



Published in final edited form as:

J Transl Sci. 2016 ; 2(3): 154–159. doi:10.15761/JTS.1000133.

A Role for the Liver in Parturition and Preterm Birth

Anthony R. Mawson, MA, DrPH

Interim Chair, Department of Epidemiology & Biostatistics, School of Public Health, Jackson State University, 350 West Woodrow Wilson Avenue, Room 229, Jackson, MS 39213, 601-991-3811

Anthony R. Mawson: amawson@gmail.com

Abstract

Neither the mechanisms of parturition nor the pathogenesis of preterm birth are well understood. Poor nutritional status has been suspected as a major causal factor, since vitamin A concentrations are low in preterm infants. However, even large enteral doses of vitamin A from birth fail to increase plasma concentrations of vitamin A or improve outcomes in preterm and/or extremely low birthweight infants. These findings suggest an underlying impairment in the secretion of vitamin A from the liver, where about 80% of the vitamin is stored. Vitamin A accumulates in the liver and breast during pregnancy in preparation for lactation. While essential in low concentration for multiple biological functions, vitamin A in higher concentration can be pro-oxidant, mutagenic, teratogenic and cytotoxic, acting as a highly surface-active, membrane-seeking and destabilizing compound. Regarding the mechanism of parturition, it is conjectured that by nine months of gestation the hepatic accumulation of vitamin A (retinol) from the liver is such that mobilization and secretion are impaired to the point where stored vitamin A compounds in the form of retinyl esters and retinoic acid begin to spill or leak into the circulation, resulting in amniotic membrane destabilization and the initiation of parturition. If, however, the accumulation and spillage of stored retinoids reaches a critical threshold prior to nine months, e.g., due to cholestatic liver disease, which is common in mothers of preterm infants, the increased retinyl esters and/or retinoic acid rupture the fetal membranes, inducing preterm birth and its complications, including retinopathy, necrotizing enterocolitis and bronchopulmonary dysplasia. Subject to testing, the model suggests that measures taken prior to and during pregnancy to improve liver function could reduce the risk of adverse birth outcomes, including preterm birth.

Introduction

Every year about 15 million babies are born prematurely and over 1 million children die each year due to complications of preterm birth. Survivors of prematurity often face a lifetime of disability, including learning disabilities and visual and hearing problems; furthermore, rates of preterm birth are rising in almost all countries with reliable data.¹ Defined as a pregnancy ending at less than 37 completed weeks of gestation, preterm birth is the leading cause of infant mortality in the United States and accounts for one-third of deaths in children under one year of age, with an annual societal cost of over \$26 billion.^{2–4}

Conflict of Interest

The author has a US patent on a “Method for diagnosing gestational diabetes, preeclampsia, and fetal growth restriction.” US Patent Number 8,883,512 B1, November 11, 2014. <http://www.google.com/patents/US8883512>

The occurrence of preterm births rose steadily from 9.4% of all pregnancies in the United States in 1981, to 12.8% in 2006, before declining to 12.7% in 2007 and 12.3% in 2008. Most of the increase was attributed to increases in multiple gestations. In the United States, a pronounced and persistent disparity exists in the rate of preterm birth, with twofold or greater increases among women of African descent compared to other race/ethnic groups. Even after decades of basic science research and public health initiatives, this disparity remains relatively unchanged and unexplained.⁵

In high-income countries, 80% of stillbirths are delivered preterm, suggesting that preterm birth and stillbirth share common pathways and mechanisms.⁶ Of the 4 million perinatal deaths per year worldwide, up to 99% occur in developing countries,⁷ with the highest maternal, fetal, and neonatal mortality rates in sub-Saharan Africa and South Asia.⁸ Despite massive research effort, the causes and mechanisms of preterm birth and stillbirth remain uncertain and effective interventions are lacking.

Poor nutritional status is suspected of being a major cause of these problems, since vitamin A concentrations are consistently low in preterm infants.^{9,10} However, even large enteral doses of vitamin A from birth do not significantly increase plasma concentrations of vitamin A or improve outcomes in preterm or extremely low birthweight infants.^{11–13} This failure of supplementation to correct vitamin A concentrations suggests impaired hepatic mobilization and secretion of the carrier protein, retinol-binding protein (RBP); moreover, low retinol (vitamin A alcohol) concentrations do not necessarily indicate a state of vitamin A deficiency and can be associated with hypervitaminosis A.¹⁴

Retinoids

Vitamin A and its congeners (collectively known as retinoids) are mainly dietary-derived fat-soluble signaling molecules that are stored principally in the stellate cells of the liver, from which they are secreted in a regulated process and delivered to the target tissues in the form of retinol-binding protein (RBP). In normal physiological concentrations, retinoids are essential for numerous biological functions such as cellular homeostasis, embryonic development, vision, tissue differentiation, growth, and mucus secretion. In higher concentration, retinoids inhibit cell growth and can be pro-oxidant, cytotoxic, mutagenic, and teratogenic.^{15–19} Retinoic acid (RA), the most biologically active metabolite of retinol, is produced from free retinol in a process that involves: 1) hydrolysis of retinyl esters in the liver and the release of retinol into the circulation and its delivery to the target tissues bound to RBP; 2) oxidation of retinol to retinaldehyde via the action of an alcohol dehydrogenase; and 3) synthesis from retinaldehyde via an aldehyde dehydrogenase reaction, primarily in the cell microsomes. RA exerts its effects by binding to two types of nuclear protein receptors: the retinoic acid receptors (RARs) and retinoid X receptors (retinoids, RXRs), both of which exist as three distinct gene products (alpha, beta, and gamma). These receptors are members of the steroid/thyroid superfamily of ligand dependent nuclear transcription factors that include the receptors for steroids, thyroid hormone, and vitamin D. Following ligand activation, the receptors function as heterodimeric transcription factors and control the expression of numerous target genes by binding to specific DNA sequences termed RA response elements (RAREs).^{20,21}

Given that about 80% of vitamin A is stored in the liver, sudden shifts in these stores to other tissues due to infection, chronic illness or trauma can result in severe vitamin A poisoning. Indeed, an endogenous form of hypervitaminosis A associated with cholestatic liver disease is recognized, due to the spillage of stored retinoids into the circulation in bile.²² These observations point to the need to reevaluate the role of vitamin A in maternal and child health and in preterm birth in particular.

