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Emerging Concepts in Functional and Molecular Photoacoustic Imaging

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Abstract

Providing the specific imaging contrast of optical absorption and excellent spatial scalability across the optical and ultrasonic dimensions, photoacoustic imaging has been rapidly emerging and expanding in the past two decades. In this review, I focus on a few latest advances in this enabling technology that hold the potential to transform *in vivo* functional and molecular imaging at multiple length scales. Specifically, multi-parametric photoacoustic microscopy enables simultaneous high-resolution mapping of hemoglobin concentration, oxygen saturation and blood flow—opening up the possibility of quantifying the metabolic rate of oxygen at the microscopic level. The pump-probe approach harnesses a variety of photoinduced transient optical absorption as novel contrast mechanisms for high-specificity molecular imaging at depth and as nonlinear excitation strategies for high-resolution volumetric microscopy beyond the conventional limit. Novel magneto-optical and photochromic probes lead to contrast-enhanced molecular photoacoustic imaging through differential detection.

Introduction

Photoacoustic imaging, presenting an elegant way to combine optics and ultrasound, is among the most rapidly growing technologies in biomedicine [1,2]. In this emerging modality, endogenous or exogenous chromophores in the biological tissue are usually excited by nanosecond laser pulses. Part of the absorbed photon energy induces transient heating of the tissue through nonradiative relaxation of the excited states. The subsequent thermoelastic expansion converts the tiny temperature rise (typically at the level of milli-Kelvin) into ultrasonic emission, which can be captured by an acoustic detector to form a high-resolution image of the chromophore distribution.

The conversion of optical excitation to acoustic emission confers two distinct advantages to photoacoustic imaging. First, it provides the specific contrast of optical absorption, which ideally complements that of fluorescence imaging—the current technology of choice for optical molecular imaging in vivo. Indeed, photoacoustic imaging has enabled highsensitivity detection of a variety of endogenous chromophores that are intrinsically weakly fluorescent and difficult to label with exogenous fluorophores—including but not limited to cytochrome [3], hemoglobin [4], myoglobin [5], melanin [6], water [7] and lipid [8]. Labelfree photoacoustic imaging of these fluorescently "dark" chromophores has opened up new

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avenues for basic and translational research in brain ischemia [9], cancer [10,11] and atherosclerosis [12]. Second, the hybrid of light and sound allows seamless scalability of the spatial resolution and penetration depth of photoacoustic imaging across the microscopic (i.e., micrometer resolution at the depth of sub-millimeter), mesoscopic (i.e., tens of micrometer resolution at the depth of 1–10 mm) and macroscopic (i.e., 100–400 μm resolution at the depth of several centimeters) scales with an excellent depth-to-resolution ratio of ~200 [2,13]. This scalability is not yet achievable with fluorescence-based molecular imaging. Although capable of providing microscopic resolution within the optical diffusion limit (i.e., \sim 1 mm in biological tissues), fluorescence imaging has a poor depth-to-resolution ratio of ~3 in the optical diffusive regime [14]. With advances in optically absorbing probes, the photoacoustic effect provides an ideal mechanism for high-resolution molecular imaging beyond the diffusion limit.

In recent years, intensive research efforts have been made to unleash the full potential of photoacoustic imaging of both endogenous nonfluorescent chromophores and exogenous molecular probes. In this short review, I focus on only a few latest and potentially transformative advances in functional and molecular photoacoustic imaging—including (1) multi-parametric quantification of hemodynamics with the contrast of endogenous hemoglobin and its potential in oxygen-metabolic imaging at the microscopic level; (2) pump-probe interrogation of photoinduced spatiotemporal changes in the generation of photoacoustic signals and its applications in both high-specificity molecular imaging at depth and high-resolution volumetric imaging beyond the conventional limit; (3) photochromic and magneto-optical probes for high-contrast molecular photoacoustic imaging based on differential detection. Comprehensive reviews on functional and molecular photoacoustic imaging can be found in [15,16].

Multi-parametric photoacoustic microscopy

Among the multiple endogenous chromophores that can be photoacoustically detected, hemoglobin is of particular importance. As the primary carrier of oxygen in the blood circulation, hemoglobin encodes vital metabolic information. Dysfunctions in oxygen metabolism has been increasingly recognized as a critical factor in the origination and progression of multiple life-threatening diseases, including cancer, ischemic stroke and neurodegeneration [17]. In vivo high-resolution imaging of the metabolic rate of oxygen $(MRO₂)$ in mice—a species with well-established disease models and genetic manipulations —is crucial for understanding metabolism-related pathogenic mechanisms and formulating new therapeutic strategies.

