

CUH or from subsequent contact with unsuspected carriers in the case-patient's family or the community. On the basis of a matching surname, case-patient E was determined to be a member of the same family as case-patients P20, P26, and likely C. Soft tissue infection was documented in all 5 case-patients, supporting the original observation that ST2371 is associated with disease. Evidence of familial transmission in the original outbreak is further supported by transmission between case-patients P20, P26, C, and E. Furthermore, 2 case-patients infected during the original outbreak, P22/A and P14/B, continued to experience disease signs and symptoms for ≥ 15 months after their initial diagnosis.

Our data highlight the role of hospitals as reservoirs of MRSA and subsequent failure to track the entry and spread of MRSA in the community. MRSA decolonization was advised in all cases in the original outbreak, but this process clearly proved ineffective for case-patients A and B. Potential explanations include not implementing or completing the course of decolonization; failed decolonization; or limiting decolonization to only some members of an affected family. Although the outbreak in the hospital ward was resolved, the lack of a systematic surveillance program to monitor the incidence of noninvasive MRSA infections among the case-patients' contacts and the community allowed this novel lineage to continue to cause disease in a group of linked persons. Considering recommendations to move from universal to targeted MRSA screening in hospitals in England (6), more active surveillance of any identified case-patients or carriers of MRSA in the community may be warranted.

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M.S.T., F.C., E.M.H., and S.R. undertook epidemiological and bioinformatic analysis of whole-genome sequence data. S.R. prepared the figure. S.J.P. supervised and managed the study. All authors were involved in compiling the report and approved the final version.

References

- Harris SR, Cartwright EJP, Török ME, Holdem MT, Brown NM, Ogilvy-Stewart AL, et al. Whole-genome sequencing for analysis of an outbreak of methicillin-resistant *Staphylococcus aureus*: a descriptive study. *Lancet Infect Dis*. 2013;13:130–6. [http://dx.doi.org/10.1016/S1473-3099\(12\)70268-2](http://dx.doi.org/10.1016/S1473-3099(12)70268-2)
- Grundmann H, Aanensen DM, van den Wijngaard CC, Spratt BG, Harmsen D, Friedrich AW, et al. Geographic distribution of

Staphylococcus aureus causing invasive infections in Europe: a molecular-epidemiological analysis. *PLoS Med*. 2010;7:e1000215. <http://dx.doi.org/10.1371/journal.pmed.1000215>

- Rajan V, Schoenfelder SM, Ziebuhr W, Gopal S. Genotyping of community-associated methicillin resistant *Staphylococcus aureus* (CA-MRSA) in a tertiary care centre in Mysore, South India: ST2371-SCCmec IV emerges as the major clone. *Infect Genet Evol*. 2015;34:230–5. <http://dx.doi.org/10.1016/j.meegid.2015.05.032>
- Bouchiat C, El-Zeenni N, Chakrakodi B, Nagaraj S, Arakere G, Etienne J. Epidemiology of *Staphylococcus aureus* in Bangalore, India: emergence of the ST217 clone and high rate of resistance to erythromycin and ciprofloxacin in the community. *New Microbes New Infect*. 2015;7:15–20. <http://dx.doi.org/10.1016/j.nmni.2015.05.003>
- Bartels MD, Larner-Svensson H, Meiniche H, et al. Monitoring methicillin resistant *Staphylococcus aureus* and its spread in Copenhagen, Denmark, 2013, through routine whole genome sequencing. *Euro Surveill*. 2015;20:pii:21112.
- Department of Health expert committee on Antimicrobial Resistance and Healthcare Associated Infection. Implementation of modified admission MRSA screening guidance for NHS (2014) [cited 2015 May 20]. https://www.gov.uk/government/uploads/system/uploads/attachment_data/file/345144/Implementation_of_modified_admission_MRSA_screening_guidance_for_NHS.pdf

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Community-Acquired *Clostridium difficile* Infection, Queensland, Australia

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To the Editor: In Queensland, Australia, a steady increase in community-acquired (CA) *Clostridium difficile* infections (CDI) during 2003–2012 could not be explained by patients' demographic characteristics or environmental

factors (1). Several risk factors have been implicated in the increased rates of CA-CDI, primarily exposure to antimicrobial drugs, gastric acid-suppression drugs, and corticosteroids (2). Given the recent rise in prescription of corticosteroids and proton pump inhibitors in Australia, we hypothesized that the observed increase in CA-CDI was associated with increased drug prescriptions.

