- 2. Howat WJ, Wilson BA. Tissue fixation and the effect of molecular fixatives on downstream staining procedures. Methods 2014; 70: 12–19.
- Turashvili G, Yang W, McKinney S et al. Nucleic acid quantity and quality from paraffin blocks: defining optimal fixation, processing and DNA/RNA extraction techniques. Exp Mol Pathol 2012; 92: 33–43.
- Gillespie JW, Best CJ, Bichsel VE et al. Evaluation of non-formalin tissue fixation for molecular profiling studies. Am J Pathol 2002; 160: 449–457.
- Vincek V, Nassiri M, Nadji M, Morales AR. A tissue fixative that protects macromolecules (DNA, RNA, and protein) and histomorphology in clinical samples. Lab Invest 2003; 83: 1427–1435.
- Srinivasan M, Sedmak D, Jewell S. Effect of fixatives and tissue processing on the content and integrity of nucleic acids. Am J Pathol 2002; 161: 1961–1971.
- Cox ML, Schray CL, Luster CN et al. Assessment of fixatives, fixation, and tissue processing on morphology and RNA integrity. Exp Mol Pathol 2006; 80: 183–191.
- Nassiri M, Ramos S, Zohourian H et al. Preservation of biomolecules in breast cancer tissue by a formalin-free histology system. BMC Clin Pathol 2008; 8: 1.
- Ahmed AA, Etemadmoghadam D, Temple J et al. Driver mutations in TP53 are ubiquitous in high grade serous carcinoma of the ovary. J Pathol 2010; 221: 49–56.
- Leroy B, Anderson M, Soussi T. TP53 mutations in human cancer: database reassessment and prospects for the next decade. Hum Mutat 2014; 35: 672–688.
- Forshew T, Murtaza M, Parkinson C et al. Noninvasive identification and monitoring of cancer mutations by targeted deep sequencing of plasma DNA. Sci Transl Med 2012; 4: 136ra168.
- Alexandrov LB, Nik-Zainal S, Wedge DC et al. Signatures of mutational processes in human cancer. Nature 2013; 500: 415–421.
- McGhee JD, von Hippel PH. Formaldehyde as a probe of DNA structure. I. Reaction with exocyclic amino groups of DNA bases. Biochemistry 1975; 14: 1281–1296.

- Quach N, Goodman MF, Shibata D. In vitro mutation artifacts after formalin fixation and error prone translesion synthesis during PCR. BMC Clin Pathol 2004; 4: 1.
- De Giorgi C, Finetti Sialer M, Lamberti F. Formalin-induced infidelity in PCRamplified DNA fragments. Mol Cell Probes 1994; 8: 459–462.
- Carrick DM, Mehaffey MG, Sachs MC et al. Robustness of next generation sequencing on older formalin-fixed paraffin-embedded tissue. PLoS One 2015; 10: e0127353.
- Hedegaard J, Thorsen K, Lund MK et al. Next-generation sequencing of RNA and DNA isolated from paired fresh-frozen and formalin-fixed paraffin-embedded samples of human cancer and normal tissue. PLoS One 2014; 9: e98187.
- Do H, Dobrovic A. Dramatic reduction of sequence artefacts from DNA isolated from formalin-fixed cancer biopsies by treatment with uracil-DNA glycosylase. Oncotarget 2012; 3: 546–558.
- Kerick M, Isau M, Timmermann B et al. Targeted high throughput sequencing in clinical cancer settings: formaldehyde fixed-paraffin embedded (FFPE) tumor tissues, input amount and tumor heterogeneity. BMC Med Genomics 2011; 4: 68.
- Ewing AD, Houlahan KE, Hu Y et al. Combining tumor genome simulation with crowdsourcing to benchmark somatic single-nucleotide-variant detection. Nat Methods 2015; 12: 623–630.
- Lin J, Kennedy SH, Svarovsky T et al. High-quality genomic DNA extraction from formalin-fixed and paraffin-embedded samples deparaffinized using mineral oil. Anal Biochem 2009; 395: 265–267.
- Wong SQ, Li J, Tan AY et al. Sequence artefacts in a prospective series of formalin-fixed tumours tested for mutations in hotspot regions by massively parallel sequencing. BMC Med Genomics 2014; 7: 23.
- Cox ML, Eddy SM, Stewart ZS et al. Investigating fixative-induced changes in RNA quality and utility by microarray analysis. Exp Mol Pathol 2008; 84: 156–172.

