

Research Paper

Multi-lineages of Shiikuwasha (*Citrus depressa* Hayata) evaluated by using whole chloroplast genome sequences and its bio-diversity in Okinawa, Japan

Ryuji Ishikawa*¹⁾, Nathan Badenoch²⁾, Kunimasa Miyagi³⁾, Kaname Medoruma⁴⁾, Toshiki Osada⁵⁾ and Masayuki Onishi⁶⁾

¹⁾ Faculty of Agriculture and Life Science, Hirosaki University, Hirosaki, Aomori 036-8561, Japan

²⁾ Center for Southeast Asian Studies, Kyoto University, Kyoto, Kyoto 606-8501, Japan

³⁾ Association of Shishigaki Network, Naha, Okinawa 902-0071, Japan

⁴⁾ Okinawa prefectural Chūbu Agricultural Development Center, Department of Agriculture, Forestry and Fisheries, Okinawa Prefecture, Nago, Okinawa 904-2155, Japan

⁵⁾ Research Institute of Humanity and Nature, Kyoto, Kyoto 603-8047, Japan

⁶⁾ Research Center for Knowledge Science in Cultural Heritage, Doshisha University, Kyoto, Kyoto 610-0394, Japan

Shiikuwasha (*Citrus depressa* Hayata) is distributed from the South-west of the Japanese archipelago to Taiwan. In this study, re-sequencing against the orange (*C. sinensis* (L.) Osbeck) chloroplast genome was applied to one superior landrace of Shiikuwasha cultivated in Oku ward, Okinawa, Japan. The chloroplast genome of the landrace was estimated to comprise 160,118 bp, including 48 indels and 71 nucleotide substitutions against the reference genome. The presumptive chloroplast indels were confirmed by subsequent experiments, and these identified multiple maternal lineages among other landraces. Some of the orange SSR markers were available for genotyping of other superior landraces and were able to distinguish among them. These molecular markers were then applied for evaluation of genetic diversity among wild and cultivated Shiikuwasha accessions. Except for Oku ward, the cultivated populations were found to have lost their genetic diversity in comparison with wild populations. Groves in Oku ward maintained, or showed even higher genetic diversity than wild accessions in the surrounding areas by the force of villagers.

Key Words: *Citrus depressa* Hayata, Shiikuwasha, re-sequencing, maternal lineage, selection.

Introduction

Wild relatives within the genus *Citrus* are distributed from East Asia to Oceania. Many horticultural species are considered to have their origins in these areas, and fruit production is very rich worldwide. For example, more than 68 Mt of oranges and 131 Mt of other citrus fruits were produced globally in 2012 (FAO Stats, <http://faostat3.fao.org>). Taxonomical relationships among *Citrus* species are, however, enormously complicated; on the basis of independent taxonomical considerations, Swingle (1943) identified 10 species, whereas Tanaka (1954) identified 147, including horticultural species.

Vegetative propagation has contributed to the spread of horticultural *Citrus* species. For example, cultivation of the sweet orange (*C. sinensis* (L.) Osbeck) has been widespread,

but details about its origin have not been clear. Based on molecular data (Xu *et al.* 2013), it is presumed that the species was created through a process of domestication involving hybridization between the pomelo (*C. maxima* (Burm.) Merr.), synonymously *C. grandis* L.) and the mandarin orange (*C. reticulata* Marcow.), followed by additional hybridization between the mandarin and the F1 hybrid. The grapefruit probably originated from cross hybridization between the orange and the pomelo. In addition to cross-hybridization breeding, bud mutation has complicated the origins of *Citrus* species. For example, detection of bud mutations might be involved in the origin of *Citrus* varieties. For example, bud mutation and mutation breeding using mutagens have enabled breeders to establish further varieties of grapefruit (Moore 2001). Most of the *Citrus* species currently found in Japan were introduced from abroad and they were subsequently improved by bud-mutation breeding and cross-hybridization as in the case of orange and grapefruit.

Okinawa is one of the regions where *Citrus* species have been reported in abundance, and two wild species are

Communicated by K. Kato

Received November 18, 2015. Accepted April 19, 2016.

First Published Online in J-STAGE on July 9, 2016.

*Corresponding author (e-mail: ishikawa@hirosaki-u.ac.jp)

present there (Tanaka 1948, 1957). One of these native species, *C. depressa* Hayata (a common name; Shiikuwasha), is distributed in the south-west part of the Japanese archipelago and in mountainous areas of Taiwan. Tachibana (*C. tachibana* (Makino) Tanaka) is another wild species distributed in Japan, but is not as popular as other *Citrus* fruits at the commercial level. As Shiikuwasha carries more heterozygous loci than Tachibana, it appears to be a hybrid between Tachibana and an unknown species (Yamamoto *et al.* 1998). However, Shiikuwasha shows a variety of morphologic features in terms of fruit size, shape, and appearance, suggesting that it may not have originated from a single cross, but rather through independent domestication from a wild population involving multiple events. In fact, for a very long time it has been tradition for local farmers to introduce wild plants into their groves. Language studies also support the idea of multiple domestication because there are various local names for specific accessions. Some are designated “Kugani”, “Kachi”, or “Tanibuta”, meaning gold, thick peel, or large seeds (Kinjo 2007, Onishi and Miyagi 2016). Despite an abundance of local names, the accessions corresponding to each are not known precisely. Therefore, there is still a chance of finding valuable resources such as a seedless accession (Medoruma *et al.* 2011).