To test our hypothesis, we analyzed a subset of data used in a previous study (1), which included fecal samples from patients seen by general practitioners in the community from January 2008 through December 2012. The samples were submitted to Sullivan Nicolaides Pathology (Taringa, Queensland, Australia) for *C. difficile* toxin gene detection. After samples submitted from healthcare facilities and nursing homes were excluded, the final dataset contained data from 14,330 fecal samples. We aggregated the data by patient sex, age categories, year, and statistical area level 4 (SA4). For each sex-age-year-SA4 group, we used as numerators the numbers of CA-CDI cases identified and as denominators the numbers of samples submitted for microbiological testing.

The Australian Department of Human Services provided data from the Pharmaceutical Benefits Scheme. The quantities of 11 anatomic therapeutic chemical drugs were accessed by patient sex, age group, year, and SA4. Corresponding with the CA-CDI data, medication data to be analyzed were then aggregated by sex, age group, year, and SA4.

For each medication, we built binomial logistic regression models, using CA-CDI status as the outcome, in a Bayesian framework, incorporating fixed effects for sex, age group, quantity of drug prescribed, year (2008–2012), and spatially unstructured random effects at the SA4 level. After performing an initial burn-in, we stored and summarized 1,000 values from the posterior distribution of each parameter by using descriptive statistics (posterior mean, 95% posterior credible interval [95% CrI], and p value). We examined multiple pairwise comparisons of CA-CDI and medication exposure; thus, we used the Holm adjustment for p values to avoid inflation and to control the familywise error rate.

Of the 14,330 fecal samples tested, 1,430 (10%) were positive for *C. difficile*. The proportion of positive fecal samples increased over the 5-year period, from 7.10% in 2008 to 12.72% in 2011 and 11.48% in 2012 ($p < 0.001$). After adjusting the regression models for sex, age group, temporal pattern, and spatial distribution, we found that exposure to antimycobacterial drugs (odds ratio [OR] 1.09; 95% CrI 1.02–1.16) and anthelmintic drugs (OR 1.07; 95% CrI 1.01–1.13) were associated with increased odds of CA-CDI. After post hoc Holm adjustments, no statistically significant association between medication exposure and CA-CDI was observed (Table).

Our findings suggest that the increase in CA-CDI proportion was not associated with population-level medication exposure in Queensland during 2008–2012. CA-CDI epidemiology in Queensland might be driven by a group of factors other than medication exposure, such as transmission of the pathogen from food, animals, or hospitals into the community. Studies have confirmed the risk for foodborne and animalborne spread of *C. difficile* into the community (3). In Australia and New Zealand, importation of onions and garlic from the United States and Mexico might be responsible for increased CDI cases during Southern Hemisphere summers (4), and high prevalence of *C. difficile* colonization in piglets has been identified (5). However, the role of these factors in leading to CA-CDI cases remains unknown.

A recent contact tracing study in the United Kingdom demonstrated that a considerable proportion of CDIs among patients in healthcare settings originated from the community (6); this finding was supported by another study, which showed that in Queensland, more than two thirds of patients with CA-CDI required hospitalization (7). Currently, there is no evidence of a reverse-infection route (healthcare-acquired CDI being transmitted to persons in the community). However, Sethi et al. documented environmental shedding of *C. difficile* by inpatients for several weeks after resolution of symptoms (8). Therefore, the possibility that asymptomatic patients might be a source of transmission after hospital discharge needs to be examined. In recent years, epidemiologic models

Table. Binomial logistic regression models for medication exposure adjusted for sex, age group, temporal pattern, and spatial distribution among patients with community-acquired *Clostridium difficile* infection, Queensland, Australia, 2003–2012

Medication exposure	Odds ratio (95% credible interval)	p value	Holm-adjusted p value
Drugs for acid-related disorders	1.052 (0.943–1.163)	0.348	0.819
Drugs for constipation	1.056 (0.963–1.151)	0.235	0.781
Antidiarrheal drugs	1.106 (0.994–1.218)	0.051	0.379
Antithrombotic drugs	1.073 (0.955–1.197)	0.224	0.781
Corticosteroids for systemic use	1.043 (0.952–1.133)	0.348	0.819
Antibacterial drugs for systemic use	1.083 (0.990–1.174)	0.067	0.425
Antimycotic drugs for systemic use	1.035 (0.944–1.126)	0.454	0.819
Antimicrobial drugs for mycobacterial infections	1.089 (1.023–1.155)	0.006	0.063
Anti-inflammatory drugs	1.070 (0.970–1.170)	0.158	0.700
Antiprotozoal drugs	1.037 (0.953–1.123)	0.394	0.819
Anthelmintic drugs	1.068 (1.008–1.127)	0.021	0.189

exploring the role of CDI coming from the community into the hospital have become increasingly popular (9); however, to the best of our knowledge, only 1 modeling study described CDI dynamics within the wider community (10). Although this approach is innovative, we acknowledge some limitations. Medication exposure was used as a proxy, based on the average prescription in the community, and it cannot be applied to the individual patient. In addition, we were unable to adjust the regression model for the presence of concurrent medical conditions and other unmeasured confounders.

Exposure to medications, particularly antimicrobial drugs, probably influences CA-CDI pathogenesis (2). However, our community-based assessment indicates that a more holistic exploration is needed to identify alternative factors driving increases in CA-CDI cases in the wider population.

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References

1. Furuya-Kanamori L, Robson J, Soares Magalhaes RJ, Yakob L, McKenzie SJ, Paterson DL, et al. A population-based spatio-temporal analysis of *Clostridium difficile* infection in Queensland, Australia over a 10-year period. *J Infect*. 2014;69:447–55. <http://dx.doi.org/10.1016/j.jinf.2014.06.014>
2. Furuya-Kanamori L, Stone JC, Clark J, McKenzie SJ, Yakob L, Paterson DL, et al. Comorbidities, exposure to medications, and the risk of community-acquired *Clostridium difficile* infection: a systematic review and meta-analysis. *Infect Control Hosp Epidemiol*. 2015;36:132–41. <http://dx.doi.org/10.1017/ice.2014.39>
3. Songer JG, Trinh HT, Killgore GE, Thompson AD, McDonald LC, Limbago BM. *Clostridium difficile* in retail meat products, USA, 2007. *Emerg Infect Dis*. 2009;15:819–21. <http://dx.doi.org/10.3201/eid1505.081071>
4. Riley T. *Clostridium difficile* infection: the Australian experience. 2013 [cited 2016 Mar 1]. <http://www.hqsc.govt.nz/assets/Infection-Prevention/CDI-workshop-Feb-2013-Riley.pdf>
5. Knight DR, Squire MM, Riley TV. Nationwide surveillance study of *Clostridium difficile* in Australian neonatal pigs shows high prevalence and heterogeneity of PCR ribotypes. *Appl Environ Microbiol*. 2015;81:119–23. <http://dx.doi.org/10.1128/AEM.03032-14>
6. Walker AS, Eyre DW, Wyllie DH, Dingle KE, Harding RM, O'Connor L, et al. Characterisation of *Clostridium difficile* hospital ward-based transmission using extensive epidemiological data and molecular typing. *PLoS Med*. 2012;9:e1001172. <http://dx.doi.org/10.1371/journal.pmed.1001172>
7. Huber CA, Hall L, Foster NF, Gray M, Allen M, Richardson LJ, et al. Surveillance snapshot of *Clostridium difficile* infection in hospitals across Queensland detects binary toxin producing ribotype UK 244. *Commun Dis Intell Q Rep*. 2014;38:E279–84.
8. Sethi AK, Al-Nassir WN, Nerandzic MM, Bobulsky GS, Donskey CJ. Persistence of skin contamination and environmental shedding of *Clostridium difficile* during and after treatment of *C. difficile* infection. *Infect Control Hosp Epidemiol*. 2010;31:21–7. <http://dx.doi.org/10.1086/649016>
9. Yakob L, Riley T, Paterson D, Clements A. *Clostridium difficile* exposure as an insidious source of infection in healthcare settings: an epidemiological model. *BMC Infect Dis*. 2013;13:376. <http://dx.doi.org/10.1186/1471-2334-13-376>
10. Yakob L, Riley TV, Paterson DL, Marquess J, Soares Magalhaes RJ, Furuya-Kanamori L, et al. Mechanisms of hypervirulent *Clostridium difficile* ribotype 027 displacement of endemic strains: an epidemiological model. *Sci Rep*. 2015;5:12666. <http://dx.doi.org/10.1038/srep12666>

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Multidrug-Resistant *Campylobacter coli* in Men Who Have Sex with Men, Quebec, Canada, 2015

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To the Editor: In 2015, an outbreak of multidrug-resistant *Campylobacter coli* was documented in Montreal, Quebec, Canada. We report results of an epidemiologic and molecular investigation suggesting a sexually transmitted enteric infection among men who have sex with men (MSM).

The ethics committee of Centre Hospitalier de l'Université de Montréal approved the research. During January 14–February 7, 2015, six men 35–62 years of age were documented with an enteric, erythromycin-, tetracycline- and ciprofloxacin-resistant *C. coli* pulsovar 15 infection. All 6 men had diarrhea; 5 had abdominal pain; 1 had fever $\geq 39^{\circ}\text{C}$; 1 had blood in feces; and 1 had vomiting. No extraintestinal focus was documented in these patients.

Five men were evaluated in the outpatient clinic or emergency department; 1 man was hospitalized for 3 days. Five patients were treated with an antimicrobial agent.