

Research Article

The Impact of Acute Matriptase Inhibition in Hepatic Inflammatory Models

Judit Pomothy,¹ Gergely Szombath,¹ Patrik Rokonál,¹ Gábor Mátis,²
Zsuzsanna Neogrády,² Torsten Steinmetzer,³ and Erzsébet Pászti-Gere¹

¹Department of Pharmacology and Toxicology, University of Veterinary Medicine, István u. 2, Budapest 1078, Hungary

²Department of Physiology and Biochemistry, University of Veterinary Medicine, István u. 2, Budapest 1078, Hungary

³Faculty of Pharmacy, Institute of Pharmaceutical Chemistry, Philipps University Marburg, Marbacher Weg 6, 35037 Marburg, Germany

Correspondence should be addressed to Erzsébet Pászti-Gere; gere.erszebet@aotk.szie.hu

Received 27 April 2016; Accepted 1 August 2016

Academic Editor: Jozef Anné

Copyright © 2016 Judit Pomothy et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Purpose. Dysfunction of matriptase-2 can be involved in iron regulatory disorder via downregulation of hepcidin expression. In the present study, we investigated the effects of 3-amidinophenylalanine-derived matriptase inhibitors on porcine hepatic inflammatory cell models. **Methods.** Hepatocyte-Kupffer cell cocultures (ratio of 2 : 1 and 6 : 1) were treated with four structurally related matriptase inhibitors at 50 μ M. Cell cytotoxicity and relative expressions of IL-6 and IL-8 and the levels of hepcidin were determined by MTS and porcine-specific ELISA. The extracellular H₂O₂ contents were analyzed by Amplex Red method. **Results.** Matriptase inhibitors at 50 μ M for 24 h did not increase cell death rate. The elevated ROS production observed after short-term application of inhibitor MI-441 could be correlated with lowered hepcidin expression. MI-460 could significantly enhance hepcidin levels in the supernatants of cocultures (by $62.21 \pm 26.8\%$ in hepatocyte-Kupffer cell, 2 : 1, and by $42.6 \pm 14.3\%$ in hepatocyte-Kupffer cell, 6 : 1, cocultures, resp.). No significant changes were found in IL-6 and IL-8 levels in cocultures exposed to matriptase inhibitors. **Conclusions.** Based on *in vitro* findings, administration of MI-460 via modulation of hepcidin expression without cytotoxic and oxidative stress inducing properties might be a reliable alternative to treat iron overload in human and veterinary clinical practice.

1. Introduction

Several type II transmembrane serine proteases (TTSPs) are associated with cell surface proteolysis and can be divided into four subfamilies such as human airway trypsin-like protease (HAT)/differentially expressed in squamous cell carcinoma (DESC), hepsin/transmembrane protease, serine 2 (TMPRSS), corin, and matriptase (MT). They also play crucial roles in tightly regulated physiological processes [1]. Expression of matriptase-1 (MT-1) is not limited to any particular organ [2], whereas matriptase-2 (MT-2) mainly accumulates in the liver in both human and mouse, which reveals its tissue-specific functions [3].

Consequently, deregulated MT-1 and MT-2 activities have been linked to various pathological conditions. It was

reported that MT-1 overproduction in osteoarthritis could promote the induction of cartilage destruction [4]. MT-1 activity is also involved in tumorigenesis of various cancer types and promotes the progression of malignant transformations via cleavage and activation of substrates like hepatocyte growth factor/scatter factor [5–7].

MT-2, also known as TMPRSS6, is highly expressed in the liver and it negatively affects the production of the hepatic hormone hepcidin, the main regulator of systemic iron homeostasis through the interaction with the bone morphogenetic protein coreceptor hemojuvelin (HJV) [8–10]. After binding, MT-2 degrades and reduces cell surface HJV and prevents hepcidin from being expressed via induction through bone morphogenetic protein 6 (BMP6) [9]. Active HJV facilitates transcription of hepcidin responsible for iron

transport to the bloodstream. This mechanism is impaired in iron-refractory iron deficiency anaemia due to the hepcidin-dependent cellular iron accumulation and loss of MT-2 proteolytic function as a result of genetic mutation [11, 12].

The elevated production rate of hepcidin triggered by high iron level can lower the iron absorption from the duodenum and reduce the iron release from the iron storages thus stabilizing the finely adjusted physiological iron concentration [13, 14]. The hepcidin secretion in the liver can be elevated due to the infections and inflammatory disorders, which can be facilitated by the proximity of pathogen-sensing Kupffer cells, to which the body reacts with iron deficiency and anaemic conditions consequently. This host defensive mechanism in response to infectious diseases can provide significant reduction in the amount of iron essential for the survival of the microbes. In case of sustained iron deficiency, the hepatocytes produce less hepcidin; thus iron concentration increases again in the blood circulation. Other influencing factors such as bleeding, application of erythropoietin (EPO), and overproduction of reactive oxygen species (ROS) can also modulate the hepcidin expression significantly [15].