Capable of measuring the concentration of hemoglobin (C_{H_b}) , oxygen saturation of hemoglobin ($SO₂$) and blood flow—the three parameters required for MRO₂ quantification, photoacoustic microscopy (PAM) is ideally suited for oxygen-metabolic imaging in vivo. Indeed, PAM of $MRO₂$ has been demonstrated first in the tumor-bearing mouse ear [18] and later in the electrically stimulated mouse brain [19] by measuring C_{Hb} , sO₂ and blood flow at selected locations in the feeding arteries and draining veins of the region of interest. Although encouraging, this method has fundamental limitations. First, it can only be applied to the regions that have well-defined feeding arteries and draining veins. Moreover, the

volumetric flow rates in the feeding and draining vessels must be equal (i.e., a closed circulation). Even if applicable, this method only quantifies the total $MRO₂$ over the entire region. The lack of the ability to resolve $MRO₂$ at the microscopic level limits its utility in studying the metabolic heterogeneity in cancer and ischemia.

A recent innovation, multi-parametric PAM, overcomes all these limitations by enabling simultaneous imaging of C_{Hb} , sO₂ and blood flow at the microvascular level [20,21]. As shown in Fig. 1A, statistical, spectroscopic and correlation analysis of successively acquired dual-wavelength (i.e., 532 and 558 nm) A- line pairs allows quantification of C_{Hb} , sO₂ and blood flow at the same spatiotemporal scale. Specifically at 532 nm, where the optical absorption of oxy- and deoxy-hemoglobin (HbO₂ and HbR) are identical, PAM is insensitive to sO2. Fluctuations in the PAM signal acquired at this wavelength encode both the Brownian motion and the flow of red blood cells (RBCs) [22]. The Brownian motioninduced statistical fluctuation in the amplitude of these A-lines depends on the RBC count within the detection volume of PAM but not the blood flow, and thus can be used to derive C_{Hb} in absolute values [23]. In parallel, the speed of blood flow can be quantified by examining the cross-correlation of the same set of A-lines [20]. The decay rate of the correlation reveals the dwell time of RBCs in the detection volume. The faster the decorrelation, the higher the flow speed. Taking advantage of bi-directional raster scan, PAM can further determine the direction of blood flow. In addition, by comparing the readouts at both wavelengths, PAM can assess the proportions of $HbO₂$ and HbR in C_{Hb} , from which $sO₂$ can be derived. With the quantitative analyses, multi-parametric PAM has demonstrated simultaneous high-resolution imaging of C_{Hb} , SO_2 and blood flow velocity (i.e., both the speed and direction) in vivo (Fig. 1B-1D) [21]. Future development of complementary algorithms to extend these hemodynamic parameters from the microvascular level to the tissue level will ultimately enable $MRO₂$ quantification at the microscopic level by using the Fick's law.

Pump-probe photoacoustic imaging

Pump-probe imaging of transient optical absorption is an emerging field in nonlinear microscopy [24,25]. Recently, this concept has been adopted by photoacoustic imaging for enriching the molecular contrast and refining the spatial resolution. This innovation has expanded the envelope of photoacoustic imaging by enabling lifetime and fluorophore imaging at depth and high-resolution volumetric imaging beyond the conventional limit. Three main mechanisms of current pump-probe photoacoustic imaging—excited-state absorption, stimulated emission and ground-state depletion—are introduced here.

Lifetime imaging based on excited-state absorption

In the case of excited-state absorption (top row of Fig. 2A), the pump laser pulse excites the molecule to be imaged from the ground state (S_0) to an excited state (S_1) . The molecule can either decay back to S_0 via radiative or nonradiative relaxation or be promoted to a higher state (S_2) by absorbing a photon from the probe pulse. Thus, populating S_1 with the pump excitation increases the molecule's absorptivity of the probe light. The differential optical absorption at the probe wavelength in the presence and absence of the pump encodes the

energy structure of the molecule and can be utilized as a contrast mechanism for molecular photoacoustic imaging with high specificity. More excitingly, dynamic adjustments of the delay between the pump and probe pulses allow photoacoustic imaging of the depopulation rate of the excited state, from which its lifetime can be estimated.