Annals of Oncology 27: 539–543, 2016 doi:10.1093/annonc/mdv598 Published online 8 December 2015

TP53 mutational status is predictive of pazopanib response in advanced sarcomas

K. Koehler¹, D. Liebner^{1,2} & J. L. Chen^{1,2*}

¹Division of Medical Oncology, Department of Internal Medicine, The Ohio State University, Columbus; ²Division of Bioinformatics, Department of Biomedical Informatics, The Ohio State University, Columbus, USA

Received 14 October 2015; revised 18 November 2015; accepted 20 November 2015

Background: To investigate whether TP53 DNA mutational status impacts progression-free survival (PFS) in patients with advanced sarcomas (soft tissue sarcoma) treated with vascular endothelial growth factor receptors (VEGFR) inhibition. **Patients and methods:** We retrospectively reviewed 19 cases of patients treated at the Ohio State James Comprehensive Cancer Center with advanced sarcoma treated with VEGFR inhibition who also had next-generation sequencing of their tumors (via FoundationOne Heme panel). We evaluated TP53 as well as mutations that were observed in at least 20% of patients and evaluated its contribution to PFS using the Kaplan–Meier survival analysis of available radiology end points.

Results: Mutations that were observed in at least 20% of patients included TP53 and Rb1. Only TP53 was predictive of PFS in the context of VEGFR inhibition. The PFS of patients with TP53 mutations was significantly greater than TP53 wild-type tumors with the median PFS of 208 versus 136 days, respectively [P = 0.036, hazards ratio 0.38 (95% confidence interval 0.09–0.83)].

*Correspondence to: Dr James L. Chen, Department Biomedical Informatics/Division of Bioinformatics, The Ohio State University, 1800 Cannon Dr/250 Lincoln Tower/Columbus, OH 43210-1267, USA. Tel: +1-614-366-1525; E-mail: james.chen@osumc.edu

© The Author 2015. Published by Oxford University Press on behalf of the European Society for Medical Oncology. All rights reserved. For permissions, please email: journals.permissions@oup.com.

Conclusions: Mutations in TP53 may serve as a predictive biomarker of response to VEGFR inhibition in patients with advanced sarcoma. Larger, prospective studies are necessary to confirm these findings. **Key words:** sarcoma, TP53, VEGFR, pazopanib, next-generation sequencing

introduction

Bone and soft tissue sarcomas (STS) comprise a diverse group of rare tumors originating from the embryonic mesoderm. They account for 1% of all new adult malignancies diagnosed annually [1, 2]. The mainstay of curative therapy is surgical resection of localized disease with or without adjuvant chemoradiation. Overall survival has changed little for patients with advanced sarcomas in the past few decades, albeit with few exceptions such as in gastrointestinal stromal tumors. The median overall survival remains 12 months in the metastatic setting, and cytotoxic chemotherapeutic options have not significantly altered this landscape.

Pazopanib, an orally available multitargeted tyrosine kinase inhibitor, has recently been shown to improve progression-free survival (PFS) for patients with non-adipocytic STS in the metastatic setting. It has activity against vascular endothelial growth factor receptors (VEGFR) 1, 2, 3 and platelet-derived growth factor receptors. EORTC 62043 was a single-arm, phase II study that characterized the activity of pazopanib in the metastatic setting in four different strata: leiomyosarcomas, adipocytic STS, synovial sarcoma and other STS. In 142 patients, the predefined primary end point of progression-free rate of 40% of patients at 12 weeks was met in the leiomyosarcoma, synovial and other STS cohorts. The predefined PFR12 weeks was not met in the adipocytic cohort [3]. The PALETTE study was a randomized, phase III, multi-institutional trial designed to confirm the efficacy of pazopanib observed in EORTC 62043. In this trial, pazopanib 800 mg once daily was directly compared with placebo in non-adipocytic STSs. The patients in the pazopanib arm experienced an increase of 3 months in PFS versus patients on placebo, thus establishing pazopanib as a new treatment option for patients with advanced STS [4].