In order to ascertain the origin of Shiikuwasha and its degree of genetic diversity, intensive molecular studies are required, but no effective tool for studying Shiikuwasha existed until the development of *Citrus* genome studies. The orange is one *Citrus* species that is dominant worldwide, and genome studies have revealed the full chloroplast genome sequence and a partial nuclear genome sequence (Bausher *et al.* 2006, Xu *et al.* 2013). In the present study, using the chloroplast genome of the orange as a reference-genome, we subjected Shiikuwasha to next generation sequencing (NGS). Nuclear SSR markers used to construct

an orange linkage map, were also applied to Shiikuwasha, although it was not possible to amplify most of them. These molecular tools demonstrated that polymorphism does exist among Shiikuwasha landraces. Accordingly, landraces and wild accessions were collected in Okinawa in order to evaluate their degree of diversity. We found that recent cultivation had resulted in a dramatic reduction of diversity, and also that genetic variability still exists in the northern parts of Okinawa island.

Materials and Methods

Plant materials

In order to screen for genetic polymorphism, domesticated and wild accessions of Shiikuwasha were collected from three areas of Okinawa prefecture, Japan, during 2013–2014: Ogimi village, Nago city, and Kunigami village (**Table 1**, **Fig. 1**). All materials were donated by local villagers and farmers, and taken from groves that were owned by them. Leaves and fruit from wild individuals that were not endangered species were collected from public areas. Ogimi village is the most well-known area for Shiikuwasha production, and has 29 domesticated accessions. Seventy-six wild accessions were collected as a wild population at Nekumachiji Mountain in Ogimi village. Katsuyama ward in Nago city is

Table 1. Shiikuwasha materials collected in Okinawa, Japan

Site	Sites	No. of accessions
Oogimi	Domesticated	29
	Wild	76
Nago	Domesticated	21
	Wild	44
Kunigami	Domesticated	190
	Wild	75
Total		435

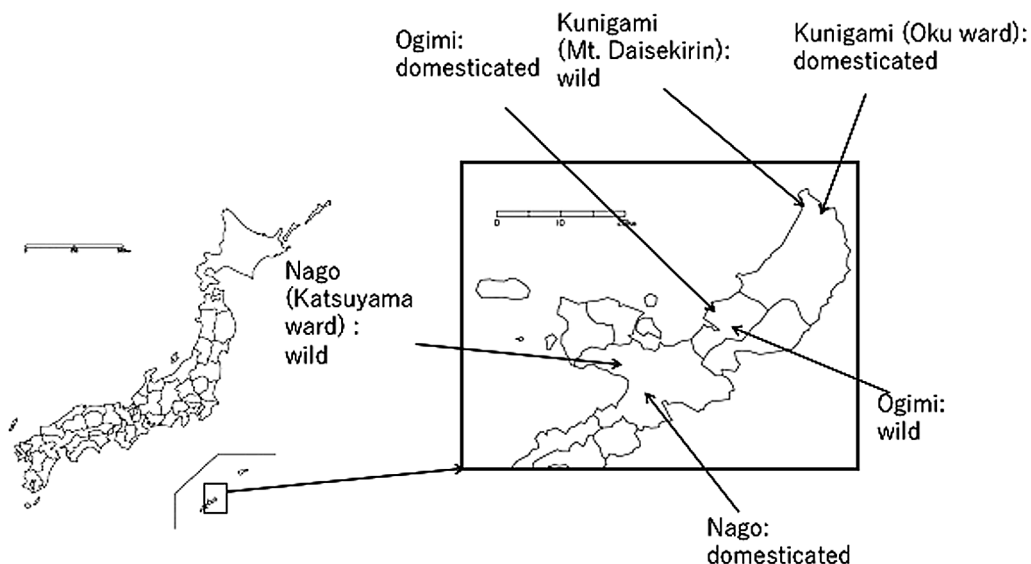


Fig. 1. Collection sites in Okinawa, Japan.

Table 2. Chloroplast and nuclear SSR markers and chloroplast INDELs (cpINDEL) to evaluate polymorphism in chloroplast and nuclear genome

Type	Markers	Abbreviation	Forward	Reverse	Genetic diversity*	
					Oku ward	Daisekirin-zan
Cytoplasmic marker	MM 2-cpINDEL1		TAATTGGAACCTAGGGCTC	ACTGCGACTGATCCTGAAAGG	0.018	0.000
	MM 2-cpINDEL2		GCTTGAAAAAGCCGCTTTAC	AATGATAGAAATGGAATTGCTG	0.018	0.000
	MM 2-cpINDEL3		GGATTCCATTCCGAGTACC	GTATCAATGGTCACTCTCTGG	0.489	0.000
	MM 2-cpINDEL4		GATCAATTGAGATATCCGGAAACC	GGTTAGTCCTTGAACCGAGTG	0.053	0.000
	MM 2-cpINDEL5		GATATGAGATCTACAAATCTCC	CCCTTGGGTCTAATACAACAAC	0.557	0.000
	MM 2-cpINDEL6		GTGGCTTCAACCTCAGCGAAACC	GCTAAATCGCAGTCCAGTATTC	0.018	0.000
	MM 2-cpINDEL7		CGAATTTTTATTCAACCCTATAG	CTATACGGTTCGAACCTATCTAC	0.413	0.080
	MM 2-cpINDEL8		CTTTTTCTAAAGGTAGTTAGTG	GAAGTACGTTTTTTTGGAACTG	0.053	0.000
Nuclear	Ma3_98	98	GATCACCACAAGCAGCACAC	TCTCAAGAGCCCAGTTCGAT	0.132	0.077
	Ma3_142	142	TTTCTTCGTCAACCCCAAAG	TAGAAGAAGGACCCCGAACC	0.266	0.186
	Ma3_168	168	GATTCTTCTTCGGCTGCTG	GCTCGACAGGTTGTTGGTTT	0.533	0.756
	168rev		GTTGGTTTGGTGAGTTTGG	GCGGCAGCATCAGAAGAATG	–	–
	Ma3_200	200	ATGTCGACGTTGACCAATGA	TTTCGTTCTCAGGTGGACT	0.375	0.484
	Ma2_582	582	GACATGTCACACAAGCAGGC	GCAAAATACAGTTGAAAATGCG	0.000	0.497
	Ma2_1710	1710	TGGAACATTGAAGTGGGTGA	ACTTGAGATTAGGGCCGGTT	0.649	0.630
	1710rev			ACACTAACCTCAGCTCATAAG	–	–