Recently, excited-state absorption-based photoacoustic lifetime imaging has been combined with oxygen-sensitive dyes (e.g., methylene blue) to measure the partial pressure of oxygen $(pO₂)$ at depths up to 12 mm [26]. Since the excited state of the dye molecule can be dynamically quenched through its collision with the surrounding oxygen molecules, photoacoustic imaging of its excited-state lifetime provides a quantitative measure of $pO₂$ (Fig. 2B) [27]. Future integration of the lifetime measurement of $pO₂$ and the spectroscopic measurement of SO_2 will equip photoacoustic imaging with the unique capability of comprehensively characterizing the oxygen dynamics in vivo.

Fluorophore imaging based on stimulated emission

In contrast to excited-state absorption, which induces transient optical absorption at the probe wavelength, stimulated emission can be utilized to modulate the transient optical absorption at the pump wavelength. This mechanism has been applied to extend the scope of photoacoustic imaging from chromophores to fluorophores [28]. As shown in the mid row of Fig. 2A, the probe pulse can stimulate fluorescence emission after the upper state (S_1) of the fluorophore is populated by the pump excitation. This process can accelerate the relaxation of the fluorophore back to the ground state (S_0) and facilitates the absorption of additional pump photons. Proper selection of the pump and probe wavelengths to match the excitation and emission peaks of the fluorophore can maximize the efficiency of stimulated emission, which results in multiple excitation-relaxation cycles within the duration of the pump pulse and thus increases the fluorophore's absorptivity of the pump photon. Moreover, increasing the pump-probe time delay reduces the stimulated emission due to the gradual depopulation of $S₁$. The differential optical absorption of the pump pulse in the presence and absence of the probe or due to different pump-probe delays can be utilized as a contrast mechanism for photoacoustic imaging of fluorophores with enhanced specificity.

Recently, the feasibility of stimulated emission-based photoacoustic imaging was experimentally demonstrated in a 6-mm-thick tissue phantom consisting of polymer capillaries filled with Atto680 and murine blood [28]. The pump and probe wavelengths were selected to be 680 and 742 nm, respectively, matching the excitation and emission peaks of Atto680. As shown in Fig. 2C, the phantom was imaged with both simultaneous (i.e., non-delayed) and delayed pump-probe pulses. Subtraction of the two images completely removes the background signal from blood, which has negligible stimulated emission in this pump-probe setting, and enables high-specificity imaging of Atto680. Providing much improved spatial resolution compared to that of deep-tissue fluorescence imaging, stimulated emission-based photoacoustic imaging has potentially broad basic and translational applications.

Isotropic-resolution PAM based on ground-state depletion

The third contrast mechanism of pump-probe photoacoustic imaging is ground-state depletion. Different from that in the excited-state absorption and stimulated emission settings, the pump and probe pulses used for photoacoustic imaging of ground-state depletion are typically of the same wavelength. In this process (bottom row of Fig. 2A), the first (i.e., pump) pulse excites the molecule from the ground state $(S₀)$ to the upper state $(S₁)$. Due to the pump-induced depopulation of the ground state, the second (i.e., probe) pulse experiences reduced optical absorption. The difference between the photoacoustic signals generated by the probe light in the presence and absence of the pump depends on the recovery time of S_0 from depletion, providing another molecule-specific contrast mechanism for photoacoustic imaging [29].

Besides, the effect of ground-state depletion provides a way to break the conventional limit on the axial resolution of PAM [30]. In conventional PAM, the axial resolution is determined by the bandwidth of the photoacoustic signal detected by the ultrasonic transducer [2]. Refining the axial resolution to the level of the optical diffraction-limited lateral resolution requires GHz acoustic bandwidth, which is impractical. In ground-state depletion-based PAM, the differential signal with the pump on and off is proportional to the product of the pump and probe fluence. This nonlinear dependence offers PAM the optical-sectioning capability equivalent to that of two-photon microscopy. Indeed, with this pump-probe strategy, the axial resolution of PAM has been refined down to 1.5 μm—approaching the lateral resolution defined by optical focusing [31]. The isotropic resolution enables threedimensional visualization of the biconcave shape of a single RBC (Fig. 2D) [31]. It is worth noting that similar optical sectioning in PAM has also been achieved through other nonlinear processes, such as photobleaching [32] and photothermal tagging [33]. Advancing these techniques to in vivo settings will excite broad new applications.

Contrast-enhanced molecular photoacoustic imaging

Molecular photoacoustic imaging in the visible and near-infrared regions often suffers from insufficient contrast due to the substantial background generated by endogenous hemoglobin. A conventional strategy to differentiate the molecular signal of interest from the background is photoacoustic spectroscopy, which however requires repeated measurements at multiple optical wavelengths, is computationally expensive, and relies on fluence compensation to account for the spectral and spatial distortion by the tissue [34].