MT-1 and MT-2 are belonging to the TTSPs having an extracellular trypsin-like serine protease domain at the C-terminus, which can be blocked by 3-amidinophenylalanine-derived inhibitors. The most potent inhibitors contain N-terminal dichloro- or dimethoxy-biphenyl-3-sulphonyl group and inhibit MT-1 in the low one-digit nanomolar range, whereas MT-2 is less potently inhibited [16]. As mentioned above, MT-2 plays an important regulatory role in iron homeostasis; thus this enzyme may act as a pharmacological target in the treatment of the systemic iron overload (hemochromatosis). This approach seems to be supported by the fact that lack of functional MT-2 can result in high hepatic hepcidin expression, which results in iron-deficiency anaemia [8].

Appropriately established cell models can be used to mimic *in vivo* pathophysiological conditions such as inflamed liver thereby enabling the replacement of some animal experiments. A porcine primary hepatocyte-Kupffer cell coculture with variable ratios of cell types has been previously developed and characterized [17]. In this model, hepatocytes were cocultured with Kupffer cells in the ratio of 6:1 (HepK6) and 2:1 (HepK2) in an attempt to mimic hepatic inflammation with different severity. This system seems to be a proper tool to investigate certain molecular mechanisms within the inflamed liver, such as to assess the possible effects of MT-2 inhibition on the regulation of iron homeostasis in hepatic inflammation.

The main aim of this research was to explore the possible relationship between coculture model of hepatocytes and Kupffer cells of swine liver and selected drug candidates. In our study, the regulatory properties of four potent 3-amidinophenylalanine-derived MT-1 inhibitors (MI-432, MI-441, MI-460, and MI-461), which also possess a moderate inhibitory potency against MT-2, were elucidated *in vitro* in hepatic inflammatory models HepK2 and HepK6 cocultures to determine how MT-1/MT-2 modulation affects cell viability and physiological redox status, possibly being in

connection with hepcidin production. The analysis of changes in the hepcidin expression was a prerequisite for exploring a link between the effect of MT-1/MT-2 suppression and iron homeostatic processes. As hepcidin-mediated disturbances in iron metabolism are often coupled to an inflammatory response, therefore, the expressions of the proinflammatory cytokines, IL-6 and IL-8, were also monitored.

2. Materials and Methods

2.1. Isolation and Cultivation of Hepatocytes and Kupffer Cells. Hepatocyte-Kupffer cell cocultures were freshly prepared from 15 kg weighing male pigs of the Hungarian Large White breed (obtained from Dunahybd Ltd., Fadd, Hungary) [17]. Briefly, hepatocytes and Kupffer cells were isolated by multi-step perfusion and collagenase-mediated enzymatic digestion of the caudate lobe of the liver. Hepatocytes were sedimented and purified by a revised low-speed centrifugation (50 ×g, 75 s); Kupffer cells were obtained from the supernatants by Percoll gradient centrifugation (500 ×g, 20 min). The viability of both cell types was assessed by trypan blue exclusion test and revealed more than 90% viable cells in all cases.

As nonparenchymal cells can attach faster, Kupffer cells were seeded first on previously collagen-coated (Type I, 10 μg/cm²) 12-well Costar TC6 cell culture dishes (Corning International, Corning, NY, USA) and on 96-well microplates, the latter being used for MTS assays on cell viability. Hepatocytes and Kupffer cells were seeded on the same dishes in the appropriate cell ratio 6:1 (hepatocyte to Kupffer cell) for modelling moderate and 2:1 for more severe inflammation. The prepared cocultures were incubated under humid atmosphere at 37°C using 10% CO₂, and culture medium was exchanged 6 h after seeding. Culturing of cells was performed in Williams medium E, supplemented with 10000 IU/mL penicillin, 10 mg/mL streptomycin, 2 mM glutamine, 10% FBS, and 0.2 IU/mL insulin. Fetal bovine serum (FBS) was applied during the first 24 h of culturing. Confluent monolayer cocultures were obtained after 24 h incubation (Figures 1(a) and 1(b)), being suitable for the further examinations.

2.2. Chemical Structures of Applied Matriptase Inhibitors. The structures of the used 3-amidinophenylalanine-derived inhibitors and their previously published K_i values against MT-1 and MT-2 [16] are summarized in Table 1. The compounds MI-432 and MI-441 are dibasic inhibitors containing a C-terminal 2-aminoethylpiperidide group, whereas the analogues MI-460 and 461 are more hydrophobic monobasic derivatives.

2.3. Exposure of HepK2 and HepK6 Cells to Matriptase Inhibitors. Stock solutions of the inhibitors were prepared with a concentration of 10 mM and kept at -20°C. Before treatment, confluent HepK2 and HepK6 cocultures were washed twice with plain medium. The solutions of the inhibitors in phenol red free Williams medium E at 50 μM were prepared freshly prior to each experiment from the stock