Retinoid Hypothesis of Parturition

Neither the mechanisms of preterm birth nor those of normal parturition are well understood. Since a common feature of term and preterm birth is rupture of the fetal membranes, understanding the process of membrane rupture could provide important clues for that of premature rupture of the membranes (PROM) in preterm birth. Preterm PROM (or PPRM) is a leading cause of preterm delivery, accounting for one third of spontaneous preterm births²³ and is strongly associated with adverse pregnancy outcomes.^{24,25} Membrane rupture is related to biochemical changes in collagen structure and formation as well as increased oxidative stress,²⁶ involving an imbalance between the synthesis and degradation of collagen within the extracellular matrix of the chorioamniotic membrane induced by matrix metalloproteinases.²⁷ Pregnancy itself is also associated with oxidative stress²⁸ and reduced antioxidant capacity.²⁹

It is proposed that human parturition is due in part to an accumulation of retinoids in the liver in the course of pregnancy, resulting in mild inflammatory changes and the progressive spillage of stored retinoid compounds into the circulation. At around nine months of gestation these compounds have accumulated to the point where they rupture the fetal membranes and initiate parturition. It is hypothesized that early disturbances in liver function and retinoid metabolism, resulting in exposure of the fetus to excess retinoid concentrations, are critically involved in preterm birth and other adverse birth outcomes. Vitamin A can be either antioxidant or pro-oxidant, depending on dosage and cellular condition. At both excessive and therapeutic doses, acute and chronic vitamin A supplementation to laboratory animals leads to increased levels of markers of oxidative stress in liver mitochondria and in the substantia nigra and to alterations in locomotor and exploratory activity.^{18,19}

Retinoids are essential for the growth and development of the mammalian fetus and placenta;^{30,31} for instance, retinoic acid receptor (RAR)-null mutant mice die in utero or shortly after birth and exhibit congenital abnormalities.³² RARs and retinoid X receptors (RXRs) also show specific spatiotemporal patterns of expression in all developing systems during embryonic development.³³ Indeed, the role of retinoic acid signaling in the genetic regulation of morphogenesis and pattern formation (fetal embryonic development) is well established.³⁴ The associated amniotic membranes are extra-embryonic structures, indispensable for normal gestation in mammals. The existence of metabolic and molecular pathways of retinoic acid signaling in human fetal membranes is shown by evidence of retinoid receptor (RAR α , β , γ and RXR α , β , γ) expression at transcript and protein levels and by enzyme activity involved in the production of retinoic acid.³¹

Natural History of Pregnancy

Retinol in maternal serum declines during pregnancy, increases at parturition and decreases again with lactation.³⁵ Retinoids also accumulate in the liver during gestation³⁴ and in breast tissue³³ in preparation for breastfeeding. It is conjectured that the accumulation of vitamin A in the liver during pregnancy induces a mild form of inflammation and cholestasis, resulting in impaired secretion of RBP and the spillage of active retinoid compounds into the circulation in bile, compounds that are normally be excreted via the duodenum. Retinyl esters also leak into the circulation from damaged hepatocytes. The overall result is an endogenous form of vitamin A intoxication associated with an increased percentage of plasma retinyl esters as a fraction of total vitamin A, as well as increased retinoic acid (RA) concentrations. Percent retinyl esters > 10% of total vitamin A (retinol plus esters) is an accepted diagnostic criterion of hypervitaminosis A.¹² Liver dysfunction can be precipitated by high concentrations of circulating and stored retinoids and is known to contribute to an endogenous form of vitamin A toxicity through the spillage of retinyl esters and acidic biliary metabolites of vitamin A into the circulation.^{35,36} Bile drains from the liver through the gall bladder and common bile duct into the duodenum. In mice, bile contains significantly higher concentrations of retinol (about four times higher) than found in serum and other extrahepatic tissues.³⁷

It is postulated that retinoids which have accumulated in the liver during the course of pregnancy begin to spill into the circulation as a result of mild hepatic inflammation and cholestasis. As the concentrations of these retinoids steadily rise they begin to weaken and eventually rupture the fetal membranes as a function of their membrane-destabilizing effect, thereby triggering the process of parturition at around nine months of gestation. Although there are no apparent morphologic changes in the liver during normal human pregnancy, functional alterations occur suggesting the presence of inflammatory changes, including a doubling of serum alkaline phosphatase activity as well as increases in serum concentrations of certain bile acids. These observations led to earlier suggestions that pregnancy is associated with sub-clinical cholestasis.³⁸ In mice, dramatic changes occur in the size of the liver during pregnancy, whereby livers double in weight from the non-pregnant state to day 18 of pregnancy. Growth of the fetal mouse liver begins following implantation and peaks at parturition.³⁹

With regard to human pregnancy, although little is known about circulating levels of RA in the human fetus and newborn, it is suggested that upon reaching a critical concentration threshold, presumably around nine months of gestation, RA begins to lyse the fetal membranes and induce parturition. Retinol is a highly surface-active, membrane-seeking and destabilizing compound. When unbound to protein, it is known to cause the degradation of the extracellular matrix of chick limb-bone rudiments via effects on lysosomal membranes and the release of lysosomal enzymes.^{40,41} Vitamin A can also cause hemolysis of erythrocytes and result in increased permeability and fluidity.⁴²