Given the modest, albeit significant, increase in PFS observed in the PALETTE trial, it would be ideal to identify patients who are likely to benefit from pazopanib before initiating therapy. Kasper et al. used the pooled data from EORTC 62043 (n = 118) and the PALETTE trial (n = 226) to compare patient characteristics among long-term survivors on pazopanib, defined as a PFS of ≥ 6 months. Based on this definition, 36% of patients were considered long-term survivors. They found that performance status, lowintermediate histology and normal baseline hemoglobin were advantageous for long-term outcome. Interestingly, 12 patients (3.5%) had a response to pazopanib for more than 2 years. These patients tended to be young, female and have low-intermediate histologies [5].

Recently, there has been a convergence of evidence to suggest that STSs with mutations in the TP53 gene may respond better to VEGFR inhibition than TP53 wild-type (WT). In the preclinical setting, Pollock et al. demonstrated that wild-type p53 suppresses angiogenesis by transcriptional suppression of vascular endothelial growth factor (VEGF) expression in human leiomyosarcoma and synovial sarcoma cell lines. As such, loss of function p53 mutant cells produced significantly more VEGF, which contribute directly to angiogenesis, metastasis and growth [6]. Farhang Ghahremani et al. subsequently found that the p53-VEGF pathway is dependent on a functional Rb1. They showed that p53 WT promotes VEGF expression and angiogenesis in the absence of an intact p21-Rb pathway in mouse embryonic fibroblasts exposed to hypoxic conditions [7].

Additionally, there is mounting clinical evidence that TP53 mutations may predict for response to VEGFR inhibition. A recent phase I study from MD Anderson demonstrated that VEGFR inhibition in combination with histone deacetylase (HDAC) inhibition appeared to be more effective in TP53 mutant versus TP53 WT tumors [8]. In their cohort of 78 patients with mostly metastatic colorectal or STS, they showed an increase in stable disease beyond 6 months for patients with mutant TP53 versus wild-type (45% versus 16%, P = 0.096), as well as trends toward increased PFS and OS (3.5 versus 2 months, P = 0.042; 12.7 versus 7.4 months, P = 0.1).

Based on these compelling preliminary data, we hypothesized that TP53 mutations resulting in loss of function in p53 would be a positive predictor of response to pazopanib as measured by PFS in advanced sarcoma patients.

materials and methods

We carried out a retrospective chart review of patients with advanced STS treated with pazopanib at the Ohio State Comprehensive Cancer Center (IRB 2014E0450). We stratified patients by TP53 and Rb1 mutational status based on next-generation sequencing (FoundationOne Heme/Sarcoma panel) and used PFS as our primary end point.

patients

From 1 January 2012 to 9 January 2014, we treated over 70 patients with advanced sarcomas with pazopanib. Of these, 19 patients had next-generation sequencing carried out through Foundation Medicine (sarcoma/heme panel) and were included in this retrospective review.

treatment

Eighteen patients were treated with oral pazopanib at the maximal tolerated doses. One patient was treated with sunitinib. Some patients had dose reductions of 400 mg daily based on tolerability. Eleven of 19 had been treated with prior doxorubicin-based therapy.

clinical assessment

Patients were evaluated for PFS based on imaging modality chosen by the treating physician (PET CT, CT or MRI). Responses were assessed using RECIST or PERCIST criteria, depending on the imaging modality. Time to progression was measured in days. One cycle of pazopanib was defined as 28 days. Patients were gauged for response after two cycles of drug and subsequently every two cycles thereafter. Patients who had to discontinue therapy due to drug toxicities were excluded from this analysis.

statistical analysis

Descriptive statistics were used to summarize the overall patient mutational statuses. The Kaplan–Meier survival estimates were generated using Graphpad Prism 6.0 by log-rank (Mantel Cox) test.

results

Patient characteristics are listed in Table 1 (male/female 8/11; mean age 49). Histologies included leiomyosarcoma (5), liposarcoma (3), mesenchymal chondrosarcoma (2), desmoblastic small round cell (1), myxofibrosarcoma (2), epithelioid (1), myxoid chondrosarcoma (2), synovial (1), angiosarcoma (1), undifferentiated pleomorphic sarcoma (1). Mutations seen in 10% or more of patients are summarized in Table 2. A total of 233 mutations were observed in this cohort of patients. TP53 mutations were the most common mutations present with 53% (10/19) of patients, predicting loss of function in p53. Thirty-two percent (6/19) patients had mutations in Rb1. All other mutations were present in <20% of cases.