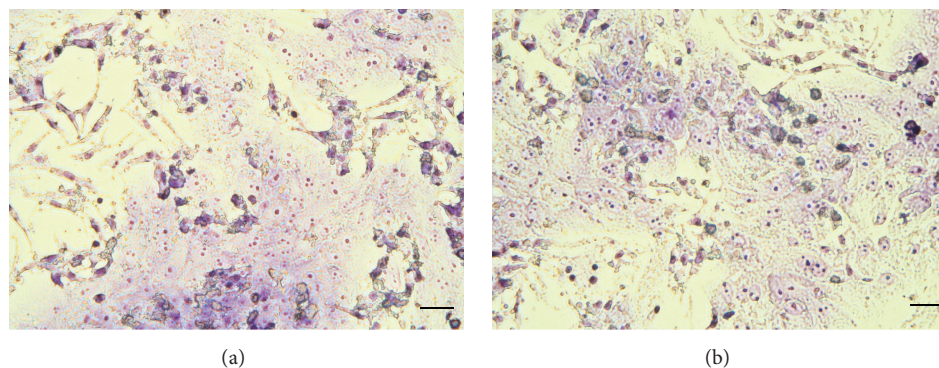
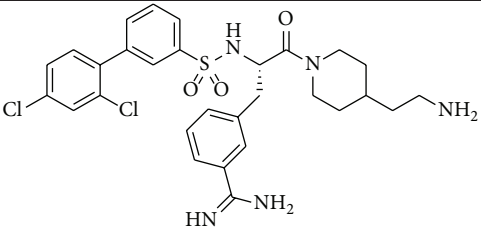
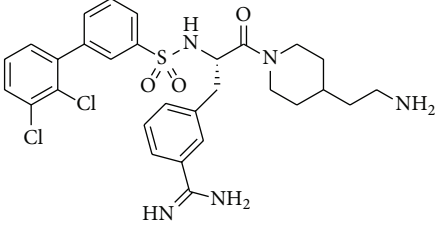
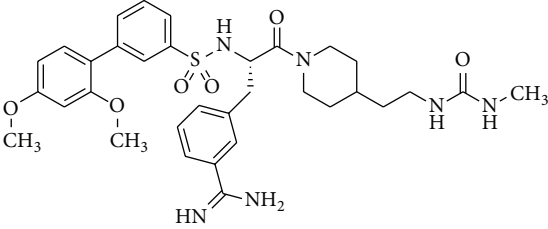
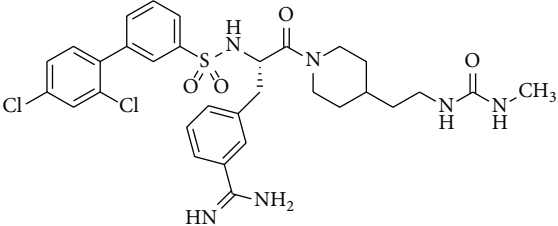


FIGURE 1: Giemsa staining of cocultures. HepK cell cocultures with the cell ratio of 2:1 (a) and 6:1 (b) (hepatocyte to Kupffer cell) after 24 h culturing following Giemsa staining (200x magnification). Scale bar = 50 μm .

TABLE 1: Chemical structures and K_i values of the used 3-amidinophenylalanine-derived inhibitors towards matriptase-1 (MT-1), matriptase-2 (MT-2), and thrombin.

Number	Structure	K_i (μM)		
		MT-1	MT-2	Thrombin
MI-432		0.002	0.11	0.02
MI-441		0.011	0.21	0.02
MI-460		0.0018	0.18	0.0008
MI-461		0.0009	0.073	0.0002

solutions. After incubation of the inhibitor for 2 h (in case of H_2O_2 , IL-6 and IL-8, and hepcidin assays) or 24 h (in case of cell viability tests), the cocultures were washed twice with plain medium before being subjected to the subsequent procedures.

2.4. *MTS Assay for Cell Viability.* Influence of matriptase inhibitors in phenol red free Williams medium E on the viability of HepK2 and HepK6 cocultures was tested. Monolayer cocultures, grown on a 96-well plate for 24 h, were incubated with matriptase inhibitors for 24 h in treated

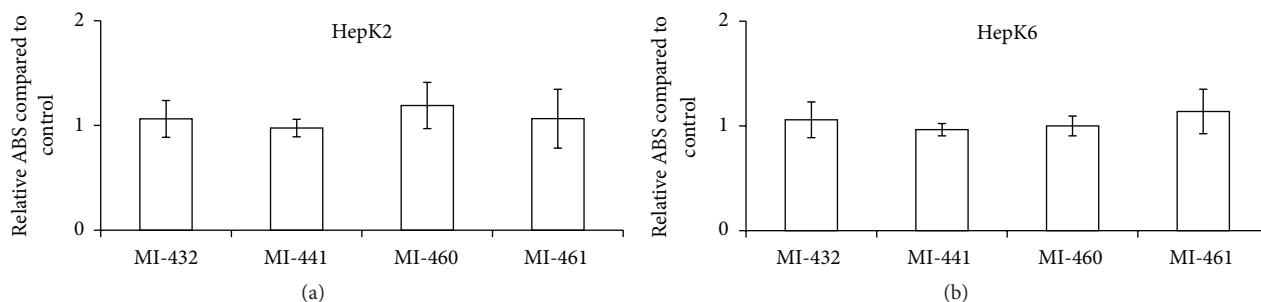


FIGURE 2: Lack of cytotoxic effects of matriptase inhibitors. (a) 24 h incubation of HepK2 cells with MI-432, MI-441, MI-460, or MI-461 at 50 μM . Values represent average relative absorbance values of produced MTS formazan in metabolically active cells expressed in control values \pm SEMs. No significant differences were found between control and cocultures exposed to matriptase inhibitors ($n = 4$). (b) HepK6 cocultures were treated with MI-432, MI-441, MI-460, or MI-461 at 50 μM for 24 h. Relative absorbance values expressed in control values \pm SEMs are indicated. No significant differences were found between control and treated cocultures ($n = 4$).