The proposed model is supported by a small study of retinol and retinoic acid during pregnancy, showing that retinol concentrations were unrelated to those of the active derivative, all-trans retinoic acid; serum retinol was lower and essentially constant in

pregnant women and parturient mothers compared to that in non-pregnant control women. In contrast, all-trans-RA concentrations were higher in the parturient mothers than in control subjects, increasing in the third and fourth month of pregnancy to reach a steady state at about 40% above the initial concentration; moreover, when all-trans-RA concentrations were corrected for changes in binding protein concentrations, i.e., albumin, about a twofold increase occurred in all-trans-RA in the second and third trimesters.⁴³

Role of Retinoids in Fetal Membranes

The fetal membranes comprise the inner amnion lining of the amniotic cavity and the outer chorion underlying the uterine lining (endometrium). The amnion comprises a single layer of epithelial cells surrounded by connective tissue stroma. These membranes are fundamental for normal gestation in mammals. Amniotic fluid (AF) is produced by the fetus throughout pregnancy, averaging from 500 to 700 ml/day. The fetal membranes are involved in the regulation of AF volume and transfer;⁴⁴ this requires aquaporins (AQPs), a family of 13 glycoproteins, which facilitate water flux across cellular membranes. The factors thought to modulate AQP expression in placental and fetal membranes include some members of the nuclear receptor superfamily, e.g., the retinoic acid receptors (RAR α , β , γ). RA is reported to regulate AQP1, 3, 5 and 9 in other cellular and tissue environments,⁴⁵ and the RAR signaling pathway regulates certain target genes in human amniotic membranes,⁴⁶ but its role in the fetal membranes has remained uncertain. To answer the question of whether RA regulates AQPs in the fetal membranes, Sapin and colleagues cultured explants and primary and established amniotic cells to determine which AQPs were transcriptionally modified by all-trans-RA (the most abundant natural form of retinoic acid). Using Immunohistochemistry and other methods to determine the impact of all-trans-RA on AQP protein expression and function, they showed that the specific transactivation of a single AQP (that is, AQP3) in the amniotic environment involves RAR α . Only RAR α (and not RAR γ) was able to transduce all-trans-RA signaling on the AQP3 coding gene. These results suggested that upregulation of AQP3 by all-trans-RA participates in maintaining amniotic fluid homeostasis across human fetal membranes. Consistent with the present hypothesis, Sapin and colleagues also concluded that retinoic acid signal dysregulation in fetal membranes could be involved in adverse obstetric outcomes.⁴⁷

Role of Liver Dysfunction and Retinoid Metabolism in Preterm Birth

A healthy liver is essential for reproduction in women. Pregnancy in patients with advanced liver disease is uncommon since most women with cirrhosis are infertile and have high rates of anovulation; gestation, if it occurs, carries high risks to both mother and fetus such as increased rates of spontaneous abortion, prematurity, pulmonary hypertension and postpartum hemorrhage. Conversely, after liver transplantation menstruation resumes and most women are able to conceive and give birth,⁴⁸ although the complication rate is high.⁴⁹ Here it is suggested that preterm birth and related adverse birth outcomes occur due to liver dysfunction, when the concentration of retinoids spilled from the liver exceed a critical threshold.

A central hypothesis of this paper is that liver dysfunction and associated alterations in retinoid metabolism – involving the spillage of stored retinoids into the maternal and fetal circulations in bile, together with the leakage of retinyl esters from damaged hepatocytes – are importantly involved in the pathogenesis of pregnancy complications and adverse birth outcomes. It is proposed that liver dysfunction and endogenous changes in vitamin A metabolism induce a spectrum of disorders, including early pregnancy loss, birth defects, preterm birth, stillbirth, and fetal growth restriction, depending on the concentration of these compounds and the timing of exposure of the fetus to retinoids. Maternal liver dysfunction itself may be due in part to preexisting conditions such as diabetes, obesity or hypertension, or develop in pregnancy associated with gestational diabetes and/or preeclampsia, due to hyperestrogenemia, which can lead to cholestatic liver dysfunction.⁵⁰

Membrane Rupture and Parturition

With regard to the mechanism of preterm birth, it is proposed that cholestatic liver dysfunction and associated increases in circulating concentrations of retinyl esters and/or retinoic acids rupture the fetal membranes, inducing preterm birth and the characteristic features of the preterm infant, including retinopathy, necrotizing enterocolitis, and bronchopulmonary dysplasia. The process leading to preterm birth is conjectured to begin with the accumulation of excess retinoids in the liver in early pregnancy due to hyperestrogenemia, following a signal from the conceptus. Pregnancy increases the sensitivity of the bile ducts to estrogen, and cholestasis often develops during the second and third trimesters of pregnancy.⁵¹ The presence of liver dysfunction is increasingly evident in the progression of preeclampsia to more severe pregnancy-related disorders such as HELLP syndrome (Hemolysis, ELevated liver enzymes, and Low Platelet count). Abnormal liver tests are reported in 10% of pregnancies overall, suggesting reduced liver metabolic capacity in late pregnancy,⁵² but conventional liver enzyme tests underestimate the true extent of liver dysfunction.⁵³