All of the mutations in TP53 were predicted to be loss of function mutations by Foundation Medicine. The most common mutations were those within the DNA binding domain (DBD) (3/10 patients) or that affected the tetramerization domain (4/10 patients). One patient had a mutation predicted to affect both the DBD and tetramerization domain. Two patients had homozygous deletion resulting in complete loss of p53.

Foundation Medicine next-generation sequencing does not differentiate between germline and somatic mutations. No patients in this cohort had been diagnosed with LiFraumeni syndrome or concurrent malignancies, however, and all TP53 mutations observed were assumed to be somatic in origin.

There were no documented CRs to pazopanib in our 19 patients included in this analysis. The best response was a partial remission, which 1 of 11 responders to pazopanib patient achieved. Ten of the 11 responders otherwise had stable disease.

discussion

To our knowledge, this retrospective review is the first study to show that response to VEGFR inhibition with single-agent pazopanib can be predicted by TP53 mutational status in advanced non-adipocytic STS.

Preclinical studies have established the importance of the p53-VEGF pathway for angiogenesis in tumor biology. It has previously been shown that wild-type TP53 suppresses angiogenesis by transcriptional suppression of VEGF expression in human leiomyosarcoma and synovial sarcoma cell lines [6]. As such, cells with loss-of-function TP53 produce significantly more VEGF, thus directly contributing to the angiogenesis, metastases and growth of tumor cells. Moreover, mutations in TP53 have been shown to directly increase the level of hypoxia-inducible factor 1α (HIF- 1α) and augment HIF- 1α -dependent transcriptional activation of the VEGF gene in response to hypoxia [9, 10].

Additionally, the p53-VEGF pathway appears to be dependent on a functional Rb1, as increased levels of VEGF have been observed in Rb1-null mouse fibroblasts with a functional p53 [7]. The loss of function mutations in either TP53 or RB1 would therefore hypothetically directly contribute to the angiogenesis of the malignant cells, thus making tumors with mutations in either gene potentially sensitive to anti-angiogenic therapy.

There is mounting clinical evidence that demonstrates the importance of TP53 mutations in predicting response to antiangiogenic therapy. Recently, Fu et al. published the results of a phase I study of pazopanib in combination with the HDAC inhibitor, vorinostat, in advanced solid malignancies. Analysis of 78 unselected patients (23 sarcoma) with combination pazopanib and vorinostat did not lead to antitumor activity (5% PR and 14% SD \geq 6 months). However, when stratified by TP53 mutational status, patients with detected mutations tended to

Table 1. Patient characteristics						
Patient	Histology	Sites of metastases	TP53	RB1	Time to progression (days)	
1	LMS	Liver, abdominal lymph nodes	MUT	WT	334	
2	Angiosarcoma	Scalp lesion	MUT	WT	447	
3	Mesenchymal chondro	Intraabdominal mesenteric tumors	WT	WT	245	
4	LMS	Lungs, mediastinum	MUT	WT	249	
5	LPS	Lungs	WT	MUT	153	
6	Synovial	Lungs	WT	WT	112	
7	Desmoplastic small round cell	Intraabdominal lymph nodes, lungs	MUT	MUT	153	
8	LMS	Peritoneum, retroperitoneum	MUT	WT	133	
9	LMS	Left pelvic soft tissue, hilar lymph nodes	MUT	MUT	169	
10	Myxofibrosarcoma	Lungs	MUT	MUT	98	
11	Epithelioid	Lungs, supraclavicular LNs, acetabulum	WT	WT	111	
12	Osteosarcoma	Pelvic mass, lungs	WT	WT	85	
13	Small blue round cell	Sacrum, lungs	WT	WT	62	
14	Мухо	Vertebral lesions	MUT	MUT	92	
15	LMS	Lungs, mediastinum, liver	MUT	MUT	132	
16	Mesenchymal chondro	Cervical and thoracic spine metastases	WT	WT	162	
17	LPS	Liver, retroperitoneum	WT	WT	136	
18	LPS-DD	Retroperitoneum	WT	WT	163	
19	UPS	Right gluteal mass, lungs	MUT	WT	168	