groups. The control cells were incubated only with phenol red free Williams medium E. After removal of the medium and washing the cells 3-fold with phosphate-buffered saline (PBS), 20 μL of CellTiter 96 aqueous one solution proliferation assay (MTS, Promega, Bioscience, Budapest, Hungary) containing tetrazolium compound and an electron coupling reagent, phenazine ethosulfate, was pipetted into each well of the 96-well assay plate containing the samples in 100 μL of culture medium. The plate was incubated with dye for 2 h. Viability of HepK2 and HepK6 cocultures was measured at 490 nm using EZ Read Biochrom 400 microplate reader.

2.5. Extracellular H_2O_2 Measurement by the Amplex Red Method. Fluorescence ROS measurement of cell supernatant was based on the detection of H_2O_2 using the Amplex Red Hydrogen Peroxide Assay Kit (Invitrogen, Molecular Probes). In the presence of horseradish peroxidase (HRP), Amplex Red reacts with H_2O_2 in a 1:1 stoichiometry producing a highly fluorescent resorufin [18]. Following the exposure of cocultures to matriptase inhibitors (50 μM , 2 h), the H_2O_2 concentrations in the medium were determined 6 h after completion of the treatment using a working solution of 100 μM Amplex Red and 0.2 U/mL HRP. After 30 min incubation with the dye at room temperature, the quantitative H_2O_2 contents were measured using a Victor X2 2030 fluorometer ($\lambda_{\text{ex}} = 560 \text{ nm}$; $\lambda_{\text{em}} = 590 \text{ nm}$).

2.6. Cytokine Measurements by Enzyme-Linked Immunosorbent Assay. After 2 h inhibitor treatment (50 μM), HepK2 and HepK6 cocultures were incubated in the medium for 6 h. Culture media were collected, centrifuged (245 $\times\text{g}$ for 10 min at room temperature), and diluted to measure the IL-6 and IL-8 levels.

Changes in IL-6 and IL-8 concentrations relative to those in controls were determined by porcine-specific IL-6 (Sigma-Aldrich, St. Louis, MO, USA) and IL-8 (Abcam, Cambridge, UK) ELISA kits according to the manufacturer's instructions. The absorbance values were detected at 450 nm using EZ Read Biochrom 400 microplate reader (Biochrom Ltd., UK).

2.7. Hepcidin ELISA Measurements. Porcine ELISA kit for hepcidin was obtained from Cloud-Clone Corp. (Houston, Texas, USA), employing the competitive inhibition enzyme immunoassay technique. After 2 h inhibitor treatment (50 μM), the HepK2 and HepK6 cell cocultures were incubated for 6 h and then the culture media were collected, centrifuged (1000 $\times\text{g}$ for 20 min at room temperature), and diluted in PBS. The assay procedure was performed according to the manufacturer's guideline. After addition of the 3,3',5,5'-tetramethylbenzidine (TMB) substrate solution for 15 min, the intensity of color developed is reverse proportional to the concentration of hepcidin in the sample. The absorbance values were detected at 450 nm using a EZ Read Biochrom 400 microplate reader (Biochrom Ltd., UK).

2.8. Statistical Analysis. For statistical evaluation R 2.11.1 software package (2010) was applied. Statistical significance of differences was assessed with one-sample Student's *t*-tests for assessment of relative values. Differences between absolute means were evaluated by one-way analysis of variance (one-way ANOVA) with *post hoc* Tukey test, where data were of normal distribution and homogeneity of variances was confirmed. Differences were considered significant if the *p* value was <0.05 marked with * (** $p < 0.01$).

3. Results

3.1. Assessment of Cell Viability after Matriptase Inhibition. Cell cytotoxicity assays were performed to evaluate if the used inhibitors at 50 μM concentration can cause significant cell death in HepK2 and HepK6 cocultures. It was found that the selected inhibitors can be used at 50 μM concentration safely for 24 h in HepK2 (Figure 2(a)) and HepK6 cocultures (Figure 2(b)), and thus this concentration could be applied in further experiments to characterize the effects of MT-1/MT-2 modulation in *in vitro* inflammatory hepatic models.