The association between liver dysfunction and preterm birth is well documented. For instance, women diagnosed with alcoholic liver disease before birth have an increased risk of preterm birth and of delivering infants small-for-gestational age.⁵⁴ Total bile acids are also associated with preterm delivery.⁵⁵ After the 12th week of pregnancy, phospholipids, cholesterol and triglycerides increase in response to estrogen stimulation and insulin resistance. Although maternal hypertriglyceridemia appears to have some positive effects, it also increases risk of preeclampsia and preterm birth and may have a role in increasing cardiovascular risk later in life.⁵⁶ Consistent with the retinoid hypothesis, hypertriglyceridemia is associated with hypervitaminosis A⁵⁷ and is a frequent complication in the therapeutic use of retinoic and acid retinoid derivatives.⁵⁸ In a study of women with preeclampsia, those who delivered preterm had significantly greater liver enzyme concentrations than those who delivered at term.⁵⁹ Menon et al.⁶⁰ studied metabolic changes associated with early spontaneous preterm birth (<34 weeks) using high-throughput metabolomics of amniotic fluid (AF) retrieved by transvaginal amniocentesis at the time of labor prior to delivery in an African American population that included a control group of women who delivered at term. They found evidence of altered liver function in many cytochrome P450-related pathways including bile acids, steroids, xanthines, heme, and

phase II detoxification of xenobiotics. The largest fold change was in pantothenol, a CoA synthesis inhibitor that was 8-fold more abundant in preterm birth samples. Several bile acids and their metabolites were elevated in preterm birth samples, inversely related to gestational age, including glycocholate, taurocholate, taurochenodeoxycholate, taurodeoxycholate, and glycodeoxycholate. High circulating levels are known to cause fetal stress. Bile acid metabolism, secretion from liver, and liver reuptake can be disrupted by hormonal changes associated with cholestasis in pregnancy.⁶¹ The hypothesis that alterations in liver function and retinoid metabolism are associated with complications of pregnancy and adverse birth outcomes was supported by the results of an animal study carried out by the author and colleagues. The drug streptozotocin was administered to rats early in pregnancy to create an animal model of gestational diabetes and preeclampsia.⁶² Pup weights of the experimental rats were significantly lower than those of controls ($\mu = 4.6$ g vs. 1.4 g, $p < 0.05$), indicating intrauterine growth restriction. Retinoid profiling indicated that retinol concentrations in the experimental dams were significantly lower than in controls. On the other hand, consistent with the model, the percentage of retinyl esters $>10\%$ (an indicator of retinoid toxicity) was significantly increased (median, 24% vs. 11%; $p = 0.008$), as was the concentration of retinoic acid (median, 0.0155 vs. 0.0075 $\mu\text{mol/L}$; $p = 0.045$). Liver enzyme levels were also significantly elevated.

Complications of Preterm Birth

The proposal that preterm birth is associated with an endogenous form of hypervitaminosis A suggests that the characteristic and usually combined complications of prematurity, e.g., retinopathy, necrotizing enterocolitis and bronchopulmonary dysplasia, are themselves due to hypervitaminosis A. In the case of retinopathy of prematurity, both prenatal and postnatal exposure to isotretinoin (13-cis-retinoic acid, a synthetic vitamin A derivative used in the treatment of severe acne) is known to cause retinopathy and optic nerve abnormalities.⁶³ Retinoic acid also contributes to light-induced retinopathy in mice via mechanisms that may include plasma membrane permeability and mitochondrial poisoning, leading to caspase activation and mitochondria-associated cell death.⁶⁴ Necrotizing enterocolitis (NEC), an inflammatory intestinal disorder primarily seen in premature infants, is characterized by variable damage to the intestinal tract ranging from mucosal injury to full-thickness necrosis and perforation.⁶⁵ Although no direct link between retinoid exposure and NEC has been described, adverse side effects attributed to isotretinoin include inflammatory bowel disease as well as depression, suicidality, and teratogenicity.⁶⁶

Bronchopulmonary dysplasia (BPD) of the preterm neonate is similar to emphysema of the adult lung in that both are characterized by increased airspace and respiratory insufficiency. They are also strikingly similar in their pathophysiology, including the precipitating effect of oxidative stress, chronic inflammation, increased apoptosis, protease-antiprotease imbalance, elastic fiber deterioration and altered microvascularization.⁶⁷ In both diseases there is evidence of low vitamin A concentrations, which have been hypothesized to indicate vitamin A deficiency. However, it has been suggested here that while vitamin A (retinol) concentrations in preterm neonates are low, due to cholestatic liver dysfunction and impaired hepatic mobilization, other vitamin A metabolites are spilled or leaked into the maternal and fetal circulation and induce the characteristic features of the preterm neonate, including

bronchopulmonary dysplasia. In support of this hypothesis, maternal vitamin A supplementation during pregnancy and lactation increases oxidative stress parameters in rat neonatal lungs, effects that may be involved in conditions associated with redox dysfunctions and free radical-induced cell damage.⁶⁸ Oxidative stress is caused by an imbalance between the production of free radicals and the ability of antioxidant system to detoxify them. These conditions are known as oxygen radical diseases of neonatology and include bronchopulmonary dysplasia, retinopathy, and necrotizing enterocolitis.⁶⁹

Thrombocytopenia and anemia are also commonly seen in the preterm infant; for instance, thrombocytopenia occurs in up to a third of preterm neonates admitted to intensive care units.⁷⁰ Anemia (hemoglobin < 10.5 g/dL) is likewise strongly associated with intrauterine growth restriction and prematurity.⁷¹ Consistent with the model, hypervitaminosis A in infants causes severe thrombocytopenia and anemia, which may be due to a direct inhibitory effect on the growth of all bone marrow cellular components, mediated by the upregulation of cyclin-dependent kinase inhibitors.⁷²

Conclusion

In 1953 Peter Medawar raised the question: “How does the pregnant mother contrive to nourish within itself, for many weeks or months, a fetus that is an antigenically foreign body?”⁷³ In other words, why does the mother fail to reject her fetus as a foreign piece of tissue? The suggestion offered here is that the mother does not reject, or rather eject the fetus, normally at around 37 weeks of gestation, as the result of a biochemical rather than an immunological mechanism. Parturition at term is hypothesized to represent an active process of “rejection of the fetus” associated with the hepatic accumulation and spillage of stored retinoids due to mild liver dysfunction, which progressively weaken the fetal membranes until they rupture at around the ninth month of gestation. Based on this template for parturition at term, it has been proposed that spontaneous preterm birth is similarly due to more severe liver dysfunction and retinoid concentrations that exceed the threshold for inducing rupture of the fetal membranes before 37 weeks of gestation.