Table 2. Frequency of mutations (n = patients)						
Mutations detected in >20% of patients	Mutations detected in >15% of patients	Mutations detected in >10% of patients				
TP53 (10), RB1 (6)	BCL6 (3), BLM (3), BRCA1 (3), CIC (3), MAP3K1 (3), NF1 (3), POT1 (3), WDR90 (3), WHSC1 (3)	APC (2), ARID2 (2), ASXL1 (2), ATRX (2), BRCA2 (2), CCT6B (2), CDK12 (2), CDKN2A (2), CHD2 (2), CHEK2 (2), CIITA (2), CPS1 (2), EP300 (2), EXOSC6 (2), FANCE (2), FRS2 (2), GRIN2A (2), HDAC7 (2), IRS2 (2), JAK1 (2), LRP1B (2), LRRK2 (2), MDM2 (2), MED12 (2), NOTCH1 (2), NTRK1 (2), PIK3R2 (2), RAD50 (2), SDHC (2), TET2 (2)				

respond more favorably with greater SD \geq 6 months (45% versus 16%, *P* = 0.096) [8]. A significantly longer median PFS (3.5 versus 2.0 months, *P* = 0.042) was also observed in the group with TP53 mutations, which supports our observation that patients with TP53 mutations are more likely to respond to VEGFR inhibition (Figure 1).

Of additional interest is whether the type of TP53 mutation relates to the sensitivity to VEGFR inhibition. All of the mutations in our small population were predicted to be loss of function. The two most common mutations in TP53 that result in loss of function are those involving the DBD or tetramerization domain. We did not observe a significant difference in PFS when stratifying between p53 mutations in the DBD versus tetramerization domain; however, our study is limited by small sample size and this should be an area of future clinical study.

The PALETTE study showed an improvement of 3 months in PFS in patients with advanced non-adipocytic STS versus placebo. Interestingly, in a subset analysis, patients with leiomyosarcoma were noted to have greater PFS on pazopanib versus other histologies (hazards ratio 0.88) [4]. We suspect that this may relate to the prevalence of TP53 mutations in this tumor group. According to the COSMIC database, 24% of LMS have mutations in TP53, making it the most common mutation in this histology (cancer.sanger.ac.uk, query date: 6 October 2015) [11]. Although liposarcomas were not included in the PALETTE trial as the adipocytic strata in EORTC 62043 did not demonstrate a clinical benefit in this group of patients, in the post-marketing survey, the PFS rate at 12 weeks was 26% (5 of 19 LPS patients) and higher than the threshold for study continuation [12]. In fact, pazopanib is approved in Japan and ongoing clinical trials are exploring these PALETTE ineligible cohorts [13]. Given these data, we included three adipocytic histologies in this retrospective review as our assumption was that response to treatment would be predicted by genetic alterations and not histology.

In this study, we were also curious as to whether the TP53 mutational status may be relevant to predicting response to doxorubicin, a common cytotoxic chemotherapy used in the treatment of advanced STSs. The mechanism of action of doxorubicin is likely highly dependent on a functional p53 since, as a topoisomerase II inhibitor, it prevents G1 to S phase. There are additionally data to suggest that tumors with TP53 mutations are less sensitive to doxorubicin. Aas et al. [14] have previously reported specific TP53 mutations associated with *de novo* resistance to doxorubicin in breast cancer patients. Therefore, we predicted that p53 WT tumors would respond better to doxorubicin. Of our 19 patients included in our analysis, 11 received doxorubicin previously. In this small patient sample, there was no observed significance



Figure 1. Progression-free survival of VEGFR inhibition stratified by TP53 mutation status. Ten patients with mutations in TP53 with longer progression-free survival than nine patients with no mutation in TP53.

difference in response to doxorubicin. Larger studies will be necessary to confirm these results.

In addition to the TP53 mutational status, we were also interested in the Rb1 status. Rb1 is relevant to the p53-VEGF pathway and was the second most common observed mutation in our cohort. Farhang Ghahremani et al. [7] have previously shown that p53 promotes VEGF expression and angiogenesis in the absence of an intact p21-Rb pathway. We therefore hypothesized that tumors with Rb1 mutations would also be sensitive to VEGFR inhibition. However, our clinical observation was contrary to this prediction. Rb1 mutants tended to do worse in the context of a p53 mutation and when patients were stratified into four groups: TP53 MUT/Rb1 WT (n = 6), TP53 MUT/Rb1 MUT (n = 4), TP53 WT/Rb1 MUT (n = 1) and TP53 WT/Rb1 WT (n = 8). However, there was no significant difference in PFS observed in our patient sample.

In considering the clinical relevance of our findings, several limitations should be borne in mind. First, the retrospective nature of the analysis limits the validity of our observations. Additionally, the small patient sample of 19 patients and the wide-range histologies we included make conclusions difficult to draw. Thirdly, this is a single-institutional study and all the patients were treated at the Ohio State University. Despite this, we observed a significant increase in the PFS of patients with advanced STS and TP53 mutations that is hypothesis-generating and supported by the emerging relevance of targeting the p53-VEGF axis as a therapeutic modality in cancer patients. Larger correlative studies should be carried out to confirm these results in the future.

funding

SARC CDA grant was awarded to Chen by SARC (no grant number).

disclosure

JLC has carried out advisory board work for Novartis. All remaining authors have declared no conflicts of interest.

references

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. CA Cancer J Clin 2015; 65 (1): 5–29.
- Clark MA, Fisher C, Judson I, Thomas JM. Soft-tissue sarcomas in adults. N Engl J Med 2005; 353(7): 701–711.
- Sleijfer S, Ray-Coquard I, Papai Z et al. Pazopanib, a multikinase angiogenesis inhibitor, in patients with relapsed or refractory advanced soft tissue sarcoma: a phase II study from the European organisation for research and treatment of cancer-soft tissue and bone sarcoma group (EORTC study 62043). J Clin Oncol 2009; 27(19): 3126–3132.
- van der Graaf WT, Blay Jy, Chawla SP et al. Pazopanib for metastatic soft-tissue sarcoma (PALETTE): a randomised, double-blind, placebo-controlled phase 3 trial. Lancet 2012; 379(9829): 1879–1886.
- 5. Kasper B, Sleijfer S, Litiere S et al. Long-term responders and survivors on pazopanib for advanced soft tissue sarcomas: subanalysis of two European

Organisation for Research and Treatment of Cancer (EORTC) clinical trials 62043 and 62072. Ann Oncol 2014; 25(3): 719–724.

- Zhang L, Yu D, Hu M et al. Wild-type p53 suppresses angiogenesis in human leiomyosarcoma and synovial sarcoma by transcriptional suppression of vascular endothelial growth factor expression. Cancer Res 2000; 60(13): 3655–3661.
- Farhang Ghahremani M, Goossens S, Nittner D et al. p53 promotes VEGF expression and angiogenesis in the absence of an intact p21-Rb pathway. Cell Death Differ 2013; 20(7): 888–897.
- 8. Fu S, Hou MM, Naing A et al. Phase I study of pazopanib and vorinostat: a therapeutic approach for inhibiting mutant p53-mediated angiogenesis and facilitating mutant p53 degradation. Ann Oncol 2015; 26(5): 1012–1018.
- Loges S, Mazzone M, Hohensinner P, Carmeliet P. Silencing or fueling metastasis with VEGF inhibitors: antiangiogenesis revisited. Cancer Cell 2009; 15(3): 167–170.
- Ravi R, Mookerjee B, Bhujwalla ZM et al. Regulation of tumor angiogenesis by p53-induced degradation of hypoxia-inducible factor 1alpha. Genes Dev 2000; 14 (1): 34–44.
- Forbes SA, Beare D, Gunasekaran P et al. COSMIC: exploring the world's knowledge of somatic mutations in human cancer. Nucleic Acids Res 2015; 43 (Database issue): D805–D811.
- Wilky BA, Meyer CF, Trent JC. Pazopanib in sarcomas: expanding the PALETTE. Curr Opin Oncol 2013; 25(4): 373–378.
- Kawai A. Post marketing surveillance in Japan for soft tissue sarcoma patients treated with pazopanib. Abstract 3438. In European Cancer Congress 2015. 2015. Vienna, Austria.
- Aas T, Borresen AL, Geisler S et al. Specific P53 mutations are associated with de novo resistance to doxorubicin in breast cancer patients. Nat Med 1996; 2(7): 811–814.