3.2. Matriptase Inhibition-Induced Oxidative Stress. Fluorescence intensities of both the control and treated samples were measured 6 h after a 2 h inhibitor application of matriptase

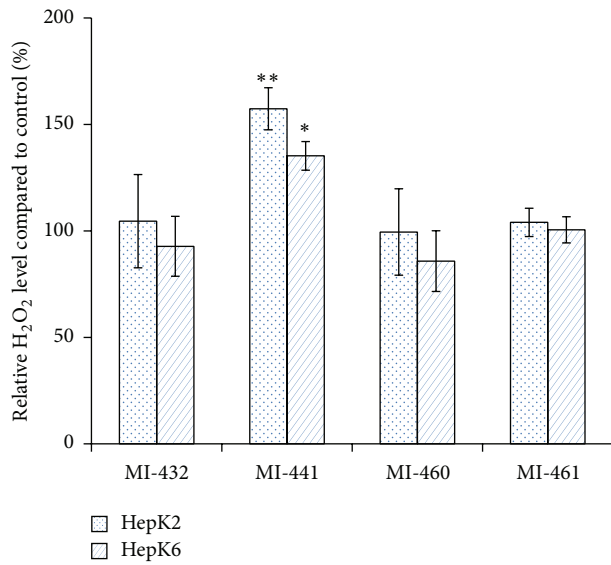


FIGURE 3: Extracellular ROS production of HepK2 and HepK6 cell cocultures control (Co) or exposed to matriptase inhibitors (MI-432, MI-441, MI-460, or MI-461) using Amplex Red method. Results are expressed as mean values relative to controls \pm SEMs. Significant differences in fluorescence intensities were found between control and MI-441-treated coculture groups (* $p < 0.05$, $n = 4$, in HepK2 and ** $p < 0.01$, $n = 4$, in HepK6 samples).

inhibitors at 50 μ M. It was found that there was an increase in the extracellular H₂O₂ levels (Figure 3) causing a significant elevation in fluorescence, when inhibitor MI-441 was applied in HepK2 and HepK6 cocultures ($p = 0.0147$ and $p = 0.00157$, resp.). However, no significant perturbations in redox balance in case of HepK2 and HepK6 cocultures were observed after treatment with the other three inhibitors under identical conditions.

3.3. Determination of IL-6 and IL-8 Levels in the Supernatants of Hepk2 and Hepk6 Cocultures. After treatment of HepK2 and HepK6 cell cocultures with matriptase inhibitors at 50 μ M concentration for 2 h, the potential changes in IL-6 expressions were monitored in the cell supernatants. It was ascertained that none of the applied inhibitors could induce elevated production of IL-6 in HepK2 and in HepK6 cells (Figure 4). The relative absorbance values of IL-8 levels were also measured to determine the putative changes introduced by short-term administration of matriptase inhibitors. It was found that the applied compounds did not cause any significant alterations in the levels of IL-8 in the supernatants of HepK2 and HepK6 cocultures compared to those in control groups (Figure 5).

3.4. Analysis of Hepcidin Levels in the Supernatants of Hepk2 and Hepk6 Cell Cocultures. In HepK2 coculture it was found that MI-432 and MI-461 at 50 μ M for 2 h did not alter hepcidin levels significantly compared to those of control samples ($p = 0.6571$ and $p = 0.2006$, resp.). However, matriptase inhibition via application of MI-441 and MI-460

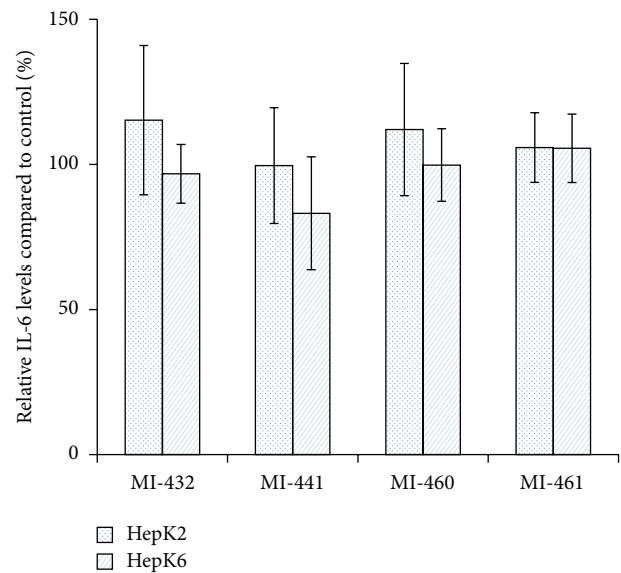


FIGURE 4: The effect of matriptase inhibition on IL-6 levels. The relative IL-6 levels after the HepK2 and HepK6 cell cocultures were treated with 50 μ M MI-432, MI-441, MI-460, and MI-461 for 2 h. The columns represent average relative absorbance values \pm SEMs ($n = 4$). No significant differences were found ($p > 0.05$) between control and matriptase inhibitor-treated groups.

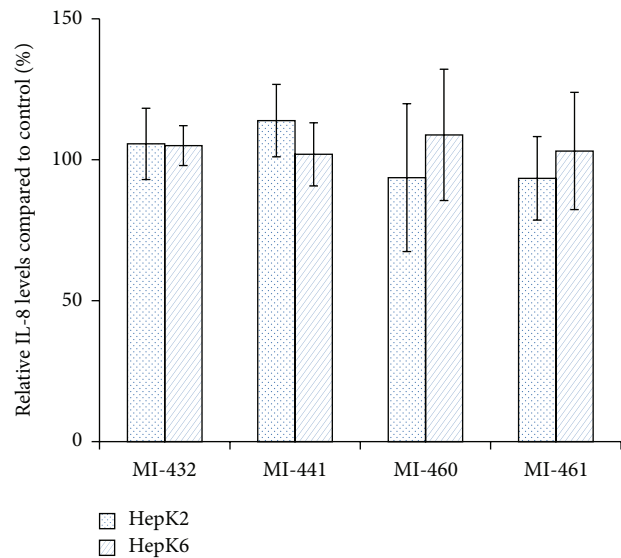


FIGURE 5: The effect of matriptase inhibition on IL-8 levels. The HepK2 and HepK6 cell cocultures were treated with 50 μ M MI-432, MI-441, MI-460, and MI-461 for 2 h. The columns represent average relative absorbance values \pm SEMs ($n = 4$). No significant differences were found ($p > 0.05$) between control and matriptase inhibitor-treated groups.