The model could be tested by determining whether 1) preterm newborns have significantly lower serum retinol (ROL) concentrations than term newborns, but higher concentrations of retinoic acid (RA), a higher percent retinyl esters (RE), and a higher RE:ROL ratio; 2) mothers of preterm newborns have higher liver enzyme levels than mothers of term neonates; and 3) an increased percentage of REs as a fraction of total serum retinol (> 10%) is a strong predictor of preterm birth. Subject to further testing in prospective studies, periodic monitoring of changes in maternal retinoid profiles throughout pregnancy could provide a biomarker for the early identification of impending adverse birth outcomes, which could be useful in prevention. Approaches to the management of pregnancies resulting in preterm birth and its complications suggested by the retinoid toxicity hypothesis could involve the adoption of measures taken prior to and after conception to improve liver function through dietary and lifestyle changes.

References

1. March of Dimes. Low Birthweight. 2013. Retrieved (February 12, 2013) from https://www.marchofdimes.com/professionals/25079_1153.asp
2. Behrman, RE.; Stith Butler, A., editors. Institute of Medicine. Societal Costs of Preterm Birth. The National Academies Press; 2007. p. 396-429.
3. Klebanoff MA, Keim SA. Epidemiology: the changing face of preterm birth. *Clin Perinatol*. 2011; 38:339–350. [PubMed: 21890013]
4. Blencowe H, Cousens S, Chou D, Oestergaard M, Say L, Moller A-B, Kinney M, Lawn J. on behalf of the Born Too Soon Preterm Birth Action Group. Born Too Soon: The global epidemiology of 15 million preterm births. *Reproductive Health*. 2013; 10(Suppl 1):S2. <http://www.reproductive-health-journal.com/content/10/S1/S2>. [PubMed: 24625129]
5. Culhane JF, Goldenberg RL. Racial disparities in preterm birth. *Semin Perinatol*. 2011; 35(4):234–9. [PubMed: 21798403]
6. Flenady V, Middleton P, Smith GC, Duck W, Erwich JJ, Khong TY, et al. Stillbirths: the way forward in high-income countries. *Lancet*. 2011; 377:1703–17. [PubMed: 21496907]
7. Lawn JE, Cousens SM, Zupan J. 4 million neonatal deaths: When? Where? Why? *Lancet*. 2005; 365(9462):891–900. [PubMed: 15752534]
8. Bhutta ZA, Yakoob MY, Lawn JE, Rizivi A, Fridberg IK, Weissmna E, et al. Stillbirths: what difference can we make and at what costs? *Lancet*. 2011; 377:1523–38. [PubMed: 21496906]
9. Mactier H, Weaver LT. Vitamin A and preterm infants: what we know, what we don't know, and what we need to know. *Arch Dis Child Fetal Neonatal Ed*. 2005; 90(2):F103–8. [PubMed: 15724031]
10. Abdel Ghany EA, Alsharany W, Ali AA, Younass ER, Hussein JS. Anti-oxidant profiles and markers of oxidative stress in preterm neonates. *Paediatr Int Child Health*. 2015 May 4. 2046905515Y0000000017.
11. Kennedy KA, Stoll BJ, Ehrenkranz RA, et al. Vitamin A to prevent bronchopulmonary dysplasia in very-low-birth-weight infants: has the dose been too low? *Early Hum Dev*. 1997; 49:19–31. [PubMed: 9179535]
12. Wardle SP, Hughes A, Chen S, et al. Randomised controlled trial of oral vitamin A supplementation in preterm infants to prevent chronic lung disease. *Arch Dis Child Fetal Neonatal Ed*. 2001; 84:F9–13. [PubMed: 11124916]
13. McCauley ME, van den Broek, Dou L, Othman M. Vitamin A supplementation during pregnancy for maternal and newborn outcomes. *Cochrane Database Syst Rev*. 2015 Oct 27.10:CD008666.doi: 10.1002/14651858.CD008666.pub3 [PubMed: 26503498]
14. Penniston KL, Tanumihardjo SA. The acute and chronic toxic effects of vitamin A. *Am J Clin Nutr*. 2006; 83:191–201. [PubMed: 16469975]
15. Ganguly, J. *The Biochemistry of Vitamin A*. Boca Raton, FL: CRC Press; 1989.
16. Hoffman, C.; Eichele, G. Retinoids in development. In: Sporn, MB.; Roberts, AB.; Goodman, DS., editors. *The Retinoids: Biology, Chemistry, and Medicine*. New York: Raven Press; 1994. p. 387-441.
17. Litwack, G., editor. *Vitamin A: Vitamins and Hormones*. Vol. 75. San Diego, CA: Elsevier Academic Press; 2007.
18. de Oliveira MR, Silvestrin RB, Mello e Souza T, Moreira JCF. Therapeutic vitamin A doses increase the levels of markers of oxidative insult in substantia nigra and decrease locomotory and exploratory activity in rats after acute and chronic supplementation. *Neurochem Res*. 2008; 33(3): 378–83. [PubMed: 17712631]
19. de Oliveira MR. The neurotoxic effects of vitamin A and retinoids. *An Acad Bras Cienc*. 2015; 87(2 Suppl):1361–73. Epub 2015 Aug 4. DOI: 10.1590/0001-3765201520140677 [PubMed: 26247148]
20. Theodosiou M, Laudet V, Schubert M. From carrot to clinic: an overview of the retinoic acid signaling pathway. *Cell Mol Life Sci*. 2010; 67:1423–45 19. [PubMed: 20140749]