* cpINDEL markers were genotyped with 110 Oku accessions and 24 Daisekiin mountain wild accessions. 168rev and 1710rev were modified to fit to score by sequencing gels with *shiikuwasha* samples.

a well-known center for manufacture of Shiikuwasha juice. Here, 21 domesticated accessions were collected. Wild accessions were collected from Katsu Mountain in Nago city, where wild Shiikuwasha was reported to be abundant nearly 60 years ago (Tanaka 1957). At Oku ward in Kunigami village, 190 domesticated accessions were collected because various phenotypes were observed in groves there. Wild accessions were also collected at Daisekirin Mountain in Kunigami village. Leaves from these collected accessions were used for extraction of DNA using the general urea method. In order to evaluate variation in fruit characteristics, three fruits per tree were harvested from 41 trees in Oku ward and 17 trees in Ogimi village in October, 2014. Fruit skin thickness was measured with a digital caliper, DCN100 (Mitsutoyo Co., Japan). Acidity (%) was measured with a PAL-ACID1 device (Atago Co., Ltd., Japan). Percentage sugar content (BRIX) was measured with a PAL1 device (Atago Co., Ltd.). Seven superior landraces (Nakamoto-seedless, Ishi-kunibu, Izumi-kugani, Katsuyama-kugani, Asahikawa-Shiikuwasha, Fusubuta, and Ogimi-kugani) were employed as control plant materials stocked at Okinawa Agricultural Research Center (Nago branch). MM2, which is a late-ripening landrace present in Oku ward, was used for detailed comparison of genotypes with a superior landrace.

Molecular markers

SSR markers developed for the orange nuclear genome were applied to Shiikuwasha (Xu *et al.* 2013). Only seven markers among 29 nuclear SSRs examined were able to genotype; the others could not be amplified (Supplemental Table 1). One of the seven markers was excluded later because it was not stably amplified (Ma3_5). These markers were then used to measure molecular diversity (Table 2). Chloroplast markers developed using other *Citrus* species were then applied to Shiikuwasha (Cheng *et al.* 2005), and four of them were useful for measurement of Shiikuwasha

diversity. These markers were amplified using PCR cycles of 94°C for 3 min, 30 rounds of 95°C for 10 s, 55°C for 30 s, and 72°C for 30 s, and finally 72°C for 5 min with Thermopol (NEB Ltd., Japan). The amplified DNA fragments were electrophoresed on 6% denaturing polyacrylamide gel at 1500 V for 2 h in 0.5X TBE. The gels were then stained with silver nitrate.

Evaluation of the full chloroplast sequence using next generation sequencing (NGS)

The superior landrace, MM2, sampled in Oku ward (GPS:N2651305/E12815160) under a permission of the owner was used for DNA extraction with a Dneasy Plant Mini Kit (QIAGEN Co., Japan). After libraries had been prepared with a 350-bp insert-TrueSeq Nano DNA LT Sample Prek kit, 100-bp pair-end reads were obtained by HiSeq 1000 (Illumina Inc., Japan), and this yielded 52,666,382 reads corresponding to 5,318 Mb in total size. The High Quality Base (30 < A) content was 92.49%. Re-sequencing analysis against the *C. sinensis* chloroplast genome (NC_008334) was performed using CLC Genomics Workbench (ver 7.0). Reads were covered for 178 times on average against the orange chloroplast genome. Indels of more than 4 bp against the reference genome were screened. Primers were designed to amplify a fragment of about 200 bp covering each indel deduced in the orange genome (Table 2). Then, Shiikuwasha accessions were genotyped with the standard SSR protocol as described above.

Results

Variations in morphological and physiological traits

Fruits were collected from 58 trees to evaluate variations in morphological and physiological traits (Fig. 2). Three fruits were measured per single individual for acidity and Brix. In general, acidity gradually decreased until January.