with the same incubation time and concentration has changed hepcidin levels in different ways. Treatment with MI-441 has lowered the hepcidin expression by $32.3 \pm 12.8\%$ ($p = 0.0487$), whereas it seems that compound MI-460 leads to an upregulation of hepcidin expression by $62.21 \pm 26.8\%$ ($p = 0.0368$) in the cell supernatants. Similar

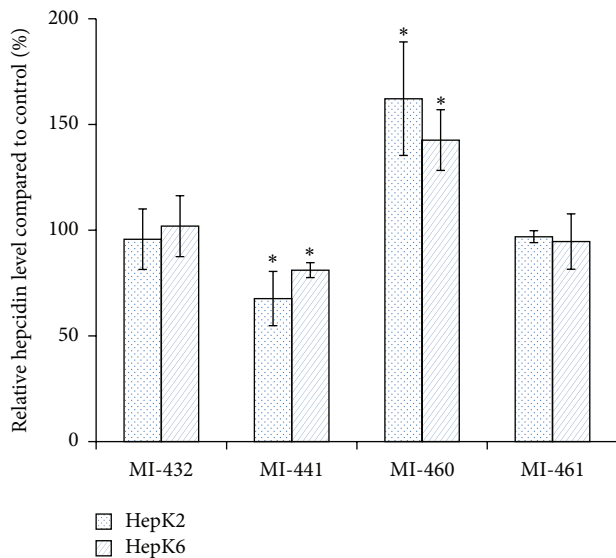


FIGURE 6: Changes in relative hepcidin levels after matriptase inhibition. Measurements were performed at 6 h after completion of 2 h 50 μ M MI-432, MI-441, MI-460, or MI-461 treatment. Each column represents the mean relative absorbance values \pm SEMs ($n = 3$). The asterisk (* $p < 0.05$) shows statistical significances for results expressed in control percentage.

findings were obtained in HepK6 cell cocultures where MI-441 administration reduced the hepcidin levels ($p = 0.0387$) by $18.9 \pm 3.5\%$ and MI-460 elevated ($p = 0.0114$) the expression of hepcidin by $42.6 \pm 14.3\%$. Application of MI-432 and MI-461 did not cause significant alterations in the levels of hepcidin in the supernatants of HepK6 compared to control samples ($p = 0.8404$ and $p = 0.549$, resp.) (Figure 6).

4. Discussion

Physiological plasma and cellular iron concentrations are provided via strict regulation of critical steps responsible for iron absorption, transport, storage, and recycling [19]. MT-2 plays a major role in inactivation of HJV thus reducing the expression of the liver-derived peptide hormone, hepcidin. Misregulation of hepcidin as a negative regulator of iron absorption has been found in many diseases such as the anemia of chronic disease, iron refractory iron deficiency anemia, cancer, hereditary hemochromatosis, and ineffective erythropoiesis such as β -thalassemia [20]. Other factors such as oxidative stress can also affect hepcidin levels. Experimental findings support that hepatitis C virus- (HCV-) induced oxidative stress inhibits hepcidin production offering a possible explanation for phenomenon of iron loading when this viral infection occurs [21, 22]. In the prevention and in the treatment of iron overload hepcidin agonists can represent therapeutical alternatives against several forms of the hemochromatosis. Small peptide analogues containing 7–9 amino acids, minihepcidins, were successfully applied in the regulation of plasma iron concentration in mice models [23]. It was reported that MT-2 could also affect the level of produced hepcidin. The loss of the serine protease

domain of MT-2 in different species caused anaemia with increased hepcidin concentration and with low plasma iron level due to the reduced iron absorption [9, 10, 19]. MT-2 mutations in humans or lack of MT-2 in mice resulted in increased hepcidin expression and iron refractory iron deficiency anemia [8].

Elevated MT-1 levels could be found in epithelial cancers suggesting that the dysfunctional matriptase activity may contribute to tumor development and metastatic events. MT-1 could trigger the degradation of extracellular matrix components and intercellular structures thus enabling cell migration [5]. Moreover, also an aberrant expression of MT-2 is observed in several forms of cancers such as breast and prostate cancer [3, 24, 25]. However, more studies should be conducted to determine the efficacy of MT-1/MT-2 inhibitors on tumor formation and migration ability of cancerous cells more precisely based on the approach that matriptase inhibition might possess beneficial effect against tumor invasion and metastasis.

Suppression of MT-2 via application of 3-amidinophenylalanine compounds can be an effective pharmacological option in the effective treatment of hemochromatosis in the future [8, 12]. The safety of our used inhibitors at 50 μ M was confirmed since their 24 h long administration did not cause any significant changes in cell viability in HepK2 and HepK6 cocultures. Our data revealed that potent suppression of MT-2 activity via 2 h acute treatment of HepK2 and HepK6 cocultures with 50 μ M MI-460 could enhance hepcidin expression to great extent. This suggests that application of MI-460 can control dysregulated iron metabolism via increasing hepcidin level present mainly in several forms of systematic iron overload. In contrast, treatment with inhibitor MI-441 significantly decreased hepcidin levels, which might be explained by the fact that hepcidin synthesis can be affected via oxidative stress and as it was shown by us that extracellular H_2O_2 production was elevated by the addition of MI-441 in both cocultures. The other matriptase inhibitors (MI-432, MI-460, and MI-461), however, did not cause any significant changes in extracellular ROS production in these cocultures. In contrast, in our previous studies it was found that significant ROS release could be detected mainly after acute administration of MI-432 at 50 μ M in nontumorigenic intestinal porcine epithelial IPEC-J2 cell line via suppression of MT-1 enzyme activity. These apparent discrepancies between extracellular peroxide productions can be explained by various responsiveness of different cells such as IPEC-J2 cells and HepK cocultures towards matriptase inhibition [26]. As MI-432 and MI-461 did not show any acute effect on hepcidin regulation, thus further studies should be performed to clarify whether long-term applications of these compounds possess any impact on pathophysiological iron conditions.

Hepatic coculture systems consist of primary hepatocytes and Kupffer cells in a ratio of 6:1 and 2:1 and they can represent milder or severe inflammatory states of liver [17, 27]. Kupffer cells can be activated by bacterial endotoxins, which results in the elevated production of several proinflammatory cytokines such as IL-6 and IL-8. It was established that this coculture system can be used to examine chemical-induced

inflammatory reactions such as shifting in cytokine profile (IL-6 and TNF- α) as a consequence of acute hepatocellular toxicity introduced by lipopolysaccharide and trovafloxacin administration [27]. In our study, 2 h inhibitor administration did not show any effects on IL-6 and IL-8 levels in HepK2 and HepK6 cocultures. This suggests that the hepatic inflammatory response is not influenced by the applied MT-1/MT-2 inhibitors.

In conclusion, taken together, the fact that MT-1 and MT-2 are membrane anchored serine proteases on the cell surface with relatively convenient accessibility for drugs and the downregulation of excessive matriptase activities can compensate severe pathological perturbations, further development and advanced biomolecular screening of more selective MT-1 and MT-2 inhibitors can be of high pharmacological and therapeutical importance.

Disclosure

The authors alone are responsible for the content and writing of this paper.

Competing Interests

The authors report no declaration of interest.

Acknowledgments

This research was supported by the I1475-4/2016/FEKUT grant of the Hungarian Ministry of Human Resources. This project was supported by the János Bolyai Research Scholarship of the Hungarian Academy of Sciences and by the Hungarian Scientific Research Fund [Grants nos. I15685 and I14033].