21. Brun PJ, Yang KJ, Lee SA, Yuen JJ, Blaner WS. Retinoids: Potent regulators of metabolism. *Biofactors*. 2013 Mar-Apr;39(2):151–63. Epub 2012 Dec 22. DOI: 10.1002/biof.1056 [PubMed: 23281051]
22. Leo MA, Lieber CS. Alcohol, vitamin A, and beta-carotene: adverse interactions, including hepatotoxicity and carcinogenicity. *Am J Clin Nutr*. 1999; 69(6):1071–85. [PubMed: 10357725]
23. Goldenberg RL, Culhane JF, Iams JD, Romero R. Epidemiology and causes of preterm birth. *Lancet*. 2008; 371(9606):75–84. [PubMed: 18177778]
24. McParland, PC.; Taylor, DJ. Preterm prelabour rupture of the membranes. In: Bonnar, J.; Dunlop, W., editors. *Recent Advances in Obstetrics and Gynecology*. Vol. 23. 2005. p. 27-38.
25. Menon R. Spontaneous preterm birth. A clinical dilemma: etiologic, pathophysiologic and genetic heterogeneities and racial disparities. *Acta Obstet Gynaecol Scand*. 2008; 87(6):590–600.
26. Kundtson EJ, Smith K, Mercer BM, Miodovnik M, Thurnau GA, et al. Serum homocysteine levels after preterm premature rupture of the membranes. *Am J Obstet Gynecol*. 2004; 191(1):537–41. [PubMed: 15343233]
27. Fortunato SJ, Menon R. Distinct molecular events suggest different pathways for preterm labor and premature rupture of membranes. *Am J Obstet Gynecol*. 2001; 184:1399–405. [PubMed: 11408859]
28. Mathews F, Neil A. Antioxidants and preterm prelabor rupture of the membranes. *BJOG*. 2005; 112(5):588–594. [PubMed: 15842282]
29. Idogun ES, Odiegwu ME, Momoh SM, Okonofua FE. Effects of pregnancy on total antioxidant capacity in Nigerian women. *Pak J Med Sci*. 2008; 24(2):292–295.
30. Wilson JG, Roth CB, Warkany J. An analysis of the syndrome of malformations induced by maternal A deficiency: effects of restoration of vitamin A at various times in gestation. *Am J Anat*. 1953; 92:189–217. [PubMed: 13030424]
31. Marceau G, Gallot D, Borel V, Lémery D, Dastugue B, Dechelotte P, Sapin V. Molecular and metabolic retinoid pathways in human amniotic membranes. *Biochem Biophys Res Commun*. 2006; 346(4):1207–16. [PubMed: 16793012]
32. Lohnes D, Mark M, Mendelshon C, Dollé P, Dierich A, Gorry P, Gansmuller A, Chambon P. Function of the retinoic acid receptors (RARs) during development. (I) Craniofacial and skeletal abnormalities in RAR double mutants. *Development*. 1994; 120:2723–2748. [PubMed: 7607067]
33. Morriss-Kay GM, Sokolova N. Embryonic development and pattern formation. *FASEB J*. 1996; 10:961–968. [PubMed: 8801178]
34. Maden M. Retinoic acid in the development, regeneration and maintenance of the nervous system. *Nat Rev Neurosci*. 2007; 8:755–65. [PubMed: 17882253]
35. Bates CJ. Vitamin A in pregnancy and lactation. *Proc Nutr Soc*. 1983; 42:65–79. [PubMed: 6340124]
34. Satre MA, Ugen KE, Kochar DM. Developmental changes in endogenous retinoids during pregnancy and embryogenesis in the mouse. *Biol Reprod*. 1992; 46:802–810. [PubMed: 1591336]
35. Leo MA, Lieber CS. New pathway for retinol metabolism in liver microsomes. *J Biol Chem*. 1985; 260(9):5228–31. [PubMed: 3988751]
36. Shirakami Y, Lee SA, Clugston RD, Blaner WS. Hepatic metabolism of retinoids and disease associations. *Biochim Biophys Acta*. 2012; 1821(1):124–36. [PubMed: 21763780]
37. Jaensson-Gyllenbäck E, Kotarsky K, Zapata F, Persson EK, Gundersen TE, Blomhoff R, Agace WW. Bile retinoids imprint intestinal CD103+ dendritic cells with the ability to generate gut-tropic T cells. *Mucosal Immunol*. 2011; 4(4):438–47. DOI: 10.1038/mi.2010.91 [PubMed: 21289617]
38. Carter J. Serum bile acids in normal pregnancy. *Br J Obstet Gynaecol*. 1991; 98(6):540–543. [PubMed: 1873244]
39. Dai G, Bustamante JJ, Zou Y, Myronovych A, Bao Q, Kumar S, Soares MJ. Maternal hepatic growth response to pregnancy in the mouse. *Exp Biol Med (Maywood)*. 2011; 236(11):1322–32. [PubMed: 21969712]
40. Dingle JT, Fell HB, Goodman DS. The effect of retinol and of retinol-binding protein on embryonic skeletal tissue in organ culture. *J Cell Sci*. 1972; 11(2):393–402. [PubMed: 4263171]