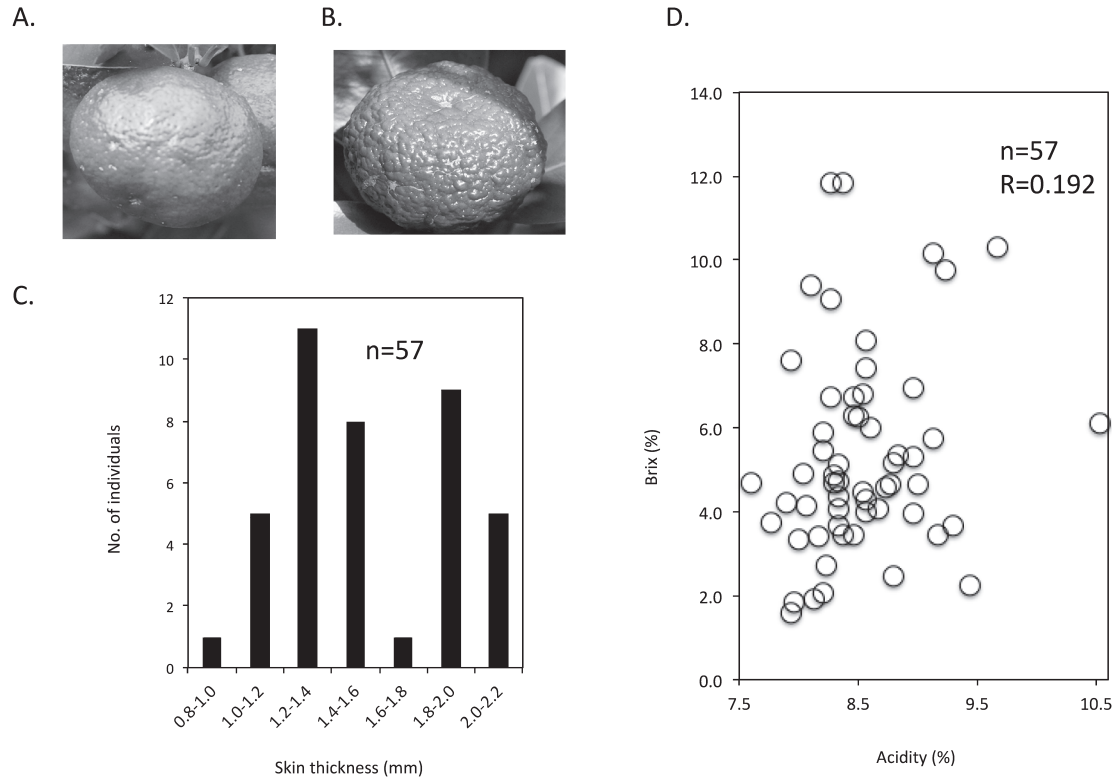


Fig. 2. A. Fruit with smooth surface called as Kugani type generated abundant fruit juice. B. Fruit with rough surface called as “Kachi” type generated fragrance. C. Fruit skin thickness represented discontinuous distribution. D. Fruit phenotypes, Acidity and Brix (%) with 57 accessions measured by three fruits of single individual trees. There was no significant correlation between the traits. D. Collection sites wild and domesticated accessions were obtained.

However, immature fruits are harvested starting in October. Compared to other *Citrus* species, consumers prefer the sour taste of Shiikuwasha. The thickness of the fruit skins showed discontinuous variation with two modes of 1.2–1.4 and 1.8–2.0 mm. These ranged from 0.9 to 2.1 mm in Oku ward and from 0.8 to 1.6 mm in Ogimi village, and thus variation in fruit skin was wider in Oku ward. The acidity scores ranged from 7.6% to 10.5%, and the Brix scores from 1.6% to 11.9%. The distributions of both scores were continuous. The Oku population showed a wider range of acidity. The minimum acidity score was 7.6% in the Oku population, which was 0.57% lower than that in the Ogimi population. The Brix score in the Oku population was higher than that in the Ogimi population.

Chloroplast markers of the orange genome

In order to develop molecular markers for *C. depressa* Hayata, re-sequencing of a superior accession from Oku ward in Kunigami village was performed. The average depth was $\times 178$ against the reference chloroplast genome, which is 160,129 bp in size (NC_008334). Indels and SNP variants were screened in comparison with the reference genome. Twenty-one deletions, 26 insertions, one multi-nucleotide variant, and 71 SNPs were detected (Supplemental Tables 2, 3). The flanking sequences of the corresponding regions tended to carry single nucleotide repeats

or duplications of short sequences. In total, the chloroplast genome of *C. depressa* Hayata was estimated to be 160,118 bp in size. The complete chloroplast genome sequence has been registered in the DDBJ as Submission ID: 5714a8902113a5b85901557c. Based on the genome sequence, more than 4 bp of chloroplast insertion/deletion (MM2-cpINDELs) were developed as INDEL markers and confirmed experimentally (Fig. 3, Supplemental Table 3). Three markers—MM2-cpINDEL3, MM2-cpINDEL4, and MM2-cpINDEL5—carried more than three alleles when wild and cultivated accessions were genotyped (data not shown). A deletion of CTCCTTTT inside MM2-cpINDEL3 was flanked with 10 thymine repeats, whereas polymorphism in MM2-cpINDEL4 was based on single adenine repeats. The MM2-cpINDEL5 was an insertion of AATTTG at a tandem duplication of the AATTTG motif. These nucleotide structures resulted in the generation of multiple alleles.

Among the cpINDELs, eight MM2-cpINDELs were further analyzed. For six out of eight MM2-cpINDELs, alternative alleles were observed among landraces (Table 3). The indel markers and categorized landraces formed two groups. Group 1 comprised Nakamoto-seedless and Ishikunibu, whereas Group 2 comprised Izumi-Kugani, Fusubuta, Katsuyama-Kugani, Asahikawa-Shiikuwasha, and Ogimi-Kugani. MM2 carried genotypes different from

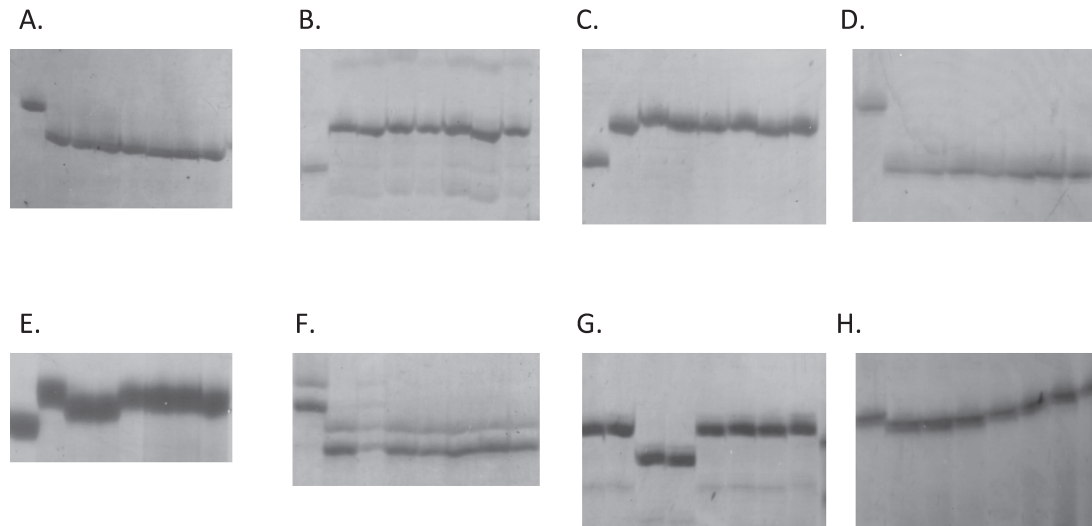


Fig. 3. Maternal traceability accessed with eight chloroplast INDELS developed from a whole genome sequence data obtained from MM2 (lane 1). From lane 1 to lane 8, MM2, genotypes of Oogimi kugani, Nakamoto seedless, Ishi kunib, Izumi kugani, Asahikawa shiikuwasha, and Fusubuta were shown.