References

- [1] É. Colombo, A. Désilets, D. Duchêne et al., "Design and synthesis of potent, selective inhibitors of matriptase," *ACS Medicinal Chemistry Letters*, vol. 3, no. 7, pp. 530–534, 2012.
- [2] R. Szabo and T. H. Bugge, "Type II transmembrane serine proteases in development and disease," *The International Journal of Biochemistry and Cell Biology*, vol. 40, no. 6-7, pp. 1297–1316, 2008.
- [3] G. Velasco, S. Cal, V. Quesada, L. M. Sánchez, and C. López-Otín, "Matriptase-2, a membrane-bound mosaic serine proteinase predominantly expressed in human liver and showing degrading activity against extracellular matrix proteins," *The Journal of Biological Chemistry*, vol. 277, no. 40, pp. 37637–37646, 2002.
- [4] J. M. Milner, A. Patel, R. K. Davidson et al., "Matriptase is a novel initiator of cartilage matrix degradation in osteoarthritis," *Arthritis and Rheumatism*, vol. 62, no. 7, pp. 1955–1966, 2010.
- [5] K. Uhlund, "Matriptase and its putative role in cancer," *Cellular and Molecular Life Sciences*, vol. 63, no. 24, pp. 2968–2978, 2006.
- [6] K. List, T. H. Bugge, and R. Szabo, "Matriptase: potent proteolysis on the cell surface," *Molecular Medicine*, vol. 12, no. 1-3, pp. 1–7, 2006.
- [7] C.-J. Ko, C.-C. Huang, H.-Y. Lin et al., "Androgen-induced TMPRSS2 activates matriptase and promotes extracellular matrix degradation, prostate cancer cell invasion, tumor growth, and metastasis," *Cancer Research*, vol. 75, no. 14, pp. 2949–2960, 2015.
- [8] X. Du, E. She, T. Gelbart et al., "The serine protease TMPRSS6 is required to sense iron deficiency," *Science*, vol. 320, no. 5879, pp. 1088–1092, 2008.
- [9] J. E. Maxson, J. Chen, C. A. Enns, and A.-S. Zhang, "Matriptase-2- and proprotein convertase-cleaved forms of hemojuvelin have different roles in the down-regulation of hepcidin expression," *The Journal of Biological Chemistry*, vol. 285, no. 50, pp. 39021–39028, 2010.
- [10] L. Silvestri, A. Pagani, A. Nai, I. De Domenico, J. Kaplan, and C. Camaschella, "The serine protease matriptase-2 (TMPRSS6) inhibits hepcidin activation by cleaving membrane hemojuvelin," *Cell Metabolism*, vol. 8, no. 6, pp. 502–511, 2008.
- [11] A. J. Ramsay, J. D. Hooper, A. R. Folgueras, G. Velasco, and C. López-Otín, "Matriptase-2 (TMPRSS6): a proteolytic regulator of iron homeostasis," *Haematologica*, vol. 94, no. 6, pp. 840–849, 2009.
- [12] K. E. Finberg, M. M. Heeney, D. R. Campagna et al., "Mutations in TMPRSS6 cause iron-refractory iron deficiency anemia (IRIDA)," *Nature Genetics*, vol. 40, no. 5, pp. 569–571, 2008.
- [13] E. Nemeth, M. S. Tuttle, J. Powelson et al., "Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization," *Science*, vol. 306, no. 5704, pp. 2090–2093, 2004.
- [14] G. J. Anderson, D. M. Frazer, S. J. Wilkins et al., "Relationship between intestinal iron-transporter expression, hepatic hepcidin levels and the control of iron absorption," *Biochemical Society Transactions*, vol. 30, no. 4, pp. 724–726, 2002.
- [15] T. Ganz, "Systemic iron homeostasis," *Physiological Reviews*, vol. 93, no. 4, pp. 1721–1741, 2013.
- [16] M. Hammami, E. Rühmann, E. Maurer et al., "New 3-amidinophenylalanine-derived inhibitors of matriptase," *Medicinal Chemistry Communication*, vol. 3, no. 7, pp. 807–813, 2012.
- [17] G. Mátyás, A. Kulcsár, J. Petrilla, P. Talapka, and Z. Neogrady, "Porcine hepatocyte-Kupffer cell co-culture as an in vitro model for testing the efficacy of anti-inflammatory substances," *Journal of Animal Physiology and Animal Nutrition*, 2016.
- [18] J. G. Mohanty, J. S. Jaffe, E. S. Schulman, and D. G. Raible, "A highly sensitive fluorescent micro-assay of H₂O₂ release from activated human leukocytes using a dihydroxyphenoxazine derivative," *Journal of Immunological Methods*, vol. 202, no. 2, pp. 133–141, 1997.
- [19] A. R. Folgueras, F. M. de Lara, A. M. Pendás et al., "Membrane-bound serine protease matriptase-2 (Tmprss6) is an essential regulator of iron homeostasis," *Blood*, vol. 112, no. 6, pp. 2539–2545, 2008.
- [20] N. Zhao, A.-S. Zhang, and C. A. Enns, "Iron regulation by hepcidin," *Journal of Clinical Investigation*, vol. 123, no. 6, pp. 2337–2343, 2013.
- [21] K. Miura, K. Taura, Y. Kodama, B. Schnabl, and D. A. Brenner, "Hepatitis C virus-induced oxidative stress suppresses hepcidin expression through increased histone deacetylase activity," *Hepatology*, vol. 48, no. 5, pp. 1420–1429, 2008.
- [22] S. Nishina, K. Hino, M. Korenaga et al., "Hepatitis C virus-induced reactive oxygen species raise hepatic iron level in mice by reducing hepcidin transcription," *Gastroenterology*, vol. 134, no. 1, pp. 226–238, 2008.

- [23] G. C. Preza, P. Ruchala, R. Pinon et al., “Minihepcidins are rationally designed small peptides that mimic hepcidin activity in mice and may be useful for the treatment of iron overload,” *The Journal of Clinical Investigation*, vol. 121, no. 12, pp. 4880–4888, 2011.
- [24] C. Parr, A. J. Sanders, G. Davies et al., “Matriptase-2 inhibits breast tumor growth and invasion and correlates with favorable prognosis for breast cancer patients,” *Clinical Cancer Research*, vol. 12, no. 12, pp. 3568–3576, 2007.
- [25] A. J. Sanders, C. Parr, T. A. Martin, J. Lane, M. D. Mason, and W. G. Jiang, “Genetic upregulation of matriptase-2 reduces the aggressiveness of prostate cancer cells in vitro and in vivo and affects FAK and paxillin localisation,” *Journal of Cellular Physiology*, vol. 216, no. 3, pp. 780–789, 2008.
- [26] E. Pászti-Gere, S. McManus, N. Meggyesházi, P. Balla, P. Gálfi, and T. Steinmetzer, “Inhibition of matriptase activity results in decreased intestinal epithelial monolayer integrity in vitro,” *PLoS ONE*, vol. 10, no. 10, Article ID e0141077, 2015.
- [27] J. A. Bonzo, K. Rose, K. Freeman et al., “Differential effects of trovafloxacin on TNF- α and IL-6 profiles in a rat hepatocyte–Kupffer cell coculture system,” *Applied in Vitro Toxicology*, vol. 1, no. 1, pp. 45–54, 2015.