41. Dingle JT, Lucy JA. Studies on the mode of action of excess vitamin A. The effect of vitamin A on the stability of the erythrocyte membrane. *Biochem J.* 1962; 84:611–621. [PubMed: 13886480]
42. Urano S, Inomori Y, Sugawara T, Kato Y, Kitahara M, Hasegawa Y, Matsuo M, Mukai K. Vitamin E inhibition of retinol-induced hemolysis and membrane-stabilizing behavior. *J Biol Chem.* 1992; 267(26):18365–70. [PubMed: 1526978]
43. Söderlund MB, Fex GA, Nilsson-Ehle P. Concentrations of retinoids in early pregnancy and in newborns and their mothers. *Am J Clin Nutr.* 2005; 81(3):633–6. [PubMed: 15755833]
44. Beall MH, van den Wijngaard JP, van Gemert MJ, Ross MG. Regulation of amniotic fluid volume. *Placenta.* 2007; 28(8–9):824–32. Epub 2007 Feb 15. [PubMed: 17303237]
45. Gao RW, Kong XY, Zhu XX, Zhu GQ, Ma JS, Liu XX. Retinoic acid promotes primary fetal alveolar epithelial type II cell proliferation and differentiation to alveolar epithelial type I cells. *In Vitro Cell Dev Biol Anim.* 2015; 51(5):479–87. Epub 2014 Dec 17. DOI: 10.1007/s11626-014-9850-2 [PubMed: 25515249]
46. Blanchon L, Marceau G, Borel V, Prat C, Herbet A, Bouvier D, Gallot D, Sapin V. Implications of retinoid pathway in human fetal membranes: study of target genes. *Gynecol Obstet Fertil.* 2011; 39(6):370–2. [PubMed: 21596610]
47. Prat C, Bouvier D, Comptour A, Marceau G, Belville C, Clairefond G, Blanc P, Gallot D, Blanchon L, Sapin V. All-trans-retinoic acid regulates aquaporin-3 expression and related cellular membrane permeability in the human amniotic environment. *Placenta.* 2015; 36(8):881–7. Epub 2015 May 19. DOI: 10.1016/j.placenta.2015.05.010 [PubMed: 26045060]
48. Hammoud GM, Almashhrawi AA, Ahmed KT, Rahman R, Ibdah JA. Liver diseases in pregnancy: liver transplantation in pregnancy. *World J Gastroenterol.* 2013; 19(43):7647–51. DOI: 10.3748/wjg.v19.i43.7647 [PubMed: 24282354]
49. Ludvigsson JF, Bergquist A, Ajne G, Kane S, Ekbohm A, Stephansson O. A population-based cohort study of pregnancy outcomes among women with primary sclerosing cholangitis. *Clin Gastroenterol Hepatol.* 2014; 12(1):95. [PubMed: 23891928]
50. Forde, K.; Kaplan, DE. Cholestatic liver disease related to systemic disorders. In: Lindor, KD.; Talwalkar, JA., editors. *Cholestatic Liver Disease.* Humana Press; 2008. p. 135-153.
51. Arrese M, Macias RI, Briz O, Perez MJ, Marin JJ. Molecular pathogenesis of intrahepatic cholestasis of pregnancy. *Expert Rev Mol Med.* 2008; 10:e9. [PubMed: 18371245]
52. Everson GT. Liver problems in pregnancy: distinguishing normal from abnormal hepatic changes. *Medscape General Medicine.* 1999; 1(3)
53. Stefan N, Kantartzis K, Häring HU. Causes and metabolic consequences of Fatty liver. *Endocr Rev.* 2008; 29(7):939–60. DOI: 10.1210/er.2008-0009 [PubMed: 18723451]
54. Stokkeland K, Ebrahim F, Hultcrantz R, Ekbohm A, Stephansson O. Mothers with alcoholic liver disease and the risk for preterm and small-for-gestational-age birth. *Alcohol Alcohol.* 2013; 48(2): 166–71. DOI: 10.1093/alcal/ags122 [PubMed: 23161891]
55. Pata O, Vardareli E, Ozcan A, Serteser M, Unsal I, Saruç M, Unlü C, Tözün N. Intrahepatic cholestasis of pregnancy: correlation of preterm delivery with bile acids. *Turk J Gastroenterol.* 2011; 22(6):602–5. [PubMed: 22287405]
56. Ghio A, Bertolotto A, Resi V, Volpe L, Di Cianni G. Triglyceride metabolism in pregnancy. *Adv Clin Chem.* 2011; 55:133–53. [PubMed: 22126027]
57. Ellis JK, Russell RM, Makrauer FL, Schaefer EJ. Increased risk for vitamin A toxicity in severe hypertriglyceridemia. *Ann Intern Med.* 1986; 105(6):877–9. [PubMed: 3777711]
58. Lilley JS1, Linton MF, Fazio S. Oral retinoids and plasma lipids. *Dermatol Ther.* 2013; 26(5):404–10. DOI: 10.1111/dth.12085 [PubMed: 24099071]
59. Phillips JK, Janowiak M, Badger GJ, Bernstein IM. Evidence for distinct preterm and term phenotypes of preeclampsia. *J Matern Fetal Neonatal Med.* 2010; 23(7):622–6. DOI: 10.3109/14767050903258746 [PubMed: 20482241]
60. Menon R, Jones J, Gunst PR, Kacerovsky M, Fortunato SJ, Saade GR, Basraon S. Amniotic fluid metabolomic analysis in spontaneous preterm birth. *Reprod Sci.* 2014; 21(6):791–803. Epub 2014 Jan 18. DOI: 10.1177/1933719113518987 [PubMed: 24440995]
61. Pathak B, Shebani L, Lee RH. Cholestasis of pregnancy. *Obstet Gynecol Clin North Am.* 2010; 37(2):269–282. [PubMed: 20685553]