Table 3. Discrimination of single landraces with eight chloroplast INDELS developed with NGS data and six nuclear SSR markers

Cultivars	MM2-cpINDEL								Nuclear markers					
	C1	C2	C3	C4*	C5	C6	C7	C8	98	142	168	200	582	1710
MM2	2	1	1	3	1	2	2	2	2/3	1/3	2/2	1/2	3/3	1/2
Nakamoto-seedless	1	2	3	1	2	1	1	1	2/2	2/2	2/2	1/2	3/3	1/3
Ishi-kunibu	1	2	3	1	2	1	1	1	2/2	2/2	2/2	1/1	3/3	1/2
Izumi-Kugani	1	2	2	1	3	1	2	1	2/2	2/2	2/3	1/1	3/3	1/4
Katsuyama-Kugani	1	2	2	1	3	1	2	1	2/2	2/2	2/4	1/1	3/3	1/4
Asahikawa-Shiikuwasha	1	2	2	1	3	1	2	1	2/2	2/2	2/4	1/1	3/3	1/4
Fusubuta	1	2	2	1	3	1	2	1	2/2	2/2	2/2	1/2	3/3	1/4
Ogimi-Kugani	1	2	2	1	3	1	2	1	2/2	2/2	2/4	1/1	3/3	1/4

* Multiple allele, 2 was observed in wild accessions (data not shown).

all the others. Although 110 Oku accessions were genotyped with eight MM2-cpINDELS, only MM2 carried alternative alleles at three loci and a few accessions shared the same alleles with MM2 (Table 2). Three markers—MM2-cpINDEL3, 5, and 7—showed relatively high genetic diversity, at 0.489, 0.557, and 0.413, respectively. The genetic diversity of other markers ranged from 0.018 to 0.053. In all cases, MM2 carried unique genotypes in comparison to most of the others, except for MM2-cpINDEL7.

Development of nuclear markers

SSR markers in *C. sinensis* (L.) Osbeck were selected randomly and applied to *C. depressa* Hayata. Twenty-one out of 26 SSR markers were successfully amplified, and of these, four markers were monomorphic among the examined accessions. The remaining 17 markers showed polymorphisms, of which six were applied in order to determine the genotypes of superior landraces. Apart from two landraces—Fusubuta and Ogimi-Kugani—the rest were genetically different and distinguishable from each other. Genotypes were then compared between landraces and wild accessions, both having been collected in Oku ward. Al-

though the Oku landraces shared a single genotype at Ma2_582, all other markers showed polymorphism in both populations. These landraces showed relatively high diversity, with averaged *He* scores ranging from 0.089 to 0.588.

Genetic diversity in Okinawa

Shiikuwasha farming is widespread in the northern half of Okinawa island. Leaf samples of cultivated Shiikuwasha accessions were collected from two villages (Ogimi and Kunigami) and one city (Nago). Wild accessions were detected in forests near these sites.

In total, 345 accessions collected from Ogimi village, Nago city, and Kunigami village were genotyped with the three MM2-cpINDELS. Domesticated accessions in Kunigami village originated only in Oku ward. Fifteen genotype combinations were recognized as plastid types (Table 4). The genotype of MM2 accessions was 1-1-2 (MM2-cpINDEL3, 5, and 7), which was not shared with other domesticated or wild individuals. Among domesticated populations, the Ogimi and Nago populations were composed of only one or two genotypic combinations. Although wild populations were composed of a greater number of genotypic combinations,

Table 4. Chloroplast genotypes measured with three chloroplast INDELs developed with NGS data of MM2 accessions, compared among domesticated and wild accessions in three regions, Ogimi village, Nago city, and Kunigami village

Maternal lineage	MM2-cpINDEL			No. of accessions					
	cpINDEL3	cpINDEL5	cpINDEL7	Ogimi		Nago		Kunigami	
				Dom.	Wild	Dom.	Wild	Dom.	Wild
Type1	1	1	1	0	0	0	0	1	0
Type2	1	1	2	0	0	0	0	1	0
Type3	1	2	1	0	0	0	0	3	0
Type4	1	2	2	0	64	0	0	8	0
Type5	1	3	2	0	8	0	0	53	0
Type6	2	1	1	0	0	0	0	5	0
Type7	2	2	1	0	1	7	1	24	17
Type8	2	2	2	29	0	13	18	15	41
Type9	2	3	1	0	0	0	2	0	0
Type10	2	3	2	0	0	0	7	4	4
Type11	3	1	1	0	0	0	0	0	1
Type12	3	2	1	0	0	0	8	0	4
Type13	3	2	2	0	0	0	3	0	0
Type14	3	3	1	0	0	0	1	0	0
Type15	3	3	2	0	0	0	2	0	0
Total				29	73	20	42	114	67

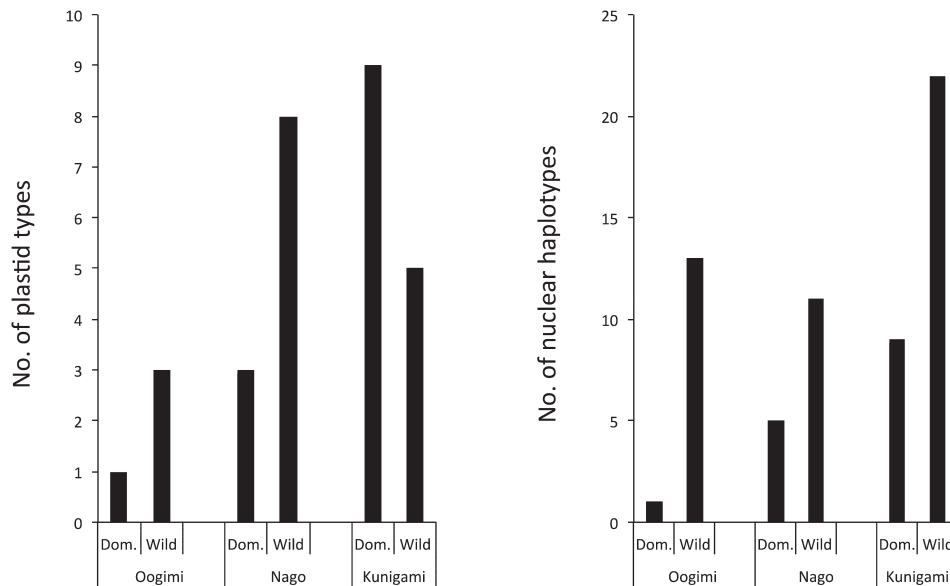


Fig. 4. Number of haplotypes detected among wild and domesticated populations at three sites. Haplotypes were determined with two loci, Ma3_168 and Ma3_200.

there were only three or eight combinations in the Ogimi and Nago populations, respectively (Fig. 4, Supplemental Table 4). The most varied combinations were identified in a population collected in Oku ward. Nine genotypic combinations including one MM2 were observed. In contrast, five genotypic combinations were detected in a wild population in Kunigami village.

Two nuclear markers—Ma3_168 and Ma3_200—were genotyped with wild and domesticated accessions. Multiple alleles at the Ma3_168 locus did not give good resolution for genotyping. A new primer pair was therefore designed based on the sequence corresponding to the Ma3_168 locus, and this primer pair yielded good resolution. The genotypes of the two markers were defined as haplotypes. The highest

number of haplotypes (nine) was found in Kunigami village (Table 5). Only one haplotype was found in Ogimi village, compared to 13 haplotypes in wild populations near the farms.

Apomixis

In the past, local farmers established their groves with stocks from wild trees or by transplantation of young seedlings from the surrounding habitats. This led to recognition of several superior accessions, which were conserved as key landraces as shown in Table 3. For this reason, trees in modern groves did not show variation. For Shiikuwasha, there had been no data about apomixis, which is one of the mechanisms by which variation can be reduced. Therefore,

Table 5. Nuclear haplotypes combined with two nuclear SSR markers, Ma3_168 (168) and Ma3_200 (200) among domesticated and wild accessions in three regions, Ogimi village, Nago city, and Kunigami village

Haplotype	Haplotype		No. of accessions					
			Ogimi		Nago		Kunigami	
	168	200	Dom.	Wild	Dom.	Wild	Dom.	Wild
Hap1	1/1	1/1	0	1	0	0	1	0
Hap2	1/1	1/2	0	0	0	2	0	7
Hap3	1/1	1/2	0	0	0	1	0	0
Hap4	1/2	1/1	0	3	0	0	0	1
Hap5	1/2	1/2	0	3	0	1	0	3
Hap6	1/2	1/2	0	1	0	0	0	2
Hap7	1/3	1/2	0	1	0	0	0	2
Hap8	1/3	2/2	0	0	0	0	0	1
Hap9	1/4	1/1	0	1	0	0	0	0
Hap10	1/4	1/2	0	1	0	1	0	1
Hap11	1/5	1/1	0	1	0	0	0	0
Hap12	2/2	1/1	0	0	1	3	10	4
Hap13	2/2	1/2	0	0	7	27	19	4
Hap14	2/2	2/2	0	1	0	0	0	0
Hap15	2/3	1/1	0	0	1	0	4	2
Hap16	2/3	1/2	0	0	2	1	6	2
Hap17	2/4	1/1	29	47	9	2	57	11
Hap18	2/4	1/2	0	8	0	2	10	10
Hap19	2/4	2/2	0	0	0	0	6	4
Hap20	2/5	1/1	0	0	0	0	1	0
Hap21	3/3	1/1	0	0	0	0	0	1
Hap22	3/3	1/2	0	0	0	1	0	4
Hap23	3/3	2/2	0	0	0	0	0	1
Hap24	3/4	1/1	0	0	0	0	0	1
Hap25	3/4	1/2	0	0	0	0	0	3
Hap26	3/4	2/2	0	0	0	1	0	1
Hap27	4/4	1/1	0	2	0	0	0	1
Hap28	4/4	1/2	0	0	0	0	0	1
Hap29	4/4	2/2	0	1	0	0	0	0
Total			29	71	20	42	114	67

progenies propagated through seeds were genotyped and compared with their parental trees. Seeds from one superior tree (MM2) and one wild accession (Oku) next to the old junior high school in Oku ward were collected. Seeds from

each tree were germinated and allowed to grow from separated poly-embryos inside the seeds (**Fig. 4**). Young leaves from single seedlings were then taken for extraction of DNA and genotyping with six SSRs separately. One of 16 young seedlings (6.3%) from the MM2 tree and two of eight seedlings (25%) differed in genotype from the mother tree. For example, a codominant marker, Ma3_168 showed a non-parental allele associated with crossing between the Oku tree and other tree(s) (**Fig. 5**). The difference in the percentage among seedlings suggested that outcrossing would have been influenced by the surrounding conditions. The tree from the Oku ward, was a wild one that stood alone, whereas MM2 stood at the top of a hilly area in a grove. Possible pollen donors could not be determined because of a lack of genetic information about the surrounding trees at present. A high rate of apomixis is one possible reason for the low diversity in Ogimi village, where extreme proliferation through seeds of particular superior accessions has occurred. In contrast, it is also suggested that careful selection to screen the progeny derived from cross-hybridization might allow breeders to improve Shiikuwasha through crossing within the species.