Recent advances in optically absorbing switchable probes have led to a promising alternative that utilizes differential detection for contrast-enhanced molecular photoacoustic imaging. One switching mechanism is magnetic field-induced motion [35]. The so-called magnetooptical probe, consisting of a magnetic core and a gold nanoshell, is both magnetically sensitive and optically absorbing. An external magnetic field can induce cyclic motion of the probe, which modulates the photoacoustic signal generated by the gold nanoshell. Motion analysis can effectively remove the magnetically insensitive background, thereby boosting the contrast by orders of magnitude over conventional photoacoustic imaging. This technique has been applied for in vivo imaging of targeted cancer cells [36]. Another switching mechanism for differential detection is photochroism [37]. BphP1—a

nonfluorescent bacterial phytochrome—can be reversibly switched between two conformational states (i.e., Pr and Pfr) upon light excitation, resulting in a shift in its optical absorption spectrum [38]. Specifically, BphP1 undergoes the conversion from Pfr to Pr when excited with 730–790-nm light and from Pr back to Pfr upon 630–690-nm excitation. Recently, this photochromic probe was genetically encoded in glioblastoma cells for in vivo tumor imaging [37]. As shown in Fig. 3, the mouse brain with a glioblastoma sitting at 3 mm beneath the scalp was repeatedly imaged at 780 nm, during which the conformational state of the BphP1 expressed by the tumor cells was photoswitched between Pfr and Pr. In the images acquired under individual states (Fig. 3A and 3B), the tumor signal is obscured by the overwhelming vascular background. Strikingly, subtracting the two images completely removes the non-photochromic vasculature and highlights the BphP1-expressing tumor (Fig. 3C). Engineering transgenic mouse models with cell or pathway-specific expression of BphP1 will expose this high-contrast deep-penetrating molecular imaging technique to more broad applications.

Summary

Centering on the two niches of photoacoustic imaging, optical absorption contrast and spatial scalability, recent research efforts have led to rapid and exciting expansion of the technology envelope. Translating these emerging concepts into robust tools will fill important gaps in basic and translational research, including microscopic imaging of oxygen metabolism *in vivo* and high-resolution high-contrast molecular imaging at depth.

Acknowledgments

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- **•** Pump-probe photoacoustic imaging enables high-specificity molecular imaging at depth and high-resolution volumetric imaging beyond the conventional limit.
	- **•** Photochromic and magneto-optical probes enables high-contrast molecular photoacoustic imaging based on differential detection.

Statistical (C_{Hb}) & correlation (blood flow) analysis (100 A-lines)

Figure 1.

Multi-parametric PAM. (A) Scanning mechanism for statistical, spectroscopic and correlation analysis. $(B-D)$ Simultaneous PAM of C_{Hb} , SO_2 and blood flow velocity in the mouse brain *in vivo* (adapted from [21]).

Figure 2.

Pump-probe photoacoustic imaging. (A) Simplified energy diagrams depicting excited-state absorption, stimulated emission and ground-state depletion. The blue and green arrows respectively denote the transition in the energy state due to pump and probe excitation. (B) Excited-state absorption-based photoacoustic lifetime imaging of $pO₂$ in the mouse hindlimb under systemic normoxia (21% O_2), hyperoxia (80% O_2) and hypoxia (10% O_2). The concurrently acquired ultrasound image is shown in gray (adapted from [27]). (C) Stimulated emission-based photoacoustic images of a tissue phantom consisting of polymer capillaries filled with Atto680 (white arrows) and murine blood (red arrows) acquired using both simultaneous and delayed pump-probe pulses. The differential image completely eliminates the blood signals and highlights the transient absorption signal from Atto680 (adapted from [28]). (D) ground-state depletion-based isotropic-resolution PAM of fixed red blood cells in a volume of 50 μ m \times 50 μ m \times 20 μ m. Blow-up view of a single RBC clearly shows the biconcave shape (adapted from [31]).

Figure 3.

Contrast-enhanced molecular photoacoustic imaging based on differential detection of photochroism. (A, B) Photoacoustic images of the tumor-bearing mouse brain acquired when the BphP1 expressed by the tumor cells was photoswitched to the Pfr state and the Pr state, respectively. The tumor is invisible in either image due to the overwhelming vascular background. (C) The differential image completely eliminates the background vasculature and clearly delineates the BphP1-expressing tumor. Images are adapted from [37].