62. Mawson, AR.; Bennett, W.; Tanumihardjo, S.; Mills, J.; Smith, A.; Kirui, P.; May, W.; Martin, JN, Jr. Altered retinoid profiles in an animal model of preeclampsia, gestational diabetes and fetal growth restriction. Presented at the 57th Annual Meeting of the Society for Gynecological Investigation; Orlando, FL. March 26, 2010; Reproductive Science 2010 Abstract 592. Vol. 17 (Supplement)
63. Lammer EJ, Chen DT, Hoar RM, Agnish ND, Benke PJ, Braun JT, Curry CJ, Fernhoff PM, Grix AW Jr, Lott IT, et al. Retinoic acid embryopathy. *N Engl J Med.* 1985; 313:837–41. [PubMed: 3162101]
64. Maeda A, Maeda T, Golczak M, Chou S, Desai A, Hoppel CL, Matsuyama S, Palczewski K. Involvement of all-trans-retinal in acute light-induced retinopathy of mice. *J Biol Chem.* 2009; 284:15173–83. [PubMed: 19304658]
65. Zani A, Pierro A. Necrotizing enterocolitis: controversies and challenges. *F1000Research.* 2015; 4:F1000. Faculty Rev-1373. doi: 10.12688/f1000research.6888.1 [PubMed: 26918125]
66. Prevost N, English JC. Isotretinoin: update on controversial issues. *J Pediatr Adolesc Gynecol.* 2013; 26(5):290–3. [PubMed: 24147278]
67. Bourbon JR, Boucherat O, Boczkowski J, Crestani B, Delacourt C. Bronchopulmonary dysplasia and emphysema: in search of common therapeutic targets. *Trends Mol Med.* 2009; 15(4):169–79. DOI: 10.1016/j.molmed.2009.02.003 [PubMed: 19303361]
68. Pasquali MA, Schnorr CE, Feistauer LB, Gelain DP, Moreira JC. Vitamin A supplementation to pregnant and breastfeeding female rats induces oxidative stress in the neonatal lung. *Reprod Toxicol.* 2010; 30(3):452–6. Epub 2010 Jun 8. DOI: 10.1016/j.reprotox.2010.05.085 [PubMed: 20679000]
69. Marseglia L, D'Angelo G, Manti S, Arrigo T, Barberi I, Reiter RJ, Gitto E. Oxidative stress-mediated aging during the fetal and perinatal periods. *Oxid Med Cell Longev.* 2014; 2014:358375. Epub 2014 Aug 18. doi: 10.1155/2014/358375 [PubMed: 25202436]
70. Chakravorty S, Murray N, Roberts I. Neonatal thrombocytopenia. *Early Hum Dev.* 2005; 81(1):35–41. Epub 2004 Nov 19. [PubMed: 15707713]
71. Breyman C. Iron deficiency anemia in pregnancy. *Semin Hematol.* 2015; 52(4):339–47. Epub 2015 Jul 10. DOI: 10.1053/j.seminhematol.2015.07.003 [PubMed: 26404445]
72. Perrotta S, Nobili B, Rossi F, Crisuolo M, Iolascon A, Di Pinto D, et al. Infant hypervitaminosis A causes severe anemia and thrombocytopenia: evidence of a retinol-dependent bone marrow cell growth inhibition. *Blood.* 2002; 99:2017–22. [PubMed: 11877274]
73. Medawar PB. Some immunological and endocrinological problems raised by the evolution of viviparity in vertebrates. *Sym Soc Exp Biol.* 1953; 7:320–328.

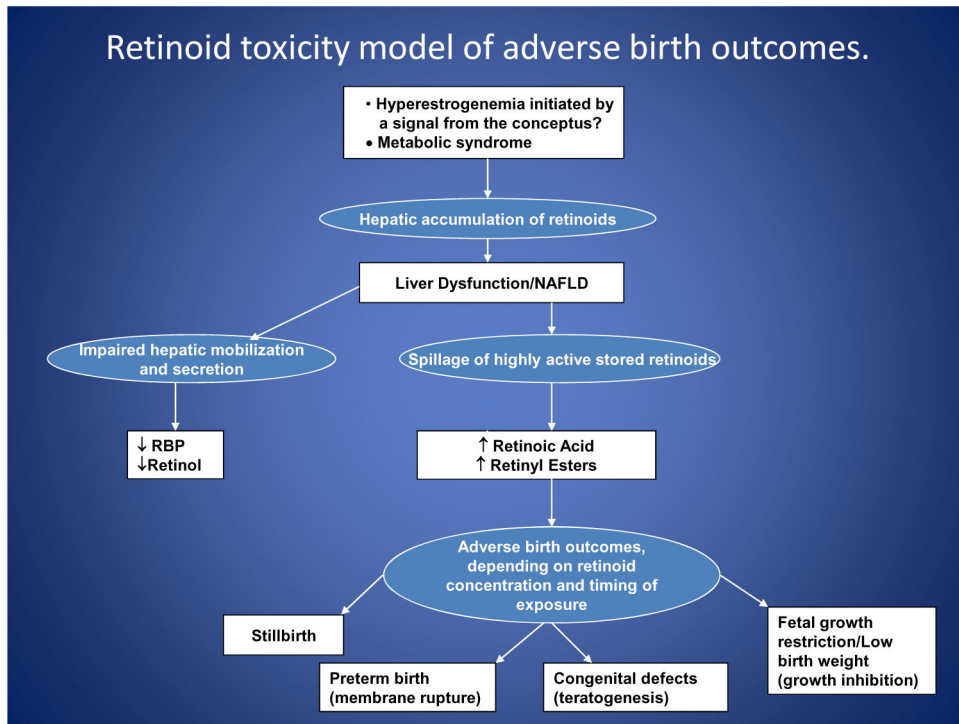


Figure 1.
Retinoid toxicity model of adverse birth outcomes

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript