjects. *J Med Food.* 2014;18(1):137–43.
28. Willoughby DS. Effects of heavy resistance training on myostatin mRNA and protein expression. *Med Sci Sports Exerc.* 2004;36(4):574–82.
29. Cappon J, Brasel JA, Mohan S, Cooper DM. Effect of brief exercise on circulating insulin-like growth factor I. *J Appl Physiol.* 1994;76(6):2490–6.
30. El Ej N, Elloumi M, Zaouali M, Latiri I, Lac G, Tabka Z. Discrepancy in IGF-1 and GH response to submaximal exercise in young male subjects. *Sci Sports.* 2007;22(3–4):155–9.
31. Schwarz AJ, Brasel JA, Hintz RL, Mohan S, Cooper DM. Acute effect of brief low- and high-intensity exercise on circulating insulin-like growth factor (IGF) I, II, and IGF-binding protein-3 and its proteolysis in young healthy men. *J Clin Endocrinol Metab.* 1996;81(10):3492–7.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