Discussion

Cultivation history and local selection processes

Shiikuwasha is the only *Citrus* species in Japan that is commercially cultivated and co-exists in a wild form in forests. Therefore it may have high tolerance to biotic or abiotic stress. No other varieties of popular cultivated fruits, such as Oto, Tsukurubu, or Kabuchi, are found under wild conditions in Okinawa. Shiikuwasha has long been consumed in the form of vinegar, and has been used as a substitute bleaching solution. To this day, it is quite common to find trees in individual home gardens and yards. The historical record of the Shiikuwasha is documented in the *Omoro-zoushi*, an Okinawan text that was edited in the mid 16th to

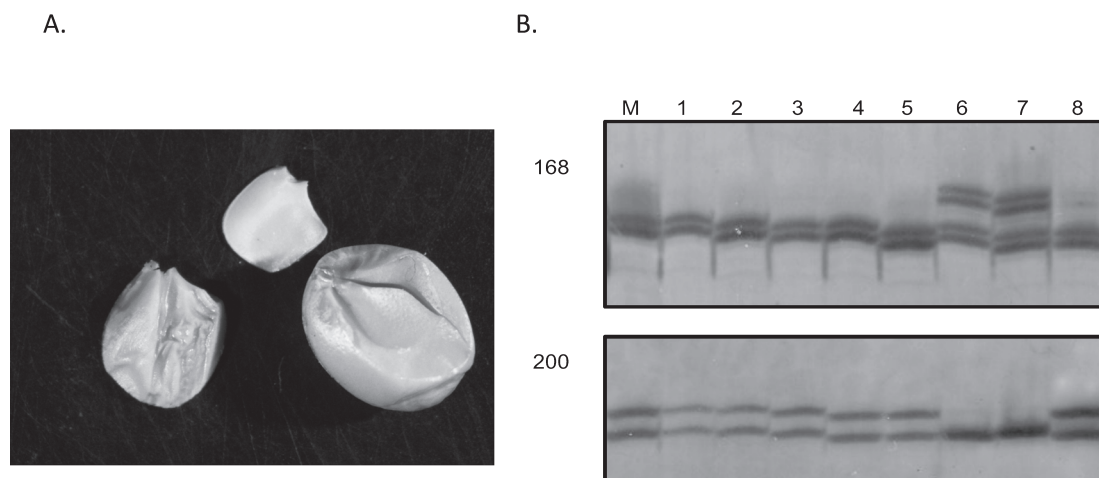


Fig. 5. Polyembryos in a single seeds and polymorphisms detected in progenies of single trees. A. polyembryos detected in a single seed. B. Polymorphisms found by using Ma3_168 and Ma3_200 markers in progeny seedling of the Oku tree. Genotypes of Ma3_168 and Ma3_200 suggested that No. 6 and No. 7 seedlings outcrossed with other plants.

17th centuries (Kinjo 2007). However, commercial cultivation began only about 50 years ago. Since then, landraces have been cultivated in restricted areas of central and northern Okinawa island. Consumption of Shiikuwasha increased gradually in the 1980s when the juice industry began to take off. Food science research subsequently identified high levels of vitamin C and other health benefits, further boosting the popularity of the Shiikuwasha (Kinjo 2007). This intensive period of commercial development necessitated the amplification of vegetative clones for expansion of cultivation in areas of central Okinawa, such as Katsuyama ward and Ogimi village. Although market forces have tended to homogenize the industry, a significant variation in cultivated Shiikuwasha is still evident. One example of this variation is fruit skin thickness, which shows a bimodal distribution. These traits may be partly affected by environmental conditions on a broader scale, but genetic factors are generally important as well. Experienced farmers have selected superior individuals at their farms. Against the interplay of these factors, breeders have succeeded in selecting superior accessions and maintained them at Okinawa Prefectural Institute of Agriculture.

Molecular markers make it possible to evaluate differences among genetic resources. However, there are few molecular markers in Shiikuwasha that yield high resolution. At present, the orange is one *Citrus* species for which a large amount of genetic information is available. In order to develop molecular tools for examination of Shiikuwasha, we obtained re-sequencing data against the orange chloroplast genome, and this helped us to develop chloroplast indel markers for distinguishing between landraces. This approach also revealed multiple maternal lineages in the cpINDELs, suggesting that the origin of Shiikuwasha is complex. Multiple domestication, or a single domestication event followed by a long period to allow accumulation of DNA mutations, are the most likely scenarios. As shown in **Tables 4, 5**, wild populations demonstrated wider variation than cultivated ones. It implies that the domestication was happened in Okinawa, Japan.

Katsuyama ward and Ogimi village showed markedly fewer different maternal lineages in comparison with the multiple lineages in Oku Ward. Nuclear genotypes also showed the same trend. The number of haplotypes was highest in a wild population collected from Daisekirin mountain in Kunigami village. In contrast, the number of maternal lineages was higher in Oku ward than in any of the wild populations. This diversity is a direct result of the introduction of other accessions from different areas into Oku ward, in addition to the ease with which clones from wild trees can be developed. Oku is located in the northern-most area of the main island of Okinawa. Local people keep their own Shiikuwasha trees in home gardens and yards, while utilizing wild plants that occur nearby. When compared with Ogimi village and Katsuyama Ward, Oku farmers allowed, and in fact encouraged, marked variation in their groves, ranging from sour to sweet fruit, and early to late

flowering individuals. These have been maintained locally through the conscious efforts of the farmers.

In attempting to clarify the extent of diversity in Shiikuwasha, it should be noted that, historically, Oku ward has remained relatively isolated. The local language, rituals and ecological knowledge that supports the people's traditional livelihood of the local population have all been preserved until very recently (Onishi and Miyagi 2016). In response to the cultural and economic pressures that accompany increased integration into contemporary Okinawan society, the village has developed several strategies for conserving their culture. A village museum has been established to document and transmit the history of the village. Community leaders explained to us how their culture is intimately related to the surrounding environment and their livelihood activities; they keep a wide range of Shiikuwasha fruits in their groves. We hypothesize that their culture, which stresses the importance of maintaining their local language and rituals, may have contributed to the maintenance of plant diversity, including Shiikuwasha. The villagers of Oku demonstrate a locally based, yet deep understanding of biocultural diversity that underpins the genetic diversity that has been observed in our study. This kind of diversity is important for screening of valuable genetic stock and making improvements to it.

NGS data and traceability

The fruits of accession MM2 are of superb quality. In addition, the significantly late maturity is unique, in comparison to other Shiikuwasha accessions. On the basis of its indels, this accession was determined as plastid Type 14. These indels were based on DNA polymorphism between the orange chloroplast genome and the re-sequenced genome based on reads of the MM2 accession. NGS is able to provide genetic tools such as indels markers for characterization of non-model plants or unique landraces such as the MM2 tree. For certification of food security or organic cultivation, genetic diversity and chloroplast genome information is cost-effective, and recent biochemical data suggest that Shiikuwasha fruit may be beneficial to human health in addition to contributing to the local economy. Molecular tools for identifying individual accessions may help to improve the agronomy of the Shiikuwasha. Although the production of this fruit is deeply influenced by environmental conditions such as soil, nutrients, water content, and probably other unknown factors, genetic identification followed by cultivation experiments may provide important insights into the necessary conditions for obtaining good fruit.

Center of diversity

The significant diversity found in local residential areas tended to be secondary centers of diversity, where various genetic components have accumulated in particular crops (Vavilov 1926). Secondary centers of biodiversity in residential areas are a potential source of further domestication for new candidate crops (Anderson 1954, Nakao 1966).

Transplanting of wild Shiikuwasha trees and detection of superior individuals are frequently performed by villagers in Oku ward. As Shiikuwasha has recently been established as a commercial fruit, wild populations and populations in secondary centers such as Oku may be a good breeding resource for a pure-selection methodology which could apply to existing varieties.

The question still remains as to why the villagers in Oku have maintained this high level of diversity, not only in terms of local culture but also Shiikuwasha production. The answer may lie the spirit of the villagers themselves, who place high value on diversity in general. Domestication activities have resulted in the introduction of genetically unique individuals such as the MM2 accession, which carries a unique plastid type with superior quality. These accessions, the relatively low rate of apomixis, and molecular markers will allow breeders to develop linkage maps and recognize linkage disequilibria on the basis of phenotypic data.

Acknowledgements

We thank local coordinators and famers in Oku Ward for their help in collecting valuable materials from local farmers.

Literature Cited

- Anderson, E. (1954) Plants, man and life. A. Melrose Ltd., London.
- Bausher, M.G., N.D. Singh, S.-B. Lee, R.K. Jansen and H. Daniell (2006) The complete chloroplast genome sequence of *Citrus sinensis* (L.) Osbeck var 'Ridge Pineapple': organization and phylogenetic relationships to other angiosperms. *BMS Plant Biol.* 6: 21.
- Cheng, Y., M.C.D. Vicente, H. Meng, W. Guo, N. Tao and X. Deng (2005) A set of primers for analyzing chloroplast DNA diversity in *Citrus* and related genera. *Tree Physiol.* 25: 661–672.
- Kinjo, H. (2007) About shiikuwasha in South-east archipelago. *Studia Citologica* 17: 137–148.
- Medoruma, K., A. Higa, H. Kinjo, H. Zukeyama, Y. Awaguni, T. Miyagi, M. Arasaki, H. Inoue, S. Onda, S. Kawano *et al.* (2011) Characteristics of seedless *Citrus depressa*, Nakamoto seedless. *Bull. Okinawa Pref. Agric. Res. Center* 5: 5–10.
- Moore, G.A. (2001) Oranges and lemons: clues to the taxonomy of *Citrus* from molecular markers. *Trends Genet.* 17: 536–540.
- Nakao, S. (1966) Origin of cultivated plants and Agriculture. Iwanami, Tokyo, Japan.
- Onishi, M. and K. Miyagi (2016) Wisdom in Shiikuwasha-Oku/ Yanbaru, Circulation among dialect, local community, life. Kyoto University Press, Kyoto, Japan, p. 529.
- Swingle, W.T. (1943) The botany of Citrus and its wild relatives of the orange subfamily. *In: Webber, H.J. and L.D. Batchelor (eds.) The Citrus Industry.* Vol. I, University of California Press, USA, pp. 129–474.
- Tanaka, T. (1954) Species problem in citrus: a critical study of wild and cultivated units of citrus, based upon field studies in their native homes (Revisio Aurantiacearum IX), Japanese Society for the Promotion of Science, pp. 141–152.
- Tanaka, T. (1957) Citrus in Ryukyu. Department of Economics, Ryukyu government, pp. 31–37.
- Tanaka, Y. (1948) An iconograph of Japanese Citrus fruits—A monographic study of species and varieties of Citrus fruits grown in Japan Vol. 2. Yokendo, Tokyo, Japan, pp. 481–488.
- Vavilov, N.I. (1926) Studies on the origin of cultivated plants. *Inst. Bot. Appl. Amelior. Plants, Leningrad*, pp. 248.
- Xu, Q., L.-L. Chen, X. Ruan, D. Chen, A. Zhu, C. Chen, D. Bertland, W.-B. Jiao, B.-H. Hao, M.P. Lyon *et al.* (2013) The draft genome of sweet orange (*Citrus sinensis*). *Nat. Genet.* 45: 59–68.
- Yamamoto, M., Y. Matsuo, T. Kuniga, R. Matsumoto and Y. Yamada (1998) Isozyme and RAPD analyses of Shiikuwashes (*Citrus depressa* Hayata). *Bull. Natl. Inst. Fruit Tree Sci.* 30/31: 39–51.